

EVALUATION OF MEASUREMENT ENDPOINTS FOR THE ASSESSMENT OF POTENTIAL  
PETROLEUM RELATED RESPONSES BY PACIFIC HERRING: EARLY DEVELOPMENT

FINAL DRAFT REPORT

JANUARY 21, 2009

SUBMITTED TO:

WALTER PEARSON  
PEAPOD RESEARCH

AND

GARY MAUSETH  
POLARIS APPLIED SCIENCES, INC.  
12525 - 131ST COURT NE  
KIRKLAND, WA 98034

SUBMITTED BY:

NEWFIELDS  
4729 NE VIEW DRIVE  
PORT GAMBLE, WA 98364

NEWFIELDS

# EVALUATION OF MEASUREMENT ENDPOINTS FOR THE ASSESSMENT OF POTENTIAL PETROLEUM RELATED RESPONSES BY PACIFIC HERRING: EARLY DEVELOPMENT

*This report is a draft of the information and conclusions made to date.*

## Introduction

On 7 November 2007 the container ship *Cosco Busan* struck a tower supporting San Francisco's Bay Bridge spilling an estimated 54,000 gallons of bunker fuel into the bay. The spill contaminated shorelines in the North Central Bay where Pacific herring would spawn in upcoming months. Historical spills in areas of Pacific herring spawning have demonstrated concordance with injury to the developing eggs and newly hatched individuals from shallow subtidal and intertidal habitats. Based on historical precedence, the importance of herring to the ecology of the bay, and their relative sensitivity to petroleum contamination, this species was selected for further examination to evaluate potential damage to natural resources in the bay. The plan was to 1) study and compare biological responses of herring embryos and larvae incubated in areas receiving different degrees of oiling from the accident and 2) to characterize the degree of exposure to polynuclear aromatic hydrocarbons (PAH), potentially resulting from the *Cosco Busan*.

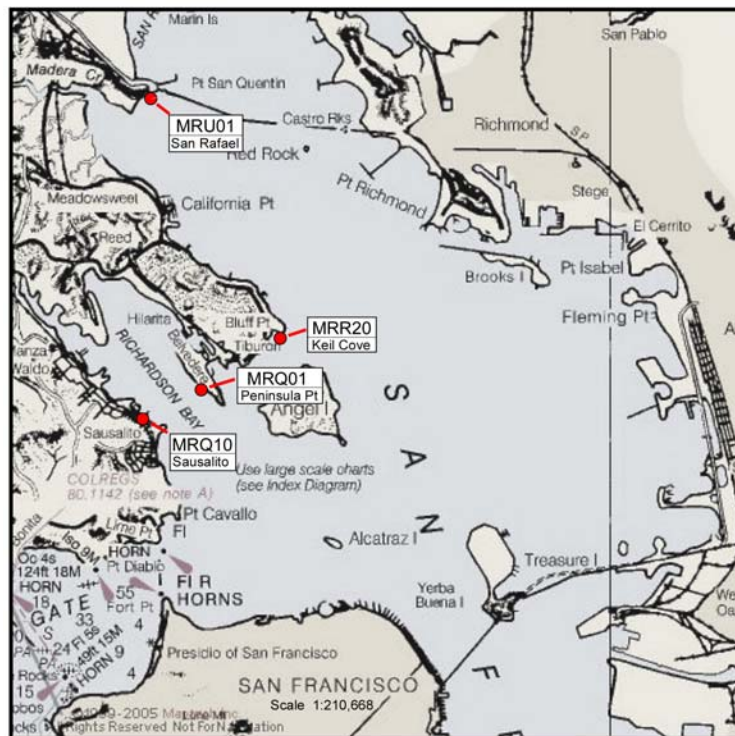
Two approaches were used to assess potential damage to herring embryos from oil exposure. The first was to sample developing eggs/embryos naturally fertilized and deposited on marine vegetation along shorelines with exposure to varying degrees of the spilled bunker fuel as well as from a control site. The second was to use laboratory-fertilized eggs deployed in cages placed in shallow subtidal areas in four locations with varying degrees of exposure to the spilled bunker fuel and two reference sites. Embryos were scored for various morphological measurements and tissues were analyzed in test organisms as well as in Polyethylene Membrane Devices (PEMD) placed with the cages. Two organizations evaluated newly hatched embryos (NOAA Fisheries and the Bodega Marine Laboratory). Data collection involved the creation of still images and video clips of live specimens. NewFields NW was asked to review the digital information and provide similar evaluations as those performed by either NOAA or BML. NewFields personnel were not informed about the locations of the samples being reviewed. Data summaries and a draft letter report were prepared by NewFields and submitted to Dr. Walter Pearson and Polaris Applied Sciences for review. After reviewing the draft letter report, NewFields was then asked to compare these data summaries to selected locations within San Francisco Bay and to evaluate whether the observations correlated with generalized descriptions of beach oiling provided by NOAA. Specific questions that were formulated as part of the review process included:

