The Vegetation of Suisun Marsh, Solano County, California: First Permanent Plot Resample Study 1999 vs. 2006

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METHODS

In keeping with the monitoring focus of the Suisun Marsh Vegetation Surveys (Keeler-Wolf and Vaghti 2000), in 2006, 100 of the original 198 Suisun Marsh vegetation plots were selected to be revisited for the first time since their establishment in the summer of 1999. To enable regular resampling into the future without any access issues, only those plots located on DFG land, Rush Ranch, or other publicly-accessible lands were considered for resampling. Of these plots, 100 were selected by the DFG Vegetation Classification and Mapping Program to capture the diversity of vegetation types that represent the Marsh's vegetation as a whole. Due to the construction of the new Benicia-Martinez Bridge, two of the selected 100 plots are no longer in existence. Therefore, 98 plots were resampled (Figure 1). Each plot was located based on careful interpretation of GPS coordinates, field sample photos, aerial photos, and plot descriptions provided in the original 1999 field data.

Figure 1. Location of resampled points and points not resampled in 2006



Relocating (Finding) the Original Suisun Marsh Relevé Plots in 2006

In 1999, the location of field plots was recorded with Trimble Geoexplorer GPS units that were differentially corrected using a nearby base station to typically ± 3 m. In 2006, these coordinates were loaded into the Garmin 12XL unit and were navigated to directly. The general location of each plot was relatively easy to find with a GPS unit, aerial photo, and the 1999 data sheet. Driving directions (and sometimes boat directions) were usually explicitly stated on the 1999 data sheets. More problems occurred when trying to determine the exact location of the plots. No standard was implemented in 1999 for the GPS recording point within each plot. Therefore, in 2006 we had to make a best guess based on the 1999 photos and the 1999 GPS point. This was not always accurate or reliable due to the error of the original GPS point, the error in the 2006 GPS reading, and the lack of precise location landmarks in many of the 1999 photos.

The orientation of the rectangular plots was also difficult to determine. It was never stated explicitly on any of the rectangular plot data sheets which way the long or short axis was oriented. Many times it was obvious (a narrow strip of vegetation along a levee or road), but sometimes field crews had to make an educated guess (especially if the ground photos were not useful).

In 2006 these standardization problems were addressed to enable future resampling to proceed more smoothly. The GPS point was almost always taken from the SW corner of the plot along with four additional photos that were taken of the cardinal directions. This was always recorded on the data sheet. Also, the orientation of each rectangular plot was stated explicitly. The corners of each plot were placed in the SW, NW, SE, and NE unless otherwise stated. If it was determined that in 1999 the plot was laid out so that the corners were in the exact cardinal position, or that the axis of the plot paralleled a road, levee, slough, etc., then this was also done in 2006 and recorded on the datasheet. To further aid future surveyors in relocating these plots, more photos were taken which were thought to give a better indication as to the position of the plots. All other elements of the original 1999 sampling protocol were replicated as best as possible.

Sampling Protocol in Both the 1999 and 2006 Efforts

Sites dominated by vegetation taller than 5 m were sampled in 1000 m sq plots. All other vegetation, including graminoids, shrubs and herbs, was sampled in 400 m sq plots. Plots were typically square but other shapes were used depending on the general dimensions of the vegetation to be sampled (e.g., long riparian corridors were typically sampled as long strips that totaled 1000 m sq). Plot size and shape were recorded on each field form. The variable size and shape of the plot based on the physiognomy of the vegetation and the fact that we collected estimates of cover for species rather than exact measurements exemplify characteristics of a phytosociological relevé (Barbour et al. 1998) rather than a fixed plot or point-intercept sample.

Global positioning systems were used to record the sample plots. Information regarding GPS file name and duration of data collection, which aided accuracy for the Trimble unit, was noted on the field form for 1999 plots; this was not necessary in 2006.

In 1999, record keeping was based on the assignment of plots to a particular vegetation polygon number. First, a preliminary number was given to the sample based on the aerial photo covering

the area of the sample and individual numbers of polygons within that photo. The polygon numbers were re-assigned following entry of all polygons into the GIS system. In 2006, the final plot number from 1999 was used.

Estimates of percent cover were required for all species greater than or equal to 1 percent cover; if less than 1 percent cover, cover was noted as "less than 1 percent." Additional estimates for total vegetation cover, and total tall, medium and low cover are included. These were thought to be important for such polygon attributes as total cover estimates. A separate entry for non-natives was noted to help with assessing impacts of invasive species.

Cover estimates for seven height classes were assigned based on a six-point scale. The dominant species for each height class was also recorded.

As with plant species, the cover values for open water, bedrock, gravel, cobble, stone, and litter were estimated in cover classes and actual percent of plot.

Appendix A contains a sample field form and the full sampling protocol.

Table 1 presents the number of person-hours required to resample the 98 plots in 2006.

	# of	# of Hrs	Total Person
Date	People	Worked/Person	Hrs/Day
17-Jul-06	4	8	32
18-Jul-06	4	8	32
19-Jul-06	5	8	40
20-Jul-06	4	8	32
24-Jul-06	2	8	16
25-Jul-06	2	8	16
26-Jul-06	2	8	16
27-Jul-06	2	8	16
31-Jul-06	2	8	16
1-Aug-06	2	8	16
2-Aug-06	3	8	24
3-Aug-06	2	8	16
7-Aug-06	2	8	16
8-Aug-06	2	8	16
9-Aug-06	2	8	16
10-Aug-06	2	8	16
5-Sep-06	2	8	16
7-Sep-06	2	8	16
11-Sep-06	2	8	16
12-Sep-06	2	8	16
13-Sep-06	2	8	16
19-Sep-06	3	6	18
I otal Persor	1 Hours		434

Table 1. Person-hours spent resampling 98 plots at Suisun Marsh

Issues of Plot Change

The original 1999 plots were located based on stand homogeneity and were thought to represent the larger stand of a particular type. However, with the combination of management-induced changes such as flooding regime, mechanical manipulation (disking, ditching, etc.), burning, or seeding of certain desirable species, some of the original stands had clearly changed internally, leading to the location of the plots currently not within homogeneous patches of vegetation. When stand boundaries had shifted from the original sample, the location was kept the same, and additional notes were taken describing the types of shifts noted from the 1999 sample.

Comparisons of the 1999 and 2006 Samples

This report compares the original 1999 samples and the 2006 resamples with regard to the following characteristics:

- Vegetation type as identified using the key developed from the 1999 field samples as displayed in Keeler-Wolf and Vaghti (2000);
- Top three dominant species comparison from 1999 to 2006;
- Shifts in non-native species composition; and
- Basic shifts in floristic diversity and water cover.

RESULTS

Vegetation Type Changes

Table 2 presents the originally classified and currently classified names for each of the resampled plots. The key to vegetation types originally developed for the marsh following the 1999 sampling (Keeler-Wolf and Vaghti 2000) was used again to identify the vegetation growing in the same plot in 2006. Names indicated in bold have shifted from their original 1999 classification at the plant association level. Those highlighted in yellow have shifted at the alliance level of the vegetation classification. Those highlighted in blue have shifted in a more fundamental level (either major life-form difference such as a shrub type to an herbaceous type, or a vegetated to a non-vegetated). Each of these signifies a more significant degree of change. Association-level changes are the most minor and tend to represent minor shifts in species composition while the major dominant species remain the same. Alliance-level changes represent a major shift in species dominance, but some of the same species are usually present. Those changes indicated by the blue highlighting indicate a fundamental shift in presence or absence of species at the life-form level and are likely to indicate major management shifts or ecological change.

Table 2. Vegetation type in resampled plots, 1999 vs. 2006

Numbers after the names are the mapping code number. Bold = shift at association level; yellow highlight = shifted at the alliance level; blue highlight = major shift, either major life-form difference such as a shrub type to an herbaceous type, or a vegetated to a non-vegetated.

Plot #	Alliance/Association 1999	Alliance/Association 2006
	Schoenoplectus (californicus and/or acutus)-	Schoenoplectus (californicus and/or acutus)-
001	Typha sp.	Typha sp. Association 157
	Schoenoplectus maritimus/Salicornia	Typha angustifolia-latifolia-domingensis
002	virginica	/Distichlis association 126

Plot #	Alliance/Association 1999	Alliance/Association 2006
		Typha angustifolia-latifolia-domingensis
003	Typha species (generic)	/Distichlis association 126
004	Salix laevigata/S. lasiolepis association	Salix laevigata/S. lasiolepis association 702
	Schoenoplectus californicus/S. acutus	Schoenoplectus californicus/S. acutus
009	association 116	association 116
010	Phragmites australis association 103	Phragmites australis association 103
011	Phragmites australis association 103	Water
012	Salix laevigata/S. lasiolepis association 702	Salix laevigata/S. lasiolepis association 702
	Centaurea solstitialis alliance (generic)	Vulpia sp. /Euthamia occidentalis association
013		
014	Salicornia/Distichlis 351	Typha species (generic) 123
015	Salicornia virginica association 346	Salicornia virginica association 346
016	Salicornia/Polypogon 355	Salicornia virginica association 346
000	Cohoononlaatus maritimus allianas 197	Schoenopiectus maritimus/Salicornia
020	Schoenoplectus maritimus alliance 137	virginica association (138)
021	schoenopiecius mantimus/Salicomia	Solicornia virginica accordiation 246
021		Distichlis/Scheenenlectus americanus
023	Atripley/Distichlis association 312	Disticility/Schoenopiectus americanus
023	Distichlis/A triangularis 143	Distichlis/Salicornia association 1/8
024	Lolium/Bromus 217	Lolium/Lenidium association 220
025	Distichlis/Cotula association 153	Distichlis/Salicornia association 1/8
020	Pubus discolor alliance 606	Pubus discolor alliance 606
034	Polygonum-Yanthium strumarium	
036	Association	Distichlis/Annual Grasses association 142
037	Cotula corononifolia alliance (generic) 342	Cotula coronopifolia alliance (generic) 342
001		Schoenoplectus maritimus/Salicornia virginica
038	Atriplex/S, maritimus association 315	association 138
039	Salicornia/Atriplex association 348	Salicornia/Sesuvium 356
040	Salicornia/Sesuvium 356	Salicornia virginica association 346
042	Distichlis/Glaux 144	Distichlis/Salicornia association 148
		Juncus balticus/Potentilla anserina
043	Distichlis/Juncus association 145	association 135
044	Rubus discolor alliance 606	Rubus discolor alliance 606
		Schoenoplectus (californicus and/or acutus)-
045	Typha species (generic) 123	Typha sp. association 157
046	Distichlis/Salicornia association 148	Distichlis/Salicornia association 148
047	Distichlis/Juncus association 145	Salicornia virginica Alliance
048	Schoenoplectus americanus (generic) 114	Schoenoplectus americanus alliance
		Typha angustifolia-latifolia-domingensis /S.
049	Typha species (generic) 123	americanus 121
050	Sesuvium verrucosum 357	Sesuvium verrucosum association 357
	Schoenoplectus californicus/S. acutus	Schoenoplectus californicus/S. acutus
051	association 116	association 116
052	Baccharis/Annual Grasses 603	Baccharis/Annual Grasses 603
053	Salicornia/Distichlis 351	Salicornia/Distichlis 351
054	Rosa/Baccharis association 605	Rosa/Baccharis association 605
056	Distichlis/Lactuca 146	Distichlis/Annual Grasses association 142
058	Bromus spp. /Hordeum spp. association 232	Distichlis spicata/Annual Grasses association 142
059	Distichlis/Salicornia association 148	Distichlis/Salicornia association 148
060	Lepidium/Distichlis stands 323	Lepidium/Distichlis stands 323
061	Distichlis/Lotus association	Distichlis/Salicornia association 148
062	Leymus triticoides alliance (generic) 215	Leymus triticoides alliance (generic) 215
063	Schoenoplectus americanus (generic) 114	Schoenoplectus americanus (generic) 114
		Distichlis-Juncus-Triglochin-Glaux
064	Distichlis/Salicornia association 148	association 160

Plot #	Alliance/Association 1999	Alliance/Association 2006
065	Leymus triticoides alliance (generic) 215	Leymus triticoides alliance (generic) 215
		Typha angustifolia-latifolia-domingensis
067	Typha species (generic) 123	alliance
		Distichlis spicata/Annual Grasses association
068	Salicornia/Lolium 354	
070	Lolium/Salicornia 224	Hordeum/Lolium association 234
074	Distichlis (Iurous accessistion 115	I ypna angustitolia-latitolia-domingensis
071	Distichils/Juncus association 145	Alliance Schoononloctus amoriconus alliance
072	Schoenopiecius americanus (generic) 114	Typha angustifelia latifelia demingensis /S
		americanus 121 or just Schoenoplectus
073	Distichlis/Juncus association 145	americanus alliance
		Frankenia salina Alliance or Lolium
074	Frankenia/Rumex crispus 319	multiflorum alliance
075	Frankenia/Distichlis 318	Distichlis/Annual Grasses association 142
076	Lolium/Rumex association 222	Lotus corniculatus alliance 344
077	Lolium (generic) 218	Salicornia virginica association 346
078	Grindelia stricta var. stricta stands 321	Lepidium latifolium alliance
079	Salicornia/Polypogon 355	Salicornia virginica association 346
	Distichlis/Schoenoplectus americanus	
083	association 149	Distichlis/Cotula association 153
086	Salicornia/Distichlis 351	Salicornia virginica association 346
097	Ananymum (Schoonenlacture, 202	Schoenopiectus (californicus or acutus)/Rosa
088	Schoonoplectus amoricanus (generic) 114	102 Typha spacies (generic) 122
089	Selicornia/Polynogon 355	Salicornia/Annual Grasses 347
000	Salicornia/Atriplex association 348	Distichlis spicata (generic) 156
091	Lolium/Bromus 217	Lolium (generic) 218
092	Salicornia/Atriplex association 348	Salicornia virginica association 346
099	Lepidium/Distichlis stands 323	Lepidium/Distichlis stands 323
107	Salicornia/Distichlis 351	Salicornia virginica association 346
108	Leymus triticoides alliance (generic) 215	Leymus triticoides alliance (generic) 215
110	Salicornia/Distichlis 351	Salicornia virginica association 346
111	Distichlis/Salicornia association 148	Distichlis/Salicornia association 148
118	Atriplex lentiformis (generic) 514	Atriplex lentiformis (generic) 514
		Distichlis spicata/Annual Grasses
136	Distichlis/Lotus association	association 142
145	Schoenopiectus californicus/S. acutus	Schoenopiectus californicus/S. acutus
145		Distichlis spicata/Appual Grasses association
147	Lolium (generic) 218	142
148	Lolium (generic) 218	Lolium (generic) 218
154	Atriplex/Distichlis association 312	Distichlis/Salicornia association 148
159	Atriplex triangularis association 311	Atriplex/Annual Grasses stands 337
	Typha angustifolia-latifolia-domingensis	Typha angustifolia-latifolia-domingensis
161	/Echinochloa-Polygonum-Xanthium	/Echinochloa-Polygonum-Xanthium 120
100		Typha angustifolia-latifolia-domingensis
168	Xanthium/Polypogon 332	/Phragmites australis 129
160	Distichlis spicata 141	Disticnlis spicata/Annual Grasses
170	Schoenoplectus maritimus/Secuvium 120	association 142 Phragmites/Schoenoplectus association 104
171	Frankenia (generic) 320	Bromus spp /Hordeum spp association 232
172	Sesuvium verrucosum 357	Sesuvium verrucosum association 357
175	Phragmites/Xanthium 104	Echinochloa-Polygonum-Xanthium 159
	Juncus balticus/Conium maculatum	Juncus balticus/Conium maculatum
176	association 133	association 133

Plot #	Alliance/Association 1999	Alliance/Association 2006
	Juncus balticus/Conium maculatum	Conium maculatum 402 or Juncus
177	association 133	balticus/Conium maculatum association 133
180	Bare ground 001	Salicornia virginica association 346
181	Rosa/Baccharis association 605	Rosa/Baccharis association 605
	Schoenoplectus californicus/S. acutus	Schoenoplectus (californicus and/or acutus)-
184	association 116	Typha sp. association 157
185	Atriplex triangularis association 311	Elytrigia pontica stands 211
186	Xanthium/Polypogon 332	Lotus corniculatus
189	Salicornia/Sesuvium 356	Salicornia virginica association 346
190	Bare ground 001	Open water
192	Frankenia/Agrostis 317	Frankenia salina Alliance
		Polypogon monspeliensis stands (generic)
194	Juncus balticus association 132	238
195	Rosa californica alliance 604	Rosa californica alliance 604
	Schoenoplectus (californicus and/or acutus)-	Schoenoplectus (californicus and/or
197	Typha sp. association	acutus)/Wetland Herbs 158

<u>Summary of Vegetation Type Changes.</u> The vegetation type of 38 plots remained unchanged. Twenty-six plots shifted at the association level, while 25 shifted at the alliance level, and 9 shifted at the fundamental life form level. Figure 2 depicts these changes graphically.

