A PETITION TO THE STATE OF CALIFORNIA
FISH AND GAME COMMISSION

For action pursuant to Section 670.1, Title 14, California Code of Regulations (CCR) and Sections 2072 and 2073 of the Fish and Game Code relating to listing and delisting endangered and threatened species of plants and animals.

I. SPECIES BEING PETITIONED:

Common Name: Klamath Trinity Spring Chinook, Upper Klamath-Trinity River spring Chinook

Scientific Name: (Oncorhynchus tshawytscha)

II. RECOMMENDED ACTION:
(Check appropriate categories)

a. List X  b. Change Status □
   As Endangered X from
   As Threatened □ to

Or Delist □

III. AUTHORS OF PETITION:

Name: Russell “Buster” Attebury, Chairman

Address: Karuk Tribe
         PO Box 1016
         Happy Camp, CA 96039

Email: battebery@karuk.us

Phone Number: 530.493.1600

I hereby certify that, to the best of my knowledge, all statements made in this petition are true and complete.

Signature: [Signature]

Date: 7-14-18
III. AUTHORS OF PETITION:

Name: Karuna Greenberg

Address: Salmon River Restoration Council
         25631 Sawyers Bar Road
         Sawyers Bar, CA 96027

Email: karuna@srrc.org

Phone Number: 530.462.4665

I hereby certify that, to the best of my knowledge, all statements made in this petition are true and complete.

Signature: [Signature]

Date: 7/20/2018
EXECUTIVE SUMMARY

Petitioners Karuk Tribe and Salmon River Restoration Council submit this petition to list the Upper Klamath Trinity River Spring Chinook (*Oncorhynchus tshawytscha*) hereinafter referred to as UKTR Spring Chinook, as an endangered species under the California Endangered Species Act (CESA) pursuant to the California Fish and Game Code §§ 2070 et seq. This petition demonstrates that the UKTR Spring Chinook warrants listing under CESA based on the factors specified in the statute.

In 2011, Center for Biological Diversity (CBD) et al. filed a Federal Endangered Species Act (ESA) listing petition (2011 Petition) with the National Marine Fisheries Service (NMFS) to address the dramatic declines of Upper Klamath- Trinity River (UKTR) spring-run Chinook salmon. The petition was denied due to NMFS’ belief that scientific evidence did not warrant reclassification of the spring-run component of UKTR Chinook as its own Evolutionarily Significant Unit (ESU) under the Endangered Species Act (ESA). However, new evidence demonstrates sufficient differentiation between the spring-run component of UKTR Chinook, referred to here as UKTR Spring Chinook, and their fall-run counterparts, to warrant the UKTR Spring Chinook’s classification as its own ESU. On that basis, the Karuk Tribe and Salmon River Restoration Council petitioned NMFS on November 2, 2017 to reconsider its decision and list the UKTR Spring Chinook as endangered. The evidence supporting the Federal listing also supports listing the UKTR Spring Chinook as an endangered species under CESA.

UKTR Spring Chinook used to be abundant in Klamath Watershed and are important to the culture, health, and economy of the Karuk Tribe. Their survival as a species in California is threatened due to the destruction of their habitat or range, construction of dams and water diversions, disease, predation, non-existent or limited regulations, and other causes. Further information on the plight of the UKTR Spring Chinook is detailed below and in the 2011 Petition. Both the 2011 Petition and the 2017 Petition to NMFS are attached hereto and incorporated by reference. The condition of the UKTR Spring Chinook has deteriorated further since the rejection of the 2011 Petition.

For purposes of this document, UKTR Spring Chinook refers to all spring run Chinook salmon in the Klamath Basin. Within this document, UKTR Spring Chinook may also be referred to by the following names: spring-run Chinook, spring run Chinook, spring Chinook, Upper Klamath spring Chinook, UKTR spring Chinook, Trinity spring Chinook.

UKTR Spring Chinook survival is threatened by any one or a combination of the following factors (as listed in Section 670.1, Title 14, CCR):

1. present or threatened modification or destruction of its habitat;

Historically, UKTR Spring Chinook over summered and spawned in the Williamson, Sprague, and Wood River systems of southern Oregon (Hamilton et al. 2005). The construction of a complex of hydropower dams between 1917 and 1962 created a barrier to fish passage near the California/Oregon border, effectively denying salmonids access to approximately half the Klamath Basin (*Klamath Facilities Removal
Final Environmental Impact Statement/Environmental Impact Report” 2012). Young’s dam on the Scott River and Dwinnell Dam on the Shasta River also serve to deny access to historic UKTR Spring Chinook habitat (Moyle et al., 2017).

Between 1870 and the 1950’s large scale placer mining, including hydraulic and dredge mining, severely altered critical spawning and rearing habitat for UKTR Spring Chinook in the middle Klamath and its tributaries. One of the most important factors leading to the decline and continued low abundance of coho and UKTR Spring Chinook is the legacy effect of historical placer mining on channel and floodplain habitat conditions throughout the mainstem and larger tributaries of the Klamath River (Stumpf 1979). Hydraulic and dredge placer mining in the Salmon River between about 1870 and 1950, for example, led to profound and lasting changes, eroding over 1,859 acres adjacent to the mainstem and larger tributary channels and delivering an estimated 20.3 million cubic yards of sediment to the river (Hawthorne 2017, de la Fuente and Haessig 1993). Placer mining denuded floodplains and adjacent river terraces and hillslopes, reduced riparian shade cover, and exposed the stream channel and surrounding areas to increased solar radiation. (Stillwater Sciences 2018)

In addition, numerous irrigation projects throughout the Klamath Basin impact fish passage, impair water quality, and impair river and stream flows, all of which contribute to decline of UKTR Spring Chinook populations.

(5) disease;

In 2014 and 2015, 81% and 90% of juvenile Chinook salmon sampled were infected with the lethal parasite Ceratonova shasta. These high rates of infection were the result of poor water quality, low flows, and prolonged absence of flushing flows necessary to scour the river bed (Hillemeier et al. 2017). These observations led Tribes and conservation groups to file suit against the Bureau of Reclamation and National Marine Fisheries Service resulting in re-consultation on the Klamath Irrigation Project operations plan.

(6) other natural events or human-related activities.

As noted above, a century of dams, diversions, and mining has been a leading cause of UKTR Spring Chinook declines.

1. POPULATION TRENDS

Long-term population abundance data are limited for anadromous Klamath River salmonids. The earliest data primarily consist of catch records for Chinook salmon from early 20th century canneries (NMFS 2009). The data and information on Chinook salmon indicate that population levels have declined significantly since the early 20th century. NMFS 2009 review of all Klamath Basin salmonids reports that, “despite the lack of cohesive long-term data sets to assess population trends, the data that do exist indicate significant population declines in all species throughout the 1900s, leading to a current state of low abundance. Currently, a significant portion of Chinook salmon and Coho salmon that return to spawn in the Klamath River Basin are fish that were spawned in hatcheries” (NMFS 2009).
**Spring run**

UKTR Spring Chinook salmon in the Upper Klamath Basin are at extremely low abundances compared to their historical status and their current low numbers make them vulnerable to extinction. This is stated clearly in the recent status review of salmon, steelhead, and trout in California:

> The numbers of spring Chinook in the Klamath and Trinity River have remained at low levels for the past 20 years with no obvious trends, but numbers are so low...that extirpation is a distinct possibility (Moyle et al. 2008).

Similarly, NMFS (2009) acknowledges the compromised status of spring runs in the Klamath Basin based on their unique life history and the resulting dangers to survival:

> Spring run Chinook salmon enter the Klamath River from April to June of each year before migrating to smaller headwater tributaries. They require cold, clear rivers and streams with deep pools to sustain them through the warm summer months. These areas have been greatly reduced in the Basin due to dams and degradation of habitat. The spring Chinook salmon run was historically abundant and may have been the dominant run prior to commercial harvest commencing in the mid-1800s. Wild spring run Chinook salmon populations are now a remnant of their historical abundance and primarily occur in the South Fork Trinity River and Salmon River Basins (NMFS 2009)

UKTR Spring Chinook were historically abundant in the Klamath River Basin and have since declined significantly due to a variety of threats. Moyle et al. (2008) state, “while it is likely that UKTR spring Chinook were historically the most abundant run in the Klamath and Trinity Rivers (Snyder 1931, LaFaunce 1967), by the time records were being kept seriously, they had been reduced to a minor component of Klamath salmon.” In the past, populations of spring-run Chinook in the Basin likely totaled over 100,000 fish (Moyle 2002). The spring run was apparently the main run of Chinook salmon in the Klamath River until it declined steeply in the 19th century as a result of hydraulic mining, dams, diversions and fishing (Snyder 1931).

In each of four main Klamath tributaries (Sprague, Williamson, Shasta, and Scott Rivers), historic run sizes were estimated by CDFG (1990) to be at least 5,000. The runs in the Sprague, Wood, and Williamson Rivers were probably extirpated in 1895 after the construction of Copco 1 Dam (Moyle et al. 2008).

In 1968, efforts to maintain a UKTR Spring Chinook run through artificial propagation of native stock at the Iron Gate Hatchery began (Klamath Task Force 1991). During the 1970s, approximately 500 fish returned each year to the hatchery but these attempts were eventually unsuccessful as the hatchery was unable to maintain the run without a source of cold summer water (Hiser 1985, Moyle et al. 2008).

The Shasta River run, probably the largest in the middle Klamath drainage, disappeared in the early 1930s as a result of habitat degradation and blockage of access to upstream spawning areas caused by Dwinnell Dam (Moyle et al. 2008). The Scott River spring run was extirpated in the early 1970s after a variety of human causes led to depleted flows and altered habitat (Moyle 2002). Along the middle Klamath River, UKTR Spring Chinook are extirpated from their historic habitat except in the Salmon River (NRC 2004). Less than ten spring-run Chinook return annually to Elk, Indian, and Clear Creeks (Campbell and Moyle 1991).

Moyle et al. state that “UKTR spring Chinook have been largely extirpated from their historic range because their life history makes them extremely vulnerable to the combined effects of dams, mining, habitat...
degradation, and fisheries, as well as multiplicity of smaller factors” (2008). By the 1980s, UKTR Spring Chinook were largely eliminated from their habitat due to the loss or lack of access to the cold, clear water and deep pools they required for survival (NRC 2004). Spring-run Chinook in particular must contend with low flows and high temperatures during up and down-river migrations that can prevent them from reaching their destinations or significantly increase mortality during migration (Moyle et al. 1995, Trihey and Associates 1996).

In the Trinity River, UKTR Spring Chinook runs above Lewiston Dam included more than 5,000 adults in the Upper Trinity River and 1,000-5,000 fish each in the Stuart Fork Trinity River, East Fork Trinity River and Coffee Creek (CDFG 1990). These runs are now extinct. Over about the last thirty years, an average of 263 fish have been counted annually in the South Fork Trinity River, with runs as low as 59 (1988, 2005) and as high as 1,097 (1996). Between 1980 and 1989, an average of 142 spring-run Chinook were counted annually in the South Fork Trinity River; 351 fish between 1990 and 1999; and most recently 232 between 2000 and 2005. Historically, 7,000-11,000 UKTR Spring Chinook entered this stream (LaFaunce 1967) and outnumbered fall-run Chinook in the watershed. Between 1980 and 2004, an average of 18,903 UKTR Spring Chinook returned above Junction City on the main stem Trinity River. In 2004, 16,147 UKTR Spring Chinook were estimated to migrate into this area with 6,019 (37%) of fish entering Trinity River Hatchery classified as spring-run Chinook (Moyle et al. 2008). Trinity River Hatchery releases over one million juvenile spring-run Chinook every year and apparently all spawners in the main stem Trinity River are of hatchery origin (NRC 2004).

Hatcheries have severe negative effects on wild populations and are considered a high threat to both spring- and fall-run Upper Klamath Chinook (NMFS 2009, J. Katz pers. comm. 2010). Interactions between wild and hatchery fish influence abundance, spatial distribution, life history diversity and productivity. For more details on the threat of hatcheries in the Basin, see “hatcheries” in the discussion of threats in this petition. The Trinity River population of UKTR Spring Chinook is highly affected by hatchery fish and cannot be considered a viable wild population. Moyle et al. explain,

Essentially, the only viable wild population today is in the Salmon River. Other populations are either small and intermittent or heavily influenced by hatchery fish, so may not be self-sustaining and are likely to be extirpated in the near future (Moyle et al. 2008). Spring run Chinook populations in the Salmon River, exhibit high variability among years. The 2005 adult count estimate was 90 fish, the lowest on record, but in 2007 the number reached 841 (Moyle et al. 2008) and in 2009, it was 643 (CDFG personal communication). In Wooley Creek, escapement has ranged between 0 and 81 during 1968-1989, but more recent surveys suggest spring run Chinook are nearly extinct in this watershed. In 2005, only 18 spring run Chinook were observed (Moyle et al. 2008).

The National Research Council (2004) also noted the low abundance and limited distribution of spring-run Chinook in the Klamath Basin, especially those of wild spawning origin:

In the Klamath River drainage above the Trinity, only the population in the Salmon River and Wooley Creek remains; it has annual runs of 150–1,500 fish (Campbell and Moyle 1991, Barnhart 1994). Numbers of fish in the area continue to decline (Moyle 2002). Because the Trinity River run of several thousand fish per year is apparently sustained largely by the Trinity River Hatchery, the Salmon River population may be the last wild (naturally spawning) population in the basin.

Moyle et al. point out the current reliance of the spring run on this dwindling Salmon River population as they make conclusions about the status of the species:
Overall, while UKTR Spring Chinook salmon are still scattered throughout the lower Klamath and Trinity basins, the only viable wild population appears to be that in the Salmon River. Trinity River fish numbers are presumably largely influenced by fish from the Trinity River hatchery. Even if Trinity River tributary spawners are considered to be wild fish, the total number of UKTR Spring Chinook in the combined rivers rarely exceeds 1000 fish and may drop to <300 in many years (2008).

In the 2008 status review, Moyle et al. report that the UKTR Spring Chinook are “vulnerable to extinction in the next 50-100 years” based on the “fluctuating nature and small size of the Salmon River population and its localized distribution in a single watershed.”

This report produced the following table:

Table 1.

<table>
<thead>
<tr>
<th>Metric</th>
<th>Score</th>
<th>Justification</th>
</tr>
</thead>
<tbody>
<tr>
<td>Area occupied</td>
<td>2</td>
<td>Multiple populations exist including hatchery populations but only Salmon River is viable</td>
</tr>
<tr>
<td>Effective population, size</td>
<td>2</td>
<td>Although there is a hatchery stock, there are few natural spawners support the population.</td>
</tr>
<tr>
<td>Dependence on intervention</td>
<td>3</td>
<td>Hatchery program in Trinity is probably maintaining the Trinity run. The Salmon River wild population is vulnerable to extinction from both local and out-of-basin events. More human intervention necessary to preserve Klamath stock by re-establishing populations.</td>
</tr>
<tr>
<td>Tolerance</td>
<td>2</td>
<td>Temperature and other factors in summer holding areas may exceed physiological tolerances.</td>
</tr>
<tr>
<td>Genetic risk</td>
<td>2</td>
<td>Hybridization may be occurring in some watersheds with fall run fish; populations are low enough so genetic problems can develop.</td>
</tr>
<tr>
<td>Climate change</td>
<td>1</td>
<td>The Salmon River has temperatures in summer (21-23°C) that approach lethal temperatures. A 1-2°C increase in temperature could greatly reduce the amount of suitable habitat.</td>
</tr>
<tr>
<td>Average</td>
<td>2.0</td>
<td>12/6</td>
</tr>
<tr>
<td>Certainty</td>
<td>3</td>
<td>Monitoring efforts by USDA Forest Service, CDFG, tribes and local organizations give us reasonable information about status.</td>
</tr>
</tbody>
</table>

Spring-run Chinook are listed as a Species of Special Concern by California Department of Fish and Wildlife and are thus qualified to be added to the state and federal lists of threatened or endangered fish (Moyle et al. 2008). They are also considered a Sensitive Species by the Pacific Southwest Region of the US Forest Service.

Should NMFS choose not to consider the spring run of Upper Klamath Trinity River Chinook as a separate ESU or DPS, the threatened status of the spring run within the current ESU is enough rationale for listing the entire current ESU under the Endangered Species Act. Protecting the spring run from extinction is essential to maintaining the diversity of the existing ESU regardless of whether the ESU is redefined or a spring-run Chinook DPS is acknowledged. By NMFS precedent, an entire ESU may be listed under the ESA based on the threat to one of the life histories that composes it. According to Bilby et al. (2005), the
loss of many of the spring-run Chinook salmon populations from the Lower Columbia River ESU was one of the factors supporting the NMFS decision to list the ESU as threatened (NOAA 2003). The same is true of the Puget Sound Chinook ESU.

In describing foreseeable long-term trends for UKTR Spring Chinook, Moyle et al. conclude:

UKTR spring Chinook have declined from being the most abundant run in the basin, to being a tiny run in danger of extinction. There are multiple possible futures for this distinctive salmon. The two extremes are extinction and restoration to a large segment of its historic range. At the present time it is headed for extinction. Climate changes will lead to increased water temperatures and fluctuations in many portions of the basin. Without drastic management measures, climate change will likely be the final blow to wild spring Chinook in the Klamath Basin. The run will then simply be a remnant hatchery run in the Trinity River for a few decades before it finally becomes so introgressed with the fall run so that it loses its genetic and life history distinctiveness. Alternately, there is potential for UKTR spring Chinook salmon to be restored to large portions of the Klamath basin through a few decades of restoration of habitat and habitat access (e.g., Shasta River, upper Klamath Basin) (2008).

UKTR Spring Chinook require immediate protections under the Endangered Species Act if they are to persist in the Klamath Basin.