1. What is the inherent variability in the pericardial edema measurements?
2. Is more than one developmental stage represented for the artificial and/or naturally spawned embryos? If so, what are the developmental stages and how many are represented?
3. Does the attribute of yolk sac flattened (anterior edge next to pericardial space) appear in the artificial and/or naturally spawned samples? If so, what is the incidence of this attribute?
4. Are notochord abnormalities associated with the coiling or bent body axes observed in the naturally spawned samples? If so, what is the incidence of this attribute?

5. Are abnormalities other than the pericardial edema observed in the artificial spawned samples?
6. Is opacity of tissues in the natural and artificially spawned embryos an artifact of varying illumination? Is there a correlation between the background illumination level and opacity of tissues of the embryos?
7. What is the frequency of arrhythmia in artificial and naturally spawned embryos? What types of arrhythmia are present?
8. Does the frequency of abnormalities in the artificially spawned samples match those from the naturally spawned embryos?

This report describes procedures used by NewFields along with summaries of the data reviewed on a subset of the information for naturally spawned and artificially spawned samples, and an evaluation of statistical significance among and between samples collected at different sites within the bay. The summary also addresses the differences in these characteristics in association with the qualitative description of oiling along shorelines. A discussion is then provided that addresses the concordance of oiling and experimental artifacts that may be associated with the observed effects.

Location: Data were evaluated from video clips and still images of individual specimens collected from artificial or natural spawning from four locations in San Francisco Bay, California. The station designations and general locations are provided in Figure 1 and Table 1.



**FIGURE 1. Location of Sites Evaluated**

**TABLE 1. Station Designations, Location and Ranked Level of Intertidal and Shallow Subtidal Oiling** (NOAA and Bodega Marine Laboratory, University of California-Davis, 2008).

Station Designation	Location	Rank Order of Oiling
MRU01	San Rafael Bay	0
MRQ10/P01	Sausalito	1
MRQ01	Peninsula Point	2
MRR20	Keil Cove	3

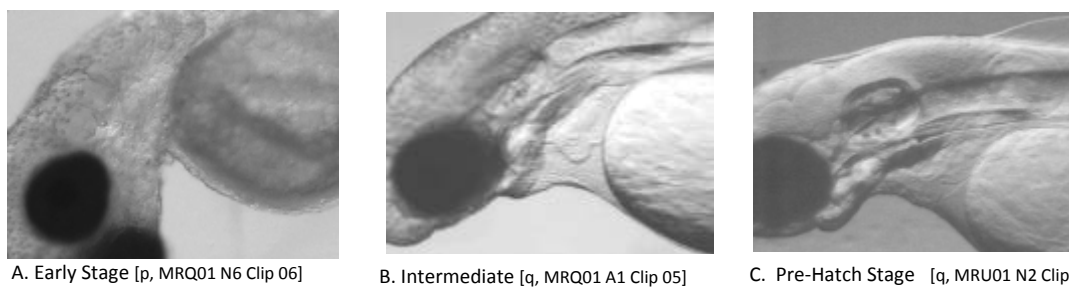
## Data Review Methods

### Morphological Characteristics

Late stage embryogenesis characteristics defined by Hill and Johnson (1997) provided the basis for assessing developmental stage through evaluation of specific characteristics such as degree of eye pigmentation, jaw and otic vesicle development, yolk sac features, pericardium shape and location, tail development, and caudal fin characteristics. Still images were previewed for selected files representing natural and artificial spawn conditions to assess gross morphological features such as planar view, degree of coiling, and relative development of tail region while video clips provided for a more detailed evaluation of head, pericardium and yolk sac regions. A description of the otic vesicle, eye pigmentation, notochord differentiation, pericardial space, heart rate and shape, heart patterns (arrhythmia), yolk sac shape and color were summarized for selected video samples and are included in Appendix A.