Note that changes from "Typha spp. Generic" to "Typha (latifolia-angustifolia-domingensis)" (Plots 003, 049 and 067) do not constitute a true change but a shift in name for the same vegetation type.

Figure 2. Vegetation Type Changes



Species Dominance Changes

Table 3 displays the 98 resampled plots compared to their 1999 original data with regard to the species with the highest cover estimates in each sample. This comparison addresses the major floristic similarities between the two sampling efforts. In the case where less than three species exist in the plot, missing species are indicated by "none." *Phragmites australis* is highlighted because even though it is treated as a native, technically the race that is predominant in the marsh is non-native and is thus important to keep track of for management purposes.

Table 3. The top species by % cover in 1999 versus 2006

Only those plots that were resampled in 2006 are listed. Those species with percent cover highlighted red indicate that there are other species within that plot with the same percent cover; the highlighted red ones were chosen for this table based on alphabetical order. Yellow highlight indicates *Phragmites australis* cover where it was recorded solely or, if in both the 1999 and 2006 paired sample, whichever plot had the higher cover.

Plot #	Percent Cover 1999	Top Species 1999	Percent Cover 2006	Top Species 2006
1	57	Schoenoplectus californicus	35	Schoenoplectus acutus var. occidentalis
1	28	Typha angustifolia	15	Schoenoplectus californicus
1	10	Schoenoplectus acutus	8	Typha angustifolia
2	85	Schoenoplectus maritimus	55	Typha angustifolia
2	3	Salicornia virginica	2	Schoenoplectus maritimus
2	2	Typha angustifolia	0.2	Distichlis spicata
3	83	Typha angustifolia	50	Typha angustifolia
3	0.2	Atriplex triangularis	12	Distichlis spicata
3	0	None	8	Schoenoplectus maritimus
4	56	Salix laevigata	40	Leymus triticoides
4	41	Leymus triticoides	25	Salix gooddingii
4	16	Salix lasiolepis	10	Salix lasiolepis
9	70	Schoenoplectus californicus	40	Schoenoplectus californicus
9	30	Spartina foliosa	5	Spartina foliosa
10	<mark>100</mark>	Phragmites australis	95	Phragmites australis
10	0.2	Lepidium latifolium	0.2	Scirpus acutus var. occidentalis
11	<mark>95</mark>	Phragmites australis	0	None
12	60	Salix laevigata	55	Salix laevigata
12	13	Cynodon dactylon	40	Salix lasiolepis
12	10	Salix lasiolepis	6	Schoenoplectus americanus
13	70	Centaurea solstitialis	30	Vulpia myuros
13	20	Distichlis spicata	22	Centaurea solstitialis
13	12	Bromus hordeaceus	8	Lolium multiflorum
14	80	Salicornia virginica	30	Typha latifolia
14	6	Distichlis spicata	2	Salicornia virginica
14	3	Schoenoplectus maritimus	1	Schoenoplectus maritimus
15	91	Salicornia virginica	65	Salicornia virginica
15	4	Frankenia salina	3	Frankenia salina
15	1	Polypogon monspeliensis	8	Distichlis spicata
16	90	Salicornia virginica	60	Salicornia virginica
16	6	Polypogon monspeliensis	2	Atriplex triangularis
16	2	Distichlis spicata	4	Distichlis spicata
20	87	Elytrigia pontica	27	Salicornia virginica
20	5	Lolium multiflorum	2	Schoenoplectus maritimus
20	2	Hordeum marinum	0.2	Typha angustifolia
21	75	Schoenoplectus maritimus	60	Salicornia virginica
21	25	Salicornia virginica	3	Sesuvium verrucosum
21	0.2	Cotula coronopifolia	0.2	Atriplex triangularis
23	35	Atriplex triangularis	28	Distichlis spicata
23	32	Distichlis spicata	24	Schoenoplectus americanus
23	12	Aster subulatus var. cubensis	20	Phragmites australis
24	87	Distichlis spicata	45	Distichlis spicata

Plot #	Percent Cover 1999	Ton Species 1999	Percent Cover 2006	Top Species 2006
24	3	Atriplex triangularis	20	Atriplex triangularis
24	3	Salicornia virginica	15	Salicornia virginica
25	45	Lolium multiflorum	60	Lolium multiflorum
25	16	Vulpia mvuros	10	Bromus diandrus
25	7	Vicia sativa	7	Raphanus sativus
26	30	Distichlis spicata	60	Distichlis spicata
26	16	Cotula coronopifolia	25	Salicornia virginica
26	15	Schoenoplectus maritimus	0.2	Atriplex triangularis
34	100	Rubus discolor	85	Rubus discolor
34	0.2	Bromus diandrus	10	Raphanus sativus
34	0.2	Distichlis spicata	1	Carduus pvcnocephalus
36	52	Polvaonum aravrocoleon	26	Distichlis spicata
36	15	Rumex conclomeratus	17	Sonchus oleraceus
36	9	Cotula coronopifolia	6	Polypogon monspeliensis
37	48	Rumex conclomeratus	33	Phragmites australis
37	45	Cotula coronopifolia	26	Cotula coronopifolia
37	20	Polvaonum aravrocoleon	16	Polypogon monspeliensis
38	82	Atriplex triangularis	32	Schoenoplectus maritimus
38	25	Schoenoplectus maritimus	18	Polypogon monspeliensis
38	3	Polypogon monspeliensis	10	Salicornia virginica
39	76	Salicornia virginica	15	Sesuvium verrucosum
39	4	Cotula coronopifolia	10	Salicornia virginica
39	7	Atriplex triangularis	7	Atriplex triangularis
40	70	Salicornia virginica	1	Salicornia virginica
40	6	Sesuvium verrucosum	0.2	Sesuvium verrucosum
42	48	Distichlis spicata	30	Salicornia virginica
42	40	Glaux maritima	25	Glaux maritima
42	20	Grindelia stricta	20	Grindelia stricta
43	75	Distichlis spicata	42	Juncus balticus
43	40	Juncus balticus	25	Distichlis spicata
43	12	Potentilla anserina	20	Grindelia stricta
44	98	Rubus discolor	75	Rubus discolor
44	0.2	Typha angustifolia	6	Distichlis spicata
44	0.2	Typha latifolia	0.2	Bromus diandrus
45	55	Typha angustifolia	70	Typha angustifolia
45	40	Schoenoplectus acutus	25	Schoenoplectus acutus
45	1	Schoenoplectus californicus	0.2	Distichlis spicata
46	64	Distichlis spicata	40	Distichlis spicata
46	26	Triglochin maritimum	25	Salicornia virginica
46	8	Jaumea carnosa	15	Jaumea carnosa
47	38	Juncus balticus	30	Salicornia virginica
47	36	Distichlis spicata	18	Juncus balticus
47	28	Glaux maritima	14	Glaux maritima
48	76	Schoenoplectus americanus	65	Schoenoplectus americanus
48	55	Calystegia sepium	30	Calystegia sepium
48	30	Euthamia occidentalis	25	Euthamia occidentalis
49	92	Typha angustifolia	55	Typha angustifolia
49	6	Schoenoplectus americanus	20	Schoenoplectus americanus

Plot #	Percent Cover 1999	Top Species 1999	Percent Cover 2006	Top Species 2006
49	1	Schoenoplectus californicus	13	Typha latifolia
50	98	Sesuvium verrucosum	30	Sesuvium verrucosum
50	1	Atriplex triangularis	12	Schoenoplectus maritimus
50	0.2	Schoenoplectus maritimus	12	Digitaria sanguinalis
51	80	Schoenoplectus californicus	98	Schoenoplectus acutus
51	0	None	1	Schoenoplectus americanus
52	60	Hordeum marinum	26	Baccharis pilularis
52	55	Baccharis pilularis	22	Hordeum marinum
52	14	Bromus diandrus	11	Avena barbata
53	65	Salicornia virginica	70	Salicornia virginica
53	19	Distichlis spicata	20	Distichlis spicata
53	5	Atriplex triangularis	2	Cotula coronopifolia
54	80	Rosa californica	70	Rosa californica
54	9	Baccharis pilularis	25	Baccharis pilularis
54	6	Schoenoplectus californicus	7	Lolium multiflorum
56	85	Distichlis spicata	50	Distichlis spicata
56	27	Lactuca serriola	25	Bromus diandrus
56	14	Frankenia salina	20	Frankenia salina
58	30	Hordeum marinum	48	Bromus diandrus
58	28	Bromus hordeaceus	26	Distichlis spicata
58	24	Bromus diandrus	5	Lactuca serriola
59	85	Distichlis spicata	55	Distichlis spicata
59	20	Salicornia virginica	50	Salicornia virginica
59	13	Cuscuta salina var. major	5	Triglochin maritimum
60	62	Lepidium latifolium	60	Lepidium latifolium
60	18	Distichlis spicata	15	Distichlis spicata
60	6	Schoenoplectus americanus	10	Salicornia virginica
61	49	Distichlis spicata	52	Distichlis spicata
61	28	Lotus corniculatus	25	Salicornia virginica
61	10	Jaumea carnosa	20	Lotus corniculatus
62	95	Leymus triticoides	52	Leymus triticoides
62	1	Bromus hordeaceus	14	Lactuca serriola
62	1	Lepidium latifolium	6	Bromus hordeaceus
63	88	Scirpus americanus	50	Schoenoplectus americanus
63	2	Atriplex triangularis	11	Juncus balticus
63	2	Frankenia salina	8	Typha latifolia
64	57	Distichlis spicata	35	Distichlis spicata
64	16	Juncus balticus	30	Jaumea carnosa
64	13	Triglochin maritimum	25	Salicornia virginica
65	85	Leymus triticoides	53	Leymus triticoides
65	22	Distichlis spicata	25	Distichlis spicata
65	7	Bromus hordeaceus	7	Bromus hordeaceus
67	80	Typha latifolia	85	Typha latifolia
67	20	Juncus balticus	2	Schoenoplectus americanus
67	10	Atriplex triangularis	0.2	Atriplex triangularis
68	48	Salicornia virginica	50	Distichlis spicata
68	30	Lolium multiflorum	20	Lepidium latifolium
68	17	Frankenia salina	10	Lolium multiflorum

Plot #	Percent Cover 1999	Top Species 1999	Percent Cover 2006	Top Species 2006
70	68	Lolium multiflorum	65	Lolium multiflorum
70	13	Frankenia salina	25	Hordeum marinum
70	10	Atriplex triangularis	20	Frankenia salina
71	71	Distichlis spicata	76	Typha latifolia
71	10	Juncus balticus	2	Atriplex triangularis
71	7	Typha	0.2	Distichlis spicata
72	80	Schoenoplectus americanus	75	Schoenoplectus americanus
72	7	Atriplex triangularis	2	Salicornia virginica
72	1	Salicornia virginica	0.2	Atriplex triangularis
73	60	Distichlis spicata	50	Schoenoplectus americanus
73	16	Juncus balticus	18	Typha latifolia
73	7	Sonchus oleraceus	18	Atriplex triangularis
74	40	Frankenia salina	60	Lolium multiflorum
74	25	Rumex crispus	40	Frankenia salina
74	20	Lolium multiflorum	4	Hordeum marinum
75	51	Frankenia salina	30	Distichlis spicata
75	45	Distichlis spicata	25	Atriplex triangularis
75	10	Atriplex triangularis	15	Frankenia salina
76	70	Rumex crispus	45	Lotus corniculatus
76	20	Lolium multiflorum	20	Lepidium latifolium
76	5	Distichlis spicata	12	Lolium multiflorum
77	45	Hordeum marinum	45	Salicornia virginica
77	40	Lolium multiflorum	10	Frankenia salina
77	12	Frankenia salina	2	Lolium multiflorum
78	77	Grindelia stricta var. angustifolia	70	Lepidium latifolium
78	10	Lolium multiflorum	35	Grindelia stricta
78	5	Lotus corniculatus	30	Frankenia salina
79	90	Salicornia virginica	70	Salicornia virginica
79	1	Polypogon monspeliensis	1	Polypogon monspeliensis
79	0.2	Hordeum marinum	0	None
83	50	Distichlis spicata	60	Distichlis spicata
83	35	Schoenoplectus americanus	17	Cotula coronopifolia
83	5	Juncus balticus	8	Polypogon monspeliensis
86	53	Salicornia virginica	60	Salicornia virginica
86	16	Distichlis spicata	20	Distichlis spicata
86	12	Polypogon monspeliensis	10	Picris echioides
87	35	Apocynum cannabinum	45	Schoenoplectus acutus
87	20	Schoenoplectus acutus	30	Rosa californica
87	18	Rosa californica	15	Apocynum cannabinum
88	60	Schoenoplectus americanus	85	Typha latifolia
88	35	Typha angustifolia	4	Schoenoplectus americanus
88	6	Polypogon monspeliensis	2	Schoenoplectus acutus
89	60	Salicornia virginica	51	Salicornia virginica
89	15	Polypogon monspeliensis	18	Poaceae
89	10	Rumex crispus	16	Salicornia virginica
90	85	Salicornia virginica	90	Distichlis spicata
90	7	Atriplex triangularis	4	Lepidium latifolium
90	1	Lepidium latifolium	7	Phragmites australis