Fall run Chinook

Compared to current numbers of Chinook salmon in the Upper Klamath and Trinity Rivers, runs were much larger historically (NRC 2004) and low abundance predictions of Klamath River fall Chinook in recent years have forced severe harvest restrictions to West Coast fisheries (NMFS 2009). The vast majority of the fish today are fall-run fish of both wild and hatchery origin" (NRC 2004) and most records of Chinook salmon abundance in the Basin were taken after the initial decline of spring-run Chinook and therefore historical estimates tend to refer primarily to the fall run (Moyle et al. 2008). NMFS (2009) refers to sizable historic estimates in the Basin: “Based on records of commercial harvest, fall run Chinook are likely to have numbered 400,000 to 500,000 in the early 1900s. Runs in the last several decades have ranged from below 50,000 to 225,000 fish. These runs are substantially lower than historic levels.” Snyder (1931) provided an early estimate of 141,000 fish, based on the 1912 fishery catch of 1,384,000 pounds of packed salmon.

Moffett and Smith (1950) then estimated the Klamath River Chinook runs to be about 200,000 fish annually, from commercial fishery data from between 1915 and 1943. USFWS (1979) combined these statistics to approximate an annual catch and escapement of about 300,000 to 400,000 fish for the Klamath River system from 1915-1928 (Moyle et al. 2008).

The National Research Council (2004) reviewed historical estimates of fall Chinook:

…the river harvest alone in 1916–1927 was 35,000–70,000 fish (as estimated from Snyder’s data showing an average weight of 14 lb/fish and a harvest of 500,000–1,000,000 lb each year). If, as Snyder’s data suggest, the river harvest was roughly 25% of the ocean harvest in this period, annual total catches were probably 120,000–250,000 fish. This in turn suggests that the number of potential spawners in the river was considerably higher than the number spawning in the river today. Since 1978, annual escapement has varied from 30,000 to 230,000 adults. In both 2000 and 2001, runs were over 200,000 fish. If it is assumed that fish returning to the hatcheries are, on the average, 30% of the population and that 30% of the natural spawners are also hatchery fish, then roughly half the run consists of salmon of natural origin (including progeny of hatchery fish that spawned in the wild).
At the Klamathon Racks, a fish counting station close to the location of Iron Gate Dam, an estimated annual average of 12,086 Chinook were counted between 1925-1949, and the number declined to an average of 3,000 between 1956-1969 (USFWS 1979). In 1965, the Klamath River Basin was reported to contribute 66% (168,000) of Chinook salmon spawning in California’s coastal basins (CDFG 1965). This production was distributed between the Klamath (88,000 fish) and Trinity (80,000 fish) basins, with approximately 30% of the Klamath Basin fish originating in the Shasta (20,000 fish), Scott (8,000 fish), and Salmon (10,000 fish) Rivers (Moyle et al. 2008). Snyder (1931) recorded the Shasta River as the best spawning tributary in the basin. It has since seen a marked decline in the number of fish returning. Leidy and Leidy (1984) estimated an annual average abundance of 43,752 Chinook from 1930-1937; 18,266 between 1938 and 1946; 10,000 between 1950 and 1969; and 9,328 from 1970-1976. A review of recent escapement into the Shasta River found an annual escapement of 6,032 fish from 1978-1995, and an escapement of 4,889 fish between 1995 and 2006 (CDFG 2006). In the Scott River, fall Chinook escapement averaged 5,349 fish between 1978 and 1996 and 6,380 fish between 1996 and 2006 (Moyle et al. 2008).

The National Research Council (2004) notes the drop in the population in the Shasta River as an important contributor to the overall decline of Upper Klamath Chinook:

Additional evidence of decline is the exclusion of salmon from the river and its tributaries above Iron Gate Dam in Oregon, where fairly large numbers spawned, and the documented decline of the runs in the Shasta River. The Shasta River once was one of the most productive salmon streams in California because of its combination of continuous flows of cold water from springs, low gradients, and naturally productive waters. The run was probably already in decline by the 1930s, when as many as 80,000 spawners were observed. By 1948, the all-time low of 37 fish was reached. Since then, run sizes have been variable but have mostly been well below 10,000. Wales (1951) noted that the decline had multiple causes, most related to fisheries and land use in the basin, but laid much of the blame on Klamath River lampreys: the lampreys preyed extensively on the salmon in the main stem when low flows delayed their entry into the Shasta River.

In the Trinity River, Coots (1967) estimated an annual run of about 80,000 fish. Hallock et al. (1970) reported about 40,000 Chinook salmon entered the Trinity River above the South Fork. Burton et al. (1977 in USFWS 1979) estimated that 30,500 Chinook below Lewiston Dam on the Trinity River escaped between 1968 and 1972. The average fall Chinook run in the Trinity River between 1978 and 1995 was 34,512. This average declined between 1996 and 2006 to 23,463 fish (CDFG 2007).

The total in river escapement into this ESU ranged from 34,425 to 245,542 fish with an average 5-year geometric mean of 112,317 fish between 1978 and 2006 (Moyle et al. 2008). A large proportion of these fish are of hatchery origin and therefore do not contribute, and even constitute a threat, to the long-term persistence of Chinook salmon in the Basin and (Bilby et al. 2005).

Hatcheries have played a major role in fall-run Chinook salmon abundance since the 1960s (Moyle et al. 2008). Approximately 67% of hatchery releases have been fall-run Chinook from Iron Gate and Lewiston hatcheries (Myers et al 1998). Between seven and twelve million juveniles have been released annually (NRC 2004). Between 1997 and 2000, an average of 61% of the juveniles captured at the Big Bar outmigrant trap were hatchery origin fish (USFWS 2001) and at the Willow Creek trap on the Trinity River, between 1997 and 2000, 53% and 67% of the Chinook captured in the spring and fall were hatchery-origin fish, respectively (USFWS 2001). Some naturally-spawning fish are actually hatchery strays. Based on coded wire tag expansion multipliers, as much as 40% (Shasta River) of annual escapement consists of hatchery strays (R. Quinones, unpublished data as cited by J. Katz, pers. comm. 2010). As this region becomes dominated by hatchery fish, wild fish are threatened by greater competition, predation, disease transmission, and reduced fitness due to interbreeding with hatchery fish. As a region becomes dependent
on hatchery fish, its ability to recover as a wild-spawning population of fish is highly compromised (ISAB 2005)

Upper Klamath-Trinity River fall-run Chinook are a US Forest Service Sensitive Species. They are managed by CDFW for sport, tribal, and ocean fisheries.

According to the Moyle et al. (2008) status review, fall-run Chinook have declined from historical numbers of between 125,000 and 250,000 fish returning annually to the Basin to an average run size of about 120,000 since 1978 (from tables compiled by CDFG). Numbers in the past 25 years have sometimes reached this historical range but lower numbers are now typical and current runs depend heavily on hatchery production. Fall-run Chinook have experienced a major downward trend in recent years, especially as a result of the 2002 fish kill in the lower river. Climate change will lead to even more threatening conditions for this ESU (Barr et al. 2010).

The Moyle et al. status review summarizes the long term trends for Klamath Basin Fall-run Chinook and reports:

There is little reason to be optimistic about long-term trends in the future without major changes in watershed management. High summer water temperatures are a major driver of UKTR Chinook survival and they are likely to increase under most climate change scenarios. Likewise, changes in ocean conditions may cause decreased survival of fish once they leave the river (Moyle et al. 2008).

The report also points out that the increased reliance of the fall run on hatchery production is “likely masking a decline of wild production in the Klamath-Trinity basins”. Moyle et al. cited a 2005 report stating, “models evaluating limiting factors and habitat availability for UKTR Chinook salmon suggest that crucial steps need to be taken soon to increase UKTR fall Chinook spawners” (citing Bartholow and Henrikson 2005).

The National Research Council acknowledges that while fall-run Chinook have declined significantly, they may be good candidates for recovery under the right management reporting, “the fishery of the Klamath is particularly important…because of the possibility of maintaining it (NRC 2004). NRC goes on to note that both adults migrating upstream and juveniles moving downstream face water temperatures that are bioenergetically unsuitable or even lethal and that the vulnerability of the run to stressful conditions was dramatically demonstrated by the mortality of thousands of adult Chinook in the lower river in late September 2002.

Both spring- and fall-run Chinook have declined in the Klamath Basin with spring-run Chinook demonstrating the most drastic trends of reduction. The spring run requires protections under the ESA in order to avoid extinction. Maintaining the spring run is essential to supporting the diversity of the current ESU and the vulnerability of this run in particular could justify listing the entire Upper Klamath-Trinity Rivers ESU according to the ESA.

2. RANGE AND DISTRIBUTION

Spring- and fall-run Chinook distributions have been affected differently by conditions in the Basin because spring-run Chinook enter freshwater earlier than fall-run Chinook, and historically traveled much greater distances upstream (Hamilton et al. 2005).

Spring-run Chinook salmon were historically found throughout the Klamath Basin. They used suitable
reaches in the larger tributaries such as the Salmon River and, flows permitting, they also accessed smaller tributaries for holding and spawning. They were once especially abundant in the major tributary basins of the Klamath and Trinity Rivers, such as the Salmon, Scott, Shasta, South Fork and North Fork Trinity Rivers (Moyle et al. 2008). Spring run Chinook were once also widely distributed throughout the Basin above the current sites of dams, attaining holding and spawning grounds on the Sprague, Williamson and Wood Rivers above Upper Klamath Lake (Moyle et al. 2008). This habitat was blocked below Klamath Falls in 1912 by construction of Copco 1 Dam (Hamilton et al. 2005). The construction of Dwinnell Dam in 1925 on the Shasta River eliminated access to UKTR Spring Chinook habitat in that watershed.

Currently, only the Salmon River, a major freshwater tributary to the Klamath River, maintains a viable population in the Klamath River Basin (Moyle et al. 2008). Approximately 177 km (110 mi) of habitat is accessible to spring-run Chinook in the Salmon River (West 1991) but most of it is underutilized or unsuitable (Moyle et al. 2008). The South Fork Salmon River holds the majority of the spawning population but smaller tributaries where spring Chinook redds have been found in the Salmon River Basin include Wooley, Nordheimer, Knownothing, and Methodist Creeks. In addition, there are dwindling populations of spring Chinook in Elk, Indian, Clear Creeks (Moyle et al. 2008).

In the Trinity River Basin, spring Chinook salmon once spawned in the East Fork, Stuart Fork, Coffee Creek, and the main stem Upper Trinity River (Campbell and Moyle 1991). The construction of Lewiston Dam in 1964 blocked access to 56 km of spawning and nursery habitat on the main stem Trinity River (Moffett and Smith 1950).

Currently, Trinity River spring Chinook are present in small numbers in Hayfork and Canyon Creek, as well as in the North Fork Trinity, South Fork Trinity and New Rivers (Moyle et al. 2008). The Trinity River Hatchery releases over 1 million juvenile spring run Chinook every year, usually in the first week of June. Apparently, all spawners in the main-stem Trinity River below Lewiston Dam are of hatchery origin (NRC 2004).

The distribution of fall-run Upper Klamath Chinook has been less affected by dam construction because of their lower reliance on upstream spawning habitat. They are found in all major tributaries above the confluence of the Klamath and Trinity rivers and in the river main stems (Moyle et al. 2008). Fall-run Chinook return to both Iron Gate and Trinity River Hatcheries.

Upper Klamath fall Chinook salmon once ascended to spawn in habit, now-blocked, in middle Klamath tributaries (Jenny Creek, Shovel Creek, and Fall Creek), and in rivers in the Upper Klamath Basin, especially in wetter years (Hamilton et al. 2005). On the lower Klamath River, tributaries providing suitable spawning habitat include Bogus, Beaver, Grider, Thompson, Indian, Elk, Clear, Dillon, Wooley, Camp, Red Cap, and Bluff Creeks (Moyle et al. 2008). The Salmon, Shasta and Scott Rivers were historically and remain among the most important spawning areas for fall-run Chinook, when sufficient flows are present. Spawning consistently occurs in the main stem Klamath River between Iron Gate Dam and Indian Creek, with the two areas of greatest spawning density typically occurring between Bogus Creek and the Shasta River and between China Creek and Indian Creek (Magneson 2006).

On the Trinity River, UKTR Spring Chinook once ascended above the site of Lewiston Dam to spawn as far upstream as Ramshorn Creek and historically, the majority of Trinity River fall Chinook spawning was located between the North Fork Trinity River and Ramshorn Creek. Currently, spawning is confined to the approximately 100 km between Lewiston Dam and Cedar Flat (Moyle et al. 2008). Important historic spawning tributaries above Lewiston Dam include the Stuart Fork, Browns and Rush Creeks (Moffett and Smith 1950). The distribution of redds in the Trinity River is highly variable (Moyle et al. 2008). The reaches closest to the Trinity Hatchery contain significant spawning but there is great variability in use of spawning habitat in reaches between the North Fork Trinity River and Cedar Flats (Quilhiullalt 1999). Additional
tributaries contain spawning fall-run Chinook salmon in the Trinity River including the North Fork, New River, Canyon Creek, and Mill Creek (Moyle et al. 2008). In the South Fork, fall-run Chinook once spawned in the lower 30 miles up to Hyampom, and in the lower 2.7 miles of Hayfork Creek (LaFaunce 1967).

The distributions of both the fall and spring runs of UKTR Chinook have contracted since the end of the 19th century. Because of the unique life history of the spring run, it has been most damaged by these changes, directly causing extirpation of several populations and making the run vulnerable to future genetic introgression with the other life history type in the Basin.

3. ABUNDANCE

Please see #1, Population Trend.

4. LIFE HISTORY (SPECIES DESCRIPTION, BIOLOGY, AND ECOLOGY)

A. Life Cycle and Physiology

The Chinook salmon life cycle begins when an adult female prepares a nest, called a "redd," by digging in a stream area with suitable gravel type, water depth and water speed (McCullough 1999). Body size, which is related to age, may be an important factor in migration and redd construction success. All Chinook salmon tend to use spawning sites with large gravel and significant water flow through the gravel. Deep water with sufficient sub-gravel flow is essential to provide oxygen to the eggs and remove metabolic waste. Thus, limited sub-gravel flow resulting in low oxygen concentrations are linked to egg mortality (Allen and Hassler 1986). Excess silt in the water can also block water flow through gravel (Healey 1991).

Female Chinook lay 2,000 to 17,000 eggs, each about nine millimeters in diameter (Healey 1991). One or more males then release sperm into the redd before females cover it with gravel (Allen and Hassler 1986). Once the eggs have been fertilized, adult Chinook guard the nest briefly (up to a month) before dying. Egg mortality can result from limited oxygenation, extreme temperatures, predation and toxic chemicals (Healey 1991). Depending on water temperature, the eggs will hatch three to five months after being laid, which ensures young salmon (termed “alevins”) emerge when river conditions are best.

Alevins remain in the spawning habitat for at least two to four weeks until their yolk sacs are completely used. Like the eggs, Alevins require adequate water flow through the gravel for growth and survival (Nawa and Frissell 1993). Once the alevin consumes its yolk sac, it enters the fry-fingerling stage and begins feeding and socializing. Some fry remain in the spawning grounds, while others begin their tail-first migration to the ocean soon after emerging from the redd. A number of factors such as water flow, food availability, temperature and competition may influence when the fry and fingerlings migrate.

The vast majority of juvenile fall Chinook migrate within one year of hatching whereas the majority of spring Chinook migrate after one year. Moyle et al. (2008) reports on a study by Sullivan (1989) which identified three distinct types of juvenile freshwater life history strategies for UKTR fall Chinook. The majority of fish fall into the first and second categories: 1) rapid migration following emergence, and 2) tributary or cool-water area rearing through the summer and fall migration. A small percentage of fish were in a third category, which remained in freshwater through winter and migrated to the estuary as yearlings.

Juvenile Chinook undergo smoltification, a physiological transformation that prepares the fish for the increased salinity in the ocean (Weitkamp 2001). Fall Chinook grow to smolt size near the end of their time in the estuary, whereas spring Chinook turn into large smolts before they reach the estuary (Healey 1991). The amount of time a juvenile salmon spends in freshwater varies. Some male Chinook salmon mature in freshwater while others spend less than a year in freshwater, depending on genetic and environmental
factors (NRC 2004). Juvenile fall-run Chinook spend less than a year in the fresh water of the Klamath River Basin, allowing the juveniles to avoid unfavorable late summer stream conditions (Healey 1991, Moyle 2002). Spring-run Chinook however, spend at least one year in freshwater before migrating to the ocean (Healey 1991).

The majority of spawners returning to the Klamath River Basin are age three fish. This reflects heavy mortality of older and larger fish in ocean fisheries. Some four, five, and six year old fish are found spawning (Moyle et al. 2008). Some fish return from the ocean within two or three months, in the case of a small number of yearling males (called jack salmon). These jack salmon constituted 2-51 percent of the annual Klamath River Chinook salmon numbers between 1978 and 2006 (Game 2006 as cited in Moyle et al. 2008).

In the ocean, Klamath River Chinook salmon are found in the California Current system off the California and Oregon coasts. Moyle et al. (2008) reports that salmon follow predictable ocean migration routes. Chinook recaptured from the Klamath River generally use ocean areas that exhibit temperatures between 8° and 12°C (Hinke et al. 2005). Chinook salmon from the Klamath and Trinity hatcheries were observed in August south of Cape Blanco (Brodeur et al. 2004).

Adult Chinook return to freshwater to spawn and die. During ocean residence, salmon build up stores of body fat and cease feeding during upstream migration. Spring-run Chinook, enter the Klamath River between March and July and spawn between late August and September, while fall-run Chinook enter the river between July and October and spawn between September and January (Myers et al. 1998).

The timing of upriver migration into freshwater and spawning of Chinook salmon is likely defined by water temperature and flow regimes. For example, data collected primarily from Columbia River migration suggests that spring Chinook migrate at 3.3-13.3°C and fall Chinook migrate at 10.6-19.4°C (McCullough 1999).