### Assessment of Variability

*Developmental Stage.* A subset of video files from each station location representing both natural and artificial spawned and dechorionated embryos was evaluated. Unfortunately, lack of still and video image linkage precludes a full evaluation of developmental characteristics. However, an overview of the selected embryos indicates a range of developmental growth transitioning between the organogenesis stages p, q, and r as described by Hill and Johnson. Figure 2 presents examples of the developmental stages (p-q) encountered in our review; Table 2 summarizes distinctive characteristics.

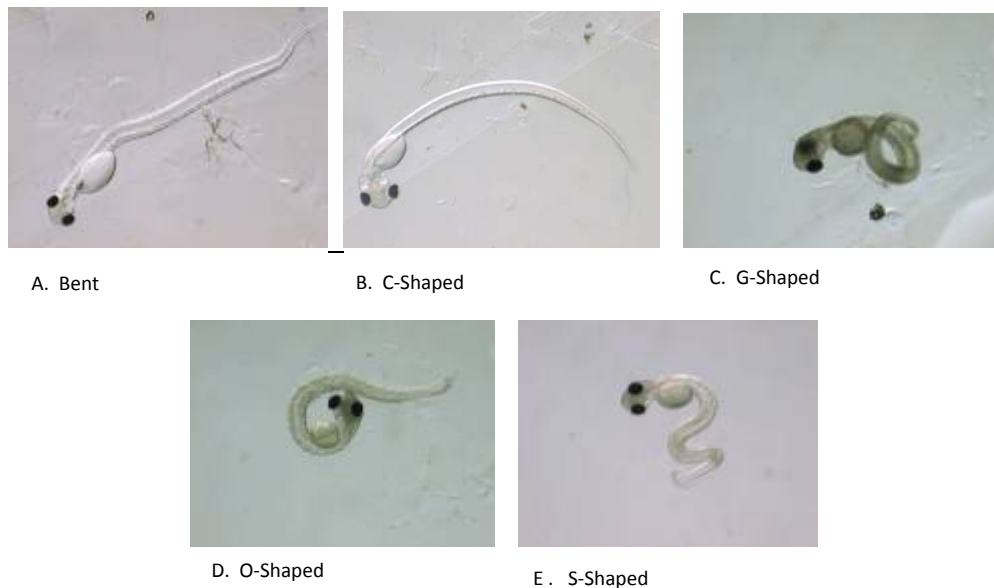


**FIGURE 2. Range of Developmental Characteristics**

**Table 2. Notable Characteristics of Pre-Hatch Developmental Stages** (based on Hill and Johnston 1997)

Diagnostic Characteristic	Early (p-Stage)	Intermediate to Pre-Hatch (q-Stage)
Days Post Fertilization (dpf)	7 - 10 dpf	11- 14 dpf
Length	3- 5 mm	5-7 mm
Eye Pigmentation	10% - 75%	85 – 100%
Head/Trunk Angle (HTA)	50°	130° - 145°
Hatching glands	Cover dorsal regions of head	Hatching glands release contents
Otic Vesicle	Otoliths developed; primordia of semi-circular canals develop	Semi-circular canals in otic vesicles are more prominent
Pectoral Fin	Pectoral fin bud develops	Pectoral fin develops into functional structure
Jaw Development	Rudimentary	Protrusion evident, may fully extend under eye
Pericardium Shape, and Orientation	Elongated; anterior portion of pericardial space	Atrium follows vertical axis and ventricle follows horizontal axis, oriented in center of pericardial region
Yolk sac shape	Rounded	Elliptical
Notochord development	Rudimentary – segmentation not clearly defined	Segmentation notable

*Specimen Orientation.* Many still images from the natural spawn groups demonstrated various degrees of coiling, generally considered a sign of abnormal development. We noted basic shape differences as shown in Figure 3. However, it should be clarified whether presence of early developmental stage at time of dechoriation may contribute to some degree of residual coiling, and/or whether salinity stress (i.e., ambient salinity at time of collection versus laboratory salinity) may have impacted progression of developmental stages.