91 75 Lolum multiflorum 68 Lolum mainum 91 7 Bromus hordeacus 16 Frankenia salima 91 4 gussonienum 10 Hordeum mainum 92 85 Salicomia virginica 85 Salicomia virginica 92 14 Atriplex triangularis 2 Polypogon monspeliensis 92 2 Rumex pulcher 3 Rumex dentatus 99 78 Lepidum latifulum 40 Distichlis spicata 99 78 Lepidum latifulum 40 Distichlis spicata 107 48 Salicomia virginica 54 Salicomia virginica 107 48 Salicomia virginica 54 Distichlis spicata 107 2 Atriplex triangularis 1 Linonium californicum 108 30 Bromus britecides 40 Leymus tritecides 109 5 Bromus britecides 10 Bronus dandrus 110 89 Salicomia virginica 75 Salicomia virginica 110 6 Cuscuta salina var. major 6 Cuscuta salina 110 2 Distichlis spicata 49 Salicomia virginica 111 <t< th=""><th>Plot #</th><th>Percent Cover 1999</th><th>Top Species 1999</th><th>Percent Cover 2006</th><th>Top Species 2006</th></t<>	Plot #	Percent Cover 1999	Top Species 1999	Percent Cover 2006	Top Species 2006
91 7 Bromus hordesceus 16 Frankenia salina 91 4 gussoniarum msp. 10 Hordeum mainum 92 85 Salicomia virginica 85 Salicomia virginica 92 14 Artiplex triangularis 2 Polyogon monspeliensis 93 2 Rumex pelcher 3 Rumex dentstus 99 53 Distichits spicata 40 Lepidum tatifolum 99 4 Salicomia virginica 3 Cotula connopifola 107 48 Salicomia virginica 54 Salicomia virginica 108 30 Bromus diandrus 11 Limonium californicum 108 45 Leymus triticoides 40 Leymus triticoides 108 30 Bromus diandrus 12 Teanisherum caput-medusee 110 6 Cusuta salina 2 Hainardia cylindrica 110 12 Distichits spicata 2 Cusuta salina 110 12 Distichits spicata <t< td=""><td>91</td><td>75</td><td>Lolium multiflorum</td><td>68</td><td>Lolium multiflorum</td></t<>	91	75	Lolium multiflorum	68	Lolium multiflorum
Hordeum marinum ssp. 10 Hordeum marinum 92 85 Saliconia virginica 85 Saliconia virginica 92 14 Atriplex triangularis 2 Polypogon monspellensis 93 78 Lepidium latifolium 40 Lepidium latifolium 99 53 Distrchis spicata 40 Distrchis spicata 107 43 Saliconia virginica 5 Distrchis spicata 107 10 Distrchis spicata 5 Distrchis spicata 107 2 Atriplex triangularis 1 Limonium calitonicum 108 45 Leyrus tritocides 40 Leyrus tritocides 108 30 Bronus diandrus 12 Taeinatherum caput-medusae 110 8 Saliconia virginica 75 Saliconia virginica 110 6 Cuscuta salina var. major 6 Cuscuta salina 110 2 Distrchis spicata 49 Distrchis spicata 111 20 Saliconia virginica 49 <td>91</td> <td>7</td> <td>Bromus hordeaceus</td> <td>16</td> <td>Frankenia salina</td>	91	7	Bromus hordeaceus	16	Frankenia salina
9 85 Salaconia virginica 85 Salaconia virginica 92 14 Atriplex triangularis 2 Polypogon monspeliensis 92 2 Rumex pulcher 3 Rumex dentatus 99 78 Legoldium taltifolum 40 Distichlis spicata 99 4 Salaconia virginica 3 Cotula cononptifola 107 48 Salaconia virginica 54 Salaconia virginica 107 10 Distichlis spicata 5 Distichlis spicata 107 2 Atriplex triangularis 1 Linonium californicum 108 30 Bromus diandrus 12 Tensitherium caput-medusae 108 30 Bromus diandrus 12 Tensitherium caput-medusae 110 6 Cuscuta salina vat. major 6 Cuscuta salina 110 6 Cuscuta salina 49 Salicornia virginica 111 70 Distichlis spicata 24 Ocium maculatum 111 20 Sa	01	4	Hordeum marinum ssp.	10	Hordoum marinum
32 33 Salicohia viginica 33 Salicohia viginica 32 14 Atriplex triangularis 2 Polyogon monspelensis 32 2 Rumex pulcher 3 Rumex dentatus 99 78 Lopidum latfolum 40 Lopidum latfolum 99 53 Distichis spicata 40 Distichis spicata 107 48 Salicornia virginica 5 Distichis spicata 107 10 Distichis spicata 5 Distichis spicata 108 45 Leyrnus triticoides 40 Leyrnus triticoides 108 45 Leyrnus triticoides 10 Baronus diandrus 110 89 Salicornia virginica 75 Salicornia virginica 110 2 Distichis spicata 2 Hairardia cylindrica 111 70 Distichis spicata 49 Salicornia virginica 111 20 Salicornia virginica 33 Atripkex lentiformis 1111 20 Salicornia virgi	91	95	Solicornia virginico	95	
32 2 Rulps A language 2 Pubplegram for special 99 2 Rumex pulcher 3 Rumex dentatus 99 78 Lepidium latifolium 40 Lepidium latifolium 99 53 Distichis spicata 40 Distichis spicata 99 4 Salicomia virginica 3 Cotula connegificia 107 48 Salicomia virginica 54 Salicomia virginica 107 2 Atriptex triangularis 1 Limonium californicum 108 45 Leyrus triticoides 40 Leyrus triticoides 108 45 Leyrus triticoides 10 Bronus diandrus 110 89 Salicomia virginica 75 Salicomia virginica 110 6 Cuscuta salina var. major 6 Cuscuta salina 1111 70 Distichis spicata 49 Salicomia virginica 1111 20 Salicomia virginica 33 Atripkex halformis 1111 20 Salicomia vi	92	00		00	Balupagan mananalianaia
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107 2 Atriplex triangularis 1 Limonium californicum 108 45 Leymus triticoides 40 Leymus triticoides 108 30 Bronus diandrus 12 Taeniatherum caput-medusae 108 15 Bronus hordeaceus 10 Bronus diandrus 110 89 Salicornia virginica 75 Salicornia virginica 110 6 Cuscuta salina var. major 6 Cuscuta salina 110 2 Distichil's spicata 49 Salicornia virginica 111 70 Distichil's spicata 49 Distichil's spicata 111 20 Salicornia virginica 49 Distichil's spicata 111 20 Conium maculatum 3 Atriplex triangularis 118	107	10	Distichlis spicata	5	Distichlis spicata
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108 15 Bromus hordeaceus 10 Bromus diandrus 110 89 Salicornia virginica 75 Salicornia virginica 110 6 Cuscuta salina var. major 6 Cuscuta salina 110 2 Distichiis spicata 2 Hainardia cylindrica 111 70 Distichiis spicata 49 Salicornia virginica 111 20 Salicornia virginica 49 Distichiis spicata 111 13 Cordylanthus mollis sp. mollis 2 Cuscuta salina 118 28 Atriplex lentiformis 33 Atriplex lentiformis 118 28 Atriplex tringularis 22 Conium maculatum 136 76 Distichiis spicata 70 Lolium multiflorum 136 8 Lolium multiflorum 22 Distichiis spicata 136 6 Lotus corniculatus 10 Frankenia salina 145 98 Schoenoplectus californicus 93 Schoenoplectus californicus 145 1 Atriplex tringularis 7 Distichiis spicata 145 0.2 Juncus balitcus 0.2 Agrostis avenacea 147 44 Lolium multiflorum 25	108	30	Bromus diandrus	12	Taeniatherum caput-medusae
110 89 Salicornia virginica 75 Salicornia virginica 110 6 Cuscuta salina var. major 6 Cuscuta salina 110 2 Distichilis spicata 2 Hainardia cylindrica 111 70 Distichilis spicata 49 Salicornia virginica 111 20 Salicornia virginica 49 Distichilis spicata 111 13 Cordylanthus mollis ssp. mollis 2 Cuscuta salina 118 28 Atriplex lentiformis 33 Atriplex lentiformis 118 28 Atriplex Intingularis 22 Conium maculatum 118 24 Conium maculatum 3 Hordeum marinum 118 24 Conium maculatum 3 Hordeum marinum 136 76 Distichilis spicata 70 Lolium multiflorum 136 8 Lolium multiflorum 22 Distichilis spicata 145 98 Schoenoplectus californicus 93 Schoenoplectus californicus 145 1 Atriplex triangularis 7 Distichilis spicata 145 0.2 Juncus balticus 0.2 Agrostis avenacea 147 44 Lolium multiflorum 25	108	15	Bromus hordeaceus	10	Bromus diandrus
1106Cuscuta salina var. major6Cuscuta salina1102Distichlis spicata2Hainardia cylindrica11170Distichlis spicata49Salicornia virginica11120Salicornia virginica49Distichlis spicata11113Cordylanthus mollis ssp. mollis2Cuscuta salina11828Atriplex lentiformis33Atriplex lentiformis11828Atriplex triangularis22Conium maculatum11824Conium maculatum3Hordeum marinum13676Distichlis spicata70Lolium multiforum1368Lolium multiforum22Distichlis spicata1366Lotus corriculatus10Frankenia salina14598Schoenoplectus californicus93Schoenoplectus californicus1451Atriplex triangularis7Distichlis spicata14744Lolium multiflorum25Distichlis spicata14723Xanthium strumarium10Lolium multiflorum1482Atriplex triangularis2Bromus diandrus1480.2Lactuca seriola1Distichlis spicata15415Distichlis spicata15Distichlis spicata15415Distichlis spicata16Distichlis spicata15428Atriplex triangularis2Bromus diandrus15415Distichlis spicata15Distichli	110	89	Salicornia virginica	75	Salicornia virginica
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1366Lotus corniculatus10Frankenia salina14598Schoenoplectus californicus93Schoenoplectus californicus1451Atriplex triangularis7Distichlis spicata1450.2Juncus balticus0.2Agrostis avenacea14744Lolium multiflorum25Distichlis spicata14735Agrostis avenacea15Polypogon monspeliensis14723Xanthium strumarium10Lolium multiflorum14896Lolium multiflorum97Lolium multiflorum1482Atriplex triangularis2Bromus diandrus1480.2Lactuca serriola1Distichlis spicata15415Distichlis spicata15Distichlis spicata15415Distichlis spicata15Distichlis spicata15415Distichlis spicata15Distichlis spicata15997Atriplex triangularis35Atriplex triangularis1591Polypogon monspeliensis30Phragmites australis1591Polypogon monspeliensis30Phragmites australis1591Phragmites australis5Polygonum16150Typha angustifolia40Xanthium strumarium	136	8	Lolium multiflorum	22	Distichlis spicata
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1450.2Juncus balticus0.2Agrostis avenacea14744Lolium multiflorum25Distichlis spicata14735Agrostis avenacea15Polypogon monspeliensis14723Xanthium strumarium10Lolium multiflorum14896Lolium multiflorum97Lolium multiflorum1482Atriplex triangularis2Bromus diandrus1480.2Lactuca serriola1Distichlis spicata15428Atriplex triangularis20Salicornia virginica15415Distichlis spicata15Distichlis spicata1548Schoenoplectus maritimus12Polypogon monspeliensis1591Polypogon monspeliensis30Phragmites australis1591Phragmites australis5Polygonum16150Typha angustifolia40Xanthium strumarium	145	1	Atriplex triangularis	7	Distichlis spicata
14744Lolium multiflorum25Distichlis spicata14735Agrostis avenacea15Polypogon monspeliensis14723Xanthium strumarium10Lolium multiflorum14896Lolium multiflorum97Lolium multiflorum1482Atriplex triangularis2Bromus diandrus1480.2Lactuca serriola1Distichlis spicata15428Atriplex triangularis20Salicornia virginica15415Distichlis spicata15Distichlis spicata1548Schoenoplectus maritimus12Polypogon monspeliensis15997Atriplex triangularis35Atriplex triangularis1591Polypogon monspeliensis30Phragmites australis1591Phragmites australis5Polygonum16150Typha angustifolia40Xanthium strumarium	145	0.2	Juncus balticus	0.2	Agrostis avenacea
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1482Atriplex triangularis2Bromus diandrus1480.2Lactuca serriola1Distichlis spicata15428Atriplex triangularis20Salicornia virginica15415Distichlis spicata15Distichlis spicata15415Schoenoplectus maritimus12Polypogon monspeliensis15997Atriplex triangularis35Atriplex triangularis1591Polypogon monspeliensis30Phragmites australis1591Phragmites australis5Polygonum16150Typha angustifolia40Xanthium strumarium	148	96	Lolium multiflorum	97	Lolium multiflorum
1480.2Lactuca serriola1Distichlis spicata15428Atriplex triangularis20Salicornia virginica15415Distichlis spicata15Distichlis spicata1548Schoenoplectus maritimus12Polypogon monspeliensis15997Atriplex triangularis35Atriplex triangularis1591Polypogon monspeliensis30Phragmites australis1591Phragmites australis5Polygonum16150Typha angustifolia40Xanthium strumarium	148	2	Atriplex triangularis	2	Bromus diandrus
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1548Schoenoplectus maritimus12Polypogon monspeliensis15997Atriplex triangularis35Atriplex triangularis1591Polypogon monspeliensis30Phragmites australis1591Phragmites australis5Polygonum16150Typha angustifolia40Xanthium strumarium	154	15	Distichlis spicata	15	Distichlis spicata
15997Atriplex triangularis35Atriplex triangularis1591Polypogon monspeliensis30Phragmites australis1591Phragmites australis5Polygonum16150Typha angustifolia40Xanthium strumarium	154	8	Schoenoplectus maritimus	12	Polypogon monspeliensis
1591Polypogon monspeliensis30Phragmites australis1591Phragmites australis5Polygonum16150Typha angustifolia40Xanthium strumarium	159	97	Atriplex triangularis	35	Atriplex triangularis
1591Phragmites australis5Polygonum16150Typha angustifolia40Xanthium strumarium	159	1	Polypogon monspeliensis	30	Phragmites australis
161 50 Typha angustifolia 40 Xanthium strumarium	159	1	Phragmites australis	5	Polygonum
	161	50	Typha angustifolia	40	Xanthium strumarium
161 30 Polygonum lapathifolium 30 Typha angustifolia	161	30	Polygonum lapathifolium	30	Typha angustifolia
161 30 Echinochloa crus-galli 16 Atriplex triangularis	161	30	Echinochloa crus-oalli	16	Atriplex triangularis
168 52 Polypogon monspeliensis 50 Phragmites australis	168	52	Polypogon monspeliensis	50	Phragmites australis
168 28 Xanthium strumarium 25 Typha angustifolia	168	28	Xanthium strumarium	25	Typha angustifolia

Plot #	Percent Cover 1999	Top Species 1999	Percent Cover 2006	Top Species 2006
168	10	Lotus corniculatus	3	Juncus balticus
169	93	Distichlis spicata	51	Lolium multiflorum
169	1	Lolium multiflorum	20	Bromus diandrus
169	1	Salicornia virginica	20	Distichlis spicata
170	62	Schoenoplectus maritimus	20	Phragmites australis
170	12	Sesuvium verrucosum	14	Rumex conglomeratus
170	7	Polygonum argyrocoleon	10	Schoenoplectus maritimus
171	85	Frankenia salina	56	Bromus diandrus
171	27	Lolium multiflorum	35	Lolium multiflorum
171	1	Juncus balticus	3	Frankenia salina
172	100	Sesuvium verrucosum	70	Sesuvium verrucosum
172	1	Schoenoplectus maritimus	14	Polygonum lapathifolium
172	0	None	2	Atriplex triangularis
175	35	Xanthium strumarium	70	Xanthium strumarium
175	<mark>30</mark>	Phragmites australis	12	Polygonum lapathifolium
175	27	Polygonum lapathifolium	9	Phragmites australis
176	34	Juncus balticus	42	Conium maculatum
176	33	Conium maculatum	40	Juncus balticus
176	8	Centaurea solstitialis	5	Centaurea solstitialis
177	80	Juncus balticus	70	Juncus balticus
177	48	Conium maculatum	53	Conium maculatum
177	7	Raphanus sativus	2	Sonchus oleraceus
180	0.2	Sesuvium verrucosum	0.2	Salicornia virginica
180	0.2	Salicornia virginica	0	None
181	60	Baccharis pilularis	60	Baccharis pilularis
181	20	Rosa californica	50	Rosa californica
181	4	Euthamia occidentalis	2	Calystegia sepium
184	50	Schoenoplectus californicus	40	Typha angustifolia
184	12	Schoenoplectus americanus	30	Schoenoplectus acutus
184	7	Typha angustifolia	15	Schoenoplectus americanus
185	88	Atriplex triangularis	90	Elytrigia pontica
185	0.2	Lactuca serriola	3	Conium maculatum
185	1	Elytrigia pontica	3	Brassica nigra
186	80	Xanthium strumarium	51	Xanthium strumarium
186	7	Cotula coronopifolia	25	Cotula coronopifolia
186	5	Polypogon monspeliensis	10	Salicornia virginica
189	33	Salicornia virginica	65	Salicornia virginica
189	1	Cotula coronopifolia	1	Schoenoplectus maritimus
189	6	Sesuvium verrucosum	0.2	Atriplex triangularis
190	1	Echinochloa crus-galli	0	None
190	0.2	Sesuvium verrucosum	0	None
190	0.2	Cotula coronopifolia	0	None
192	92	Frankenia salina	30	Frankenia salina
192	35	Agrostis avenacea	20	Bromus diandrus
192	7	Distichlis spicata	16	Distichlis spicata
194	92	Juncus balticus	85	Polypogon monspeliensis
194	12	Salicornia virginica	1	Rumex conglomeratus
194	1	Frankenia salina	3	Cotula coronopifolia

Plot #	Percent Cover 1999	Top Species 1999	Percent Cover 2006	Top Species 2006
195	76	Rosa californica	49	Rosa californica
195	7	Frankenia salina	37	Poa
195	4	Lepidium latifolium	12	Frankenia salina
197	70	Schoenoplectus acutus	40	Schoenoplectus acutus
197	70	Typha angustifolia	30	Typha angustifolia
197	60	Euthamia occidentalis	15	Rubus discolor

<u>Summary of Table 3.</u> A total of 32 plots shared a single top species; 50 plots shared two top species; 9 shared 3 top species; and 7 did not share any of the 3 top species. Figure 3 represents this graphically. Note that those species shared are not necessarily in the same order of dominance.





Non-Native Species Comparison

Another way to compare the data is to focus on the non-native plants. The marsh is home to a high diversity of non-native species. These include "new natives" that have become well integrated into the California ecosystems over the past 200 years (including grasses such as Bromus spp., Avena spp., Lolium multiflorum, and Vulpia myuros) and also include more recent introductions such as Arundo donax, Lepidium latifolium, and Elytrigia spp.). These latter species are actively increasing in many parts of the state and are especially invasive in wetlands such as Suisun Marsh. Table 4 displays the top (up to 3) species of non-native species on each of the resampled plots compared to the original top (up to 3) species sampled in 1999. Lepidium latifolium, one of the most worrisome non-natives, is highlighted in yellow where it attains its highest value in the plot by plot comparison. Comparisons between cover of the partially nonnative species *Phragmites australis*, which is represented by both native and non-native forms, was not made because the native and non-native genotypes were not differentiated in the 1999 surveys. This will now be possible because field crews identified the non-native P. australis when encountered in the 2006 surveys. However, assuming that the majority of *P. australis* in Suisun Marsh is the non-native European ecotype, referring back to Table 1, there appears to be a sobering increase in the cover of this species. In 1999, 226% Phragmites cover was noted in 4 plots. In 2006, 264% cover was noted in 7 plots for a net increase of 3 plots and 58% cover. Plot

11 was the only sample where a substantial reduction in *Phragmites* was noted and this was a result of complete flooding of the plot.

Table 4. Top non-native species by percent cover for each plot in 1999 versus 2006

Only those plots that were resampled in 2006 are listed. Not all plots contained exotic species and are, therefore, not included in the table. Those species with percent cover highlighted red indicate that there are other non-native species within that plot with the same percent cover and they were chosen for this table based on alphabetical order. Presence of *Lepidium latifolium* and its highest cover in the pair-wise plot comparisons are highlighted in yellow.