In general, salmon runs today occur later than they did historically. The current fall run of Chinook occurred earlier and was known as the summer run in the past (Snyder 1931). For example, Moyle et al. (2008) reports that run timing on the Shasta and Klamathon Racks appears to occur one to four weeks later than historic run timing. Although run timing has responded to accommodate warmer stream conditions, temperatures are likely still stressful to migrating salmon and may result in increased mortality of spawning adults (NRC 2004).

Chinook rely primarily on olfaction memory and partially on sight to find their way back to their natal stream. Some evidence suggests that fall Chinook seem to have a stronger homing instinct than spring Chinook (Healey 1991). Adults primarily migrate during the day, which exposes them to higher temperatures that may inhibit their migration or increase mortality. After spawning, adult females defend their eggs; thereafter both male and female salmon deteriorate rapidly, often developing a fungal disease, and die within 2-4 weeks (Allen and Hassler 1986).

Spring Chinook

The variation of life history between spring and fall Chinook is relevant to the difference in status between the runs. Many of these are shown below, in Table 1. Unlike fall Chinook, spring Chinook in the Klamath River Basin utilize streams and tributaries a great deal during their life cycle. Juveniles usually reside in streams for at least one year before migrating to the ocean (Healey 1991). These juveniles are much more dependent on freshwater stream ecosystems because of their extended residence in these areas.

Spring Chinook adults return to the Klamath River between March and July before their gonads have fully developed (Moyle et al. 2008). The majority of late entry spring Chinook in the Klamath system are of
hatchery origin (Barnhardt 1994, NRC 2004). Moyle et al. (2008) note a study which identified adult Trinity River spring Chinook migration continuing until October. They argue however that given this late timing, it is unclear if these fish are sexually mature and able to spawn with spring Chinook adults already in the system. Also, they report, that because this late spring run is limited to the Trinity River, it is possible these fish represent hybrid spring and fall Chinook created by hatchery practices (Moyle et al. 2008).

Spring adults typically hold in deep (greater than two meters) freshwater pools for 2-4 months to allow their gonads to develop before spawning (NRC 2004). These behaviors allow spring Chinook salmon to spawn much further upstream than fall Chinook, who must contend with higher temperatures and lower flows in the lower Klamath during the late summer months (Moyle 2002). Spring Chinook spawning peaks in October.

After emerging from the redds between March and early June, spring Chinook fry remain in the same cold headwaters as holding adults for the summer (West 1991). Some juveniles migrate downstream beginning in October, but most remain in the headwaters until the spring (Trihey and Associates 1996).

Spring Chinook typically spend more time in freshwater streams, both during their downriver and spawning migrations. They are therefore more vulnerable to adverse stream conditions. The increased time spent in streams and greater distance of migration are disadvantages to survival in the current system because spring Chinook experience low flows and high temperatures during migration that can prevent them from reaching their destinations and significantly increase mortality during migration (Moyle et al. 1995, Trihey and Associates 1996).

| Summary of Life Cycle and Physiological Differences between Spring and Fall Chinook in the Upper Klamath River Basin |
|-------------------------------------------------|-------------------------------------------------|-------------------------------------------------|
| Adult migration immigration | Spring Chinook: Between March and May between March and July with a peak between May and early June. Spring Chinook migrate before reaching sexual maturity and holdover in deep (greater than two meters) freshwater pools for 2-4 months prior to spawning. | Fall Chinook: Between mid July and late October. Migration and spawning occur under decreasing temperature regimes. | Citations: Barnhart 1994, NRC 2004, Myers et al. 1998, Moyle et al. 2008 |
| Holding elevation | Historically, overlap of spawning areas was rare between spring and fall Chinook because spring Chinook spawned well upstream of fall Chinook before the construction of dams. Spatial separation between the two runs in the Klamath-Trinity system occurs at approximately 1,700 feet. | Downstream of 1,700 feet elevation (must contend with higher temperatures and lower flows during migration in the late summer months. | Citations: Moyle 2002, Moyle et al. 2008 |
| Spawning | Begins between late August and September, peaks in October. | Between September and January. | Citations: Myers et al. 1998, Moyle et al. 2008 |
| Emergence from gravel | Between March and early June, remain in the same cold headwaters as holding | Late winter or spring, timing dictated by water temperature. | Citations: Trihey and Associates 1996, Moyle et al. 2008 |
### B. Diet

Chinook salmon diet varies depending on growth stage. As alevins, the young fish rely on nutrients provided by the yolk sack attached to the body until leaving the redd after a few weeks. After emerging from the gravel, young fry begin to feed independently. Juveniles feed in streambeds before gaining strength to make the journey to the ocean. During this time, fry feed on terrestrial and aquatic insects and amphipods.

As juveniles migrate toward the ocean, they may spend months in estuarine environments feeding on plankton, small fish, insects, or mollusks. Small fry feed primarily on zooplankton and invertebrates, while larger smolts feed on insects and other small fish (i.e., chironomid larvae, chum salmon fry and juvenile herring; Healey 1991).

Juvenile Chinook salmon can feed and grow at continuous temperatures up to 24°C when food is abundant and conditions are not stressful (Myrick and Cech 2001). In the late summer, juveniles seek out cooler temperatures in refuge pools along the Lower Klamath River, where they may experience intraspecies competition for food.

At sea, where the bulk of feeding and growth is done, adult Chinook typically feed on small marine fish, crustaceans, and mollusks (i.e., squid). Adult Chinook grow quickly in the estuary and gain body mass during their time at sea, building fat reserves that are required for upstream migration and spawning. During the upstream migration, Chinook do not feed and rely on stored energy while traveling hundreds of miles.

### 5. KIND OF HABITAT NECESSARY FOR SURVIVAL

The variety of habitats Chinook salmon encounter means that they require a number of particular conditions in order to survive and reproduce. Chinook salmon in the Klamath-Trinity River Basin occupy the main stem rivers and tributaries during migration, spawning, and rearing. They also occupy the estuary and open ocean for variable time periods during maturation. Chinook salmon habitat use and requirements are best studied for their time spent in freshwater although ocean conditions are also significant to the survival and viability of these populations.
Upper Klamath Chinook salmon migrate from the open ocean to spawning habitat, typically to the same place where they hatched. During this time, they are in a stressed condition due to their reliance on stored energy to complete the long journey upstream, leaving them highly susceptible to additional environmental stressors. This was clearly a factor during the 2002 fish kill when inadequate stream flows, temperature conditions, and the resultant crowding of fish led to disease outbreaks and mass mortality. Chinook salmon require access to spawning habitat in the main stem rivers and tributaries, cold water, cool pools in which to hold, clean spawning gravel, and particular dissolved oxygen levels, water velocities, and turbidity levels in order to successfully migrate and spawn. Access to spawning habitat is threatened by physical conditions including the existence of impassable dams, which caused the extirpation of several populations of spring run Chinook. Also, the ongoing variability in water flows does not allow Chinook salmon to access certain streams for spawning.

During migration and spawning, low water temperatures are crucial to success of Chinook salmon. Under warm conditions, salmon cease their upstream migration and instead hold in cooler pools. Upper Klamath spring Chinook enter the Klamath estuary during a period when river water temperatures are at or above optimal holding temperatures (Moyle et al. 2008). In June, temperatures in the Lower Klamath River typically rise above 20°C and can be as high as 25°C in August (Moyle et al. 2008). Prior to entering fresh water, Spring Chinook use thermal refuges in the estuarine salt wedge and associated near shore ocean habitat (Strange 2003). Strange (2005) found that when daily water temperatures were on the rise, Chinook migrated upstream until temperatures reached 22°C; when temperatures were decreasing, fish continued to migrate upstream at water temperatures of up to 23.5°C. Optimal adult holding habitat for spring Chinook is characterized by pools or runs greater than one meter deep with cool summer temperatures (<20°C), all day riparian shade, little human disturbance, and underwater cover such as bedrock ledges, boulders, or large woody debris (West 1991). Because the Salmon River and its forks regularly warm to summer daytime peaks of 21-22°C, presumably the best holding habitats are deep pools with cold water sources, such as those at the mouths of tributaries, or are deep enough to be subject to thermal stratification (Moyle et al. 2008). Due to the typically higher flows during spring Chinook migration, Salmon River spring Chinook are typically able to move high into the system, allowing them to reach areas with more optimal river temperatures, however this is not as feasible during drought years. UKTR fall Klamath fall Chinook salmon enter the Klamath estuary for only a short period prior to spawning. However, unfavorable temperatures can be found in the Klamath estuary and lower river during this period and chronic exposure of migrating adults to temperatures of even 17°-20°C is detrimental (Moyle et al. 2008). Optimal spawning temperatures for Chinook salmon are less than 13°C (McCollough 1991) and fall temperatures are usually within this range in the Trinity River (Quilhillalt 1999). Magneson (2006) reported water temperatures up to 14.5°C during spawner surveys in 2005. The Shasta River historically was the system’s most reliable spawning tributary from a temperature perspective (Snyder 1931), but diversions of cold water have greatly diminished its capacity to support salmon (Moyle et al. 2008).

According to McCullough (1999), adults are more sensitive to higher temperatures than juveniles, as higher temperatures can increase the adults’ metabolic rate and deplete their energy reserves, weaken their immune system, increase exposure to diseases, and prevent migration. Also, temperatures at or above 15.6°C can increase the onset of diseases (Allen and Hassler 1986). Riparian vegetation is critical as it provides much needed shade to cool the water (Moyle 2002) and creating “thermal refugia” in which fish can escape high temperatures. The presence of cold water in the Basin is threatened by dams, water withdrawals, as well as logging and grazing which decrease riparian vegetation.

Spring Chinook migrate earlier before their gonads are fully developed and then hold in deep cool pools before spawning. Therefore, the presence of deep cold-water pools is essential to the survival of spring-run fish in particular. Dams, water withdrawals, logging, mining, and grazing all contribute to lower water levels
in the Basin and threaten the presence of deep pools essential for spring Chinook. Spring Chinook are also more sensitive to high temperatures than fall Chinook (Allen and Hassler 1986).

According to the National Research Council (2004), Migrating adults also need dissolved oxygen levels above five mg/l, deep water (deeper than 24 cm), breaks from high water velocity, and water turbidity below 4,000 ppm (NRC 2004).

Spawning gravel also must be free of excessive sediment such that water flow can bring dissolved oxygen to the eggs and newly hatched fish. With too much sediment, incubating eggs are smothered and reproductive success rate declines significantly. In a study on the Shasta River (Ricker 1997), six out of seven locations, had levels of fine sediment high enough to significantly reduce fry emergence rates and embryo survival. Logging, mining, and grazing increase sediment in Chinook spawning habitat in the Basin. Spawning occurs primarily in habitats with large cobbles loosely imbedded in gravel and with sufficient flows for subsurface infiltration to provide oxygen for developing embryos (Moyle et al. 2008). In a survey of Trinity River Chinook redds, Evenson (2001) found embryo burial depths averaged 22.5-30cm suggesting minimum depths of spawning gravels needed. Regardless of depth, the key to successful spawning is having adequate flows of water (Moyle et al. 2008).

Rearing

During rearing and migration, Chinook require certain temperatures, habitat diversity, and water quality characteristics.

After hatching, juvenile Chinook require rearing habitat before making their migration to the estuary and to the ocean. Ideal fry rearing temperature is estimated at 13°C and temperatures above 17°C are linked with increased stress, predation, and disease. High water temperatures can prevent smoltification, an essential process that prepares fish to leave freshwater habitat (McCullough 1999).

Stream temperature during migration is critical, as prolonged exposure to temperatures of 22-24°C has resulted in high mortality for migrating smolts, and juveniles who transform into smolts above 18°C may have low survival odds at sea (Baker et al. 1995, Myrick and Cech 2001). Vegetation provides relief from high temperatures, as well as shelter from predators (Moyle 2002). Logging, mining, and grazing all have reduced streamside vegetation in the Basin.

Habitat diversity is important for juvenile Chinook survival, as juveniles face predation by fish and invertebrates, as well as competition for rearing habitat from other salmonids (hatchery Chinook and Steelhead; Healey 1991, Kelsey et al. 2002). Chinook require the correct grades of gravel, the right depths and prevalence of deep pools as well as the existence of large woody debris and the right incidence of riffles (Montgomery et al. 1999). This allows for a variety of habitats which are required by Chinook at different life stages.

Chinook fry may compete for shallow water rearing habitat with hatchery fish and steelhead. Increased river flows mitigate this competition and help Chinook survival by increasing habitat on the river’s edge, where fry (under 50 mm) feed and hide from predators (NRC 2004).

As juvenile Chinook migrate down river, they prefer boulder and rubble substrate, low turbidity and water velocity slower than 30 cms⁻¹ (Healey 1991). These conditions allow juveniles to use the faster-moving water in the center of the river for drift feeding, while resting in the slower areas (Trihey and Associates 1996). Smaller fish tend to stay in the slower-moving water near the banks of the river. High water turbidity threatens Chinook (Bash et al 2001) and in the Klamath Basin, logging and grazing both serve to increase turbidity.
Juvenile Chinook require high levels of dissolved oxygen (DO). Low DO levels decrease alevin and fry survival; decrease successful Chinook egg incubation rates; decrease the growth rate for surviving alevins, embryos, and fry; force alevins and juveniles to move to areas with higher DO; and negatively impact the swimming ability of juvenile Chinook (NCWCB 2010). If DO levels average lower than 3-3.3 mg/L, 50% mortality of juvenile salmonids is likely, while in water above 20˚C, daily minimum DO levels of 2.6 mg/L are required to avoid 50% mortality (NCWCB 2010). Factors in the Basin which contribute to sub-optimal DO levels include chemical pollution, logging, and dams.

Chinook salmon also require pH levels that are not too high. Even high pH levels which are not directly lethal to salmonids can cause severe harms to Upper Klamath Chinook (NCWCB 2010), including decreased activity levels, increased stress responses, a decrease or cessation of feeding, and a loss of equilibrium (NCWCB 2010). The Klamath River’s pH in the summer often rises above 8.5, and sometimes reaches 9. At the Miller Island Boat Camp in 2008, the river’s pH in early July, measured daily, had several consecutive days with pH values ranging from 9.06-9.53 (USGS 2009, Appendix B). Few studies directly examine the effects of high pH values on Chinook salmon. However, rainbow trout are stressed by pH values above 9 and generally die if the pH value rises above 9.4 (NCWCB 2010). Nutrient loading of stream systems including those caused by agricultural runoff can lead to higher pH in river systems (NCWCB 2010).

Once juvenile Chinook reach the estuary, less developed fall-run fry remain and seek out the tidal channel where the banks are low, while larger spring run smolts prefer near shore areas near the mouth of the river (Healey 1991). Juveniles change location with the tide as the salinity of the water changes. Larger Chinook smolts seek out deeper pools to avoid light.

Ocean

Once Chinook enter the ocean, most reside at depths of 40-80 meters (Healey 1991). Some research suggests that spring Chinook migrate further offshore, while fall Chinook tend to stay near the shore and close to their river (Allen and Hassler 1986). In the marine environment, Chinook salmon require nutrient-rich, cold waters associated with high productivity and higher rates of salmonid survival. Warm ocean regimes are characterized by lower ocean productivity which can affect salmon by limiting the availability of nutrients regulating the food supply and increasing the competition for food. Climate and atmospheric conditions can affect these conditions (NMFS 1998). In order to survive in the marine environment, Chinook salmon also require favorable predator distribution and abundance. This can be affected by a variety of factors including large scale weather patterns such as El Niño. NMFS (1998) cites several studies which indicate associations between salmon survival during the first few months at sea and factors such as sea surface temperature and salinity.

6. FACTORS AFFECTING ABILITY TO SURVIVE AND REPRODUCE

Discuss the basis for the threats to the species or subspecies, or to each population, occurrence or portion of range (as appropriate) due to one or more of the following factors:

(1) present or threatened modification or destruction of its habitat;

Dams

Dams in the Klamath Basin have destroyed Chinook habitat and forced modifications to the UKTR Chinook’s range. Most fisheries biologists rate dams as being a “high” threat to both spring and fall Klamath Chinook salmon (NMFS 2009, J. Katz, pers. comm. 2010). The sequestration of habitat behind
dams has acted as a major limiting factor to Klamath Basin Chinook populations, especially spring-run Chinook and the presence of these dams has likely inhibited recovery in years when conditions would otherwise have permitted it. In addition, dams affect the quality of habitat downstream by preventing spawning gravel from traveling downstream (Moyle et al. 2008), releasing limited, warm, and sometimes toxic water, and dictating unnatural stream morphology or structure.

Dams have been a barrier for Upper Klamath Chinook since 1912, when construction of Copco 1 Dam began (Hamilton 2016), closely followed by Copco 2 Dam in 1925. Iron Gate Dam represents the current extent of upstream migration for Chinook on the Klamath River. It was built in 1962 to produce hydroelectric power as well as to regulate the wildly varying flows released by the Copco 1 and 2 Dams. In 1963, Lewiston Dam was built and became the current upstream limit to Chinook migration in the Trinity River.

UKTR spring Chinook have been particularly affected by dams, as they spawned largely in areas that are now unavailable (Moyle et al. 2008). Above Iron Gate Dam, there are approximately 970 km of blocked Chinook habitat (Hamilton et al. 2005). The construction of Dwinell Dam in 1926 on the Shasta River blocked habitat that led to the disappearance of the Shasta River spring run (NRC 2004). Half of the available spawning habitat in the Trinity River Basin was blocked by Lewiston Dam (Myers et al. 1998). These restrictions to Chinook spawning range have been widely implicated in the decline of Upper Klamath Chinook populations, particularly spring run populations, throughout the Klamath Basin. Another result of limits to upstream habitat has been the introgression of the spring and fall runs, leading to a decline in genetic variability and further threatening the long-term viability of the ESU (Moyle et al. 2008).

Dams also contribute to a reduction in spawning gravel. Gravel can be caught in reservoirs behind dams and is unable to travel downstream to spawning habitat. Limited access to spawning gravel has been reported to affect spawning prevalence in both the Shasta and Klamath Rivers (Kondolf 2000).