**Figure 3. Various Specimen Orientations**

Some instances of anomalous notochord development were observed (Figure 4); however these images were not directly associated with corresponding video recordings, and thus a full assessment of abnormality could not be made. Based on the description of abnormality of notochord associated with a 90° bend in the notochord beyond the dorsal, ventral, or lateral axis (Hershberger et al. 2005), or a constriction of the notochord, the specimens presented in Figure 4 are probably anomalous. We reviewed the still images from each site and summarized the frequency of body axis deviations from the horizontal plane, commonly encountered in review of naturally spawned larvae. This is a qualitative assessment, and caution should be exercised when interpreting these data. A quantitative evaluation of frequency of anomalies occurring to the notochord is very restricted due to the lack of appropriate resolution of the full specimen still images.



**Figure 4. Anomalous Notochord Development**

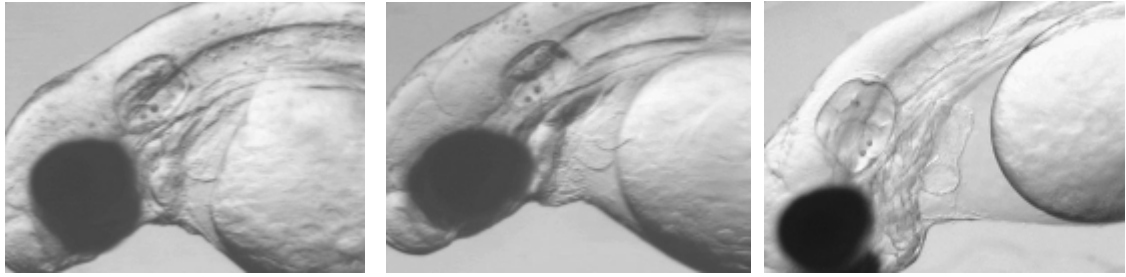
*Illumination.* An assessment of the variability of illumination was made on a select subset of video clips. Points were measured from the background, the head region (including the otic vesicle), the eye, and the yolk sac using ImageJ software to analyze median gray values. The degree of illumination and pigmentation in these regions of the individual specimens were compared using regression analysis.

A cursory scan of the full image collection indicated that the still and video recordings were made under various lighting conditions. This variation in illumination makes an evaluation of relative opacity of the various areas of the specimen problematical. Variation in illumination encountered during review of the still images is exemplified in Figure 5. Very frequently, a gold-tone cast to specimen captured in still photos was noted, particularly if there was some degree of coiling in body orientation (e.g., photo at far right below).



**Figure 5. Variation in Background Illumination in Still Images**

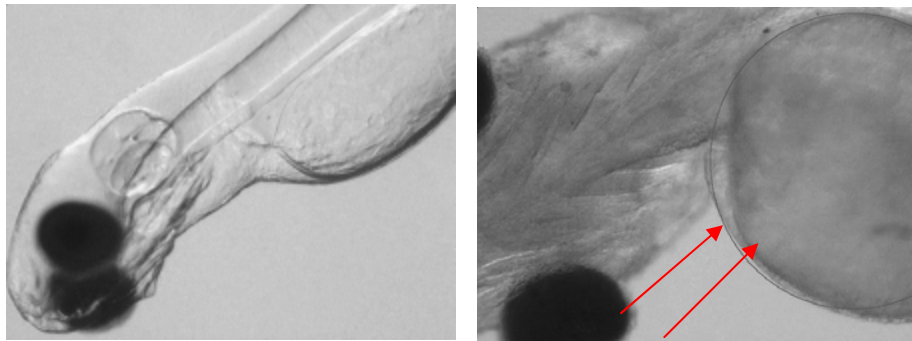
*Position of the Heart Relative to Yolk Sac* A large variation in the size of the pericardium and position of the heart relative to the yolk sac was observed in the subset of video recordings included in this review (Figure 6; flattening of the yolk sac may or may not be evident).