F	Plot #	% Cover 1999	Top Non-Natives 1999	% Cover 2006	Top Non-natives 2006
	1	0	None	0.2	Apium graveolens
	4	7	Echinochloa crus-galli	1	Bromus diandrus
	4	2	Foeniculum vulgare	1	Malva neglecta
	4	1	Polypogon monspeliensis	1	Paspalum dilatatum
	12	13	Cynodon dactylon	4	Agrostis viridis
	12	0.2	Acacia	1	Rubus discolor
	12	0.2	Cotula coronopifolia	0.2	Hedera
	13	70	Centaurea solstitialis	30	Vulpia myuros
	13	12	Bromus hordeaceus	22	Centaurea solstitialis
	13	3	Hordeum marinum	8	Lolium multiflorum
	14	2	Cotula coronopifolia	0.2	Cotula coronopifolia
	14	0.2	Polypogon monspeliensis	0	None
	15	1	Polypogon monspeliensis	1	Polypogon monspeliensis
	15	0	None	0.2	Cotula coronopifolia
	15	0	None	0.2	Rumex pulcher
	16	6	Polypogon monspeliensis	0.2	Cotula coronopifolia
	16	1	Cotula coronopifolia	0.2	Bromus hordeaceus
	16	0.2	Rumex pulcher	0.2	Bromus diandrus
	20	87	Elytrigia pontica	0	None
	20	5	Lolium multiflorum	0	None
	20	2	Hordeum marinum	0	None
	21	0.2	Cotula coronopifolia	0.2	Crypsis schoenoides
	23	7	Lactuca serriola	2	Polypogon monspeliensis
	23	6	Cotula coronopifolia	1	Rumex conglomeratus
	23	0.2	Sonchus oleraceus	0.2	Cotula coronopifolia
	24	1	Lactuca serriola	0.2	Lepidium latifolium
	24	1	Cotula coronopifolia	0.2	Cotula coronopifolia
	24	0.2	Polypogon monspeliensis	0.2	Lactuca serriola
	25	45	Lolium multiflorum	60	Lolium multiflorum
	25	16	Vulpia myuros	10	Bromus diandrus
	25	7	Vicia sativa	7	Raphanus sativus
	26	16	Cotula coronopifolia	0.2	Cotula coronopifolia
	26	15	Polypogon monspeliensis	0	None
	26	0.2	Rumex conglomeratus	0	None
	34	100	Rubus discolor	85	Rubus discolor
	34	0.2	Bromus diandrus	10	Raphanus sativus
	34	0.2	Lepidium latifolium	1	Carduus pycnocephalus
	36	52	Polygonum argyrocoleon	17	Sonchus oleraceus
	36	15	Rumex conglomeratus	6	Polypogon monspeliensis

Plot #	% Cover 1999	Top Non-Natives 1999	% Cover 2006	Top Non-natives 2006
36	9	Cotula coronopifolia	4	Cotula coronopifolia
37	48	Rumex conglomeratus	28	Polypogon monspeliensis
37	45	Cotula coronopifolia	26	Cotula coronopifolia
37	20	Polygonum argyrocoleon	1	Chenopodium album
38	3	Polypogon monspeliensis	22	Polypogon monspeliensis
38	1	Rumex conglomeratus	1	Rumex conglomeratus
38	0.2	Cotula coronopifolia	0	None
39	4	Cotula coronopifolia	2	Polypogon monspeliensis
39	1	Rumex pulcher	1	Cotula coronopifolia
39	0.2	Polypogon monspeliensis	1	Rumex conglomeratus
42	3	Apium graveolens	<mark>3</mark>	Lepidium latifolium
42	1	Lepidium latifolium	1	Apium graveolens
42	0.2	Sonchus oleraceus	0.2	Polypogon monspeliensis
43	0.2	Lepidium latifolium	1	Lepidium latifolium
43	0.2	Cirsium	0.2	Rumex crispus
43	0.2	Lotus corniculatus	0.2	Polygonum sp.
44	98	Rubus discolor	75	Rubus discolor
44	0.2	Lepidium latifolium	0.2	Lepidium latifolium
44	0	None	0.2	Bromus diandrus
45	0	None	0.2	Lepidium latifolium
46	0	None	0.2	Lotus corniculatus
46	0	None	0.2	Apium graveolens
47	0	None	1	Apium graveolens
47	0	None	0.2	Potentilla anserina ssp. pacifica
47	0	None	0.2	Lotus corniculatus
48	0.2	Sonchus oleraceus	5	Apium graveolens
48	0	None	0.2	Polypogon monspeliensis
48	0	None	0.2	Asparagus officinalis
50	0	None	12	Digitaria sanguinalis
50	0	None	0.2	Xanthium strumarium
52	60	Hordeum marinum	22	Hordeum marinum
52	14	Bromus diandrus	11	Bromus diandrus
52	9	Hypochaeris radicata	11	Avena barbata
53	0.2	Polypogon monspeliensis	2	Cotula coronopifolia
53	0.2	Rumex crispus	1	Polypogon monspeliensis
54	3	Bromus diandrus	7	Lolium multiflorum
54	0.2	Lactuca serriola	4	Foeniculum vulgare
54	0	None	3	Cynodon dactylon
56	27	Lactuca serriola	25	Bromus diandrus
56	6	Bromus diandrus	3	Lolium multiflorum
56	0.2	Sonchus oleraceus	2	Centaurea solstitialis
58	30	Hordeum marinum	48	Bromus diandrus
58	28	Bromus hordeaceus	5	Lactuca serriola
58	24	Bromus diandrus	4	Centaurea solstitialis
59	0	None	2	Apium graveolens
59	0	None	1	Lotus corniculatus
59	0	None	0.2	Polypogon monspeliensis

Plot #	% Cover 1999	Top Non-Natives 1999	% Cover 2006	Top Non-natives 2006
60	<mark>62</mark>	Lepidium latifolium	60	Lepidium latifolium
60	0.2	Lactuca serriola	0.2	Apium graveolens
60	0	None	0.2	Polypogon monspeliensis
61	28	Lotus corniculatus	20	Lotus corniculatus
61	2	Rumex crispus	1	Polypogon monspeliensis
61	0.2	Picris echioides	0.2	Apium graveolens
62	1	Bromus hordeaceus	14	Lactuca serriola
62	1	Lepidium latifolium	6	Bromus hordeaceus
62	0.2	Rumex crispus	4	Avena
63	1	Cirsium vulgare	<mark>6</mark>	Lepidium latifolium
63	1	Polygonum argyrocoleon	1	Picris echioides
63	1	Picris echioides	0.2	Apium graveolens
65	7	Bromus hordeaceus	7	Bromus hordeaceus
65	5	Convolvulus arvensis	5	Bromus diandrus
65	1	Bromus diandrus	3	Convolvulus arvensis
68	30	Lolium multiflorum	20	Lepidium latifolium
68	1	Rumex crispus	15	Lolium multiflorum
68	0.2	Lepidium latifolium	5	Rumex crispus
70	68	Lolium multiflorum	65	Lolium multiflorum
70	5	Hordeum marinum ssp. gussonianum	25	Hordeum marinum
70	3	Bromus hordeaceus	2	Rumex crispus
71	5		0	None
71	0.2	Sonchus oleraceus	0	None
71	0.2	Rumex crispus	0	None
73	7	Sonchus oleraceus	2	Sonchus oleraceus
73	5	Lactuca serriola	0.2	Rumex crispus
73	0	None	0	None
74	25	Rumex crispus	60	Lolium multiflorum
74	20	Lolium multiflorum	4	Hordeum marinum
74	3	Bromus diandrus	3	Rumex crispus
75	0.2	Lactuca serriola	15	Lolium multiflorum
75	0.2	Sonchus oleraceus	6	Lactuca serriola
75	0.2	Polygonum arenastrum	1	Polygonum baldschuanicum
76	70	Rumex crispus	45	Lotus corniculatus
76	20	Lolium multiflorum	20	Lepidium latifolium
76	1	Lepidium latifolium	12	Lolium multiflorum
77	45	Hordeum marinum	2	Lolium multiflorum
77	45	Lolium multiflorum	0.2	Cotula coronopifolia
77	0	None	0.2	Polypogon monspeliensis
78	10	Lolium multiflorum	70	Lepidium latifolium
78	5	Lotus corniculatus	1	Lotus corniculatus
78	5	Lepidium latifolium	1	Apium graveolens
79	1	Polypogon monspeliensis	1	Polypogon monspeliensis
79	0.2	Hordeum marinum	0.2	Lolium multiflorum
83	5	Polypogon monspeliensis	17	Cotula coronopifolia
83	0.2	Sonchus oleraceus	8	Polypogon monspeliensis
83	0.2	Rumex crispus	3	Lotus corniculatus

Plot #	% Cover 1999	Top Non-Natives 1999	% Cover 2006	Top Non-natives 2006
86	12	Polypogon monspeliensis	10	Picris echioides
86	6	Apium graveolens	2	Cressa truxillensis
86	5	Picris echioides	2	Apium graveolens
87	2	Foeniculum vulgare	5	Foeniculum vulgare
87	0.2	Sonchus oleraceus	3	Cynodon dactylon
87	0.2	Rumex crispus	3	Picris echioides
88	8	Polypogon monspeliensis	0.2	Lotus corniculatus
88	0.2	Cotula coronopifolia	0.2	Polypogon monspeliensis
88	0	None	0.2	Cirsium vulgare
89	15	Polypogon monspeliensis	18	Poaceae
89	10	Rumex crispus	7	Lolium multiflorum
89	5	Lolium multiflorum	0	None
90	1	Lepidium latifolium	<mark>4</mark>	Lepidium latifolium
90	0.2	Rumex pulcher	0	None
90	0.2	Rumex crispus	0	None
91	75	Lolium multiflorum	68	Lolium multiflorum
91	7	Bromus hordeaceus	10	Hordeum marinum
91	4	Hordeum marinum ssp. gussonianum	1	Bromus hordeaceus
92	2	Rumex pulcher	3	Rumex dentatus
92	0.2	Rumex conglomeratus	2	Polypogon monspeliensis
92	0.2	Cotula coronopifolia	0.2	Sonchus oleraceus
99	<mark>78</mark>	Lepidium latifolium	40	Lepidium latifolium
99	1	Polypogon monspeliensis	3	Cotula coronopifolia
99	0.2	Sonchus oleraceus	2	Polypogon monspeliensis
107	0	None	0.2	Polypogon monspeliensis
108	30	Bromus diandrus	12	Taeniatherum caput-medusae
108	15	Bromus hordeaceus	10	Bromus diandrus
108	5	Taeniatherum caput-medusae	5	Bromus hordeaceus
110	1	Polypogon monspeliensis	2	Hainardia cylindrica
110	0.2	Lolium multiflorum	1	Polypogon monspeliensis
110	0	None	0.2	Bromus diandrus
111	0.2	Picris echioides	2	Lotus corniculatus
111	0.2	Polypogon monspeliensis	0.2	Apium graveolens
111	<mark>0.2</mark>	Lepidium latifolium	0.2	Lolium multiflorum
118	24	Conium maculatum	22	Conium maculatum
118	6	Polypogon monspeliensis	3	Hordeum marinum
118	5	Bromus diandrus	3	Vulpia bromoides
136	8	Lolium multiflorum	73	Lolium multiflorum
136	6	Lotus corniculatus	3	Raphanus sativus
136	4	Sonchus oleraceus	0	None
145	0.2	Sonchus oleraceus	0.2	Agrostis avenacea
145	0	None	0.2	Cotula coronopifolia
145	0	None	0.2	Conium maculatum
147	44	Lolium multiflorum	15	Polypogon monspeliensis
147	35	Agrostis avenacea	10	Lolium multiflorum
147	23	Xanthium strumarium	1	Cotula coronopifolia
148	96	Lolium multiflorum	97	Lolium multiflorum

Plot #	% Cover 1999	Top Non-Natives 1999	% Cover 2006	Top Non-natives 2006
148	0.2	Xanthium strumarium	2	Bromus diandrus
148	0.2	Lactuca serriola	1	Lactuca serriola
154	7	Polypogon monspeliensis	15	Polypogon monspeliensis
154	2	Lolium multiflorum	3	Cotula coronopifolia
154	1	Cotula coronopifolia	1	Lolium multiflorum
159	1	Polypogon monspeliensis	5	Polygonum
159	0.2	Rumex crispus	2	Sonchus oleraceus
159	0.2	Sonchus oleraceus	2	Rumex crispus
161	30	Echinochloa crus-galli	40	Xanthium strumarium
161	3	Xanthium strumarium	4	Raphanus sativus
161	2	Polypogon monspeliensis	2	Lactuca serriola
168	52	Polypogon monspeliensis	2	Cotula coronopifolia
168	28	Xanthium strumarium	0	None
168	10	Lotus corniculatus	0	None
169	1	Lolium multiflorum	51	Lolium multiflorum
169	1	Sonchus oleraceus	30	Bromus diandrus
169	0	None	1	Lactuca serriola
170	7	Polygonum argyrocoleon	14	Rumex conglomeratus
170	2	Rumex pulcher	2	Raphanus sativus
170	0.2	Polypogon monspeliensis	0.2	Chenopodium album
171	28	Lolium multiflorum	56	Bromus diandrus
171	0.2	Polypogon monspeliensis	35	Lolium multiflorum
171	0.2	Raphanus sativus	1	Hordeum murinum ssp. Ieporinum
172	0	None	2	Rumex conglomeratus
172	0	None	0.2	Chenopodium album
172	0	None	0.2	Cotula coronopifolia
175	38	Xanthium strumarium	70	Xanthium strumarium
175	30	Phragmites australis	1	Rumex
175	2	Echinochloa crus-galli	1	Echinochloa crus-galli
176	33	Conium maculatum	42	Conium maculatum
176	8	Centaurea solstitialis	5	Centaurea solstitialis
176	6	Cirsium vulgare	5	Raphanus sativus
177	48	Conium maculatum	53	Conium maculatum
177	7	Raphanus sativus	2	Sonchus oleraceus
177	3	Picris echioides	1	Raphanus sativus
181	0	None	<mark>0.2</mark>	Lepidium latifolium
181	0	None	0.2	Asparagus officinalis
184	0.2	Polygonum arenastrum	<mark>0.2</mark>	Lepidium latifolium
184	0.2	Polypogon monspeliensis	0	None
185	1	Elytrigia pontica	90	Elytrigia pontica
185	0.2	Sonchus oleraceus	3	Conium maculatum
185	0.2	Lactuca serriola	3	Brassica nigra
186	80	Xanthium strumarium	51	Xanthium strumarium
186	7	Cotula coronopifolia	25	Cotula coronopifolia
186	5	Polypogon monspeliensis	2	Polypogon monspeliensis
189	1	Cotula coronopifolia	0.2	Rumex dentatus
189	0	None	0.2	Cotula coronopifolia

Plot #	% Cover 1999	Top Non-Natives 1999	% Cover 2006	Top Non-natives 2006
189	0	None	0.2	Rumex crispus
192	35	Agrostis avenacea	20	Bromus diandrus
192	6	Bromus diandrus	8	Lactuca serriola
192	1	Lolium multiflorum	2	Centaurea solstitialis
194	0	None	85	Polypogon monspeliensis
194	0	None	3	Cotula coronopifolia
194	0	None	1	Rumex conglomeratus
195	<mark>4</mark>	Lepidium latifolium	37	Poa
195	3	Bromus hordeaceus	2	Conium maculatum
195	3	Lolium multiflorum	0.2	Bromus diandrus
197	25	Rubus discolor	15	Rubus discolor
197	1	Lepidium latifolium	<mark>4</mark>	Lepidium latifolium
197	0.2	Rumex conglomeratus	0.2	Agrostis avenacea

<u>Summary of Table 4.</u> There are 85 total plots represented in Table 4; of these, 75 plots in 1999 had non-natives and 83 plots have non-natives in 2006. Six 1999 plots had higher *Lepidium* and twelve 2006 plots had higher *Lepidium*. Based on simple additive cover, there was 45.5 % greater cover in the six 1999 plots that had higher cover relative to 2006. Conversely, there was a total percentage cover of 119.2 % more in the twelve 2006 plots with greater cover. Thus, *Lepidium* appears to have had an overall increase of 73.7% (112.2% minus 45.5%) cover on the re-sampled plots in the 7-year interval. This same kind of comparison can be made with other species. A summary of the number of plots containing all 56 top non-native species and their average covers when present is provided in Table 5.