Dams have negative effects on downstream water quality. The water which is held behind dams is both stagnant and warm and serves to dramatically increase the prevalence of Harmful Algal Blooms (HABs) in reservoirs and downstream (Humborg et al. 2000, Anderson et al. 2002). Dams also decrease levels of dissolved silicon in the water, leading to changes and imbalances in downstream phytoplankton communities and increased human water use causes raised levels of nitrogen and phosphorus in reservoirs, all contributing to the prevalence and severity of HABs (Humborg et al. 2000, Anderson et al. 2002). HABs have been noted at abnormally high levels in both the Copco and the Iron Gate Reservoirs, such that the EPA demanded that California include microcystin toxin (released by HABs) as a cause of impairment in the Klamath River (EPA 2008). In 2006, microcystin toxins were measured in those reservoirs at 600 times the World Health Organization’s recommended levels (EPA 2008). Higher levels of algal productivity also leads to increased decomposition, which in turn leads to lower levels of dissolved oxygen in the water (Correll 1998). In addition to causing HABs, reservoirs are also environments that harbor high levels of certain parasites affecting Upper Klamath Chinook (Bartholomew et al. 2007), and Chinook downstream from dams have been observed to have heightened infection rates from those parasites due to higher exposure doses (Bartholomew et al. 2007).

Channel morphology is altered by dams as well. Chinook salmon need a variety of different stream features to host a complicated interplay of biological and physical processes; they need the correct grades of gravel, the right depths and prevalence of deep pools, the existence of large woody debris, and the right incidence of riffles (Montgomery et al. 1997). Dams alter stream morphologies greatly, leading to a much narrower channel and a less complicated environment (Van Steeter & Pitlick 1998), which in turn leads to lower Chinook salmon populations (Montgomery et al. 1997). Meanwhile, reservoir morphology contributes to lower levels of dissolved oxygen (Cole & Hannan 1990). Low levels of dissolved oxygen have been noted on the Shasta River below the Dwinell Dam, (CRWQCB 1993). The presence of dissolved oxygen is
critical for the health of downstream fish populations. The particular effects of dissolved oxygen on Upper Klamath Chinook include serious problems with egg and embryo survival, as well as changes in behavior.

Dams have had a major impact on Upper Klamath Chinook populations. They have blocked off habitat throughout the Basin, prevented essential spawning gravel from traveling downstream, damaged water quality and changed channel morphologies of Klamath Basin streams. Dams both decrease available habitat and add to significant existing water quality problems in the Klamath.

Water withdrawals

Water withdrawals also pose a significant risk to UKTR Spring Chinook (NMFS 2009, J. Katz, pers. comm. 2010). Since 1906 and the start of the Bureau of Reclamation’s Klamath Project, a large portion of Klamath Basin surface and ground water has been withdrawn for agricultural uses. For decades this was done without considering the effects on anadromous fish in the Basin, and on Upper Klamath Chinook in particular (Foster 2002, Hecht & Kamman 1996). Agricultural water withdrawals have had a major impact on Upper Klamath Chinook populations, as resulting low flows and high temperatures cause stress and direct mortality of fish, contribute to disease prevalence and severity, and decrease Chinook egg survival.

The Project was constructed in order to reshape the dry hills of the Klamath Basin into agricultural land (Foster 2002), and wildlife have long played an inferior role in shaping land use policies in the Basin (Foster 2002). Historically, the Klamath Basin hosted a vast system of wetlands, shallow lakes, and marshes that effectively stored water during the wet season and released water in the main stem rivers during dry summer months, providing cool, clean water to fish and wildlife (Foster 2002). Today, over 80% of these wetlands have been drained in the interest of agriculture (Doremus & Tarlock 2003), eliminating key natural water storage resources in the basin. Without increased water storage and with intense competing uses, water withdrawals for agricultural use are, in their ongoing inefficient form, incompatible with the survival of Upper Klamath Chinook (Doremus & Tarlock 2003).

Water withdrawals in the Basin have increased steadily since they began and threaten fish survival in the Basin. In the Trinity River, from 1964-2004, 75-90% of the River's water was rerouted to the Central Valley for agricultural purposes (Moyle et al. 2008). Diversions into the A Canal (the primary diversion channel to the Klamath Project) increased from approximately 190,000 acre feet in 1929 to 290,000 acre feet in 1989 (Hecht & Kamman 1996), and 350,000 in 2010 (NMFS 2010). Under the pending Klamath Basin Restoration Agreement, farmers would be guaranteed levels close to the current average and significantly higher than historical rates, at 330,000 acre-feet (KBRA 2010), an amount incompatible with Chinook recovery and survival. The 2010 NMFS Biological Opinion on the Klamath Project stated that the lowered summer flows are undoubtedly connected to decreasing coho populations (NMFS 2010). Because Upper Klamath Chinook live in the same habitat as the species addressed in the Biological Opinion, the effects of withdrawals may be extended to Chinook salmon as well (NRC 2004). Since the listing of coho, stream flows in the Klamath Basin increased only briefly in 2001, before political pressure from irrigators forced the Bureau of Reclamation to resume irrigation in 2002 (Doremus & Tarlock 2003). The Ninth Circuit decision revising the NMFS ruling has supported resident coho, but has not resolved the Basin’s overall crisis (NMFS 2009).

The Shasta and the Scott rivers are currently all but uninhabitable for Upper Klamath Chinook (Chandler 2009). In the summers of 2008 and 2009, both the Scott and Shasta rivers were at their lowest levels since flow recording began, with the Scott River’s flow falling to two cfs on August 14th 2009, despite the fact that precipitation that year was at 77%. The Shasta River shared the Scott’s predicament, with its flows almost reaching six cfs on October 11, 2008, when fall Chinook normally spawn.
Water withdrawals have altered the natural hydrograph of the river and increased the seasonal variability by decreasing summer flows, which are most essential for the fall run of Upper Klamath Chinook (Hecht & Kamman 1996). The Upper Klamath Basin, with its porous volcanic rock and numerous wetlands and lakes, was historically a natural storage facility, contributing a large proportion of stream flows during drought years as well as late-summer months (Hecht & Kamman 1996), with the snowpack contributing to flows mostly during the spring and summer (Hecht & Kamman 1996). One major effect of the combination of water withdrawals and dams is that the snowmelt peak that increased flows in spring and early summer is greatly reduced (Hecht & Kamman 1996). In 2010, the NMFS Biological Opinion stated that the altered hydrograph from the Klamath Project was harming coho (NMFS 2010). Chinook fry require water flow rates above certain levels (Allen 1986), and it is likely that this seasonal reduction in water flows arrives to the detriment of Upper Klamath Chinook populations.

High temperatures caused by water withdrawals and resulting low flows are a serious threat to Upper Klamath Chinook, causing increased stress levels and mortality. The temperatures in three Klamath Basin tributaries were measured every day in August and September of 2002. Average temperatures during September 2002, before the fish kill, ranged from 23°C to 17°C (Guillen 2003). Research shows that water temperatures in the Shasta exceeded 21°C on a daily basis for the entire summer season and through September during both 2002 and 2003 (Flint et al. 2005). Maximum temperatures in the Shasta reached nearly 30°C in mid-July, far above temperatures which can lead to Chinook stress and mortality (Flint et al. 2005). Increased water temperatures due to low instream flows have affected spring Chinook in particular (NRC 2004). Spring Chinook generally need temperatures below 16°C due to disease prevalence and loss of egg viability; but the deep pools holding spring Chinook in the Salmon river have temperatures often exceeding 20°C (NRC 2004).

Low flows and warm temperatures caused by water withdrawals also inhibit migration and cause crowding which create ideal conditions for disease outbreaks (McCullough 1999, NRC 2004). This was demonstrated during the Klamath Basin fish kill of 2002. Withdrawals above Iron Gate Dam in September of this year, immediately before the fish kill, reduced flows from the dam from an estimated 1441-1470 cfs (cubic feet per second) to 759 cfs (Guillen 2003) and these low flows were implicated as a cause for the rapid spread of Ich and Columnaris.

Other diseases thrive under warmer conditions as well. Many diseases that affect the Upper Klamath Chinook population are dormant at temperatures below 15.6°C (McCullough 1999). Increased levels of Ceratonova shasta infection in Klamath and Trinity Chinook populations Chinook were noted in 2009, with especially high rates immediately below the Iron Gate Dam where high temperatures are most apparent, upstream of major tributaries (True et al. 2010). This effect is no doubt also partly due to the fact that the stagnant, warm waters of reservoirs are ideal environments for C. shasta and their polychaete hosts (True et al. 2010).

Water withdrawals which lead to lower flows and warmer stream temperatures drastically decrease Chinook egg survival (McCullough 1999). The EPA has determined that temperatures above 13°C are unsuitable for Chinook spawning (EPA 2003). Temperatures above 15.6°C result in near total mortality for Chinook eggs (McCullough 1999). Higher water temperatures also result in smaller alevins and fry, as well as higher rates of alevin abnormality (McCullough 1999). The increased temperatures in the Klamath River in September and October have narrowed the available incubation period for Chinook eggs (Hecht & Kamman 1996) and may limit the species’ overall reproductive success.

Water withdrawals are prevalent throughout the region and have caused dramatic changes to Upper Klamath Chinook habitat. This represents a persistent and ongoing threat to the long-term survival of this species in the Klamath Basin.
Logging

Historically, the Klamath Basin was heavily forested, with forest covering approximately 80% of the Upper Klamath Lake watershed alone (NRC 2004), providing stability and shade for streams. Logging in the Klamath Basin, after its beginning in the 1850s, expanded rapidly starting in the 1910s (NRC 2004); 120 million board feet of timber were logged in the upper Basin in 1920, and by 1941 timber harvesting increased to 808.6 million board feet in the upper Basin alone (NRC 2004). As of 2004, approximately 400 million board feet of timber were logged in the upper Basin annually (NRC 2004). Logging also involves the construction of road systems. In the Scott River watershed alone, more than 288 miles of logging roads were constructed as of 2004, as well as more than 191 miles of skid trails (NRC 2004). Logging is a particularly high threat for spring Chinook (J. Katz pers. comm. 2010). Logging poses a significant threat to Chinook habitat by increasing stream erosion, sedimentation and turbidity, blocking Chinook access to habitat, decreasing riparian shade, decreasing the presence of large woody debris, and leading to complications with wild fire.

Erosion and the resulting sedimentation of streams is likely the largest threat to Upper Klamath Chinook caused by deforestation. The Klamath Basin’s geomorphology is particularly vulnerable to erosion, because of the steep and unstable slopes of the region (Moyle et al. 2008), and the particularly erosive soils that underlie much of the Basin, particularly in the Scott and Trinity River watersheds (NRC 2004). In the Upper Klamath Lake watershed, more than 73% of forest land is subject to severe erosion caused by logging (NRC 2004). Logging and associated road construction has long-lasting effects on the sedimentation and turbidity of nearby streams (Klein et al. 2008). Indeed, the sediment contribution to streams by roads is often greater than that from all other land-use activities combined (NMFS 1996). The construction of roads and skidtrails in the lower Klamath Basin has been a “major source” of fine sediment in the Basin (NRC 2004). One study found that in the Scott River, average erosion for a road surface alone is 11 tons per acre; including the entire road prism, this figure rises to 149 tons per acre (Sommerstram et al. 1990). Skid trails, created during logging projects, are even more erosive, with skid-trails in the Scott averaging an annual 239 tons of soil loss per acre (Sommerstram et al. 1990). It is estimated that 10%-55% of the eroded soil makes it into the Scott River as sediment (Sommerstram et al. 1990).

Furthermore, sediment is added to streams in logged areas long after the initial logging project has been completed (Klein et al. 2008). Indeed, the timber harvest rate seems to be the biggest factor contributing to high levels of turbidity measured in a stream, with an unlogged area made up of highly erosive geology, near the Klamath Basin, showing low turbidity levels (Klein et al. 2008), while logged streams nearby, with less erosive geology, showed higher turbidity levels (Klein et al. 2008).

Increased turbidity and sedimentation create adverse conditions for Chinook. The particular effects of fine sediment on Chinook and its habitat include lowered levels of dissolved oxygen, suffocation of eggs and alevins, and lowered ecosystem productivity, which results in lower levels of food available for juveniles (Cordone & Kelley 1961).

Logging has resulted in blocked and destroyed habitat for Chinook in the Basin. Spawning habitat has been restricted in the Klamath Basin during periods of low flows by aggradations due to erosion (USBR 2001) as well as through the creation of impassable barriers such as culverts (Hoffman & Dunham 2007). Shallow landslides caused by logging and road construction scour streambeds and decrease stream complexity, destroying Upper Klamath Chinook habitat (Dietrich & Real de Asua 1998). The incidence of shallow landslides is greatly increased by the presence of logging (Dietrich & Real de Asua 1998). Habitat is also undermined as sediment leads to fewer deep pools (Quigley & Arbelbide 1997).

Logging and associated roads have also been shown to lead to decreases in riparian vegetation (Quigley & Arbelbide 1997) which leads to increased stream temperatures (Bartholow 2000). Indeed, it is likely that the
largest contribution to stream temperatures in most rivers is linked to decreased riparian vegetation (Bartholow 2000). The Shasta River, due to its structure—a relatively narrow channel—is particularly vulnerable to the lack of riparian shade (NRC 2004), and it is estimated that mature riparian vegetation would lower average maximum temperatures from 31.2°C to 24.2°C (NRC 2004).

Another effect of logging is reduced presence of large woody debris (LWD) in streams (Moyle et al. 2008). LWD is an essential element of Upper Klamath Chinook habitat (Rinella et al. 2009), as it helps form and maintain the deep pools necessary for juvenile Chinook, while aiding the recruitment of spawning gravel and creating cover for Chinook from predation (Rinella et al. 2009). LWD also contributes to stream productivity by adding habitat and food for the macrobenthic invertebrates that serve as food for juvenile Chinook (Rinella et al. 2009). Studies have shown that streams with LWD tend to harbor more salmonids, while LWD removal has been shown to lead to salmonid population decline (Rinella et al. 2009). In the Klamath Basin, logging on the Shasta River watershed has resulted in particularly low levels of LWD (NRC 2004). However, the 2010 coho Biological Opinion has found that lack of LWD is an issue in a “variety” of northern California and southern Oregon coho streams, many of which are also used by Upper Klamath Chinook (NMFS 2010).

As logging increases, so does the prevalence of wildfires (NRC 2004). The logging of old, large trees, especially when combined with fire suppression, results in more dense undergrowth, susceptible to fires (NRC 2004). Loggers often leave behind unsellable branches and detritus, which increase fire prevalence and severity (Donato et al. 2006). Since the early 1900s, the Salmon River, the last remaining viable habitat for Upper Klamath spring Chinook, has been battered by damaging crown fires, and now more than 50% of the Basin has burned (NRC 2004) with devastating effects. The extent and severity of large scale fires in the Salmon River watershed has increased over time, largely as the result of fire suppression efforts over the past century and an overall increase in heating and drying trends. In less than 15 years, from 2000 to 2014, over 43% of the Salmon River watershed has burned in mostly large fire events, with some areas burning multiple times at high severity (SRRC 2018). Short-term effects of wildfires on stream habitat include direct increases in stream temperatures, changes in stream pH, and the addition of toxic chemicals to the water (Engstrom 2010). Longer term effects include chronic and pulse erosion, channel reconfiguration, decreases in quality and quantity of large woody debris, reductions in streamside vegetation, and increases in both turbidity and stream sedimentation (Engstrom 2010).

After a fire has swept through the forest, permits are often granted for “post fire” or “salvage” logging, in an attempt to reduce future fires by taking out dead trees (Donato et al. 2006). However, there is evidence that post fire logging actually increases the risk of future fires (Donato et al. 2006), while also significantly reducing the regeneration rate of the forest (Donato et al. 2006). Studies on post fire logging after the Biscuit fire in the nearby Siskiyou National Forest (Donato et al. 2006, Thompson et al. 2007), found increased fire severity and decreased levels of regeneration in areas that have been “salvage” logged in comparison to areas left intact. Both scenarios have adverse effects on sediment levels in rivers as well as water temperatures, driving both effects upwards and consequently increasing the harm done to Upper Klamath Chinook populations.

Indirectly, logging roads also lead to habitat damage by providing access for forms of recreation that are harmful for Chinook (Quigley & Arbelbide 1997).

A significant portion of land in the Klamath River Basin remains open to logging. Land ownership in the Basin is 35 percent private, which is largely open to logging and urban and agriculture development with few protections in place for Chinook salmon or their habitat. In addition, there are over 700,000 acres, or roughly 16% of the basin, of Bureau of Land Management and the U.S. Forest Service lands that are designated as matrix lands under the Northwest Forest Plan, which are largely open to logging. See Table 3 for additional land ownership information:
Table 3.

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<th>Agency</th>
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<tr>
<td>Total Watershed Area</td>
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*Other land owners include California Department of Fish and Game, California Department of Forestry and Fire Protection, California Department of Parks and Recreation, California State Lands Commission, City of Etna, Happy Camp Community Services District, Lake Shastina Community Services District, Other State Land, The Nature Conservancy, County of Trinity, U.S. Bureau of Reclamation, U.S. National Park Service, City of Weed, City of Yreka, and Weaverville-Douglas City Recreation District.

Logging remains a serious issue for Upper Klamath Chinook. Despite the legacy of sediment-choked streams, dangerously warm waters, and fire-vulnerable forests left by 100 years of heavy logging, forest management has continued in a destructive and unsustainable direction (NRC 2004). In combination with elements like water withdrawals and mining, what once might have been a mere irritant to Upper Klamath Chinook populations is further aggravating existing and serious threats to survival.

**Mining**

Historic mining in the Klamath Basin has caused damage to Upper Klamath Chinook habitat through the rearrangement of the landscape, increased sediment and mercury pollution. These legacy affects persist to
this day in the form of greatly degraded habitat that is resistant to recovery through natural processes. More recently, suction dredge mining has continued to affect Chinook in the Basin through the entrainment of fish and their food, increased erosion and the associated complications with sediment and turbidity. Also, suction dredge mining causes the destabilization of spawning and downstream habitat.

