Determining sexual maturity in male leopard sharks in San Francisco Bay, California

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The leopard shark (*Triakis semifasciata*) is endemic to the eastern North Pacific ranging from Mazatlan, Mexico and the Gulf of California to Oregon. It is an inshore species generally found in shallow water < 91 m deep (Smith 2001; Ebert 2003). This species is one of the most researched elasmobranchs along the Pacific Coast of North America with several aspects of its biology well defined (Ackerman 1971; Russo 1975, 2013, 2015, 2018, 2019; Talent 1976, 1985; Smith 1984, 2001, 2005; Smith and Abramson 1990; Cailliet 1992; Kusher et al. 1992; Au and Smith 1997; Smith et al. 2003; Hight and Lowe 2007; Lewallen et al. 2007; Carlisle and Starr 2009, 2010; Nosal et al. 2013a,b, 2014; Launer 2014; Barker et al. 2015). Various reproductive studies have occurred for leopard sharks (Ackerman 1971; Talent 1985; Ebert and Ebert 2005; Smith 2005; Nosal et al. 2013a, b, 2014; Launer 2014; Russo 2015, 2018, 2019). Although these studies have found sexual segregation the specifics of male sexual maturity remain unknown. Additionally, little detail is known concerning the process of sexual maturation in males from known breeding locations within its range.

Male leopard sharks may reach sexual maturity at a total length (TL) ranging from 70 to 120 cm (Ebert 2003), a range currently thought to be based on various known breeding locations and research methods. However, details regarding the length at which calcification of claspers and production of mature spermatozoa occur in male leopard sharks are poorly known.

Several reports on calcification of claspers and production of spermatozoa of various species have been published (Pratt 1979; Joung and Hsu 2005; Huveneers et al. 2007; Awruch et al. 2008). Clark and Von Schmidt (1965) and Chen et al. (1988) have suggested that calcified claspers and the ability of the rhipidion to splay is a measure of sexual maturity. Yet, other studies maintain that a more reliable indicator of sexual maturity involves clasper length and calcification in combination with swollen testes and the presence of spermatozoa (Peres and Vooren 1991; Jensen et al. 2002; Lucifora et al. 2005; Conde-Moreno and Galvan-Magaña 2006; Awruch et al. 2008; Dharmadi and Wiadnyana 2013; Natanson and Gervelis 2013; Gracan and Lackovic 2016). The purpose of the present study was to determine whether calcification of claspers is correlated with production of spermatozoa and thus an indicator of sexual maturity in leopard sharks. Data first reported herein, shows that in male leopard sharks in South San Francisco Bay, there is a time delay between the

length at which claspers calcify and spermatozoa is produced.

Between 1970 and 2001, data collection was conducted monthly primarily between the San Francisco Bay Bridge (37.800 N, 122.3667 W) and the entrance of Alviso Slough (37.450 N, 122.017 W) at the south end of San Francisco Bay (Figure 1). There were 224 catch events (one technique, location, date, and time) using long-lines (n = 146), rod and reel (n = 36), and otter trawl (n = 42) (Russo 2019).



Figure 1. Map of the study area of San Francisco Bay with all catch events (red circles) in this study restricted to South San Francisco Bay and close to known parturition or mating sites. Map courtesy of the East Bay Regional Park District.

I collected data on sex, external parasites, and general condition of every shark. All measurements of individual sharks were made in centimeters total length (cm TL), whereas groups of sharks were calculated in centimeters mean total length (cm MTL). I physically examined male claspers for flexibility, extent of calcification and length from the tip to the posterior margin of the cloacal opening. I also examined clasper tips for spurs, hooks, or spines used in holding the clasper in the female cloaca, but none were found in the field and samples were not removed for microscopic examination later (Pratt and Carrier 2004). Clasper length was plotted against total length for analysis with color designations at the points at which calcification of claspers and production of spermatozoa occurred. Both sperm sacs and seminal vesicles were examined for presence of spermatozoa in the field along with the condition of the vas deferens, which are coiled in mature sharks. I took samples back to the laboratory for microscopic examination.