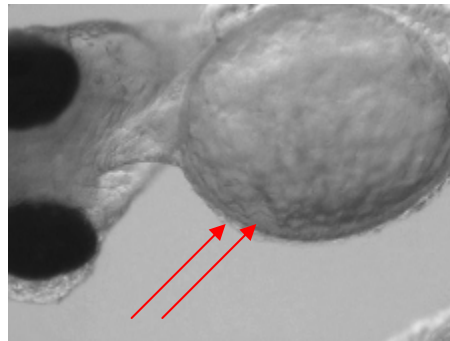
A. Compressed Pericardium with Flattening [MRQ10 A3 Clip 32]      B. Overlap of Heart onto Yolk Sac without Flattening [MRQ10 A5 Clip 20]      C. Large Pericardium with Anterior Orientation of Heart [MRR20 N4 Clip 09]

**FIGURE 6. Variations in Pericardial Region**

In addition, instances of secondary membranes were noted in the pericardial region as well as the yolk sac as shown in Figure 7. The determination of flattened yolk sac was based on a deviation from the round or elliptical shape. In cases where flattening was not easily observed, Image J software was used to make a rounded or elliptical tool to follow the outline of the yolk sac. There are two associations of flattening with an extra space around the yolk sac. In the first instance, flattening occurs in an absence of an extra clear space around the yolk sac, and in the later case it occurs in the presence of a extra space around the yolk sac “edema”. Figure 7B illustrates yolk sac flattening associated with an extra space around yolk sac. The observations are recorded in Appendix A5.



A. Pericardial Membrane [MRR20 N7 Clip 57]      B. Yolk Sac Membrane with Flattening [MRQ N6 Clip 09]

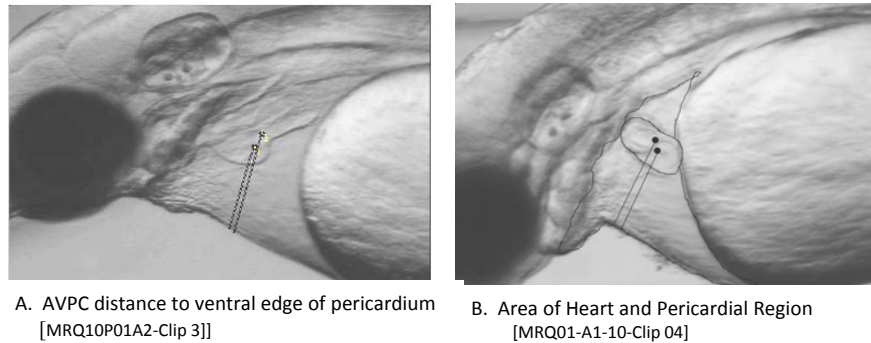


C. Yolk Sac Membrane without Flattening [MRQ01 N6 Clip 22]

**FIGURE 7. Additional Observations**

The distance of the heart atrioventricular junction (AV) to the ventral edge of pericardial space was measured using a perpendicular line for subset of videos using ImageJ software. Measurements were taken on the downbeat and the upbeat of the heart and differences in measurement were assessed statistically. In addition, the area of the heart and pericardial space

were measured using ImageJ software. The data have been summarized and provided in Appendix A and are illustrated in Figure 8 A and B. The variation in distance based on position of the heartbeat was determined for selected video clips for each of the station/areas indicated.



**Figure 8. Measurement of Distance from Upbeat/Downbeat AV to PC (A); Measurement of Area of Heart and Pericardium (B)**

*Heart Beat and Arrhythmia.* The number of heart beats for 20 second intervals was counted and recorded for each video clip evaluated. Incidence of arrhythmia was determined as the presence of an irregular beating or compression of the atrium or ventricle during a 20 second video clip.

*Heart and Pericardial Space Areas.* Computer based planimetry was employed using ImageJ to determine the area of heart, the area of pericardial space, and the percentage of pericardial space occupied by the heart.

#### Statistical Evaluation Procedures

ANOVA was performed on each of the quantified data sets. Linear regression was performed on the associations between background illumination and density of pigmentation in tissues as well as regressions of the measurements to the rank order of qualitative oiling.