	# Plots species present 1999 (of	Frequency of species in 1999 plots	# Plots species present 2006 (of	Frequency of species in 2006 plots	1999 mean % cover when	2006 mean % cover when
Species	78 total)		84 total)		present	present
Polypogon monspeliensis	28	0.36	28	0.33	5.1	7
No non-natives	20	0.26	14	0.25	n/a	n/a
Lolium multiflorum	20	0.26	21	0.17	25.3	27.6
Cotula coronopifolia	16	0.21	21	0.25	6.2	4.2
Sonchus oleraceus	16	0.21	5	0.06	0.9	4.6
Lepidium latifolium	14	0.18	15	0.18	11.1	15.3
Rumex crispus	12	0.15	6	0.07	9.1	1.3
Bromus diandrus	10	0.13	16	0.19	8.8	13.7
Lactuca serriola	10	0.13	7	0.08	4.1	5.2
Bromus hordeaceus	8	0.10	5	0.06	17.75	3.8
Xanthium strumarium	7	0.09	4	0.05	24.6	40.3
Hordeum marinum	6	0.08	5	0.06	23.4	12.8
Rumex conglomeratus	6	0.08	5	0.06	10.7	3.8
Picris echioides	5	0.06	3	0.04	1.9	4.7
Rumex pulcher	5	0.06	1	0.01	1.1	0.2

Table 5. Summary information for 56 non-native species with major cover Note: statistics for plots with no non-native species are indicated in red.

Species	# Plots species present 1999 (of 78 total)	Frequency of species in 1999 plots	# Plots species present 2006 (of 84 total)	Frequency of species in 2006 plots	1999 mean % cover when present	2006 mean % cover when present
Polygonum argyrocoleon	4	0.05	0	0.11	. 20	n/a
Lotus corniculatus	4	0.05	9	0	9.8	8.1
Rubus discolor	3	0.04	4	0.1	74.3	44
Conium maculatum	3	0.04	8	0.05	35	15.3
Echinochloa crus-galli	3	0.04	1	0.01	13	1
Elytrigia pontica	2	0.03	1	0.14	44	90
Centaurea solstitialis	2	0.03	6	0.07	39	5.8
Agrostis avenacea	2	0.03	2	0.02	35	0.2
Apium graveolens	2	0.03	12	0.02	4.5	1.1
Hordeum marinum ssp.	2	0.03	1	0.01	15	1
Giroium undere	2	0.03	1	0.01	4.5	0.2
	2	0.03	2	0.01	2	4.5
Poeniculum vulgare	2	0.03	2	0.01	0.2	4.5 n/a
	1	0.01	1	0.06	16	30
Curpia myuros	1	0.01	2	0.00	10	3
	1	0.01	2	0.02	15	
	1	0.01	5	0.01	3	5.2
	1	0.01	0	0.01	7	0.2 D/2
	1	0.01	1	0.01	5	11/a
Taeniatherum caput-	<u> </u>	0.01	I	0	5	
medusae	1	0.01	1	0	5	12
Acacia sp.	1	0.01	0	0	0.2	n/a
Cirsium sp.	1	0.01	0	0	0.2	0.2
Agrostis viridis	0	0	1	0.02	n/a	4
Asparagus officinalis	0	0	2	0.02	n/a	0.2
Avena sp.	0	0	1	0.02	n/a	4
Avena barbata	0	0	1	0.02	n/a	11
Brassica nigra	0	0	1	0.01	n/a	3
Carduus pycnocephalus	0	0	1	0.01	n/a	1
Chenopodium album	0	0	2	0.01	n/a	0.2
Crypsis schoenoides	0	0	1	0.01	n/a	0.2
Digitaria sanguinalis	0	0	1	0.01	n/a	12
Hainardia cylindrica	0	0	1	0.01	n/a	2
Hedera helix	0	0	1	0.01	n/a	0.2
Malva neglecta	0	0	1	0.01	n/a	1
Paspalum dilatatum	0	0	1	0.01	n/a	1
Poa sp.	0	0	1	0.01	n/a	37
Poaceae	0	0	1	0.01	n/a	18
Polygonum sp.	0	0	2	0.01	n/a	5
Polygonum baldschuanicum	0	0	1	0.01	n/a	1
Rumex dentatus	0	0	2	0.01	n/a	1.6
Rumex sp.	0	0	1	0.01	n/a	1
Vulpia bromoides	0	0	1	0.01	n/a	3

<u>Summary of Table 5</u>. Interestingly, only one species, *Polypogon monspeliensis*, occurred in more than 1/3 of all the samples in either 1999 or 2006. This species averaged similar cover (5 - 7 %) both years. In 1999, 15 species averaged > 10 percent cover in the plots in which they occurred. However only 3 of these, *Lolium multiflorum, Lepidium latifolium*, and *Bromus hordeaceus*, were present in more than 10 percent of the sample plots. Similarly, in 2006, 14 species averaged >10 percent cover in the plots in which they occurred, and only three species, *Lolium multiflorum, Lepidium latifolium*, and *Bromus diandrus*, were in more than 10 percent of the sample plots. Individual species with the highest cover included *Elytrigia pontica, Rubus discolor, Xanthium strumarium*, and *Conium maculatum*, all averaging more than 15% when they occurred in both 1999 and 2006. However, these species are restricted to only a few sample plots. Substantial decreases appeared to occur in *Centaurea solstitialis, Bromus hordeaceus*, and *Agrostis avenacea*. Increases were recorded for *Elytrigia pontica* and *Vulpia myuros*.

Probably the most worrisome exotic species in this group, *Lepidium latifolium*, appeared to increase slightly since 1999 occurring in 15% of the plots (versus 14% in 1999) and averaging about 15% cover as opposed to 11 % in 1999. This species has been actively managed with herbicides and other treatments in some of the DFG managed areas. It is uncertain if any treatments were applied in the resampled areas.

Overall, based on these samples, there is no clear increase or decrease in the percentage of exotic cover in Suisun Marsh.

Vegetative Cover, Total Non-Native Cover, Species Richness, and Inundation

Table 6 presents statistics for comparison between plots sampled in 1999 and 2006, and Table 7 summarizes these statistics. Table 8 shows the statistics for the plots that showed more than one cover class difference in total vegetation cover in the two sample years.

	% Veg	% Vea	% Non-	% Non-				
Plot #	Cover 1999	cover 2006	cover 1999	cover 2006	# of Species/ Plot 1999	# of Species/ Plot 2006	% Water 1999	% Water 2006
001	90	65	0	0.2	5	8	10	22
002	88	57	0	0	5	3	0	90
003	83	65	0	0	2	3	0	78
004	95	60	15	5	27	34	10	10
009	85	40	0	0	2	2	15	96
010	100	95	100	95	2	2	0.2	8
011	95	0	95	0	1	0	3	100
012	95	75	0	5	33	19	1	18
013	99	70	77	52	13	17	0	0
014	91	32	2	0.2	7	6	0	93
015	96	70	1	1	4	8	0	97
016	97	65	7	2	7	20	0	50
020	95	28	95	0	6	3	18	99
021	85	63	0	3	4	5	15	0
023	95	81	15	21	10	9	5	0
024	96	70	3	1	11	13	3	6
025	98	82	97	82	16	14	0	0

Table 6. Total percent (%) vegetation cover, non-native cover, species richness, and water cover, 1999 vs. 2006 Only those plots that were resampled in 2006 are listed. Those figures in red indicate which year has highest value.

Plot #	% Veg Cover 1999	% Veg cover 2006	% Non- native cover 1999	% Non- native cover 2006	# of Species/ Plot 1999	# of Species/ Plot 2006	% Water 1999	% Water 2006
026	94	70	32	0.2	9	6	0	25
034	100	95	100	95	4	14	0	0
036	90	62	80	30	18	21	0	0
037	88	85	99	55	16	14	0	4
038	100	70	5	20	9	9	0	0
039	85	45	7	5	11	10	0	0
040	76	1	6	0	2	2	0	0
042	100	95	12	4	23	17	0	40
043	100	99	0	1	17	19	0	15
044	98	80	98	75	4	6	0	0
045	95	95	0	0.2	4	4	5	96
046	100	90	0	0.2	8	12	0	36
047	100	96	0	1	12	15	0	4
048	92	99	30	6	8	17	0	0
049	100	99	0	0	3	9	0	4
050	99	45	98	35	3	5	0	0
051	80	99	0	0	1	2	20	0
052	93	92	85	50	13	17	0	4
053	94	85	1	3	11	6	0	4
054	100	95	3	11	11	22	0	15
056	95	96	30	36	9	17	0	0
058	99	60	60	55	12	14	0	0
059	100	92	0	3	6	12	0	65
060	100	85	60	61	14	15	0.2	30
061	99	92	31	21	12	16	0	15
062	96	90	2	35	8	18	0	0
063	96	85	4	8	14	13	0	82
064	98	99	4	0	9	9	1	72
065	96	75	7	10	9	12	0	0
067	100	87	1	0	6	5	0	0
068	100	88	31	45	8	12	0	78
070	97	96	75	75	8	6	0	0
071	97	78	4	0	10	4	0	0
072	85	77	0.2	0	5	4	0	90
073	*	80	0	2	11	7	0	0
074	80	95	51	70	9	11	0	0
075	96	85	0.2	23	6	8	0	0
076	95	95	90	75	14	18	0	20
077	98	62	90	4	7	8	0	0
078	98	88	20	70	12	17	0	30
079	91	70	1	1	3	4	0	0
083	93	82	6	28	12	14	7	0
086	98	96	24	15	15	20	, 0	0
0.87	08	85	2	20	11	33	0	20
007	92	05	0	1	6	11	7	
000	02	02	3	0	11	0	· ·	0
000	95	92	<u> </u>	7	14	9	0	0
090	95	90	<u>3</u>	1	14	8	0	0
091	90	75	80	82	1	9	0	0

Plot #	% Veg Cover 1999	% Veg cover 2006	% Non- native cover 1999	% Non- native cover 2006	# of Species/ Plot 1999	# of Species/ Plot 2006	% Water 1999	% Water 2006
092	97	92	3	4	6	6	0	0
099	98	85	80	47	13	13	0	0
107	60	62	0	0.2	10	13	0	53
108	98	75	57	40	7	11	0	0
110	92	80	7	4	6	15	0	1
111	100	100	5	3	18	16	0	96
118	92	60	42	30	14	14	0	2
136	100	95	22	74	14	16	0	0
145	99	98	0	1	6	12	0	4
147	100	75	65	43	16	13	0	0
148	98	100	97	99	10	9	0	3
154	67	50	12	15	11	11	0	0
159	100	75	1	5	12	19	0	3
161	98	92	35	60	13	15	0	0
168	96	75	40	2	16	5	0	90
169	99	92	4	90	13	11	0	0
170	88	48	2	32	15	18	0	0
171	93	90	27	90	14	6	0	0
172	100	85	0	71	2	16	0	0
175	95	72	40	1	12	11	0	0
176	88	91	50	40	13	13	0	0
177	98	85	60	51	17	13	0	0
180	0.2	1	0.2	0	2	1	0	99.5
181	88	99	0	0.2	8	13	0	3
184	75	87	0.2	0.2	7	5	10	95
185	90	97	1	97	7	5	0	0
186	90	90	89	30	7	10	1	0
189	39	65	1	0.2	3	8	0	0
190	1	0	1	0	4	0	0	100
192	100	86	42	35	15	13	0	0
194	98	92	0	91	4	17	0	0
195	97	99	11	39	9	7	0	0
197	95	88	0	20	15	28	5	10

*Total cover was not entered for this plot in the field in 1999, so this plot has been omitted from comparison of vegetation cover.

Table 7. Summary statistics for Table 6

Criteria	Ν	Percent	Range
Number of plots with total cover > 1 cover-class difference	8	8%	
Number of plots with total cover within one cover class			
difference	29	30%	
Number of plots with total cover in same cover class			
between years	60	62%	
Number of plots with more cover in 2006 (of 97 total)	16	16%	
Number of plots with less cover in 2006 (of 97 total)	77	79%	
Number of plots with same cover in both years (of 97 total)	4	4%	
Mean percent cover of non-natives per plot 1999 (n=98)		27.3	0 - 100
Mean percent cover of non-natives per plot 2006 (n=98)		25.8	0 - 100

Criteria	Ν	Percent	Range
Number of all species per plot 1999	9.7		1 - 33
Number of all species per plot 2006	11.3		1 - 34
Mean percent water cover in 1999 (n=98)		1.4	0 - 20
Mean percent water cover in 2006 (n=98)		21.2	0 - 100
Mean total vegetation cover in 1999 (n=97)		93.2	0- 100
Mean total vegetation cover in 2006 (n=98)		78.1	0- 100

Table 8. Plots with more than one cover class difference in total vegetation cover between sample years

Plot #	% Veg Cover 1999	% Veg cover 2006	% Exotic Cover 1999	% Exotic Cover 2006	# of Species/ Plot 1999	# of Species/ Plot 2006	% Water 1999	% Water 2006	Explanation for major shift
9	85	40	0	0	2	2	15	96	Much more water
11	95	0	95	0	1	0	3	100	Completely inundated recently
14	91	0	2	0.2	7	6	0	93	Completely inundated recently
20	95	28	95		6	3	18	99	Almost entirely flooded
39	85	45	7	5	11	10	0	0	Change in flooding regime? Much dead pickleweed.
40	76	1	6	0	2	2	0	0	Change in flooding regime? Pickleweed in 1999, in 2006 it was loose dirt with salty crust.
50	99	45	98	35	3	5	0	0	Site burned in 2001.
170	88	48	2	32	15	18	0	0	Change in flooding regime?

<u>Discussion of Tables 6, 7, and 8.</u> Water cover in 2006 increased many-fold compared to 1999 (Table 7). It is uncertain if this signifies a management shift to maintaining inundation for longer periods in the managed areas sampled or if it is simply an artifact of the plots selected for resampling.

Average species richness per plot was higher by about 2 species in 2006. It is unclear why this might be or if it is significant. Possible influencing factors could include: greater heterogeneity of stands in 2006, temporal or yearly effects in the ability to detect species, variation in field crews sampling intensity, etc.

Relative percent cover of non-natives remains the about the same from 1999-2006. In 2006, 43 plots had higher non-native cover than in 1999, while 46 had lower than in 1999.

After a 7-year period, the average total vegetation cover appears to have dropped by about 15%, but estimates of cover tended to be higher in 1999 because of different estimating techniques (crown versus foliar cover estimation). If cover-classes rather than actual percent cover are compared, then there is no difference in 62% of the plots. The 7-point CNPS cover-class scale (see Appendix A) was used to classify the vegetation initially in 1999 and is the conservative tool often used for data analysis by vegetation ecologists. Cover-class is often used to cancel out the effects of an individual's likelihood of minor variance in estimating cover of plants.

Eight of the plots had greater than one cover class difference in total cover between the two years (Table 8). These are unlikely to be the result of variation in the field crew's ability to accurately estimate cover. For four (and possibly seven) of the plots, the difference appeared to be the result of a management change in flooding regime, where drops in cover resulted from complete inundation in 2006 versus little or no inundation in 1999. One plot burned in 2001 and its cover and composition have not changed much since, according to the observer, who is familiar with the site.

Visual Comparison of Selected Plots Surveyed in 1999 and 2006

The following photo pairs are examples of those taken for each of the plots resampled in 2006 compared to 1999. They were selected to show clearly the value of repeat photography in distinguishing similarities and differences between the sample years. The complete set of 2006 and 1999 plot photos are on file at Vegetation Classification and Mapping Program of the Biogeographic Data Branch of DFG, as are the field survey forms, electronic database, and associated GIS layers.



Suisun Marsh relevé #024 in August 1999 (top) and 2006 (bottom). Despite increased flooding in 2006 (6% standing water compared to 0% in 1999) this plot has remained a *Distichlis spicata* Alliance.



Suisun Marsh relevé #014 in 1999 (top) and 2006 (bottom). Due to the increased flooding in 2006 most of the *Salicornia virginica* (which was the dominant species in 1999 at 80% cover) has died and *Typha angustifolia* has moved in and has become the dominant species at 30% cover.



Suisun Marsh relevé #063 in October 1999 (top) and September 2006 (bottom). This plot has remained a *Schoenoplectus americanus* Alliance, even though in 2006 this plot contained 82% standing water vs. 0% in 1999.



Suisun Marsh relevé #021 in September 1999 (top) and August 2006 (bottom). Schoenoplectus maritimus Alliance in 1999 and a Salicornia virginica Alliance in 2006. The switch in dominant cover species is likely due to a shortened inundation period in 2006 (0% standing water compared to 15% standing water in 1999).



Suisun Marsh relevé #002 in 1999 (top) and 2006 (bottom). In 1999 this was a *Schoenoplectus maritimus* (77%) Alliance and in 2006 it was a *Typha angustifolia* (55%) Alliance. *T. angustifolia* had a cover of only 2% in 1999.



Suisun Marsh relevé #171 in 1999 (top) and 2006 (bottom). In 1999 this plot was a *Frankenia salina* alliance. In 2006 it was a *Bromus diandrus* alliance. In 2006 the *F. salina* cover was 3% compared to 85% in 1999.