In this study, 4,121 elasmobranchs were captured mostly from South San Francisco Bay including 2,478 (60.1%) leopard sharks composed of 1,299 males and 1,179 females (Russo 2019). I selected a group of 99 male leopard sharks as a representative sample of all potential stages of maturity (Table 1, Figure 2A). These individuals ranged in size from 44.4 to 124.4 cm TL (86.3 cm MTL \pm 19.6 SD) and were captured in six male-dominated long line events during the months of expected sexual activity from April to early July (1977 - 1990) (Ebert and Ebert 2005, Russo 2015, 2018, 2019) and close to known parturition sites defined in Russo (2019) (Figure 1). Clasper length ranged from 2.5 to 14.6 cm (5 cm MTL \pm 1.3 SD). This analysis broadly defined the length "markers" at which calcification of claspers occurred along with the production of spermatozoa. Calcified claspers ranged in length from 6.3 cm to 14.6 cm (12.4 cm MTL \pm 2.04 SD). Of these, 44.4% (44/99) possessed calcified claspers and 32.3% had possessed spermatozoa (Table 1, Figure 2a). Of the 44 males with calcified claspers, 72.7% (n = 32) were sexually mature, while the remaining 27.3% (n = 12) with calcified claspers were sexually immature.

Additionally, there was a familiar relationship between clasper elongation and TL. This relationship shows rapid clasper elongation from 6.4 cm to 14 cm (+7.6 cm) between 86 and 101 cm TL, which is where calcification begins between 86 and 92.7 cm TL (Table 1), along with weight gain from 2 kg (86 cm TL) to > 6.3 kg (> 100 cm TL; Russo, unpublished data; Kusher et al. 1992). Similar patterns in rapid clasper elongation coinciding with weight gain prior to or during calcification have been found in school sharks (tope) (*Galeorhinus galeus*; Peres and Vooren 1991), bonnethead shark (*Sphyrna tiburo*; Kajiura et al. 2005), shortfin mako shark (*Isurus oxyrinchus*; Joung and Ysu 2005) and the blackspotted smoothhound (*Mustelus punctulatus*; Gracan and Lackovic 2016). Whereas most of these studies focused

Table 1. Summary of 99 males captured in six male-dominated catch events between 1977 and 1990. Examination	on
of body and clasper lengths were used to determine length "markers" when calcification of claspers occurs a	nd
onset of spermatozoa. n = no calcification or spermatozoa found at these size ranges.	

Size ran	ze range Clasper length		Number			
cm/cm	MTL	cm/cm	MTL	Specimens	Calcified	Spermatozoa
44.4 - 86.3	68.5	2.5 - 7	4.4	50	п	n
86.4 - 91.4	90.7	6.4 - 10.1	7.6	12	7	n
92.7 - 124.4	104.1	8.9 - 14.6	12.7	37	37	32



Figure 2. (a)Analysis of 99 male leopard sharks indicating the calcification of claspers (red) and the onset of spermatozoa with calcified claspers (yellow). (b) Analysis of 25 male leopard sharks from 85.7 cm to 101.6 cm TL indicating lengths at which calcification (red) and spermatozoa/calcification (yellow) occur. Blue indicates immature sharks without calcified claspers. Duplicate data points may be superimposed, one over another.

on clasper length, body length and testes weight, no study found or distinguished between the length at which calcification begins and the onset of spermatozoa production indicating any delay between the two episodes.

I conducted a separate analysis of 25 males, a subset of the larger sample of 99, ranging in TL from 85.7 cm (prior to calcification herein) to 101.6 cm (just beyond the 100% mature benchmark of 99 cm TL; Figure 2b). Here, claspers ranged from 6.3 to 12.7 cm long (9 cm MTL \pm 1.9 SD), including 7 (28%) juveniles along with 18 (72%) males with calcified claspers and only 8 (32%) of those possessing spermatozoa. These results showed that claspers grew rapidly until individuals approached >103 cm TL when growth slowed as maximum size limits in this study approached (124.4 cm TL), along with a delay in the production of spermatozoa between calcification at 86.4 cm and 93.9 cm TL when the first spermatozoa were found.