## **Results**

### Morphological Characteristics: Still Image Documentation (Low Resolution)

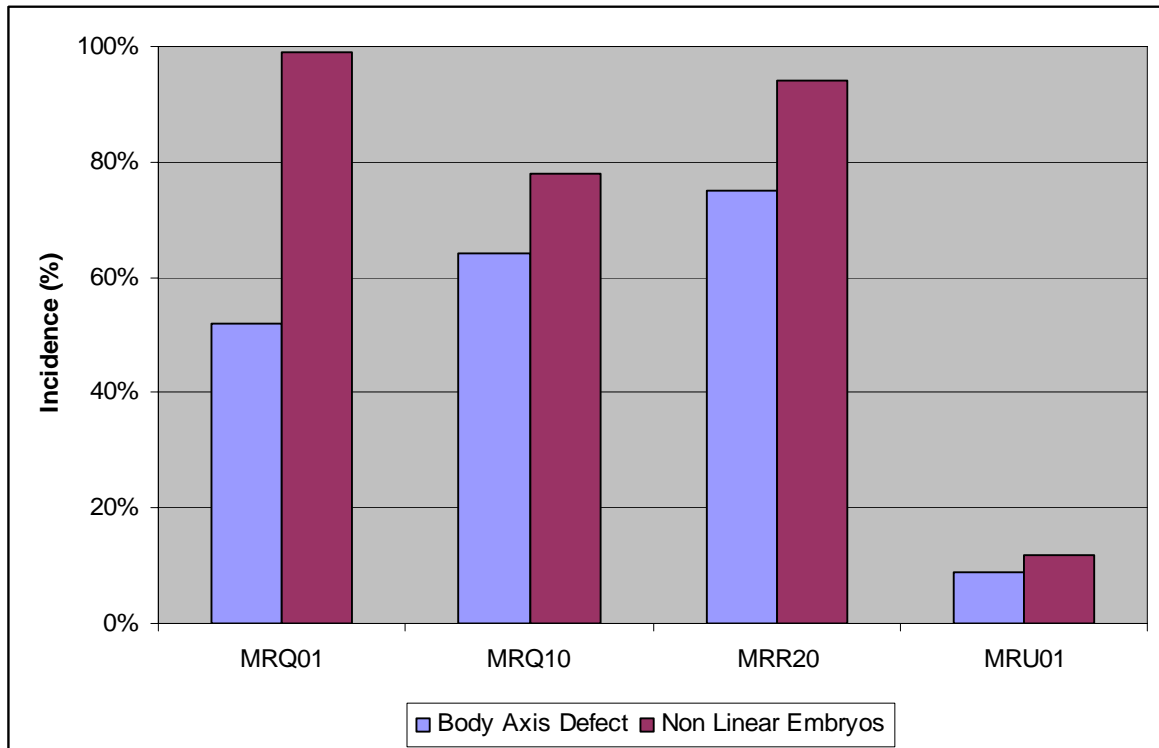
The full set of still images for naturally spawned embryos was assessed for body axis deviations. We recorded the various degrees of coiling as C-, O-, or S-shaped; non-coiled specimen were considered linear; and those with  $>90^\circ$  bends along the notochord or other anomalous aspects to notochord development were grouped as “body axis defect”. These results are summarized below in Table 3, and illustrated in Figure 9A. Figure 9B shows results of the NewFields assessment compared to results reported from a similar analysis conducted by NOAA (NOAA/BML 2008). It should be noted that all non-straight embryos were classified as having body axis defect for the NOAA/BML study, whereas the NewFields assessment included only those specimens with verifiable notochord irregularities.