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Keeler-Wolf, T. and M. Vaghti. 2000. Vegetation Mapping of Suisun Marsh, Solano County, California. Report to the California Department of Water Resources. California Department of Fish and Game, Wildlife and Habitat Data Analysis Branch. Sacramento, CA.

Appendix A. Sample Field Form and Survey Protocol

CALIFORNIA NATIVE PLANT SOCIETY RELEVÉ FIELD FORM

(Revised 4/2/04)

Page_____ of Relevé # _____ See code list for italicized fields

	FOR OFFICE USE OF	NLY				
Polygon # or Relevé #	Permanent Number:					
Date Airphoto #	Community Name:					
MM DD YYYY	Community Number:	Occurrence Number:				
County	Source Code:					
USGS Quad. 7.5' or 15' (Circle one)	Quad Code: Map Index Number:	Quad Name:				
CNPS Chapter	Update: Yes	No (Circle one)				
Landowner						
Contact Person						
Address						
City	Zip	Phone number				
Observers						
Relevé plot shape (square, rectangle, triangle, circle, entir Relevé plot size (length and width of rectangle, or circle-a Study Plot Revisit? Yes or No (Circle one) Other polygons of same type? Yes or No Is plot represe	Relevé plot shape (square, rectangle, triangle, circle, entire stand) NOTE: All forest and woodland plots should be 1000m ² Relevé plot size (length and width of rectangle, or circle-diameter) (m.) Study Plot Revisit? Yes or No (Circle one) Other polygons of same type? Yes or No Is plot representative of whole polygon? Yes or No (Circle one)					
GPS File # GPS name (or points in file)	Start Time:	(am or pm) GPS Datum (from GPS setup) (e.g. WGS 84, NAD 27)				
File type: Point or Polygon (circle one) Releve: UTMN _		UTME Error ±ft/m UTM Zone				
Transect: Start UTME UTMN UTMN		End: UTME UTMN				
Elevation (ft.) Slope (°)	Aspect (°)	Topography: Macro Micro				
VEGETATION DESCRIPTION						
Dominant Layer 0-0.5 m, 0.5-4 m,4 m	Preliminary Alliance Nam	ie				
Stand Size<1 acre,1-5 acres,>5 acres	Dominant Vegetation Gro	pup (use codes from code list)				
Structure: Ground Shrub (1. Continuous 2. Intermittent 3. Open)	Tree	Phenology: GroundShrubTree (Early, Peak, Late)				
Wetland Community Type		(Wetland or Upland)				
If Community Type = Wetland (see Artificial Keys to Cov	wardin Systems and Names	s)				
Cowardin System	Subsystem	nClass				
Distance to water (m): Vertical	Horizontal	Channel form (if riverine) (Straight, Meandering, Braided)				
Adjacent Alliance Location (e.g., North, South, E	ast, or West of stand)	Description (up to 4 species by layer)				

Photograp	hs – Note pos	ition and dir	rection of ph	oto(s) relative	e to plot					
0	*				ł					
		CAL	IFORNIA N	NATIVE PLA	ANT SOCIE	TV RELE	VÉ FIELD F	ORM		
		CAL	IF OKINA I	Page	_ of Relevé # _		VETIELD T	OKM		
STAND A	ND ENVIR	ONMENTA	L DESCRI	PTION						
Trond and		Site I	immont and an							
1. Increasing 2.	Stable 3. Decrea	sing Site I	ntensity		(List cod	es in order, with r	nost significant first	t)		
4. Pluctuating	5. UIKIIOWII	Shell			1. Light 2. Mode	rate 3. Heavy (Li	st beneath each imp	pact code)		
Site Locat	ion and Plot	Description	n							
Site Histor	ry – includin	g observatio	ns of fire sca	urs, insect/dis	ease damage	e, grazing/br	owsing, humo	ın disturban	се	
Sensitive S	Species – Lis	t species obs	erved and G	PS UTM's; E	Estimate size	and extent o	of local popula	ations		
Unknown	Specimens -	- List code, i	dentification	notes (e.g. G	Genus, condit	ion of specir	men) of unkno	owns		
Additiona	l Comments	– Including	animal obse	rvations, antl	hropological	observation	ıs, abiotic fea	tures		
Surface C	oarse Fragn	ents and So	oils Informa	tion (see cov	er class inter	vals- below ·	₩)			
Type:	Fines	Gravel	Cobble	Stone	Boulders	Bedrock	Litter	Water	Living	Other (Specify):
Descriptor:	Including sand, mud	2mm-7.5 cm diameter	7.5-25 cm diam	25-60cm diam.	>60cm diam.	Including outcrops	Organic matter covering ground	Standing or running water	At ground surface	(
Cover class (see below):							0000			
% Cover*:										
*note all s	urface fragme	ents, non-veg	getation, livii	ng stems, etc.	, should add	up to 100%	<u> </u>		I	
		Soil Te	exture			Pare	nt Material _			

Height (Height Classes for Vegetation Strata & Cover Estimates (see cover class intervals - above 1)									
Layer name:	Cryptogam Layer	0-25 cm	25-50 cm	0.5-1 m	1-2 m	2-5 m	5-10 m	10-20 m	20-30 m	>30 m.
Main species:										
Cover class:										

CALIFORNIA PLANT COMMUNITIES RELEVÉ FIELD FORM (PART 2)

SPECIES SHEET (Revised 5/17/01)

Page_____ of Relevé # _____

Cover Class Intervals: 1 (<1%), 2 (1-5%), 3a (>5-15%), 3b (>15-25%), 4 (>25-50%), 5 (>50-75%), 6 (>75%)

L=Low herbs and subshrubs (<0.5 m.), M=Medium height (0.5 m.-4.0 m.), T=Tall height (>4.0 m.)

L	М	Т	Vascular plant name or moss/lichen cryptogamic crust cover	Final species determination or Tree dbh	Cover Class	%
		Total	Vegetation Cover (Class): Total Tall Total Medium	Total Low Total No	on-Native	
			percent cover of above:			

CALIFORNIA PLANT COMMUNITIES RELEVÉ FIELD FORM (PART 2)

SPECIES SHEET (Revised 5/17/01)

Page_____ of Relevé # _____

Cover Class Intervals: 1 (<1%), 2 (1-5%), 3a (>5-15%), 3b (>15-25%), 4 (>25-50%), 5 (>50-75%), 6 (>75%)

L=Low herbs and subshrubs (<0.5 m.), M=Medium height (0.5 m.-4.0 m.), T=Tall height (>4.0 m.)

L	М	Т	Vascular plant name or moss/lichen cryptogamic crust cover	Final species determination or Tree dbh	Cover Class	%
		Total	Vegetation Cover (Class): Total Tall Total Medium	Total Low Total No	on-Native	
			percent cover of above:			

CALIFORNIA NATIVE PLANT SOCIETY RELEVÉ PROTOCOL CNPS VEGETATION COMMITTEE October 20, 2000 (Revised 4/2/04)

Introduction

In *A Manual of California Vegetation* (Sawyer and Keeler-Wolf 1995), CNPS published a Vegetation Sampling Protocol that was developed as a simple quantitative sampling technique applicable to many vegetation types in California. Investigators use an ocular estimation technique called a relevé to classify and map large areas in a limited amount of time.

The relevé method of sampling vegetation was developed in Europe and was largely standardized by the Swiss ecologist Josias Braun-Blanquet. He helped classify much of Europe's vegetation, founded and directed a synecology center in France, and was editor of *Vegetatio* for many years. The relevé was, and is, a method used by many European ecologists, and others around the world. These ecologists refer to themselves as phytosociologists. The use of relevé in the United States has not been extensive with the exception of the US Forest Service.

The relevé is particularly useful when observers are trying to quickly classify the range of diversity of plant cover over large units of land. In general, it is faster than the point intercept technique. One would use this method when developing a classification that could be used to map of a large area of vegetation, for example. This method may also be more useful than the line intercept method when one is trying to validate the accuracy of mapping efforts.

The relevé is generally considered a "semiquantitative" method. It relies on ocular estimates of plant cover rather than on counts of the "hits" of a particular species along a transect line or on precise measurements of cover/biomass by planimetric or weighing techniques.

Selecting a stand to sample:

A stand is the basic physical unit of vegetation in a landscape. It has no set size. Some vegetation stands are very small, such as alpine meadow or tundra types, and some may be several square kilometers in size, such as desert or forest types. A stand is defined by two main unifying characteristics:

- 1) It has <u>compositional</u> integrity. Throughout the site the combination of species is similar. The stand is differentiated from adjacent stands by a discernable boundary that may be abrupt or indistinct, and
- 2) It has <u>structural</u> integrity. It has a similar history or environmental setting that affords relatively similar horizontal and vertical spacing of plant species throughout. For example, a hillside forest originally dominated by the same species that burned on the upper part of the slopes, but not the lower, would be divided into two stands. Likewise, a sparse woodland occupying a slope with very shallow rocky soils would be considered a different stand from an adjacent slope with deeper, moister soil and a denser woodland or forest of the same species.

The structural and compositional features of a stand are often combined into a term called <u>homogeneity</u>. For an area of vegetated ground to meet the requirements of a stand it must be homogeneous.

Stands to be sampled may be selected by assessment prior to a site visit (e.g. delineated from aerial photos or satellite images), or may be selected on site (during reconnaissance to determine extent and boundaries, location of other similar stands, etc.). Depending on the project goals, you may want to select just one or a few representative stands for sampling (e.g., for developing a classification for a vegetation mapping project), or you may want to sample all of them (e.g., to define a rare vegetation type and/or compare site quality between the few remaining stands).

Selecting a plot to sample within in a stand:

Because most stands are large, it is difficult to summarize the species composition, cover, and structure of an entire stand. We are also usually trying to capture the most information with the least amount of effort. Thus, we are typically forced to select a representative portion to sample.

When sampling a vegetation stand, the main point to remember is to select a sample that, in as many ways possible, is representative of that stand. This means that you are not randomly selecting a plot; on the contrary, you are actively using your own best judgement to find a representative example of the stand.

Selecting a plot requires that you see enough of the stand you are sampling to feel comfortable in choosing a representative plot location. Take a brief walk through the stand and look for variations in species composition and in stand structure. In many cases in hilly or mountainous terrain look for a vantage point from which you can get a representative view of the whole stand. Variations in vegetation that are repeated throughout the stand should be included in your plot. Once you assess the variation within the stand, attempt to find an area that captures the stand's common species composition and structural condition to sample.

Plot Size

All releves of the same type of vegetation to be analyzed in a study need to be the same <u>size</u>. It wouldn't be fair, for example, to compare a 100 m2 plot with a 1000 m2 plot as the difference in number of species may be due to the size of the plot, not a difference in the stands.

A minimal area to sample is defined by species/area relationships; as the sampler identifies species present in an area of homogeneous vegetation, the number will increase quickly as more area is surveyed. Plot shape and size are somewhat dependent on the type of vegetation under study. Therefore general guidelines for plot sizes of tree-, shrub-, and herb-dominated upland, and fine-scale herbaceous communities have been established. Sufficient work has been done in temperate vegetation to be confident the following conventions will capture species richness:

Alpine meadow and montane wet meadow: 100 sq. m
Herbaceous communities: 100 sq. m plot or 400 sq. m plot (Consult with CNPS, and use one consistent size)
Grasslands and Shrublands: 400 sq. m plot
Forest and woodland communities: 1000 sq. m plot
Open desert vegetation: 1000 sq. m plot

Plot Shape

A relevé has no fixed shape, plot shape should reflect the character of the stand. If the stand is about the same size as a relevé, you need to sample the entire stand. If we are sampling a desert wash, streamside riparian, or other linear community our plot dimensions should not go beyond the community's natural ecological boundaries. Thus, a relatively long, narrow plot capturing the vegetation within the stand, but not outside it would be appropriate. Species present along the edges of the plot that are clearly part of the adjacent stand should be excluded.

If we are sampling broad homogeneous stands, we would most likely choose a shape such as a circle (which has the advantage of the edges being equidistant to the center point) or a square (which can be quickly laid out using perpendicular tapes). If we are trying to capture a minor bit of variety in the understory of a forest, for example a bracken fern patch within a ponderosa pine stand, we would want both bracken and non-bracken understory. Thus, a rectangular shape would be appropriate.

GENERAL PLOT INFORMATION

The following items appear on each data sheet and are to be collected for all plots. Where indicated, refer to attached code sheet.

Polygon or Relevé number: Assigned either in the field or in the office prior to sampling.

Date: Date of sampling.

County: County in which located.

<u>USGS Quad</u>: The name of the USGS map the relevé is located on; note series (15' or 7.5').

<u>CNPS Chapter</u>: CNPS chapter, or other organization or agency if source is other than CNPS chapter.

Landowner: Name of landowner or agency acronym if known. Otherwise, list as private.

Contact Person: Name, address, and phone number of individual responsible for data collection.

Observers: Names of individuals assisting. Circle name of recorder.

Plot shape: indicate the sample shape as: square, rectangle, circle, or the entire stand.

Plot size: length of rectangle edges, circle radius, or size of entire stand.

NOTE: See page 2 for standard plot sizes.

Study Plot Revisit: If the relevé plot is being revisited for repeated sampling, please circle "Yes".

<u>Photo interpreter community code</u>: If the sample is in area for which delineation and photo interpretation has already been done, the code which the photointerpreters applied to the polygon. If the sample site has not been photointerpreted, leave blank.

<u>Other polygons of same type</u> (yes or no, if applicable), if yes, mark on map: Other areas within view that appear to have similar vegetation composition. Again, this is most relevant to areas that have been delineated as polygons on aerial photographs as part of a vegetation-mapping project. If one is not working from aerial photographs, draw the areas as on a topographic map.

<u>Is plot representative of whole polygon?</u> (yes or no, if applicable), if no explain: Detail what other vegetation types occur in the polygon, and what the dominant vegetation type is if there is more than one type.

<u>Global Positioning System Readings</u>: Due to the recent availability of very accurate and relatively low cost GPS units, we highly recommend obtaining and using these as a standard piece of sampling equipment. Now that the military intentional imprecision (known as "selective availability") has been "turned off" (as of July 2000), it is typical for all commercial GPS units these units to be accurate to within 5 m of the actual location. Also note that the GPS units can be set to read in UTM or Latitude and Longitude coordinates and can be easily translated. Thus, the following fields for Latitude, Longitude, and legal description are now optional. In order for all positional data to be comparable within the CNPS vegetation dataset We request using UTM coordinates set for the NAD 83 projection (see your GPS users manual for instructions for setting coordinates and projections).

Caveat: Although GPS units are valuable tools, they may not function properly due to the occasionally poor alignment of satellites or due to the complexity of certain types of terrain, or vegetation. We thus also recommend that you carry topographic maps and are aware of how to note your position on them in the event of a non-responsive or inaccurate GPS.

<u>UTMN and UTME</u>: Northing and easting coordinates using the Universal Transverse Mercator (UTM) grid as delineated on the USGS topographic map, or using a Global Positioning System.

<u>UTM zone</u>: Universal Transverse Mercator zone. Zone 10S for California west of the 120th longitude; zone 11S for California east of 120th longitude.

<u>Legal Description</u>: Township/Range/Section/Quarter Section/Quarter-Quarter section/Meridian: Legal map location of the site; this is useful for determining ownership of the property. California Meridians are Humboldt, Mt. Diablo, or San Bernardino. (This is optional, see above discussion of GPS units)

Latitude and Longitude: Degrees north latitude and east longitude. This is optional (see above)

Elevation: Recorded in feet or meters. Please indicate units.

Slope: Degrees, read from clinometer or compass, or estimated; averaged over relevé

<u>Aspect</u>: Degrees from true north (adjust declination), read from a compass or estimated; averaged over relevé.

<u>Macrotopography</u>: Characterize the large-scale topographic position of the relevé. This is the general position of the sample along major topographic features of the area. *See attached code list*.

<u>Microtopography</u>: Characterize the local relief of the relevé. Choose the shape that mimics the lay of the ground along minor topographic features of the area actually within the sample. *See attached code list*.

VEGETATION DESCRIPTION

<u>Dominant layer</u>: Indicate whether the community is dominated by the Low layer (L), Mid-layer (M), or Tall (T) layer.

<u>Preliminary Alliance name</u>: Name of series, stand, or habitat according to CNPS classification (per Sawyer and Keeler-Wolf 1995); if the type is not defined by the CNPS classification, note this in the space.

<u>Adjacent alliance</u>: Adjacent vegetation series, stands or habitats according to CNPS classification; list in order of most extensive to least extensive.

<u>Structure</u>: Characterize the structure of each layer.

Continuous = greater than 2/3 (67%) cover; crowns touching Intermittent = between 1/3 and 2/3 cover (33% to 66%); interlocking or touching crowns interrupted by openings. Open = less than 1/3 (33%) cover; crowns not touching or infrequently touching.