The first indication of calcification occurred at 86.4 cm TL (claspers at 7.6 cm long), which indicated that clasper length compared to body length was highly variable. For example, the largest specimen measured at 124.4 cm TL had claspers at 10.1 cm, which was equivalent to another male at 91.4 cm TL, a difference of 33 cm in TL length. Between 86.4 cm TL and 91.4 cm TL (n = 12), calcification was inconsistent appearing in only 7 (58.3%) males in this size range (Table 1). Claspers of males > 92.7 cm TL (n = 35, 106.7 cm MTL ± 7.8 SD) had a median clasper length of 12.7 cm (± 1.4 SD) and were all calcified.

The production of spermatozoa was also inconsistent beginning at 93.9 cm TL and occurring among only 50% (n = 3) of the males between 93.9 cm TL and > 98.0 cm TL (n = 6) whereupon 100% of the males possessed spermatozoa. One male at 91.4 cm TL with 6.3 cm claspers represented the smallest claspers at which calcification occurred, yet it was still immature. The largest juvenile male with uncalcified claspers and no spermatozoa measured 91.4 cm TL. The smallest adult male with calcified claspers and spermatozoa was 93.9 cm TL.

Sexual maturity, as evidenced by presence of mature spermatozoa in seminal vesicles and sperm sacs as well as the coiled condition of the vas deferens, generally did not occur until males were > 93.9 cm TL with claspers at a minimum > 10.1 cm (Figure 2). Once calcification occurred there was a growth of 7.5 cm in body length combined with an increase in clasper length of 3.1 cm indicating a delay prior to sexual maturity as first reported here. Calcification and production of spermatozoa in this study occurred at a larger body length size (+16.4 cm) than the lower end "maturity" size (70 cm TL) given by Ebert (2003) but within his overall range. The range for sexual maturity in this study is relatively narrow (93.9 - 99 cm TL) compared to published estimates (100 - 105 cm TL [Kusher et al. 1992], 70 - 120 cm TL [Ebert 2003]) based on potential breeding locations, environmental conditions, changes in climate, and research methods. There is considerable variability in body length related to clasper length of individual leopard sharks possibly attributable to availability of food, parasites, general health, genetics and other environmental factors (Figure 2).

Based on data presented herein, sexual maturity cannot be inferred from length or the calcification of claspers alone in leopard sharks, given the potential variation in critical physical factors among individuals from one locality to another. Instead, clasper length and calcification in combination with presence of spermatozoa are the only reliable measure of sexual maturity for this species. Comparisons with male leopard sharks in other known breeding locations including an increased number of larger specimens should be made to refine our understanding of the reported 50 cm TL range for sexual maturity with leopard sharks, as well as factors that influence such differences.