Beginning in the 1850s, miners arrived in the Basin in great numbers and major human-caused changes to Klamath Basin geography and ecology became widespread (NRC 2004). During the midnineteenth century, gold rush miners used environmentally harmful methods of extracting gold from streams without regard for consequences (NRC 2004). One method, implemented in 1853, involved using high pressure water to blast away dirt and uncover placer deposits (NRC 2004). Many creeks were diverted into reservoirs for this purpose, and the jets of water unleashed sometimes washed away entire hillsides (NRC 2004). Much of the landscape in the Klamath Basin has been rearranged by this form of mining (NRC 2004). In California, before a court order mitigated some of the most harmful practices in 1884, hydraulic miners washed an estimated $1.6 \times 10^9$ yd$^3$ of sediment into the streams, hard rock miners created $3 \times 10^7$ yd$^3$ of mine tailings, and dredge miners left behind $4 \times 10^9$ yd$^3$ of debris, largely in the Klamath Basin (NRC 2004). Using the Salmon River sub-basin as an example, the Salmon River Floodplain Habitat Enhancement and Mine Tailing Remediation Project, Phase 1: Technical Analysis of Opportunities and Constraints, summarizes the legacy mining effects as follows (Stillwater, 2018):

One of the most important factors leading to the decline and continued low abundance of anadromous salmonids in the Salmon River, and in particular spring-run Chinook, is the legacy effect of historical placer mining on channel and floodplain habitat conditions throughout the mainstem and larger tributary reaches (Stumpf 1979, SRRC 2017). Hydraulic and dredge placer mining in the Salmon River between about 1870 and 1950 led to profound and lasting changes, eroding over 1,859 acres adjacent to the mainstem and larger tributary channels and delivering an estimated 20.3 million cubic yards of sediment to the river (Hawthorne 2017, de la Fuente and Haessig 1993). Placer mining denuded floodplains and adjacent river terraces and hillslopes, reduced riparian shade cover, and exposed the stream channel and surrounding areas to increased solar radiation.

Delivery of hydraulic mine debris resulted in as much as 5 meters of channel aggradation, on average, throughout the predominantly alluvial reaches within the Project area. Aggradation by hydraulically mined sediment widened and shallowed alluvial reaches, filled pools, reduced the complexity and connectivity of floodplain habitats, and led to coarsening and armoring of the channel bed. Coarse sediment stored in the bankfull channel, denuded floodplains, and mine tailings on terraces along the river corridor continues to prevent riparian vegetation establishment, and due to the increased exposure to solar radiation and thermal mass, creates a significant heating effect. These impacts significantly reduce the amount and quality of spawning, oversummering, and over-wintering habitat and decrease the cumulative channel length that remains thermally suitable for salmonids during the summer, thereby constraining population productivity and increasing extinction risk. These legacy impacts to the channel and floodplain inhibit natural recovery and require intervention to recover within human and salmon population time scales.

Historically, gold mining involved the use of mercury, large quantities of which was released back into the Klamath River (NRC 2004). It is estimated that with hydraulic mining, approximately one pound of mercury was released for every three to four ounces of gold recovered (NRC 2004). Much of that mercury remains in Klamath Basin soils and sediments, affecting Upper Klamath Chinook through leaching, as well as any animal or human that consumes them (NRC 2004). Even in the 19th century, the California government acknowledged the effects of mining on Klamath Basin salmon, and in 1852, it enacted its first salmon statute, though this piece of legislation had little practical effect (NRC 2004).
Much of the mining activity in the 19th century still affects whole streams in the Klamath Basin, and some areas, such as the Scott River, have been permanently damaged (Moyle et al. 2008). Even the Salmon River, now the last bastion for UKTR Spring Chinook, has approximately 20 million cubic yards of sediment, unleashed by mining between 1870 and 1950, slowly making its way downstream (Hawthorne 2017, de la Fuente and Haessig 1993). This sediment harms juvenile habitat, fills in the deep pools needed for adult Chinook, and degrades spawning habitat by eliminating the correct grade of gravel (Moyle et al. 2008). According to the findings of a recent and extensive assessment of mining effects on floodplains and anadromous fish habitat in the Salmon River, “Channel and floodplain aggradation resulting from historical hydraulic mining widened and shallowed alluvial reaches, filled pools, reduced the complexity and connectivity of floodplain habitats, and led to coarsening and armoring of the channel bed. Coarse sediment stored in the river channel, denuded floodplains, and mine tailings along the river corridor continue to create a significant heating effect. These legacy impacts to the channel and floodplain inhibit natural recovery and require intervention to recover within human and salmon population time scales” (Stillwater 2018). Old gold mining practices have also left their mark on the Trinity River, an area of particular concern for mercury contamination (Alpers et al. 2005).

More recently, suction dredge mining has been used for extracting gold from the Basin. Dredge mining has been operating in California continuously since the invention of the suction dredge in the 1960s (CDFG 2009), and Upper Klamath Chinook populations have been directly impacted by this activity. Effects of suction dredge mining include the entrainment of juvenile fish and eggs (Harvey & Lisle 1998), as well as the entrainment of macrobenthic invertebrates that serve as food for juvenile Chinook (Moyle et al. 2008). Apart from entrainment of macrobenthic invertebrates that serve as an important food source for juveniles, the exposure of new substrate and the deposition of sediment in the streams causes localized reductions in both macrobenthic invertebrate presence and diversity (Harvey & Lisle 1998).

Dredging has long-term erosive consequences, increasing the sediment load of streams and altering habitat by filling deep pools and eroding stream banks that formerly served as shelter for the Chinook. Effects can last for years after the dredgers have left (Harvey & Lisle 1998). Similarly, dredging of riffle crests can cause them to erode, potentially destabilizing spawning habitats, filling deep holes, and destabilizing downstream reaches (Harvey & Lisle 1998). Furthermore, dredge mining that has disturbed riffle crest tends to channel the streamwater towards a stream bank, increasing streambank erosion (Harvey & Lisle 1998).

Suction dredge mining also stirs up sediment, adding to a stream’s turbidity (Harvey & Lisle 1998). Increased turbidity resulting from dredge mining can have negative effects on Upper Klamath Chinook, particularly juveniles. Increased levels of suspended solids in the water seem to result in increased foraging time by juvenile Chinook, as it reduces their reactive distance and prey capture success rate (Harvey & Lisle 1998). Higher levels of suspended sediment can also reduce primary production in a stream, as the sediment blocks off light needed for photosynthesis (Henley et al. 2000). This limits food available for organisms at higher trophic levels (Henley et al. 2000), including juvenile Chinook.

Suction dredge mining can also increase deposition of fine sediment downstream (Harvey & Lisle 1998), reducing both the benthic invertebrate populations that serve as food for Chinook (Harvey & Lisle 1998), and the availability of habitat for alevins inhabiting the benthic zone (Harvey & Lisle 1998). Increased fine sediment deposition also reduces dissolved oxygen levels by filling interstices between gravel and reducing water circulation in the hyporheic zone (Henley et al. 2000). The hyporheic zone is the zone of gravel and sediment that composes the streambed, where groundwater and surface water interact (Findlay 1995), and where Upper Klamath Chinook deposit their eggs. Increased fine sediment deposition due to mining is of particular concern in the Trinity and Salmon rivers (NRC 2004).
Suction dredge mining leads to the destruction of Chinook redds (Harvey & Lisle 1999). Miners dredge up and then deposit gravel that is seemingly the perfect size and density for Chinook redds, attracting spawning Chinook. The tailings placed back into the stream are unsupported however, and during the high flow period in winter after the Chinook have used the sediment for spawning, the gravel is swept downstream, killing any eggs present (Harvey & Lisle 1999). The same instability kills Chinook alevins inhabiting the gravel substrate (Harvey & Lisle 1998).

Mine tailings from suction dredge mining also reduce deep pools (Harvey & Lisle 1999) that are essential habitat for both juvenile and adult Chinook. The presence of unstable mine tailings used by Chinook as spawning grounds has been noted throughout the Klamath, Salmon, and Scott rivers and their tributaries (Moyle et al. 2008).

Other general effects include the loss of channel complexity, the loss of pool habitat, and the loss of effective large woody debris (NMFS 1998). Finally, the constant noise and turbidity caused by suction dredge mining raises the stress of Upper Klamath Chinook, increasing the possibility of premature death (Moyle et al. 2008).

Suction dredge mining currently poses a threat to Upper Klamath Chinook. Recently, California recognized the threat posed to salmonids by suction dredge mining and temporarily banned it in California streams, pending environmental review. The long-term damage has already occurred to Upper Klamath Chinook habitat, and with the very limited budget California can put towards enforcing the ban, many suction dredge miners are able to continue their activities with impunity. Mining has historically caused major damage to Chinook habitat in the Klamath Basin and remains a threat to their continued existence.

**Chemicals**

Land use in the Klamath Basin has resulted in the contamination of the region’s waters by a variety of chemicals including pesticides, herbicides, and insecticides. Basin agricultural lands discharge chemical and fertilizer-contaminated wastewater, and municipal wastewater also enters the system through the Lost River. Combined, these wastewater discharges result in harmful algal blooms, higher aquatic pH levels, lower levels of dissolved oxygen, and high concentrations of ammonia (NCWQCB 2010), all of which are destructive for Chinook populations (Moyle et al. 2008).

Pesticides, insecticides, and herbicides have been used in the Klamath Basin for at least 60 years (Dileanis et al. 1996). This includes the heavy use of dangerous organochlorine pesticides such as DDT in the 1950s and 1960s, which are found in Tule Lake and elsewhere in the Basin (Dileanis et al. 1996). In the early 1990s, 16 pesticides were reported in the waters of Tule Lake Refuge, with higher concentrations measured near agricultural drains (Dileanis et al. 1996). Between 1997 and 2001, approximately 27,000 pounds of the active ingredients of four forestry herbicides were used in the Klamath Basin. In 2002, research determined that some of the forestry herbicides were drifting into waterways (Wofford et al. 2003). So far in 2010, pesticide use proposals for 81 pesticides (including those known to be dangerous to wildlife) have been granted for lease lands within the Tule Lake and Lower Klamath National Wildlife Refuges (USBR 2010).

In long term studies, USGS (2009) found high levels of a variety of pollutants especially in the 20 miles between Link River and Keno Dam. Given the high levels of toxicity, the State of Oregon classifies this 20 mile reach as “water quality limited,” as required by Section 303(d) under the Clean Water Act (USGS 2009). Water quality in this region affects the quality of the entire main stem of the Klamath River. (Sullivan et al. 2010).

In 2008 the EPA issued a Biological Opinion on “the effects of the U.S. Environmental Protection Agency’s (EPA) proposed registration of pesticide products containing the active ingredients chlorpyrifos, diazinon,
and malathion on endangered species, threatened species, and critical habitat that has been designated for those species" (NMFS 2008). The Opinion assesses the effects of these pesticides on 28 listed Pacific salmonids and determines that the continued use of these chemicals is likely to jeopardize the continued existence of 27 listed Pacific salmonids and to destroy or adversely modify critical habitat for 25 of 26 listed Pacific salmonids, with critical habitat, including the Klamath Basin’s Southern Oregon/Northern California Coast Coho (NMFS 2008). The population-level consequences of pesticide use discussed in this report included impaired swimming and olfactory-mediated behaviors, starvation during a critical life stage transition, death of returning adults, additive toxicity, and synergistic toxicity. Upper Klamath Chinook also negatively affected by these pesticides.

Diazinon, an organophosphate insecticide commonly used for general pest control, has been found to affect the olfactory nervous system of Chinook (Scholz et al. 2000). As Chinook depend largely on their olfactory system for homing, reproductive behavior, and pheromone activated anti-predator behavior, disruption of the sense of smell has wide-ranging negative effects on Chinook populations (Scholz et al. 2000). This disruption likely increases occurrence of Chinook “straying” (spawning fish returning to nontraditional spawning grounds), with results ranging from hybridization between hatchery and wild fish (Scholz et al. 2000) to lower densities of spawning Chinook in streams, leading to reproductive failure. Diazinon also negatively affects anti-predator behavior and the reproductive behavior of male Chinook (Scholz et al. 2000).

Other chemicals such as carbaryl, the third most commonly used insecticide in the United States, have been shown to neurologically affect salmonids (Labenia et al. 2007). Furthermore, pesticides seem to act synergistically, such that sub-lethal doses of two different pesticides may have effects greater than when they are encountered individually (Laetz et al. 2009). In one study, every pesticide tested acted synergistically with every other pesticide, and malathion and chlorpyrifos proved to be a particularly harmful combination (Laetz et al. 2009); both of those pesticides have been approved for use on Klamath Basin National Wildlife Refuge lease lands (USBR 2010), and are likely used to a much greater extent throughout the Klamath Irrigation Project.

Fertilizer and organic nutrients from agriculture and municipal wastewater present a serious threat (USGS 2009) by fueling algal blooms, depleting dissolved oxygen levels, and elevating pH levels (Smith et al. 1999). Algal blooms and subsequent fish die-offs are also linked to the presence of ammonia in the water (Rykbost & Charlton 2001). In the United States, eutrophication caused by agricultural runoff is the nation’s largest water pollution problem (Smith et al. 1999) and the Klamath Basin is no exception. The Klamath Straits Drain, a concrete canal which collects the upper Basin’s agricultural, refuge, and municipal wastewater and discharges it into the main stem of the Klamath River, has been designated “water quality limited” on Oregon’s 303(d) list for dissolved oxygen and ammonia levels year round and for the water’s pH and chlorophyll concentrations during the summer (USGS 2009). Discharge from the Klamath Straits Drain is impacted by high concentrations of total phosphates, biochemical oxygen demand, total solids, and ammonia and nitrate nitrogen throughout the year (ODEQ 1995).

Lowered dissolved oxygen (DO) levels due to impaired water quality as a result of agricultural and/or municipal inputs inflict harm on Upper Klamath Chinook (NCWQCB 2010). During July of 2008, the levels of DO measured above the Keno Dam were far below levels recommended for salmonids; if DO levels average lower than 3-3.3 mg/L, 50% mortality of juvenile salmonids is likely, while in water above 20˚C, daily minimum DO levels of 2.6mg/L are required to avoid 50% mortality (NCWQCB 2010). However, in 2008 from mid-July to mid-September at the Keno Dam, DO levels repeatedly dropped below one mg/L (sometimes to as low as .38 mg/L), and rarely rose to three mg/L (USGS 2009, Appendix B).

Nutrient loading of stream systems can lead to higher pH in river systems (NCWQCB 2010). The effects of a high pH on Upper Klamath Chinook are exacerbated by high temperatures (NCWQCB 2010), which is
already a major water quality problem in the Klamath Basin. Due to impaired water quality as a result of agricultural, municipal, and other inputs as discussed, the Klamath River’s pH in the summer often rises above 8.5, and sometimes reaches 9. At the Miller Island Boat Camp in 2008, the river’s pH in early July, measured daily, had several consecutive days with pH values ranging from 9.06-9.53 (USGS 2009, Appendix B). Few direct studies examine the effects of high pH values on Chinook but rainbow trout are stressed by pH values above 9 and generally die if the pH value rises above 9.4 (NCWQCB 2010).

Nutrient loading in the Klamath River can increase ammonia levels as higher concentrations of nitrogen enter the water (NCWQCB 2010). High nitrogen concentrations, a product of water runoff from fertilized agricultural fields, also increases the toxicity of the ammonia present, as higher pH levels result in most of the ammonia morphing into its deadlier, un-ionized form (NCWQCB 2010). Ammonia in the Klamath River has been noted at levels high enough to harm Chinook through a reduction in hatching success; reductions in growth rate and morphological development; and pathologic changes in tissues of gills, livers, and kidneys (NCWQCB 2010). Ammonia also reduces Chinook disease resistance and has been termed an exacerbating factor in Klamath River fish kills (NCWQCB 2010). The presence of high levels of un-ionized ammonia was noted in the Upper Klamath Lake in both 2007 and 2008 (USGS 2010).

In the Upper Klamath Lake, the combination of high pH (sometimes between 9 and 9.5 in late August) and temperatures (around 20°C at the same time; USGS 2010) with high levels of ammonia can be dangerous. On August 25th, 2008, ammonia was measured at 0.933 mgN/L (USGS 2010), far above “acute” levels of ammonia for salmonids (0.885 mgN/L when the pH is 9; NCWQCB 2010). The USGS found that ammonia concentrations in the Klamath River actually increased in the downstream direction, with significantly higher levels found at the Keno Dam when compared to the Link River Dam (USGS 2009).

Agricultural and municipal wastewater delivered into the Klamath River is a severe threat to Chinook. Pesticides, even at sub-lethal doses, can combine to alter Chinook behavior, with major consequences for Chinook survival and reproduction. The eutrophication of traditional Upper Klamath Chinook habitat in the Klamath Basin results not only in levels of dissolved oxygen low enough to cause serious harm to Chinook populations, but also causes elevated pH levels, high concentrations of ammonia, and the presence of toxins produced by algal blooms.

Grazing

Grazing threatens UKTR Spring Chinook in the Basin because of the loss of riparian vegetation, loss of large woody debris, increased sediment in streams, the addition of excessive nutrients to streams, and lowered water tables.