**Table 3. Assessment of Body Axis Irregularities**

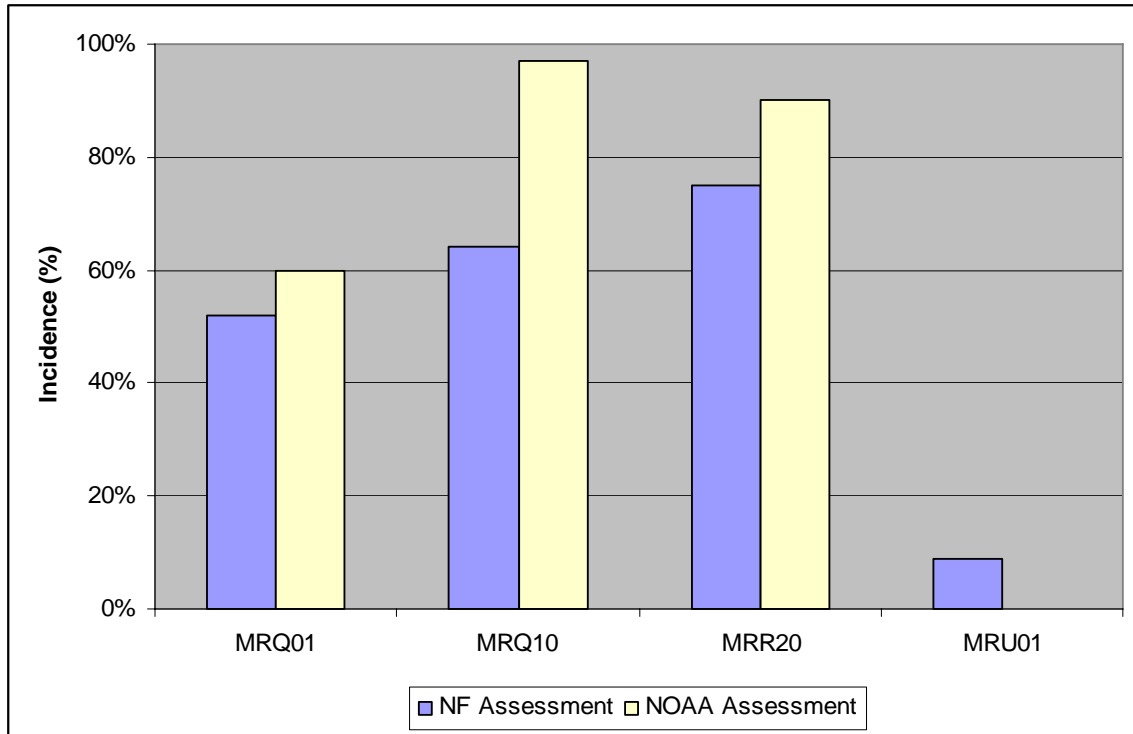
Site	Body Axis Defect	C- Shaped	G- Shaped	O- Shaped	S- Shaped	Linear	NM *	Σ Non Linear/ Total #	Total # Assessed
MRQ01--N1- N8	84	2	4	20	23	2	26		135
Mean	52%							99%	
STD	6%							2%	
CV%	12%							2%	
MRQ10- N1 - N8	107	5	2	5	2	36	9		157
Mean	64%							78%	
STD	22%							23%	
CV%	34%							29%	
MRR20- N1 -N8	122	1	2	15	5	11	7		156
Mean	75%							94%	
STD	17%							10%	
CV%	22%							11%	
MRU01-N1 -N8	15	5	0	0	0	138	0		158
Mean	9%							12%	
STD	13%							16%	
CV%	142%							129%	

\*NM = not measured \* % non-linear is equivalent to the determinations made by NOAA



**Figure 9A. Assessment of Body Axis Irregularities.**

(blue bars represent % with distinct body defects; maroon bars represent total % non-linear embryos)



**Figure 9B. Comparison of Incidence of Notochord Irregularities**

*[blue bar presents data from this study; yellow bars represent data records presented by NOAA (NOAA and BML, 2008)]*

#### Morphological Characteristics: Video Documentation

Results of the descriptive statistics (range, mean, standard deviation and coefficient of variation) and the statistical differences within and between stations were recorded for a subset of available videos representing each site location. The following parameters were evaluated and are summarized in Table 4.

1. Qualitative description of larval stages in natural and artificial spawned specimens
2. Measurement of the length and difference in length (pixels) of upbeat and downbeat AV to PC (reported as Potential % Difference) calculated by comparing the maximum length – the minimum length divided by the mean length.
3. Heart Rate per 20 seconds
4. Heart Area (pixels)
5. Pericardial Region (pixels)
6. Heart to Pericardial Region Ratio
7. Yolk Flattening (% Incidence)
8. Arrhythmia (% Incidence)

Table 5 contains the results of the ANOVA designating statistically significant differences in mean measurements for the stations; Figures 10-15 provide visual displays of the data.