<u>Phenology</u>: Based on the vegetative condition of he principal species, characterize the phenology of each layer as early (E), peak (P), or late (L).

WETLAND COMMUNITY TYPES

<u>Community type</u>: Indicate if the sample is in a wetland or an upland; note that a site need not be officially delineated as a wetland to qualify as such in this context.

<u>Dominant vegetation form</u>: This is a four letter code which relates the vegetation of the plot to the higher levels of the NBS/NPS National Vegetation Classification System hierarchy. *See attached code list*.

<u>Cowardin class</u>: See "Artificial Keys to Cowardin Systems and Names" (attached). If the plot is located in a wetland, record the proper Cowardin system name. Systems are described in detail in Cowardin et al. 1979. Classification of wetlands and deepwater habitats of the United States. US Dept. of the Interior, Fish and Wildlife Service, Office of Biological Services, Washington, D.C.

Marine: habitats exposed to the waves and currents of the open ocean (subtidal and intertidal habitats).

Estuarine: includes deepwater tidal habitats and adjacent tidal wetlands that are usually semi-enclosed by land but have open, partly obstructed, or sporadic access to the open ocean, and in which ocean water is at least occasionally diluted by freshwater runoff from the land (i.e. estuaries and lagoons).

Riverine: includes all wetlands and deepwater habitats contained within a channel, excluding any wetland dominated by trees, shrubs, persistent emergent plants, emergent mosses, or lichens. Channels that contain oceanic-derived salts greater than 0.5% are also excluded.

Lacustrine: Includes wetlands and deepwater habitats with all of the following characteristics: 1) situated in a topographic depression or a dammed river channel; 2) lacking trees or shrubs, persistent emergents, emergent mosses or lichens with greater than 30% aerial coverage; and total area exceeds 8 ha (20 acres). Similar areas less than 8 ha are included in the lacustrine system if an active wave-formed or bedrock shoreline feature makes up all or part of the low tide boundary, of if the water in the deepest part of the basin exceeds 2 m (6.6 feet) at low tide. Oceanic derived salinity is always less than 0.5%.

Palustrine: Includes all nontidal wetlands dominated by trees, shrubs, persistent emergents, emergent mosses or lichens, and all such wetlands that occur in tidal areas where salinity derived from oceanic salts is less than 0.5%. Also included are areas lacking vegetation, but with all of the following four characteristics: 1) areas less than 8 ha (20 acres); active waveformed or bedrock shoreline features lacking; 3) water depth in the deepest part of the basin less than 2 m (6.6 feet) at low water; and 4) salinity due to ocean-derived salts less than 0.5%.

<u>Vertical distance from high water mark of active stream channel</u>: If the plot is in or near a wetland community, record to the nearest meter or foot the estimated vertical distance from the middle of the plot to the average water line of the channel, basin, or other body of water.

<u>Horizontal distance from high water mark of active stream channel</u>: If the plot is in or near a wetland community, record to the nearest meter or foot the estimated horizontal distance from the middle of the plot to the average water line of the channel, basin, or other body of water.

<u>Stream channel form</u>: If the plot is located in or near a community along a stream, river, or dry wash, record the channel form of the waterway. The channel form is considered S (single channeled) if it consists of predominately a single primary channel, M (meandering) if it is a meandering channel, and B (braided) if it consists of multiple channels interwoven or braided.

<u>Photographs</u>: Describe the number of color photographs taken at the relevé, and the camera's view direction from compass bearings. It is helpful to take a photograph of the relevé from the intersection of the tapes (if tapes were used to define the plot), and another from inside the relevé. Additional photos of the stand may also be helpful. If using a digital camera or scanning in the image into a computer, relevé numbers and compass directions can be recorded digitally. If using a 35mm camera, please note the roll number, frame number, compass direction, and the initials of the person whose camera is being used. (e.g. Roll 5, #1, to the NW, SS)

STAND AND ENVIRONMENTAL INFORMATION

<u>Vegetation trend</u>: Based on the regenerating species and relationship to surrounding vegetation, characterize the stand as either increasing (expanding), stable, decreasing, fluctuating, or unknown.

<u>Impacts</u>: Enter codes for potential or existing impacts on the stability of the plant community. Characterize each as either 1. Light, 2. Moderate, of 3. Heavy. *See attached code list*.

<u>Site location and plot description</u>: A concise, but careful description that makes locating and/or revisiting the vegetation stand and plots possible; give landmarks and directions. Used in conjunction with the GPS position recorded earlier, this should enable precise re-location of the plot. Indicate where the GPS reading was taken within the plot. In general, the location of the GPS reading should be on the Southeastern corner of the plot, if the plot is square or rectangular, or in the center if the plot is circular. It is also helpful to briefly describe the topography, aspect, and vegetation structure of the site. If you can't take the GPS reading at the Southeast corner (an obstacle in the way) then note where the GPS point was taken. If you can't get a GPS reading, then spend extra time marking the plot location as precise as possible on a topo map.

<u>Site history</u>: Briefly describe the history of the stand, including type and year of disturbance (e.g. fire, landslides or avalanching, drought, flood, or pest outbreak). Also note the nature and extent of land use such as grazing, timber harvest, or mining.

<u>Unknown plant specimens</u>: List the numbers of any unknown plant specimens, noting any information such as family or genus (if known), important characters, and whether or not there is adequate material for identification. Do not take samples of plants of which there are only a few individuals or which you think may be rare. Document these plants with photographs.

<u>Additional comments</u>: Feel free to note any additional observations of the site, or deviations from the standard sampling protocol. If additional data were recorded, e.g. if tree diameters were measured, please indicate so here.

COARSE FRAGMENTS AND SOIL INFORMATION

<u>Coarse fragments, litter</u>: Estimate the cover class of each size at or near the ground surface averaged over the plot. Always remember to estimate what you actually see on the surface as opposed to what you think is hiding under, organic litter, big rocks, etc. However, rocks, organic litter, or fine material visible under the canopy of shrubs or trees should be included in the cover estimate.

One way to consider this is to assume that all of the components of coarse fragments plus the basal cross-section of living plant stems and trunks (at ground level) will add up to 100%. Thus, estimate the cover value of each of the items in the box on the form for coarse fragments (including the basal area of plant stems) so that they will add up to 100%. Remember that the basal area of plant stems is usually minimal (e.g., if there were 10 trees, each 1 m in diameter at ground level on a 1000 square meter plot, they would cover less than 1% {0.79%} of the plot).

These data are asked for because certain categories of coarse fragments of rock and other materials have been shown to correlate with certain vegetation types and are thus likely influencing the type of vegetation that is growing in a given area. These estimates should be made quickly with the main point to keep in mind being a rough estimate of the relative proportions of different coarse fragments on the plot.

Fines: Fine mineral fragments including sand, silt, soil, "dirt" < 2 mm in diameter

Gravel: rounded and angular fragments 0.2-7.5 cm (0.08 -3 in.) diameter

Cobble: rounded and angular fragments >7.5-25 cm (3 -10 in.) in diameter

Stone: rounded and angular coarse fragments >25 cm-60 cm (10 -24 in.) in diameter

Boulder: rounded and angular coarse fragments >60 cm (>24 in.) in diameter

Bedrock: continuous, exposed, non-transported rock

Litter: extent of undecomposed litter on surface of plot (this includes all organic matter, e.g. fallen logs, branches, and twigs down to needles and leaves).

<u>Soil texture</u>: Record the texture of the upper soil horizon, below the organic layer if one is present. *See attached key and code list.*

Parent Material: Geological parent material of site. See attached code list.

VEGETATION DATA

Assessment of Layers

This first step is described in the CNPS point-intercept transect protocol. Estimates the maximum height for the low and mid layers and the minimum height for the tall layer are recorded. These estimates are made after a quick assessment of the vegetation and its structure. The estimates need not be overly precise and will vary among vegetation types. A caveat: if several relevés are being sampled within the same vegetation type, it is important to be consistent when assigning layers. Some types will have more than three layers (e.g. two tree layers of different maximum height); this should be indicated in the relevé description. However, data are recorded for only three layers (low, mid, and tall). The layer a species occupies will often be determined by growth form, but exceptions do occur. For example, with trees young seedlings may occupy the low layer, saplings the mid layer, and mature individuals the tall layer for some taxa, for example.

Species List

The collection of vegetation data continues with making a comprehensive species list of all vascular plants within the relevé. This list is achieved by meandering through the plot to see all microhabitats. During list development, observers document each taxon present in each layer in

which it occurs separately, recording it on a different line of the data form and noting which layer is represented. This is important for data entry because each layer of each represented taxon will be entered separately. Each individual plant is recorded in only one layer, the layer in which the tallest portion of the individual is found. One should reach a point at which new taxa are added to the list only very slowly, or sporadically. When one has reached that point, the list is probably done.

The following sections explain how to perform the actual relevé, the Estimation of Cover Values. The sections prefaced by bold-faced titles explain the technique, and the sections with regular font titles refer to the steps needed to complete the accompanying Field Form.

Tree dbh (optional)

The CNPS protocol does not require observers to record the diameter at breast height (dbh) of each tree species in the plot. However, the dbh is important in certain studies and may be recorded next to the each tree species name, in the column labeled "Final species determination or Tree dbh". You should measure the tree dbh of every tree trunk that has diameter > or = 10 cm at breast height in the plot, and each measurement should be in centimeters (cm) using a dbh tape measure. For trunks that may be fused below breast height and branched at breast height, each trunk at breast height gets a separate measurement.

Depending on the density of trees in each plot, you can record dbh of trees for every tree trunk in the plot, or you can sub-sample the trunks to estimate dbh for every tree species in relatively dense plots. If you opt to sub-sample, you should do it for each tree species in a representative "quarter" or quadrant of the plot, and then you will come up with an estimated dbh for the entire plot (once data is processed).

When sub-sampling, make sure to denote this as a sub-sample (can note in the Additional comments field) and record the sub-sample of dbh's for each tree species in the appropriate row on the Field Form. Once the data are post-processed and entered into a database, then you will need to record each sub-sampled dbh reading three additional times to come up with a full sample of dbh readings. For example, with a sub-sampled tree dbh of 15 cm, this value of 15 should be entered four times (not just once) when it is entered in the database.

Estimating Cover:

There are many ways to estimate cover. Many people who have been in the cover estimation "business" for a long time can do so quickly and confidently without any props and devices. However, to a novice, it may seem incomprehensible and foolhardy to stand in a meadow of 50 different species of plants and systematically be able to list by cover value each one without actually "measuring" them in some way.

Of course, our minds make thousands of estimates of various types every week. We trust that estimating plant cover can be done by anyone with an open mind and an "eye for nature." It's just another technique to learn.

It is very helpful to work initially with other people who know and are learning the technique. In such a group setting, typically a set of justifications for each person's estimate is made and a

"meeting of the minds" is reached. This consensus approach and the concomitant calibration of each person's internal scales is a very important part of the training for any cover estimate project.

An underlying point to remember is that estimates must provide some level of reliable values that are within <u>acceptable</u> bounds of accuracy. If we require an accuracy level that is beyond the realm of possibility, we will soon reject the method for one more quantitative and repeatable. As with any scientific measurement, the requirement for accuracy in the vegetation data is closely related to the accuracy of the information needed to provide a useful summary of it. Put into more immediate perspective - to allow useful and repeatable analysis of vegetation data, one does not need to estimate down to the exact percent value the cover of a given plant species in a given stand.

This point relates to two facts: there is inherent variability of species cover in any environment. For example, you would not expect to always have 23% *Pinus ponderosa*, 14% *Calocedrus decurrens*, and 11% *Pinus lambertiana* over an understory of 40% *Chamaebatia foliosa*, 3% *Clarkia unguiculata*, and 5% *Galium bolanderi* to define the Ponderosa pine-Incense cedar/mountain misery/bolander bedstraw plant community. Anyone who has looked at plant composition with a discerning eye can see that plants don't space themselves in an environment by such precise rules. Thus, we can safely estimate the representation of species in a stand by relatively broad <u>cover classes</u> (such as <1%, 1-5 %, 5-25%, etc.) rather than precise percentages.

The data analysis we commonly use to classify vegetation into different associations and series (TWINSPAN and various cluster analysis programs, for example) is likewise forgiving. When analyzed by quantitative mutivariate statistics information on species cover responds to coarse differences in cover and presence and absence of species, but not to subtle percentage point differences. This has been proven time and again through quantitative analysis of vegetation classification. Many of the world's plant ecologists estimate cover rather than measure it precisely. Some of the seminal works in vegetation ecology have been based on cover estimates taken by discerning eyes.

With this as a preamble, below we offer some suggestions on estimating cover that have proven helpful. These are simply "tricks" to facilitate estimation, some work better for different situations. You may come up with other methods of estimation that may seem more intuitive, and are equally reliable in certain settings. All values on the relevé protocol that require a cover class estimate, including coarse fragment and vegetation layer information, may rely on these techniques. Just make the appropriate substitutions (using the coarse fragment example substitute, bedrock, stone, cobbles, gravel, and litter for vegetation).

Method 1: The invisible point-intercept transect:

This method works well in relatively low, open vegetation types such as grasslands and scrubs where you can see over the major stand components. For those who have worked with the original CNPS line intercept methodology it's like counting hits along an imaginary line at regular intervals of the 50 m tape. Here's how it goes:

Envision an imaginary transect line starting from your vantage point and running for 50 m (or however many meters you wish, as long as you are still ending up within the same

stand of vegetation you're sampling - <u>never</u> keep counting outside of your homogeneous stand). Now "walk" your eye along this tape for 50 m and visually "take a point" every 0.5 m. Don't worry about precision, just try to "walk" your eye along the line and stop every 0.5 m or at any other regular interval until you reach its end and mentally tally what species you hit. Once you come up with a number of hits for each major species in one imaginary transect, take another transect in another direction and estimate the number of hits on that one. Do this several times (usually 3-4 is enough if you are in a homogeneous stand), then average your results.

This can go quickly in simple environments and in environments where the major species are easily discernable (chaparral, bunch-grassland, coastal scrub, desert scrub). Your average number of hits need not be a total of 100 as in the original transect method, but could be 50 along a 25 m imaginary line (in which case you would multiply by two to get your estimated cover), or 25 along a 12.5 m line (multiply average by 4), etc.

Method 2: Subdivision of sample plot into quadrants:

Many plots, whether they are square, circular, or rectangular, may be "quartered" and have each quadrant's plant cover estimated separately. If the plot is a given even number of square meters (such as 100, 400, or 1000 m²) then you know that a quarter of that amount is also an easily measurable number. If you can estimate the average size of the plants in each of the quarters (e.g, small pinyon pines may be 5 m^2 (2.2m x 2.2m), creosote bush may be 2m^2 (or 1.41 m x 1.41 m), burrobush may be 0.5m^2) then you simply count the number of plants in each size class and multiply by their estimated size for the cover in a given quadrant. Then you average the 4 quadrants together for your average cover value.

This method works well in vegetation with open-to-dense cover of low species such as grasses or low shrubs, in open woodlands, and desert scrubs.

Method 3; "Squash" all plants into a continuous cover in one corner of the plot :

Another way to estimate how much of the plot is covered by a particular species is to mentally group (or "march", or "squash") all members of that species into a corner of the plot and estimate the area they cover. Then calculate that area as a percentage of the total plot area. This technique works well in herb and shrub dominated plots but is not very useful in areas with trees.

Method 4: How to estimate tree cover:

Cover estimates of tall trees is one of the most difficult tasks for a beginning relevé sampler. However it is possible to do this with consistency and reliability using the following guidelines.

- 1. Have regular sized and shaped plots that you can easily subdivide.
- 2. Estimate average crown spread of each tree species separately by pacing the crown diameter of representative examples of trees of each species and then roughly calculating the crown area of each representative species.

3. Add together the estimated crown area of each individual of each species of tree on the plot for your total cover.

Method 5: The process of elimination technique:

This method is generally good for estimating cover on sparsely vegetated areas where bare ground, rocks, or cobbles cover more area than vegetation. In such a situation it would be advisable to first estimate how much of the ground is not covered by plants and then subdivide the portion that is covered by plants into rough percentages proportional to the different plant species present. For example, in a desert scrub the total plot not covered by plants may be estimated at 80%. Of the 20% covered by plants, half is desert sunflower (10% cover), a quarter is California buckwheat (5% cover), an eighth brittlebush (2.5% cover), and the rest divided up between 10 species of herbs and small shrubs (all less than 1% cover).

Any of these techniques may be used in combination with one another for a system of checks and balances, or in stands that have characteristics lending themselves for a different technique for each layer of vegetation.

In a relevé, cover estimates, using the techniques described above, are made for each taxon as it is recorded on the species list. Estimates are made for each layer in which the taxon was recorded. For example, if individuals of coast live oak occur in the tall, the mid, and the low layer, an estimate is made for Tall CLO, for mid CLO, and for low CLO.