Grazing in the Klamath Basin has occurred since the late 1800s. As early as 1880, overgrazed fields caused a disastrous winter for plant life resulting in the mass mortality of cattle across the Basin (NRC 2004). More widespread effects were quickly noted, as a geologist in the early 1900s found formerly flat streams cutting channels in the land, as run-off increased due to overgrazing (NRC 2004). In an effort to save the nascent Klamath cattle industry, government agents recommended that wetlands be drained and planted with hay to provide feed for cattle, and in the 1890s, ranchers obliged, draining wetlands along the borders of the Upper Klamath Lake to provide increased forage (NRC 2004). In addition to lost water storage capacity and lower water quality caused by wetland draining, the flood irrigation of pastures to create cattle feed as well as the switch to nonnative species of hay severed healthy riparian connections to the landscape (NRC 2004). Because cattle are attracted to riparian areas for grazing, damage caused by intense cattle presence is often concentrated in sensitive riparian areas (Belsky et al. 1999). The Scott and Trinity rivers have been degraded by under-regulated grazing and ranching, as have numerous small tributaries that contribute their flows to the Klamath River (NRC 2004). In the South Fork Trinity River, unsustainable grazing and farming practices, combined with large floods in 1964, have resulted in long-term
loss of viability to salmon populations (NRC 2004). Populations in the South Fork Trinity River have made little progress recovering in the intervening decades (NRC 2004).

One major effect of grazing in riparian habitats is the decrease in riparian vegetation. Throughout the Klamath Basin, there is evidence that unfenced grazing results in the loss of vegetation through animal consumption and trampling (NRC 2004). Grazing is the primary contributor to the lack of riparian vegetation in the upper Shasta River (NRC 2004). Loss of riparian vegetation leads to increased stream temperatures as well as a decrease in the quality of Chinook habitat through the loss of large woody debris (NRC 2004) increased erosion and sedimentation, all of which have highly damaging consequences to Chinook salmon.

Cattle also cause increased levels of nutrients to be added to river systems. The effects of season-long grazing in the past in the Sprague River (a major tributary to the Upper Klamath Lake) have resulted in the Oregon Department of Environmental Quality labeling the Sprague River in the Upper Klamath Basin as one of the worst streams in Oregon for non-point-source pollution (NRC 2004). Animal waste from grazing adds nutrients to water systems that can result in HABs (Belsky et al. 1999). The Sprague River is a contributor of extremely high levels of phosphorus due to poor land use practices (NRC 2004), including grazing. As phosphorus is the primary factor limiting algal blooms in freshwater systems (Anderson et al. 2002), its input is likely to be a major cause of HABs, which can have large effects on downstream Chinook populations, through the release of toxins (EPA 2008) and lowered levels of dissolved oxygen (Correll 1998).

Grazing has also been implicated in lowering water tables; as water flows downhill during floods, it is trapped by riparian plants, slowing flows and allowing the water to percolate through the sub-soil to become groundwater (Belsky et al. 1999). Extensive grazing, combined with groundwater withdrawals and sprinkler irrigation is a significant contributor to the problem of low water tables in the Scott River watershed (NRC 2004, Van Kirk & Naman 2008). The impact of low water tables in these critical Klamath River tributaries and throughout the upper Basin translates directly to limited river flows and impaired water quality for Upper Klamath Chinook downstream.

The legacy effects of grazing have permanently harmed Upper Klamath Chinook habitat and current ranching practices continue to impair the viability of populations through impacts on water quality. For every cattle herd grazing on upper Basin rangeland, water quality for downstream Upper Klamath Chinook populations is further degraded.

(2) overexploitation;

Commercial, recreational and tribal fishing have had a combined effect on Klamath River salmonids that have contributed to their decline since the 19th century (NMFS 2009; Snyder 1931). Both legal and illegal harvest combined pose a high threat for both spring and fall Upper Klamath Trinity River Chinook (J. Katz pers. comm. 2010). Harvest of Upper Klamath Chinook salmon has added to the decline of both the spring and fall runs and continues to threaten the long-term persistence of Chinook in the Basin (Moyle et al. 2008). Moyle et al. (2008) identifies legal and illegal harvest as a major limiting factor affecting both spring and fall runs of Upper Klamath Chinook. Both illegal harvest of holding adults and legal, ocean and river harvests contribute to reduced spawning populations. Adults holding upstream in deep pools are especially vulnerable to illegal take; although these numbers are largely undocumented, it can be assumed that UKTR Spring Chinook holding in pools in the Klamath River and elsewhere in the Basin are affected by harvest from pools where they are holding prior to spawning. There is a general absence of UKTR Spring Chinook from populated areas in the Klamath, and in areas with easy access to humans, further suggesting that
illegal harvest is occurring. The illegal removal of even a small number of UKTR Spring Chinook likely has an intense effect on spawning populations (Moyle et al. 2008).

Because managing agencies do not treat UKTR Spring Chinook differently from UKTR Fall Chinook, UKTR Spring Chinook are taken legally in commercial and sport fisheries (Moyle et al. 2008). Harvest rates are defined based on combined spring- and fall-run numbers of both hatchery and natural origins; therefore, the dwindling populations of spring-run Chinook, especially wild-spawning populations are particularly vulnerable to being overfished under current management (Bilby et al. 2005). In fact, current management actions neglect to protect spring-run Chinook even when protections have been put in place to restrict fall-run Chinook harvest, essentially increasing pressure on the much smaller and more imperiled populations of spring-run Chinook. For example, after the final stock projections developed by the Pacific Fishery Management Council for Klamath River fall-run Chinook (which included spring-run return numbers) were projected to be the lowest on record, “the Fish and Game Commission adopted regulations on April 13, 2017 for a full closure of the 2017 Klamath River Fall-Run Chinook Salmon fishery in the Klamath and Trinity rivers” (CDFW 2017). The regulations went into effect August 8, 2017, after the spring-run Chinook had already entered the Klamath Basin and its tributaries. Even though low spring-run Chinook return numbers were counted as part of these projections, they were not granted equal protections to fall-run Chinook, and the daily bag limits on the Klamath River remained the same for the period of time that they were present in the river before fall-run Chinook entered the basin. During this time period, the only allowable salmon sport fishing on the Klamath River was spring-run Chinook, effectively increasing the pressure on dwindling spring-run Chinook during this year with the lowest projected returns.

(5) disease; or

Several diseases affect the Upper Klamath Trinity River Chinook salmon and will likely continue to pose a threat to this ESU in the future. Salmon are exposed to a variety of bacterial, viral and parasitic organisms throughout their life cycle, contracting diseases through both waterborne pathogens and through mingling with infected hatchery fish (NMFS 1998). It is possible for a fish to be infected with one or more pathogen but not to show signs of disease. Hatchery Chinook salmon appear to be more susceptible to disease than naturally spawning Chinook (NMFS 1998). Because Chinook salmon in the Klamath River Basin emigrate as juveniles and return to spawn when water temperatures and flows approach their limits of tolerance, they are particularly susceptible to disease (Moyle et al. 2008, NMFS 2009).

In 2002, a major fish kill occurred in the second half of September in the lowermost 40 miles of the Klamath River main stem. At least 33,000 Chinook died out of a total estimated run of 130,000 fish (NRC 2004). Although the original FWS report of estimated mortality claimed about 33,000 fall Chinook died in this fish kill, a more updated report by CDFG explains that the estimate was “conservative and DFG analyses indicate actual losses may have been more than double that number” (CDFG 2004). This was the largest known pre-spawning die-off recorded for the region and possibly the whole Pacific coast (Guillen 2003). Stressful environmental conditions in 2002 allowed columnaris and ich to sweep through a population of already stressed fish (Guillen 2003). Factors which combined included high temperatures, crowded conditions and low flows. In response to high water temperatures and low flows, fish stopped migrating and instead concentrated in cooler deep pools, creating optimal conditions for the proliferation of pathogens. All of the specimens examined during the fish kill were infected by ich and/or columnaris (Guillen 2003).

Columnaris is a bacterial infection affecting Upper Klamath Chinook salmon and is caused by Flavobacterium columnare. The disease is associated with pre-spawn mortality of spring-run Chinook especially when they are exposed to above-optimal water temperatures (Moyle et al. 2008). Columnaris is usually pathogenic at temperatures above 15° C and outbreaks are common in adult populations held at hatcheries in water at 15-18° C (Guillen 2003). The earliest sign of columnaris is a thickening of the mucus at various spots on the fish (Guillen 2003). When it becomes more developed, fish will show small bloody
spots on the skin. Eventually, respiratory and osmoregulatory function is lost at the gill surface and the fish dies (Post 1987). Although typically widespread, columnaris only causes widespread mortality when associated with high degrees of stress. This occurred during the 2002 fish kill in which columnaris was one of the two diseases implicated as a direct cause of mortality. By 2004, only 2.4% of fish examined were infected with *F. columnare* suggesting that it was not a significant problem in these fish in 2004 (Nichols and Foott 2005).

The other pathogen which directly caused the major fish kill in 2002 is ich disease, caused by the ciliated protozoan, *Ichthyophthirus multifilis*. The optimal temperature for ich development is 21.1-23.9º C and within this range, higher temperatures cause faster replication of the parasite (Guillen 2003). Ich disease reduces the capacity for fish to absorb oxygen and excrete ammonia and mortality occurs when gills become too damaged to function (Post 1987). Studies show that higher water velocities reduce and may prevent ich disease outbreaks completely because of a decreased probability of the parasite finding a host before being swept downstream (Guillen 2003).

The USFWS and CDFG monitored the health and physiology of salmonids in the Klamath and Trinity River Basins from 1991-1994 and identified *Ceratonova shasta* as the most significant disease affecting juvenile salmon in the Klamath Basin (Nichols and Foott 2005). *C. Shasta* is a myxozoan parasite that appears in the mainstem and Upper Klamath River, Copco Reservoir, both Klamath and Agency Lakes and the lower reaches of the Williamson and Sprague Rivers (Moyle et al. 2008). It is often found in reservoir environments so that dams on the Klamath River have contributed to the spread of this parasite. Soon after Iron Gate Hatchery was established, operational problems associated with *C. shasta* began to occur and significant outbreaks continued to occur into the early 1980s (NMFS 1998). A 1989 study found that Chinook salmon at Iron Gate Hatchery had a 4% susceptibility to *C. shasta* and a 19% susceptibility at the Trinity River Hatchery (Carlton 1989 as cited in NMFS 1998). *C. shasta* infection appears to be accelerated when high densities of infected fish are combined with warm water temperatures (Foott et al. 2003).

Nichols and Foott monitored the health of juvenile Klamath River Chinook Salmon. They estimated that 45% of the population was infected with *C. shasta* (Nichols and Foott 2005). Of the fish infected with *C. shasta*, 98% were also infected with another myxozoan infection, *Parvicapsula minibicornis*. The dual infection suggested that the majority of fish infected with *C. shasta* as juveniles would not survive.

More recent studies have revealed some of the factors affecting incidence of *C. shasta* infections and identified this parasite as a potentially limiting factor to the survival of Klamath River Chinook. Petros et al. (2007) studied the effect of water flows on the incidence of *C. shasta* to find out whether drought exacerbated fish health issues by concentrating spores in reduced flows and compromising resistance through increased stress from warm water temperatures. The years 2005 and 2006 had higher flows than 2004 and exposure to *C. shasta* was less severe in the years with higher flows. However, the 2006 results were not as pronounced as expected given the magnitude of the spring 2006 water levels (Petros et al. 2007).

Bjork and Bartholomew (2009) investigated the effects of water velocity on presence of *C. shasta* in *Manayunkia speciosa*, the pathogen’s intermediate polychaete host. In faster water velocities, the polychaete density was higher but the prevalence of *C. shasta* was lower and the severity of infection in fish was also decreased. Another study by Bjork (2010) showed that temperature had no effect on polychaete survival but that higher temperatures caused actinospore release in *C. Shasta* to occur earlier and in greater abundance. *C. shasta* infections can be expected to grow more severe in conditions of low flows and high temperatures.

*Parvicapsula minibicornis* the other myxozoan parasite common to the Klamath River and although often present, like *C. Shasta* it is not always abundant nor do the conditions always exist for large numbers of
Chinook salmon to be infected (Moyle et al. 2008). *P. minibicornis* appears to be highly infectious. It was estimated to infect 94% of the population of juvenile Chinook in the Klamath River in 2004 (Nichols and Foott 2005).

Another prevalent pathogen in the Klamath River Basin is Bacterial Kidney Disease (BKD) caused by the Bacterium, *Renibacterium salmoninarum*. In 1994, BKD was cited along with the trematode parasite, *Nanophyetus salmicola*, as one of the most significant pathogens affecting both natural and hatchery smolt health in the Basin (NMFS 1998). The pathogen can prevent fish from making the necessary changes in kidney function during smoltification (NMFS 1998). Also, the stress of migration can cause BKD to come out of remission (Schreck 1987).

Climate change is expected to cause increased water temperatures and therefore higher stress conditions that can be expected to increase the occurrence and severity of disease outbreaks among Chinook salmon in the Klamath Basin. Warmer temperatures favor disease outbreaks (Moyle et al. 2008). Disease has been a direct cause of mass mortalities in the Klamath Basin in the past and will present further challenges for their continued survival due to changing conditions in the future.

(6) other natural events or human-related activities.

As noted above, a century of dams and diversions has been a leading cause of UKTR Spring Chinook declines.

7. DEGREE AND IMMEDIACY OF THREAT

Please see #1, population trend

8. IMPACT OF EXISTING MANAGEMENT EFFORTS

As abundantly documented in this petition, Upper Klamath Chinook face severe threats from multiple factors. Existing regulatory mechanisms are entirely inadequate to address these threats and ensure the survival of the species. By considering Upper Klamath spring- and fall Chinook as part of the same ESU, NMFS has limited adequate protection of spring Chinook under the ESA so that they are directly at risk of extinction. Current federal and state regulations which may indirectly affect these fish lack the protection needed by Upper Klamath Chinook.

*Federal Regulatory Mechanisms: U.S. Forest Service*

In the United States, the National Environmental Policy Act (NEPA) requires Federal agencies, including agencies within the Department of Interior, Department of Agriculture (e.g. United States Forest Service), and beyond, to consider the effects of management actions on the environment. NEPA does not, however, prohibit Federal agencies from choosing alternatives that may negatively affect Upper Klamath Chinook salmon.

Upper Klamath Chinook are listed as a sensitive species by the Forest Service in Region 5, requiring analysis of impacts to the salmon from management actions or changes under NEPA. Because NEPA does not require avoidance of harm, this affords little protection. The Forest Service must analyze the impacts of their actions on the species, but as above are not required to select alternatives that avoid harm to Chinook. Indeed, the Forest Service regularly plans timber sales, maintains and utilizes roads, allows livestock grazing and conducts other actions that harm Upper Klamath Chinook.
Relevant National Forest Plans include Six Rivers National Forest, Shasta-Trinity National Forest and Klamath National Forest. The forests are responsible for maintaining suitable fish habitat that will support well-distributed, viable populations of native fish. Forest service sensitive species including the Upper Klamath Chinook are considered in planning decisions such as habitat improvement and restoration. Sensitive species are considered when establishing key watersheds within National Forest Plans. Standards and guidelines for key watersheds include analysis prior to management activities, prioritization of sensitive species during restoration activities and restrictions on the building of new roads. National Forest Plans do not have the authority to maintain fish habitat on private lands nor to regulate actions by private parties which are destructive to Upper Klamath Chinook (mining, agriculture and timber operations) and the plans are therefore insufficient to protect Chinook salmon in the Basin.

The NWFP, signed and implemented in April 1994, represents a coordinated ecosystem management strategy for Federal lands administered by the USFS and BLM within the range of the Northern spotted owl (which overlaps considerably with the freshwater range of Chinook salmon).

The most significant element of the NWFP for anadromous fish is its Aquatic Conservation Strategy (ACS). This regional scale conservation strategy includes: (1) Special land allocations, such as key watersheds, riparian reserves, and late-successional reserves, to provide aquatic habitat refugia; (2) special requirements for project planning and design in the form of standards and guidelines; and (3) new watershed analysis, watershed restoration, and monitoring processes. These components are designed to ensure that Federal land management actions achieve a set of nine Aquatic Conservation Strategy objectives, which include salmon habitat conservation. In recognition of over 300 “at-risk” Pacific salmonid stocks within the NWFP area (Nehlsen et al., 1991), the ACS was developed by aquatic scientists, with NMFS participation, to restore and maintain the ecological health of watersheds and aquatic ecosystems on public lands. The ACS attempts to maintain and restore ecosystem health at watershed and landscape scales to protect habitat for fish and other riparian-dependent species and resources and to restore currently degraded habitats. The approach seeks to prevent further degradation and to restore habitat on Federal lands over broad landscapes.

The overall effectiveness of the NWFP in conserving Upper Klamath Chinook salmon is limited by the extent of Federal lands and the fact that Federal land ownership is not uniformly distributed in the ESU. In some areas, particularly Bureau of Land Management (BLM) ownership, Federal lands are distributed in a checkerboard fashion, resulting in fragmented landscapes. This factor places constraints on the ability of the NWFP to achieve its aquatic habitat restoration objectives at watershed and river basin scales.

In addition, a significant portion of land in the Klamath River Basin remains open to logging under the NWFP. Land ownership in the Basin is 35 percent private, which is largely open to logging and urban and agriculture development with few protections in place for Chinook salmon or their habitat. In addition, there are over 700,000 acres, or roughly 16% of the basin, of Bureau of Land Management and the U.S. Forest Service lands that are designated as matrix lands under the Northwest Forest Plan, which are largely open to logging.

Under the National Forest Management Act, the Forest Service is required to “maintain viable populations of existing native and desired nonnative vertebrate species in the planning area” (36 C.F.R. §219.19). As with NEPA, this requirement does not prohibit the Forest Service from carrying out actions that harm species or their habitat, stating only that “where appropriate, measures to mitigate adverse affects shall be prescribed” (36 C.F.R. §219.19(a)(1)). This clause does little to limit long term impacts to salmonid habitat in the Klamath Basin. Also, these regulations are currently under review and any protection they afford may be removed at any time.
Despite all of these laws and plans, federal land managers have continued to plan and implement projects that harm Upper Klamath-Trinity River Chinook salmon. Destructive actions have included timber sales on steep slopes, logging of riparian reserves, failure to maintain, fix and remove roads as necessary, and problems with grazing, including inadequate and unenforced best management practices (BMPs). Also, the U.S. Forest service has failed to advocate for stream flows in the lower Scott River which is under their jurisdiction. Federal land managers in the Basin are not taking sufficient actions to manage for the persistence of Chinook salmon and better practices are necessary for conservation of these fish.