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LITERATURE CITED

- Ackerman, L. T. 1971. Contributions to the biology of leopard sharks, *Triakis semifasciata* (Girard) in Elkhorn Slough, Monterey, California. Thesis, Sacramento State College, Sacramento, California, USA.
- Au, D. W., and S. E. Smith. 1997. A demographic method with population density compensation for estimating productivity and yield per recruit of the leopard shark (*Triakis semifasciata*). Canadian Journal of Fisheries and Aquatic Sciences 54:415-420.
- Awruch, C. A., S. D. Frusher, N. W. Pankhurst, and J. D. Stevens. 2008. Non-lethal assessment of the reproductive characteristics for management and conservation of sharks. Marine Ecology Program Series 355:277-285.
- Barker, A. M., A. P. Nosal, E. A. Lewallen, and R. S. Burton. 2015. Genetic structure of leopard shark (*Triakis semifasciata*) populations along the Pacific Coast of North America. Journal of Experimental Marine Biology and Ecology 472:151-157.
- Cailliet, G. M. 1992. Demography of the central California population of the leopard shark (*Triakis semifasciata*). Australian Journal of Marine Freshwater Research 43:183-193.
- Carlisle, A. B., and R. M. Starr. 2009. Habitat use, residency, and seasonal distribution of female leopard sharks *Triakis semifasciata* in Elkhorn Slough, California. Marine Ecology Progress Series 380:213-228.
- Carlisle, A. B., and R. M. Starr. 2010. Tidal movements of female leopard sharks (*Tria-kis semifasciata*) in Elkhorn Slough, California. Environmental Biology of Fish 89:31-45.
- Chen, C. T., T. C. Leu, and S-J. Joung. 1988. Notes on reproduction in the scalloped hammerhead, *Sphyrna lewini*, in northeastern Taiwan waters. Fishery Bulletin 86:389-393.
- Clark, E., and K. Von Schmidt. 1965. Sharks of the central gulf coast of Florida. Bulletin of Marine Science 15:13-83.
- Conde-Moreno, M. and F. Galvan-Magaña. 2006. Reproductive biology of the Mako shark *Isurus oxyrinchus* on the south-western coast of Baja California, Mexico. Cybium 30:75-83.
- Dharmadi, F., and N. N. Wiadnyana. 2013. Biological aspects and catch fluctuation of the pelagic thresher shark, *Alopias pelagicus* from the Indian Ocean. Pages 77-84 in N. Arai, editor. Proceedings of the design symposium on conservation of the ecosystem. The13th Seastar 2000 Workshop.
- Ebert, D. A. 2003. Sharks, rays, and chimaeras of California. California Natural History Guide 71. University of California Press. Berkeley, California, USA.
- Ebert, D. A., and T. B. Ebert. 2005. Reproduction, diet, and habitat use of leopard sharks, *Triakis semifasciata* (Girard), in Humboldt Bay, California, USA. Marine and

Freshwater Research 56:1089-1098.

- Gracan, R., and G. Lackovic. 2016. Histological and morphological aspects of reproduction in male blackspotted smooth-hound *Mustelus punctulatus* in the Adriatic Sea (eastern Mediterranean Sea). Journal of Marine Biology 2016:1-6.
- Hight, B. V., and C. G. Lowe. 2007. Elevated body temperatures of adult female leopard sharks, *Triakis semifasciata*, while aggregating in shallow nearshore embayments: evidence for behavioral thermoregulation? Journal of Experimental Marine Biology and Ecology 352:114-128.
- Huveneers, C., T. I. Walker, N. M. Otway, and R. G. Harcourt. 2007. Reproductive synchrony of three sympatric species of wobbegong shark (genus *Orectolobus*) in New South Wales, Australia: reproductive parameter estimates necessary for population modeling. Marine and Freshwater Research 58:765-777.
- Jensen, C. F., L. J. Natanson, H. L. Pratt Jr., N. E. Kohler, and S. E. Campana. 2002. The reproductive biology of the porbeagle shark (*Lamna nasus*) in the western North Atlantic Ocean. Fishery Bulletin 100:727-738.
- Joung, S-J., and H-H. Hsu. 2005. Reproduction and embryonic development of the shortfin mako, *Isurus oxyrinchus* Rafinesque 1810, in the northwestern Pacific. Zoological Studies 44:487-496.
- Kajiura, S. M., J. P. Tyminski, J. B. Forni, and A. P. Summers. 2005. The sexually dimorphic cephalofoil of bonnethead sharks, *Sphyrna tiburo*. Biology Bulletin 209:1-5.
- Kusher, D. I., S. E. Smith, and G. M. Cailliet. 1992. Validated age and growth of the leopard shark, *Triakis semifasciata*, with comments on reproduction. Environmental Biology of Fishes 35:187-203.
- Launer, A. J. 2014. Sex specific distributions of leopard sharks (*Triakis semifasciata*) in an estuarine environment. M.A. Thesis. San Jose State University, San Jose, California. USA.
- Lewallen, E. A., T. W. Anderson, and A. J. Bohonak. 2007. Genetic structure of leopard shark (*Triakis semifasciata*) populations in California waters. Marine Biology 152:599-609.
- Lucifora, L. O., R. C. Menni, and A. H. Escalante. 2005. Reproduction, abundance and feeding habits of broadnose sevengill shark *Notorynchus cepedianus* in north Patagonia, Argentina. Marine Ecology Program Series 289:237-244.
- Natanson, L. J., and B. J. Gervelis. 2013. The reproductive biology of the common thresher shark in the western north Atlantic Ocean. Transactions of the American Fisheries Society 142:1546-1562.
- Nosal, A. P., E. A. Lewallen, and R. S. Burton. 2013a. Multiple paternity in leopard shark (*Triakis semifasciata*) litters sampled from a predominantly female aggregation in La Jolla, California, USA. Journal of Experimental Marine Biology and Ecology 446:110-114.
- Nosal, A. P., D. C. Cartamil, J. W. Long, M. Lührmann, N. C. Wegner, and J. B. Graham. 2013b. Demography and movement patterns of leopard sharks (*Triakis semifas-ciata*) aggregating near the head of a submarine canyon along the open coast of southern California, USA. Environmental Biology of Fishes 96:865-878.
- Nosal, A. P., A. Caillat, E. K. Kisfaludy, M. A. Royer, and N. C. Wegner. 2014. Aggregation behavior and seasonal philopatry in male and female leopard sharks *Triakis semifasciata* along the open coast of southern California, USA. Marine Ecology