In a traditional relevé, cover is estimated in "cover classes," not percentages, because of the variability of plant populations over time and from one point to another, even within a small stand. This protocol uses the following 6 cover classes:

Cover Class 1: the taxon in that layer covers < 1 % of the plot area Cover Class 2: the taxon in that layer covers >1 % - 5 % of the plot area Cover Class 3a: the taxon in that layer covers >5 - 15 % of the plot area Cover Class 3b: the taxon in that layer covers >15 - 25 % of the plot area Cover Class 4: the taxon in that layer covers >25 - 50 % of the plot area Cover Class 5: the taxon in that layer covers >50 - 75 % of the plot area Cover Class 6: the taxon in that layer covers >75% of the plot area

Percentages (optional)

This CNPS protocol also encourages observers to estimate percentages if they feel confident in their estimation abilities. This optional step allows the data to be compared more easily to data collected using different methods, such as a line or point intercept. It also instills confidence in the cover estimate of borderline species that are close calls between two cover classes (e.g., a cover class 2 at 5% as opposed to a cover class 3 at 6%). It is particularly useful for calculating cover by the process of elimination techniques and for estimating total vegetation cover (see below) and coarse fragment cover.

Total Vegetation Cover by Layer

In addition to cover of individual taxa described above, total cover is also estimated for each vegetation layer (e.g. tall, medium, low). This is done using the same cover classes as described above but combines all taxa of a given category. They can be calculated from the species percent cover estimates, but please make sure to disregard overlap of species within each layer. These estimates should be absolute aerial cover, or the "bird's eye view" of the vegetation cover, in which each category cannot be over 100%.

National Vegetation Classification height Classes for Vegetation Strata

The relevé method just described calls for estimates of plant cover for each taxon. It is strongly floristically oriented. Another way of considering the relationships between plants in vegetation is by evaluating structure, or physiognomy. The underlying thinking is that life forms within a stand of vegetation occur in response to similar ecological pressures (TNC 1998). Estimation of cover within predetermined height classes is one way to describe the structure of vegetation. Structure of a stand of vegetation also is used in modeling wildlife use of the vegetation (WHR).

For information gathered using this CNPS protocol to be comparable with the wealth of information being gathered by the National Park Service and the Biological Resources Division (BRD) of the USGS it is also necessary for CNPS to estimate vegetation cover according to predefined vegetation strata. The following height classes are defined by the USGS/NPS:

High Tree	>30 m
Medium High Tree	20-30 m
Medium Low Tree	10-20 m
Low Tree	5-10 m
High Shrub	2-5 m
High Herb/ Medium Shrub	1-2 m
Low Shrub	0.5-1 m
Medium Herb	25-50 cm
Low Herb	0-25 cm
Moss/Lichen	

Cover in these vegetation strata is estimated using the same cover classes as were used for cover of individual taxa. Again, estimation of percentages is optional. Please note that although these strata have names in the national classification, they don't necessarily have to be populated by the type of species that are their namesake (e.g., tall herbaceous species may be diagnostic of the tall shrub category in the case of a giant reed stand). For this reason we have simply listed the strata by their height classes and have opted not to name them.

We have also requested that you list the diagnostic species for each layer. In this case the diagnostic species is the single species that seems to best characterize that layer it may be the only species found in a given layer, it may be as common as other species in that layer but is more restricted to that single layer, or it may be less common than other species in that layer, but so representative of that layer that it can't be ignored. The cover of the diagnostic species in that layer does not have to be re-estimated as it is estimated in the individual species tally already.

Caveats:

Please consult with the members of the vegetation committee for advice and feedback on proposed vegetation surveys prior on initiating projects.

<u>Notes on the Order and Division of Labor for Data Collection</u>: As with every procedure there are always more and less efficient ways to collect the information requested. Although we respect each field crews' option to choose in what order they collect the data, we suggest the following general rules:

- Work with teams of two for each plot collected.
- Both team members can determine the plot shape and size and lay out the tapes and mark the edges for the plot boundary (see below).
- The two person teams can also divide up tasks of data collection with one member collecting location, environmental (slope, aspect, geology, soil texture, etc.) and plot description information while the other begins the species list. Thus, two clipboards are useful and data sheets that are at first separated (not stapled).
- Following the making of the initial species list and collection of location and environmental data both team members convene to do the estimation of plant cover by species followed by the estimation of total vegetation cover and cover by layer.
- Following that process, the estimation of cover by the up to 10 height strata classes and the listing of the diagnostic species for each is done collaboratively.
- This is followed by the estimation of the coarse fragment information, again done collaboratively.

For egalitarian and familiarization purposes we suggest that the roles be switched regularly between the team members and that if multiple teams are being used in a larger project, that each team member switches frequently between teams, building all-important calibration, and camaraderie among the whole group.

<u>Suggestions for Laying out Plots</u>: If you are laying out a circular plot, work with two or more people. One person stands at the center of the plot and holds the tape case while the other walks the end of the tape out to the appointed distance (radium 5.6 for 100 m^2 circle, radius 11.3 m for a 400 m^2 circle, and radius 17.6 m for a 1000m^2 circle). The walker then fixes the tape end with a pin flag and walks back to the center where he/she instructs the center person to walk in the opposite direction of the already laid out tape radius, stretching the rest of the tape to an equal length (another 11.3 or 17.6 m) to the opposite edge of the plot, where he/she affixes it with another pin flag. This process is again repeated with another tape laid out perpendicular to the

first so that an "+" shape is created. The margins of the circle can be further delineated by

measuring to the center of the circle with an optical tape measure (rangefinder) and marking mid points between the four ends of the crossed tapes.

When laying out square or rectangular plots work with two or more people per team. If doing a rectangle, determine the long axis of the plot first and have one person be stationed at the zero m end of the tape while the other person walks the unrolling tape case out to the appropriate length. The stationary end person can guide the walker, keeping them moving in a straight line. Once that tape is laid out and the far end staked, the team lays out another tape perpendicular to the first, either at one end, using the same type of process. This establishes the width of the rectangle (or square). Using an optical rangefinder and pin-flags, or colored flagging the team can further mark additional points along the other parallel long axis and short axis of the plot (every 5 m for shorter plots or every 10 m for longer plots is suggested) so that the entire plot boundary can be easily visualized.

References:

Barbour M.G., J.H. Burk, and W.D. Pitts 1987. Terrestrial Plant Ecology, Second Edition. Benjamin/Cummings Publishing Co. Menlo Park, CA. 634 pages.

Sawyer and Keeler-Wolf. 1995. Manual of California Vegetation. California Native Plant Society, Sacramento, CA. 471 pages

The Nature Conservancy and Environmental Systems Research Institute. 1994. Final Draft, Standardized National Vegetation Classification System. Prepared for United States Department of the Interior, National Biological Survey, and National Park Service. Arlington, VA. Complete document available at the following website: http://biology.usgs.gov/npsveg/fieldmethods.html

Suggested Equipment:

Equipment List: Prices as of May 2000, toll free orders from Forestry Suppliers (1-800-647-5368) (item numbers in parentheses)

Chaining pins, surveyor steel (#39167) \$2.	1.50
Fiberglass tapes 2 - 165'/50 m (#39972) \$42	2.90
Logbook cover 8 ¹ / ₂ " x 12" (#53200) \$23	3.95
Perforated flagging (#57960) \$1.	.95
UTM Coordinate Grid (#45019) \$10	5.95
Rangefinder, 10-75m (#38973) \$5	1.60
Silva Compass w/ clinometer (#37036) \$43	3.90
Garmin GPS 12XL (#39095, #39111) \$24	44.90

Simplified Key to Soil Texture (Brewer and McCann, 1982)

Place about three teaspoons of soil in the palm of your hand. Take out any particles >2mm in size, and use the following key to figure out the soil texture (e.g. loamy sand). Then figure out the texture subclass by using the Code List attached (e.g. coarse loamy sand).

A1	Soil does not remain in a ball when squeezed sand
A2	Soil remains in a ball when squeezed B
B1	Add a small amount of water. Squeeze the ball between your thumb and forefinger, attempting to make a ribbon that you push up over your finger. Soil makes no ribbonloamy sand
B2	Soil makes a ribbon; may be very shortC
C1	Ribbon extends less than 1 inch before breakingD
C2	Ribbon extends 1 inch or more before breakingE
D1	Add excess water to small amount of soil; soil feels very gritty or at least slightly grittyloam or sandy loam
D2	Soil feels smoothsilt loam
E1	Soil makes a ribbon that breaks when 1–2 inches long; cracks if bent into a ringF
E2	Soil makes a ribbon 2+ inches long; does not crack when bent into a ringG
F1	Add excess water to small amount of soil; soil feels very gritty or at least slightly grittysandy clay loam or clay loam
F2	Soil feels smoothsilty clay loam or silt
G1	Add excess water to a small amount of soil; soil feels gritty or at least slightly grittysandy clay or clay
G2	Soil feels smoothsilty clay

M	ACRO TOPOGRAPHY
00	Bench
01	Ridge top (interfluve)
02	Upper 1/3 of slope
03	Middle 1/3 of slope
04	Lower 1/3 of slope (lowslope)
05	Toeslope (alluvial fan/bajada)
06	Bottom/plain
07	Basin/wetland
08	Draw
09	Other
10	Terrace (former shoreline or floodplain)
11	Entire slope
12	Wash (channel bed)
13	Badland (complex of draws & interfluves
14	Mesa/plateau
15	Dune/sandfield
16	Pediment
17	Backslope (cliff)
MI	CRO TOPOGRAPHY
01	Convex or rounded
02	Linear or even
03	Concave or depression
04	Undulating pattern
05	Hummock or Swale pattern
06	Mounded
07	Other

SITE IMPACTS

01	Development
02	ORV activity
03	Agriculture
04	Grazing
05	Competition from exotics
06	Logging
07	Insufficient population/stand size
08	Altered flood/tidal regime
09	Mining
10	Hybridization
11	Groundwater pumping
12	Dam/inundation
13	Other
14	Surface water diversion
15	Road/trail construction/maint.
16	Biocides
17	Pollution
18	Unknown
19	Vandalism/dumping/litter
20	Foot traffic/trampling
21	Improper burning regime
22	Over collecting/poaching
23	Erosion/runoff
24	Altered thermal regime
25	Landfill
26	Degrading water quality
27	Wood cutting
28	Military operations
29	Recreational use (non ORV)
30	Nest parasitism
31	Non-native predators
32	Rip-rap, bank protection
33	Channelization (human caused)
34	Feral pigs
35	Burros

36 Rills37 Phytogenic mounding

RO TOPOGRAPHY	PARENT	MATERIAL	SOI
nch	ANDE	Andesite	COS
lge top (interfluve)	ASHT	Ash (of any origin)	ME
per 1/3 of slope	GRAN	Granitic (generic)	FISI
udie 1/5 of slope (lowslope)	DIOP	Diorite	ME
eslone (alluvial fan/baiada)	BASA	Basalt	MC
ttom/plain	OBSI	Obsidian	ME
sin/wetland	PUMI	Pumice	ME
aw	IGTU	Igneous (type unknown)	ME
ner	MONZ	Monzonite	ME
rrace (former shoreline or floodplain)	PYFL	Pyroclastic flow	MF
tire slope	QUDI	Quartz diorite	MFS
ish (channel bed)	RHYO	Rhyolite	MFS
dland (complex of draws & interfluves)	VOLC	General volcanic extrusives	FISA
sa/plateau	VOFL	Volcanic flow	FISC
liment	BLUE	Blue schist	SAN
ckslone (cliff)	CHER	Chert	LOA
ekslope (enn)	DOLO	Dolomite	CLA
O TOPOGRAPHY	FRME	Franciscan melange	UNI
nvex or rounded	INTR	General igneous intrusives	PEA
lear or even	GNBG	Gneiss/biotite gneiss	MU
ncave or depression	HORN	Hornfels	
dulating pattern	MARB	Marble	DO
mmock or Swale pattern	METU	Metamorphic (type unknown)	Tree
ounded	PHYL	Phyllite	TBS
ner	SCHI	Somi sobist	TNI
	SLAT	Slate	1111
	BREC	Breccia (non-volcanic)	CDI
IMPACTS	CACO	Calcareous conglomerate	MN
velopment	CASA	Calcareous sandstone	
Vactivity	CASH	Calcareous shale	TBE
riculture	CASI	Calcareous siltstone	
azing	CONG	Conglomerate	TNE
mpetition from exotics	FANG	Fanglomerate	
gging	GLTI	Glacial till, mixed origin, moraine	EXI
ufficient population/stand size	LALA	Large landslide (unconsolidated)	CDI
ered flood/tidal regime	SAND	Limestone	EVI
ning	SAND	Sedimentary (type unknown)	LAI
origization	SHAL	Shale	MB
m/inundation	SILT	Siltstone	
her	DIAB	Diabase	MN
face water diversion	GABB	Gabbro	
ad/trail construction/maint.	PERI	Peridotite	Shri
ocides	SERP	Serpentine	TBE
lution	ULTU	Ultramafic (type unknown)	
known	CALU	Calcareous (origin unknown)	NLE
ndalism/dumping/litter	DUNE	Sand dunes	MIE
ot traffic/trampling	LUSS	Loess Mixed improve	EXI
proper burning regime	MIG	Mixed igneous Mixed metamorphic	CDS
er collecting/poacning	MIRT	Mix of two or more rock types	ME
ared thermal regime	MISE	Mixed sedimentary	XM
adfill	CLAL	Clayey alluvium	
grading water quality	GRAL	Gravelly alluvium	Dwa
ood cutting	MIAL	Mixed alluvium	NM
litary operations	SAAL	Sandy alluvium (most alluvial fans	
creational use (non ORV)		and washes)	XEI
st parasitism	SIAL	Silty alluvium	
n-native predators	OTHE	Other than on list	DDI
-ran bank protection			ME

DE LIST	(revised 7/8/02)				
OIL TEXTURE					
OSA	Coarse sand				
ESN	Medium sand				
ISN	Fine sand				
OLS	Coarse, loamy sand				
ELS	Medium to very fine, loamy sand				
CSL	Moderately coarse, sandy loam				
ESA	Medium to very fine, sandy loam				
ELO	Medium loam				
ESL	Medium silt loam				
ESI	Medium silt				
FCL	Moderately fine clay loam				
FSA	Moderately fine sandy clay loam				
FSL	Moderately fine silty clay loam				
ISA	Fine sandy clay				
ISC	Fine silty clay				
ICL	Fine clay				
AND	Sand (class unknown)				
OAM	Loam (class unknown)				
LAY	Clay (class unknown)				
NKN	Unknown				
EAT	Peat				
UCK	Muck				
OMINAN	T VEGETATION GROUP				
rees:					
BSE	Temperate broad-leaved seasonal				
	evergreen forest				
NLE	Temperate or subpolar needle-leafed				
	evergreen forest				
DF	Cold-deciduous forest				
NDF	Mixed needle-leafed evergreen-cold				
	deciduous. forest				

MNDF	Mixed needle-leafed evergreen-cold deciduous forest				
TBEW	Temperate broad-leaved evergreen				
TNEW	Temperate or subpolar needle-leaved				
EXEW	Extremely veromorphic evergreen				
LALW	woodland				
CDW	Cold-deciduous woodland				
EXDW	Extremely xeromorphic deciduous				
	woodland				
MBED	Mixed broad-leaved evergreen-cold				
	deciduous woodland				
MNDW	Mixed needle-leafed evergreen-cold				
	deciduous woodland				
Shrubs:					
TBES	Temperate broad-leaved evergreen				
	shrubland				
NLES	Needle-leafed evergreen shrubland				
MIES	Microphyllus evergreen shrubland				
EXDS	Extremely xeromorphic deciduous				
	shrubland				
CDS	Cold-deciduous shrubland				
MEDS	Mixed evergreen-deciduous shrubland				
XMED	Extremely xeromorphic mixed evergreen-				
	deciduous shrubland				
Dwarf Shru	ibland:				
NMED	Needle-leafed or microphyllous evergreen				
VEDC	dwarf shrubland				
XEDS	Extremely xeromorphic evergreen dwarf				
DDDC	Shrubland				
MEDD	Mixed everyment cold decidence dworf				
MEDD	shrubland				
Harbacoou	sin ubland				
TSPG	Temperate or subpolar grassland				
TGST	Temperate or subpolar grassland with				
1051	sparse free				
TGSS	Temperate or subpolar grassland with				
	sparse shrublaver				
TGSD	Temperate or subpolar grassland with				
	sparse dwarf shrub layer				
TFV	Temperate or subpolar forb vegetation				
THRV	Temperate or subpolar hydromorphic				
	rooted vegetation				
TAGF	Temperate or subpolar annual grassland or				
	forb vegetation				
Sparse Vegetation:					
SVSD	Sparsely vegetated sand dunes				
SVCS	Sparsely vegetated consolidated substrates				