Federal Regulatory Mechanisms: FERC

The Federal Energy Regulatory Commission (FERC) is charged with relicensing the Klamath Hydroelectric Project (FERC P-2082-000) on the Klamath River every 20 to 50 years. The FERC license for operation of the Klamath Project expired in 2006 and FERC produced an Environmental Impact Statement (EIS) for the Project in 2007. In a new national era of dam removal, FERC has supported negotiations regarding removal of antiquated hydroelectric projects like on the Klamath River in place of intensive and costly dam improvements to comply with modern environmental laws. PacifiCorp and the Klamath River Renewal Corporation (KRRC) recently filed applications with FERC to transfer the dams to KRRC for license surrender and dam removal. FERC’s decision on the application is pending.

When considering whether or not to list a species, NMFS is not to consider promised, pending or future management actions, but instead only the current management and status of the species. In numerous ESA listing cases, the USFWS has been forced by judicial action to reverse decisions not to list species because they relied on promised management actions; this includes decisions over the Barton Spring’s salamander, Queen Charlotte goshawk, jaguar, Alexander Archipelago wolf, and coho salmon. It is imperative that NMFS consider only the current management and species status. States, federal agencies, and private interests can easily promise to protect and recover species in order to avoid or delay a potentially controversial listing; unfortunately, there are not means to ensure management agencies will follow through on promises, or that their actions will result in recovery. To protect species from ongoing destruction, modification or curtailment of habitat or range, listing under the ESA is required while management actions are being tested. If promised management actions result in substantial recovery, then such actions should be incorporated into a recovery plan for the species.

In response to the noted court decisions on various species’ listings, USFWS developed a policy for evaluating the contribution of conservation efforts while considering the potential need for listing. This policy identifies criteria for determining the certainty a conservation effort and whether it is likely to be effective. (68 Fed. Reg. No. 60, 28 Mar. 2003). We have considered this policy when evaluating pending agreements in the Klamath Basin, and understand that NMFS should do the same when considering listing of the Upper Klamath Trinity River spring Chinook salmon. Clearly, the UKTR Spring Chinook is experiencing ongoing threats, placing it in danger of extinction and thus requiring protection as an endangered species, regardless of pending, untested, or promised management actions.

The most recent genetic work on spring-run Chinook in the Klamath Basin suggest that even with dam removal, the lack of the spring-run timing allele in Upper Klamath Chinook source populations within a reasonable distance below the current dams will hinder restoration and natural spring-run Chinook recovery after dam removal. “These results highlight the need to conserve and restore critical adaptive genetic variation before the potential for recovery is lost.” (Thompson, et al. 2018)

State Regulatory Mechanisms: TMDL
State mechanisms which affect Upper Klamath Chinook and their habitat include the establishment of Total Maximum Daily Loads (TMDLs) for chemical pollution in the Klamath River. The Klamath River is listed as a water quality impaired river under Section 303(d) of the Clean Water Act and as required by the Act, states are required to establish TMDLs for instate impaired waterways. Enforceability of TMDLs is difficult and insufficient. The continued occurrence of dangerous algal blooms in reservoirs in this river system clearly illustrates the inadequacy of this regulation. Federal regulators recently adopted new TMDLs calling for a 57% reduction in phosphorous and a 32% reduction in nitrogen and a 16% cut in carbonaceous biochemical oxygen from wastewater. Although the new TMDLs are intended to protect salmon resources, there are no implementation programs in place for controlling pollutant inputs from land use. Without these implementation plans, standards are unlikely to be met.

**State Regulatory Mechanisms: Mining**

California instated a ban on suction dredge mining in 2009 in response to a lawsuit from the Karuk Tribe referencing damage to fish habitat and water quality. This ban is clearly beneficial for Upper Klamath Chinook. However, the ban is temporary until the California Department of Fish and Game completes an environmental review of suction dredge mining. There is no guarantee that this mining practice will not be reintroduced after the environmental review occurs.

**Federal and State Regulatory Mechanisms: Fishing**

Fishing harvest allocations are decided annually based on input from federal, state, regional, and tribal bodies. In general, tribes maintain the right to fifty percent of the total annual harvest. Within tribal and non-tribal fishing, further allocations are assigned for commercial ocean fisheries, sport, and subsistence fishing. Harvest quotas are based on projections for run size each year and attempt to maintain a minimum spawning escapement of 35,000 fish to protect the runs for the long-term. Overfishing is an aggravating factor to the grim future of Upper Klamath Chinook; fishing regulations alone will not provide for the continued existence of this ESU. As noted above in section 6.2 over-exploitation, because managing agencies do not treat spring-run Chinook differently from fall-run Chinook, spring-run fish are taken legally in commercial and sport fisheries (Moyle et al. 2008). Further enhancing the problem, current management actions neglect to protect spring-run Chinook even when protections have been put in place to restrict fall-run Chinook harvest, essentially increasing pressure on the much smaller and more imperiled populations of spring-run Chinook, as took place in 2017 when fall-run Chinook harvest was closed on the Klamath River to all fishing while bag limits remained the same during the spring run period.

**Federal and State Regulatory Mechanisms: California Forest Practices Rules**

California Forest Practices Rules are developed under the California Forest Practices Act of 1943 which governs logging practices on all private lands. These rules are inadequate to prevent harm to Upper Klamath Chinook.

**Regulatory Mechanisms: Climate Change**

Current global, national, and state climate change legislation and agreements are entirely inadequate to prevent ocean acidification and the variability of other ocean conditions aggravated by climate change. As noted, these conditions pose a significant threat to the long-term survival of salmonids in their marine environment.

Greenhouse gas emissions and resulting climate change is among the least regulated threats to Upper Klamath Chinook. The primary international regulatory mechanisms addressing greenhouse gas emissions and global warming are the United Nations Framework Convention on Climate Change, the Kyoto Protocol,
and the Copenhagen Accord. While the entering into force of the Kyoto Protocol on February 16, 2005 and the development of the Copenhagen accord in December, 2009 mark significant partial steps towards the regulation of greenhouse gases, they do not and cannot adequately address the impacts of global warming that threaten the Upper Klamath Chinook.

Choices about emissions now and in the coming years will have far-reaching consequences on the magnitude of climate change impacts. The longer greenhouse gas emissions reductions are delayed, the more severe the global impacts will be (Karl et al. 2009). If global warming is going to be limited to 2°C above pre-industrial values, global emissions need to peak between 2015 and 2020 and then decline rapidly (Allison et al. 2009). This will require average annual per-capita emissions to shrink to under one metric ton CO₂ per capita. This is 80-95% below the per capita emissions in developed nations in 2000 (Allison et al. 2009).

There are currently no legal mechanisms regulating greenhouse gases on a national level in the United States. The immediate reduction of greenhouse gas pollution is essential to slow global warming and ultimately stabilize the climate system in order to maintain and restore Upper Klamath Chinook habitat.

For the reasons discussed, existing and proposed regulatory mechanisms are indisputably inadequate to ensure the continued survival of the Upper Klamath Chinook salmon.

9. SUGGESTIONS FOR FUTURE MANAGEMENT

The steeper decline of UKTR Spring Chinook relative to fall-run UKTR Chinook stems in great part from their need to spend more time as an adult in fresh water during summer months when flows are low. Historically, the Klamath Basin offered unfettered access to higher elevation flood plain habitat and spring fed cold water refugia for adult UKTR Spring Chinook. Today, access to much of these habitats is blocked by dams, cold water springs are diverted for agricultural purposes and flood plains physically altered by mining or sedimentation associated with poor logging practices and road maintenance. Hatchery practices both at the Trinity at Iron Gate hatcheries may negatively impact genetic integrity, variability, and fitness of UKTR Spring Chinook. In addition, UKTR Spring Chinook are particularly susceptible to the warming trends associated with global warming and prolonged droughts.

In light of these facts, we suggest the following future management actions be considered:

i. Remove the lower four Klamath River dams consistent with the terms of the Klamath Hydroelectric Settlement Agreement and PacifiCorp’s pending application before FERC.

ii. Currently, the Salmon River and South Fork Trinity sub-basins offer the largest spawning populations of the UKTR Spring Chinook in the Klamath system. These sub basins should be managed explicitly for the restoration, protection, and management of UKTR Spring Chinook.

iii. The Shasta River should be managed as a cold-water refuge, restrictions should be placed on agricultural diversions affecting flow and temperature, ground water extraction should be limited and removal of Dwinnell dam should be considered.

iv. The Scott River should be managed for UKTR Spring Chinook, which means restrictions should be placed on agricultural diversions affecting flow and temperature, ground water extraction should be limited, and removal of Young’s Dam should be considered.

   a. Manage the Salmon River as a UKTR Spring Chinook refuge and prioritize restoration
projects aimed to restore floodplain habitat affected by historic mining and minimizing impacts associated with logging projects and grazing. Implementation of the Salmon River Floodplain Habitat Enhancement and Mine Tailing Remediation Plan and recommended restoration projects.

b. Potential restoration and enhancement actions for the Salmon River include the following:

i. Protecting and expanding cold water refuges at summer baseflow within the mainstem channels and lower reaches of major tributaries to improve holding and summer rearing habitat conditions;

ii. Adding structure within simplified channel reaches (e.g., plane-bed morphology) that promotes hydraulic complexity and pool depth, increasing the amount and quality of low velocity rearing habitat, and sorting spawning gravel;

iii. Manipulating (e.g., grading and/or adding structure) and revegetating floodplains to improve hydrologic function and processes, primarily by increasing flow connectivity (e.g., frequency and duration of inundation) and hyporheic exchange between the winter baseflow channel (20% exceedance flow), bankfull side channels (1.5- to 2-year flow), and high flow side channels (≥5-year flow);

iv. Adding structural complexity to side channels to improve rearing habitat;

v. Creating, enhancing, and connecting off-channel ponds and wetlands to improve rearing habitat; and

vi. Grading and revegetating mine tailings on floodplains and adjacent terraces to increase riparian shading, reduce heating, and improve hyporheic exchange.

c. Implement key actions from the collaboratively developed Salmon River In-stream Candidate Action Table and the Middle Klamath In-stream Candidate Action Table.

v. Develop limiting factors analysis for Klamath River spring-run Chinook for the Klamath River and all tributaries within the historic range of spring-run Chinook.

vi. Conduct assessments and develop restoration action plans to address the impacts of historic mining throughout key tributaries and the mainstem of the Klamath Basin.

vii. Develop on comprehensive Klamath Basin spring-run Chinook recovery plan and associated restoration action plan.

viii. Develop restoration actions and priorities for reducing the impacts of sediment inputs from roads, logging, and other activities into rivers of the Klamath-Trinity system, especially on public lands.

ix. Prevent dewatering of habitats and limit effects of pesticides/herbicides associated with legal marijuana cultivation through permitting programs.

x. Develop a program to investigate impact(s) of the Trinity River Hatchery on UKTR Spring Chinook populations (e.g., number of hatchery-reared fishes spawning in the wild, genetic shifts in population) and manage hatchery production accordingly. Rates of hybridization between spring-run and fall-run Chinook and relative fitness of the offspring should be paid particular attention.

xi. Investigate whether a conservation hatchery can play a role in facilitating re-colonization of Klamath River tributaries by UKTR Spring Chinook after dam removal occurs. If such an approach is explored, efforts must be made to reduce genetic impacts of founder’s effects and inbreeding/outbreeding depression.
xii. Limit recreational in-river harvest to a mark-selected fishery for 100% adipose fin clipped Trinity River Hatchery produced spring-run Chinook to keep them separate from wild fish.

xiii. Ban suction dredge mining in all areas deemed current or potential habitat.

xiv. Restore headwaters and high mountain meadow systems throughout the basin and in particular in key spring-run Chinook watersheds to maximize cold water storage, lengthen cold water releases, and promote resiliency in the face of climate change.

xv. Restore healthy fire process at a landscape scale on the Klamath Basin through increased use of prescribed fire, managed wildfire, and associated fuels treatments.

xvi. Implement the Western Klamath Restoration Partnership Plan (Harling, Tripp, 2014)

10. **AVAILABILITY AND SOURCES OF INFORMATION**

   Please see bibliography at the end of attached NMFS petition
Klamath Trinity spring-run Chinook current and historic distribution map, created by SRRC from available data, 2015.

12. ADDITIONAL INFORMATION

Legal/Regulatory Background

Recognizing that certain species of plants and animals have become extinct “as a consequence of man’s activities, untempered by adequate concern for conservation,” (Fish & G. Code § 2051 (a)) that other species are in danger of extinction, and that “[t]hese species of fish, wildlife, and plants are of ecological, educational, historical, recreational, esthetic, economic, and scientific value to the people of this state, and the conservation, protection, and enhancement of these species and their habitat is of statewide concern.”
The California Legislature enacted the California Endangered Species Act (CESA).

The purpose of CESA is to “conserve, protect, restore, and enhance any endangered species or any threatened species and its habitat....” (Fish & G. Code § 2052). To this end, CESA provides for the listing of species as “threatened” and “endangered.” The Commission is the administrative body that makes all final decisions as to which species shall be listed under CESA, while the Department is the expert agency that makes recommendations as to which species warrant listing. The listing process may be set in motion in two ways: “any person” may petition the Commission to list a species, or the Department may on its own initiative put forward a species for consideration. In the case of a citizen proposal, CESA sets forth a process for listing that contains several discrete steps.

Upon receipt of a petition to list a species, a 90-day review period ensues during which the Commission refers the petition to the Department, as the relevant expert agency, to prepare a detailed report. The Department’s report must determine whether the petition, along with other relevant information possessed or received by the Department, contains sufficient information indicating that listing may be warranted. (Fish & G. Code § 2073.5).

During this period interested persons are notified of the petition and public comments are accepted by the Commission. (Fish & G. Code § 2073.3). After receipt of the Department’s report, the Commission considers the petition at a public hearing. (Fish & G. Code § 2074). At this time the Commission is charged with its first substantive decision: determining whether the Petition, together with the Department’s written report, and comments and testimony received, present sufficient information to indicate that listing of the species “may be warranted.” (Fish & G. Code § 2074.2). This standard has been interpreted by as the amount of information sufficient to “lead a reasonable person to conclude there is a substantial possibility the requested listing could occur.” If the petition, together with the Department’s report and comments received, indicates that listing “may be warranted,” then the Commission must accept the petition and designate the species as a “candidate species.” (Fish & G. Code § 2074.2.)

Once the petition is accepted by the Commission, then a more exacting level of review commences. The Department has twelve months from the date of the petition’s acceptance to complete a full status review of the species and recommend whether such listing “is warranted.” Following receipt of the Department’s status review, the Commission holds an additional public hearing and determines whether listing of the species “is warranted.” If the Commission finds that the species is faced with extinction throughout all or a significant portion of its range, it must list the species as endangered. (Fish & G. Code § 2062.) If the Commission finds that the species is likely to become an endangered species in the foreseeable future, it must list the species as threatened. (Fish & G. Code § 2067.)

Notwithstanding these listing procedures, the Commission may adopt a regulation that adds a species to the list of threatened or endangered species at any time if the Commission finds that there is any emergency posing a significant threat to the continued existence of the species. (Fish & G. Code § 2076.5).

Unlike ESA, CESA does not contain a definition of “species” or “subspecies” in its text, nor does it determine whether or not an Evolutionarily Significant Unit (ESU), as defined in the Federal Endangered Species Act (ESA) and detailed below, may be listed as an Endangered Species under CESA. However, in California Forestry Assn. v. California Fish & Game Comm., it was determined that “the [California]
legislature did not want to limit the term ‘species or subspecies’ to the federal definition. Instead the legislature likely may have wanted to leave the interpretation of that term to the Department…and to the Commission”.3 Further, the decision elaborated that the Department and the Commission have a “longstanding adherence to the policy that the CESA allows listings of evolutionary significant units”.4 Thus, if there is sufficient evidence to show that a subset of a species should be considered an ESU under ESA, the Commission and Department should consider a petition for listing that subset as its own Endangered Species under CESA.

The Federal Endangered Species Act defines “species” to include “any subspecies of fish or wildlife or plants, and any distinct population segment of any species of vertebrate fish or wildlife which interbreeds when mature.” 16 USC § 1533(16), see also California State Grange v. National Marine Fish, 620 F.Supp 2d 1111, 1121 (ED Cal 2008). The ESA does not define the term “distinct population segment.” Grange at 1121.

In 1991 the National Marine Fisheries Service (“NMFS”) promulgated its “Policy on Applying the Definition of Species Under the Endangered Species Act to Pacific Salmon” or “ESU Policy.” (56 Fed.Reg.58612 (Nov. 20, 1991)). The ESU Policy provides that a population of Pacific salmonids is considered to be an ESU, and therefore considered for listing under the ESA, if it meets the following two criteria:

(i.) It must be substantially reproductively isolated from other nonspecific population units; and
(ii.) It must represent an important component in the evolutionary legacy of the species. Isolation does not have to be absolute, but it must be strong enough to permit evolutionarily important differences to accrue in different population units. The second criterion would be met if the population contributes substantially to the ecological/genetic diversity of the species as a whole (Waples 1991). Grange at 1123-24.

NMFS uses all available lines of evidence in applying those criteria, including specifically data from DNA analyses (“…data from protein electrophoresis or DNA analysis can be very useful because they reflect levels of gene flow that have occurred over evolutionary time scales.”), ESU Policy, 56 Fed. Reg. at 58518; see also Definition of “Species” Under the Endangered Species Act: Application for Pacific Salmon, NOAA Tech Memo NMFS F/NWC-194 (Waples 1991) at p.8 (“The existence of substantial electrophoretic or DNA differences from other conspecific populations would strongly suggest that evolutionarily important, adaptive differences also exist.”)