Progress Series 499:157-175.

- Pratt, Jr., H. L. 1979. Reproduction in the blue shark, *Prionace glauca*. Fishery Bulletin 77:454-470.
- Pratt, Jr., H. L., and J. C. Carrier. 2004. Elasmobranch courtship and mating behavior. Pages 129-169 in N. Arai, editor. Reproductive behavior and phylogeny of elasmobranchs. Chapter five. Science Publishers Inc, Queensland, Australia.
- Peres, M. B., and C. M. Vooren. 1991. Sexual development, reproductive cycle, and fecundity of the school shark, *Galeorhinus galeus* off Southern Brazil. Fishery Bulletin 89:655-667.
- Russo, R. A. 1975. Observations on the food habits of leopard sharks, *Triakis semifasciata* and brown smoothhounds, *Mustelus henlei*. California Fish and Game 61:68-81.
- Russo, R. A. 2013. Observations on the ectoparasites of elasmobranchs in San Francisco Bay, California. California Fish and Game 99:233-236.
- Russo, R. A. 2015. Observations of predation and loss among leopard sharks and brown smoothhounds in San Francisco Bay, California. California Fish and Game 101:149-157.
- Russo, R. A. 2018. Prey of neonate leopard sharks in San Francisco Bay, California. California Fish and Game 104:173-179.
- Russo, R. A. 2019. Primary and secondary nursery areas for leopard sharks and brown smoothhound sharks in San Francisco Bay, California. California Fish and Game 105:20-29.
- Smith, S. E. 1984. Timing of vertebral-band deposition in tetracycline-injected leopard sharks. Transactions of the American Fisheries Society 113:308-313.
- Smith, S. E. 2001. Leopard shark. Pages 252-254 in W. Leet, C. Dewes, R. Klingbeil, and E. Larson, editors. California's Living Marine Resources: A Status Report. California Fish and Game Resources Agency, Sacramento, USA.
- Smith, S. E. 2005. Leopard shark mating observed off La Jolla, California. California Fish and Game 91:128-135.
- Smith, S. E. and N. J. Abramson. 1990. Leopard shark *Triakis semifasciata* distribution, mortality rate, yield, and stock replenishment estimates based on a tagging study in San Francisco Bay. Fishery Bulletin 88:371-381.
- Smith, S. E., R. A. Mitchell, and D. Fuller. 2003. Age-validation of a leopard shark, *Triakis semifasciata*, recaptured after 20 years. Fishery Bulletin 101:194-198.
- Talent, L. G. 1976. Food habits of the leopard shark, *Triakis semifasciata*, in Elkhorn Slough, Monterey Bay, California. California Fish and Game 62:286-298.
- Talent, L. G. 1985. The occurrence, seasonal distribution, and reproductive condition of elasmobranch fishes in Elkhorn Slough, California. California Fish and Game 71:210-219.

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