The ESU Policy is an interpretation by NMFS of what constitutes a “distinct population segment,” and is a “permissible agency construction of the ESA.” Grange at 1124, citing Alsea Valley Alliance v. Evans, 161 F.Supp2d 1154, 1161 (D.Or. 2001).

When considering whether a species or subspecies, including an ESU, is endangered, NMFS must consider:

i. The present or threatened destruction, modification, or curtailment of its habitat or range;
ii. Overutilization for commercial, recreational, scientific, or educational purposes;
iii. Disease or predation;
iv. The inadequacy of existing regulatory mechanisms; or
v. Other natural or manmade factors affecting its continued existence.

3 California Forestry Assn. v. California Fish & Game Comm. 156 Cal. App. 4th 1535 at 1549.
4 Ibid at 1546.
The species shall be listed where the best available data indicates that the species is endangered because of any one, or a combination of, those five factors. 50 CFR § 424.11(c).

Any interested person may submit a written petition to list a species or subspecies as threatened or endangered. 50 CFR § 424.14(a).

The newly proposed 50 CFR §424.14(g)(1)(iii) states that petitions filed after an adverse ruling will be considered only where "new information or analysis such that a reasonable person conducting an impartial scientific review would conclude that the action proposed in the petition may be warranted, despite the previous determination." 81 Fed. Reg. 23454-55. NMFS states further that the proposed §424.14(f) will "clarify" the Service’s position that any supplemental petition will be considered with the previous petition, and they together will reset the statutory periods for response—constructively the same as filing a new petition. 80 Fed. Reg. 29289 (21 May 2015).

Factual Background

Chinook salmon in the upper Klamath and Trinity Rivers are currently regulated and managed as a single ESU referred to as Upper Klamath Trinity River (UKTR) Chinook, with no distinction between seasonal runs. The Klamath Trinity spring (KTS) Chinook is not defined as its’ own unique ESU, and is not listed as threatened or endangered. Water management, fisheries management, and other regulatory activities are generally conducted without consideration of potential impacts on KTS Chinook, instead considering impact to UKTR Chinook generally. This approach may be having an adverse impact on KTS Chinook especially when hatchery practices are considered.

In an effort to explain differences in run timing observed in Chinook salmon populations, conservation geneticists offer two possible explanations for the evolution of spring, or “premature,” migration patterns for salmonids: a monophyletic pattern of evolutionary history versus a polyphyletic pattern of evolutionary history. These models are based on a comparison of the DNA structure of fall and spring run individuals within the same watershed versus nearby watersheds using a variety of genetic techniques.

In evaluating whether to list seasonal runs as Evolutionarily Significant Units ("ESU") for purposes of the Endangered Species Act, the National Marine Fisheries Service ("NMFS") considers which of these two evolutionary models apply to the given population. Because spring and fall run fish fitting the polyphyletic pattern evolve from a common ancestor based on environmental factors, the genetic material for both seasonal runs are contained in fish from both runs. The evolutionary changes necessary to give rise to the phenotype are relatively easy to reproduce since, according to this model, it has happened many times in closely related populations. NMFS has argued that even if spring run migrating subpopulations were extirpated by flow diversions, barriers, or other factors, the spring migration phenotype could easily re-emerge if appropriate habitat was later restored. On that basis, polyphyletic pattern fish runs typically do not meet NMFS guidance requirement to qualify as an ESU. According to Waples, “Although the failure of most stock transfers indicates that local populations may be largely irreplaceable on human time frames, at least some patterns of Chinook salmon life history diversity appear to be evolutionarily replaceable, perhaps over time frames of a century or so. The evidence for repeated parallel evolution of run timing in Chinook salmon indicates that such a process is likely, provided that habitats capable of supporting alternative life-history trajectories are present and sufficient, robust source populations are maintained” (Waples et al. 2004).

In contrast, seasonal fish runs that evolved via the monophyletic pattern evolved from a separate ancestor, and are genetically distinct from other fish runs in that river system. Thus if extirpated, monophyletic seasonal fish runs are likely gone forever, and thus warrant classification as an ESU, as well as the protections that result from such a listing.
Until now, most conservation geneticists considered most spring run Chinook populations to fit the polyphyletic model. This would mean that fish from a common ancestor evolve genetic differences due to the reproductive isolation and natural selection driven by the unique features of their respective watersheds. According to this explanation, these separate populations later evolved the early migration or ‘spring run’ phenotype independently from each other. In other words, the spring run phenotype evolved many times over in neighboring populations. The application of the polyphyletic model to these populations stems from studies that show that the genetic structures of spring and fall run individuals within a watershed are more genetically similar than spring run individuals from different watersheds. Examples of runs thought to be a product of this process include spring and fall run Chinook in the Rogue and Umpqua (Waples et al. 2004).

However, in some fish populations the DNA structure of fall and spring run individuals within the same watershed are less similar to one another than those in neighboring watersheds. These observations suggest an alternative explanation for the evolutionary basis for the early migration phenotype. In these cases, the difference in run timing is attributed to a monophyletic pattern of evolutionary history. Under this model the genetic changes that give rise to differences in run timing predate the genetic differences that arise as a consequence of geographic isolation. Until now, the only known examples of monophyletic based premature migration are among spring run and fall run Chinook salmon in the mid and interior Columbia and Snake River basins, and winter, spring and fall run Chinook populations in California’s Central Valley. The fish in each of these seasonal runs are more closely related to each other than to Chinook salmon in any other basin, or to other Chinook salmon runs in the same tributary river (Meyers et al 1998; Banks et al 2000a; Garza et al 2007). Some researchers argue that the differences observed in the Central Valley spring and fall populations stem more from anthropogenic factors associated with hatchery management than with a true evolutionarily event.

In summary, conservation biologists consider most populations of spring Chinook salmon to be a product of polyphyletic evolution, except in a few rare exceptions where it is not.

In a memo summarizing the finding of the Biological Review Team (BRT) report on the 2011 Petition, the Science Director of the National Marine Fisheries Service Southwest Fisheries Science Center, Francisco Werner, noted that “One reviewer expressed the personal view that there is evidence for reproductive isolation and adaptive divergence between Klamath River spring-run and fall-run Chinook salmon and thus merit their own ESU. However, the reviewer found that spring-run Chinook salmon in the UKTR basin do not represent a unique component of the evolutionary legacy of the species, and therefore, do not meet one of the two requirements for recognition as an ESU under NMFS’ ESU policy (the other requirement being long-term reproductive isolation resulting from an unique evolutionary event that is unlikely to re-evolve over ecological time-scales)”(Werner 2011). However, recently published work challenges the assertion that spring run Chinook does not meet the other requirement. The study shows that a unique evolutionary event was the cause for the spatial and temporal reproductive isolation that spring and fall run exhibit in the UKTR, and shows that spring run life type Chinook are unlikely to re-evolve over ecological time scales (Prince et al. 2017).

2011 Petition for Listing UKTR Chinook

In 2011, Center for Biological Diversity (CBD) et al. filed an Endangered Species Act (ESA) listing petition (“2011 Petition”) with NMFS to address the dramatic declines of Klamath River spring Chinook salmon. CBD et al. suggested 3 alternatives for NMFS to consider: 1) list spring run Chinook as their own evolutionary significant unit (ESU); 2) list spring run Chinook as a distinct population segment (DPS) within the previously recognized UKTR Chinook ESU; or 3) list the entirety of the UKTR Chinook ESU (Center for Biological Diversity et al. 2011).
In its initial response to the 2011 Petition, the NMFS Southwest Region (SWR) determined that “… the literature cited in the petition, and other literature and information available in our files, we found that the petition met the criteria in our implementing regulations at 50 CFR 424.14(b)(2) that are applicable to our 90-day review and determined that the petition presented substantial information indicating that the petitioned action may be warranted” (National Marine Fisheries Service 2011) (76 FR 20302; April 12, 2011).

In that 90-day finding, NMFS narrowed the scope of their pending further review. In particular, the agency explained that it would not consider Petitioners’ second alternative for listing Chinook salmon in the UKTR ESU as a DPS. Instead, NMFS determined that the analysis would consider whether the KTS Chinook constitutes an ESU. NMFS noted that their Policy on Applying the Definition of Species Under the Endangered Species Act to Pacific Salmon, “…explains that a Pacific salmon stock will be considered a distinct population segment, and hence a “species” under the ESA, if it represents an ESU of the biological species” (ESU Policy; 56 FR 68612; November 20, 1991).

2011 Biological Review Team Determination

After determining that the petition actions met the appropriate criteria and may be warranted, NMFS convened a Biological Review Team (BRT) which considered the 2011 Petition and over 50 written comments from the public. Specifically, the BRT considered two fundamental issues: 1) the extent to which the new information supports the current UKTR Chinook Salmon ESU delineation, or the separation of spring-run and fall-run Chinook salmon into separate ESUs, and 2) assessment of the biological status of the supported ESU configuration using the viable salmonids population framework (Williams et al. 2011).

In the 2011 Petition, CBD et al. argued that the KTS Chinook evolved via the monophyletic pattern, and thus qualified for listings as an ESU. CBD pointed to new genetic data, and argued that KTS Chinook show genetic and life history divergence from fall run UKTR Chinook equal or greater than those of the Central Valley spring and fall run Chinook ESUs.

The BRT reviewed the new genetic data brought forth by CBD et al. The BRT did not agree based on the data that a monophyletic evolutionary model best described the prevalence of the KTS Chinook. Rather, the BRT argued that a polyphyletic evolutionary history best explained the ‘premature’ migration pattern observed within the UKTR Chinook ESU. While acknowledging some genetic differences between various UKTR Chinook runs, the BRT concluded that the genetic and life history differences of the KTS Chinook were not great enough to warrant the designation of ESU status. The BRT stated,

“The BRT concluded that the new information supports the ESU delineation of Myers et al. (1998) in which UKTR spring-run and fall-run Chinook salmon populations constitute a single ESU, and that the expression of the spring-run life-history variant is polyphyletic in origin in all of the populations for which data are available.”

The BRT went on to conclude that considered as a whole population, UKTR Chinook were not threatened or endangered, stating:

“As to the status of the UKTR Chinook Salmon ESU, the BRT found that the ESU is currently at low risk of extinction within the next 100 years” (ibid.)

The results and conclusions of the BRT report was the basis of the 12 month finding published in the Federal Register on April 2, 2012 which rejected the 2011 Petition of CBD et al. to list KTS Chinook salmon (National Marine Fisheries Service 2011).
NMFS’ 2011 conclusion was consistent with the large body of literature based on genetic analyses performed using microsatellites. While these studies often revealed genetic differences between geographically isolated populations, they failed to consistently demonstrate significant differentiation between premature and mature migrating phenotypes within a watershed (Kinziger et al. 2013; Waples 1991; Nielsen, Crow, and Fountain 1999). As a consequence, early migration phenotypes, including the KTS Chinook, have been largely grouped into the same ESU or DPS as mature migration phenotypes.

Until recent advances in genetic analysis, researchers were limited by the available technology in how they could study the genetic differences between closely related populations. Previously, researchers looked for relatively large differences in genetic structure, which often appear in genomic regions not influence by environmental pressures and natural selection, because the available technology allowed this sort of analysis. These genomic regions vary due to gene flow and genetic drift, as opposed to being driven by environmental pressures and natural selection. The weakness of this approach is that it lacks the molecular resolution necessary to detect evolutionarily significant adaptations that may stem from changes in sequence and structure in specific genomic regions, particularly in regions that encode genes.

Although the relatively large body of data is indeed consistent with the hypothesis that polyphyletic evolution explains premature run timing (at least in most cases), the evidence is also consistent with another explanation – that premature run timing is the result of a changes in genetic sequence or structure of specific regions of the genome that predates the polyphyletic changes brought on by geographic isolation. Until recently conservation geneticists lacked the tools necessary to fully explore the latter hypothesis. However, recent advances in technology now allow researchers to comb through genomes at a much higher resolution cheaply and quickly. Previously, researchers would rely on dozens or maybe hundreds of molecular markers to search for genetic differences between subpopulations. Today, researchers can quickly compare millions of genetic regions to look for differences.

Based on the technical limitations of genetic analysis, the previous approach to determining the evolutionary history of the premature migration phenotype was inferential. In other words, conservation geneticists inferred the evolutionary history of the phenotype based on demography not adaptation. The new technology now allows researchers to locate individual genomic regions that are the actual cause of evolutionary change, and reconstruct the evolutionary history of these regions directly. This direct reconstruction of the evolutionary history of the spring run Chinook versus fall run Chinook has now been performed and recently published in a peer reviewed journal (Prince et al. 2017).

Prince et al. created a high-resolution genomic library from samples of spring and fall migrating adult Chinook and steelhead from several Pacific Northwest watersheds, including the Klamath. The researchers then created high-resolution restriction-site associated DNA (RAD) libraries, sequenced them, and aligned the sequences to a recent salmonid genome draft. The genomic libraries generated from individual fish where then compared using a probabilistic framework to discover small nuclear polymorphisms (SNPs). Although Prince et al. notes that the initial analysis was consistent with current DPS and ESU delineations, the sheer volume of genomic positions they went on to compare (nearly 10 million) allowed a thorough comparison of premature and mature migrating individuals. This revealed several SNPs within a couple hundred thousand base pairs of one another. Further analysis revealed this region to be within the GREB1L gene. This result was then repeated in other populations including UKTR Chinook. Prince et al. notes that this finding makes biological sense in that this gene is implicated in foraging and fat storage in mammals. In salmon, premature migrating Chinook have a significantly higher fat content than mature migrating individuals, consistent with the fact that early migrating individuals are destined to climb higher into watersheds before spawning and thus need more stored energy.
Prince et al. went on to sequence the GREB1L region in all of their samples and created a gene tree based on parsimony. The tree revealed two monophyletic groups corresponding to migration phenotype. All samples, regardless of watershed of origin, separated into the appropriate migratory clade. In other words, Prince et al. found that all premature migrating individuals evaluated grouped together in the same monophyletic group. Thus, genetic differences in this single gene explain the difference between premature and mature migrating phenotypes. Although NMFS has argued that “some patterns of Chinook salmon life history diversity appear to be evolutionarily replaceable, perhaps over time frames of a century or so…” (Waples et al. 2004), premature migration clearly does not fall into this category as explained in greater detail below.

Without the advent of molecular tools that allow for the cheap and quick creation of detailed DNA libraries (collectively referred to as Next Generation Sequencing or NGS), the identification of a single gene that is responsible for such a complex phenotype would have been nearly impossible. Now that the technology is available and has been applied, however, the monophyletic nature and evolutionary significance of UKTR Spring Chinook must be acknowledged.

**UKTR Spring Chinook**

Myers et al. (1998) recommended that their determination, that spring-run and fall-run Chinook salmon populations in the UKTR ESU constitute a single ESU, should be revisited if substantial new genetic information from natural spring-run populations were to become available (Williams et al. 2011). This Petition presents precisely that genetic information for the upper Klamath Trinity River system Chinook populations. For spring run and fall run populations of Chinook salmon to be considered separate ESUs, as defined by Waples (1991) and later elaborated on by Waples (1995), it must be shown that these populations are substantially reproductively isolated from other conspecific population units and that they represent an important component in the evolutionary legacy of the species. Prince et al. makes that demonstration.

It is well established that spring Chinook, by virtue of entering fresh water rivers during snow melt, reach spawning areas that are, generally, reproductively isolated from their fall run counterparts (Quinn 2005). Waples’ concept of evolutionary legacy implies that there would need to be a monophyletic pattern of the evolutionary history of the two run-types within the UKTR. For spring run Chinook, Prince et al. demonstrate that the molecular basis for the spring run phenotype is associated with a defined allele that evolved long ago in Chinook evolutionary history. Prince et al. found evidence of only two allelic evolutionary events that produced a premature migration allele, one in Chinook and one in steelhead, even though the species diverged approximately 15 million years ago. This is in contrast to the assertion by the BRT review of the previous KTS Chinook petition which concluded, without the benefit of Prince et al.’s recent findings, that the spring run phenotype is polyphyletic in origin and evolved independently in many locations.

Prince’s recently published data clearly demonstrate that contrary to prevailing dogma, Klamath-Trinity spring Chinook exhibit a monophyletic pattern of evolutionary history, and meet Waples’ and NMFS’ criteria for a separate ESU.

A more recent publication (Thompson et al. 2018) further strengthens this argument and calls into question any assertion that Klamath spring-run Chinook will reemerge from Chinook heterozygotes once the spring-run phenotype is lost:

> “using a new marker identified through a high-resolution, multi-population analysis of GREB1L suggests that 1) the association of migration type with variation at GREB1L is extremely robust and 2) heterozygotes have an intermediate migration phenotype.
Therefore, while phenotypic variation within each genotype (e.g., precise freshwater entry and spawning dates) is yet to be explained, migration type (i.e., premature/spring-run or mature/fall-run) appears to have a strikingly simple genetic architecture. Furthermore, the association of a single haplotype with the spring-run phenotype in diverse locations supports a previous conclusion that spring-run alleles arose from a single evolutionary event and cannot be expected to readily re-evolve (Prince et al., 2017; Miller et al., 2012). Thus, simple modes of inheritance and rare allelic evolutionary events can underpin complex phenotypic variation.”

Citing evidence that heterozygotes are selected against, Thompson et al. conclude that that, “where the spring-run phenotype is lost, spring-run alleles should not be expected to be maintained in the heterozygous state… both theory and empirical evidence suggest heterozygotes are not a sustainable reservoir for spring-run alleles, and human factors can eliminate important adaptive variation regardless of total population size.”

As previously noted, the criteria for an ESU designation are that 1) it must be substantially reproductively isolated from other nonspecific population units; and 2) it must represent an important component in the evolutionary legacy of the species.

Prince et al. 2017 demonstrates that KTS Chinook are an important component in the evolutionary legacy of UKTR Chinook and that the reproductive isolation between spring and fall run populations is strong enough to permit evolutionarily important differences to accrue. Thompson et al. 2018 further demonstrate the point.
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