**Table 4. Summary of Statistical Evaluation of Dechorionated Embryo Metrics**

Range								
Measurement	MRU01		MRQ10/P01		MRQ01		MRR20	
	Natural	Artificial	Natural	Artificial	Natural	Artificial	Natural	Artificial
AVPC Length (min and max)	36 to 81	35 to 92	45 to 94	80 to 165	NM	34 to 148	49 to 100	51 to 116
AVPC Potential % Difference	7 TO 18	7 to 24	6 TO 27	17 to 34	NM	13 to 21	9 TO 20	0 to 23
Heart Rate per 20 seconds	34 TO 42	31 to 42	34 TO 48	28 to 36	11 TO 31	28 to 34	34 TO 50	25 to 45
Heart Area (pixels)	8063 TO 12744	4914 to 10898	6464 TO 10713	6603 to 11108	NM	5973 to 10752	8194 TO 8913	5606 to 11145
Pericardial Region (pixels)	19242 TO 42087	14576 to 31419	13612 TO 31904	21986 to 54305	NM	15977 to 35196	26509 TO 35472	10652 to 30132
Heart to Pericardial Region (%)	26 TO 61	33 to 46	29 TO 60	18 to 39		17 to 51	24 TO 34	29 to 73
Yolk Sac Flattening (Incidence %)	Not applicable							
Arrhythmia (Incidence %)	Not applicable							
Larval Stage	q	q	p and q	p and q	p	q	p and q	p and q
Oiling	No oiling		Very light to light		Light		Heavy to light	
Location	San Rafael		Sausalito		Peninsula Pt		Keil Cove	
Oiling Rank	0		1		2		3	
Mean								
Measurement	MRU01		MRQ10/P01		MRQ01		MRR20	
	Natural	Artificial	Natural	Artificial	Natural	Artificial	Natural	Artificial
AVPC Length (min and max)	56	60	62	111	NM	72	70	86
AVPC Potential % Difference	28	29	26	19	NM	28	22	19
Heart Rate per 20 seconds	37	36	39	31	22	31	42	36
Heart Area (pixels)	11269	8885	8642	9040	NM	8234	8518	8521
Pericardial Region (pixels)	31325	23781	22677	35205	NM	24205	30665	20930
Heart to Pericardial Region (%)	38	38	40	27	NM	37	28	44
Yolk Sac Flattening (Incidence %)	37	28	38	31	NM	18	56	27
Arrhythmia (Incidence %)	11	0	11	0	0	17	33	33
Larval Stage	100 q	100q	33 p; 67 q	100 q	100 p	100 q	67 p; 33q	56 p; 44 q
Oiling	No oiling		Very light to light		Light		Heavy to light	
Location	San Rafael		Sausalito		Peninsula Pt		Keil Cove	
Oiling Rank	0		1		2		3	
NM = Not Measurable								

**Table 4. Summary of Statistical Evaluation of Dechorionated Embryo Metrics (Continued)**

STD								
Measurement	MRU01		MRQ10/P01		MRQ01		MRR20	
	Natural	Artificial	Natural	Artificial	Natural	Artificial	Natural	Artificial
AVPC Length (min and max)	12.2	13.8	13.7	22.2	NM	35	21.3	8.4
AVPC Potential % Difference	6	10	12	5	NM	9	12	12
Heart Rate per 20 seconds	2	4	5	2	8	2	6	7
Heart Area (pixels)	1328	2030	1554	1189	NM	1606	364.8	1946
Pericardial Region (pixels)	7417	5544	5785	10168	NM	7786	4517	7262
Heart to Pericardial Region %	12.7	5	10	7	NM	12	5	13
Yolk Sac Flattening (Incidence %)	2.9	8	10	4	NM	2		7
Arrhythmia (Incidence %)	19	0	19	0	0	23.4	47	57.7
Larval Stage	Not applicable							
Oiling	No oiling		Very light to light		Light		Heavy to light	
Location	San Rafael		Sausalito		Peninsula Pt		Keil Cove	
Oiling Rank	0		1		2		3	
CV%								
Measurement	MRU01		MRQ10/P01		MRQ01		MRR20	
	Natural	Artificial	Natural	Artificial	Natural	Artificial	Natural	Artificial
AVPC Length (min and max)	22	23	22	20	NM	49	30	22
AVPC Potential % Difference	23	34	45	25	NM	33	56	62
Heart Rate per 20 seconds	5.4	10	12	8	38	7	15	18
Heart Area (pixels)	12	23	18	13	NM	20	4	23
Pericardial Region (pixels)	24	23	26	29	NM	32	15	45
Heart to Pericardial Region %	33	14	25	25	NM	32	18	4
Yolk Sac Flattening (Incidence %)	7.9	30	35	12	NM	13	0	25
Arrhythmia (Incidence %)	173	0	173	0	0	141	141	173
Larval Stage	Not applicable							
Oiling	No oiling		Very light to light		Light		Heavy to light	
Location	San Rafael		Sausalito		Peninsula Pt		Keil Cove	
Oiling Rank	0		1		2		3	

**Table 5. Results from ANOVA Analyses (Stations underlined with the same color are not significantly different).**

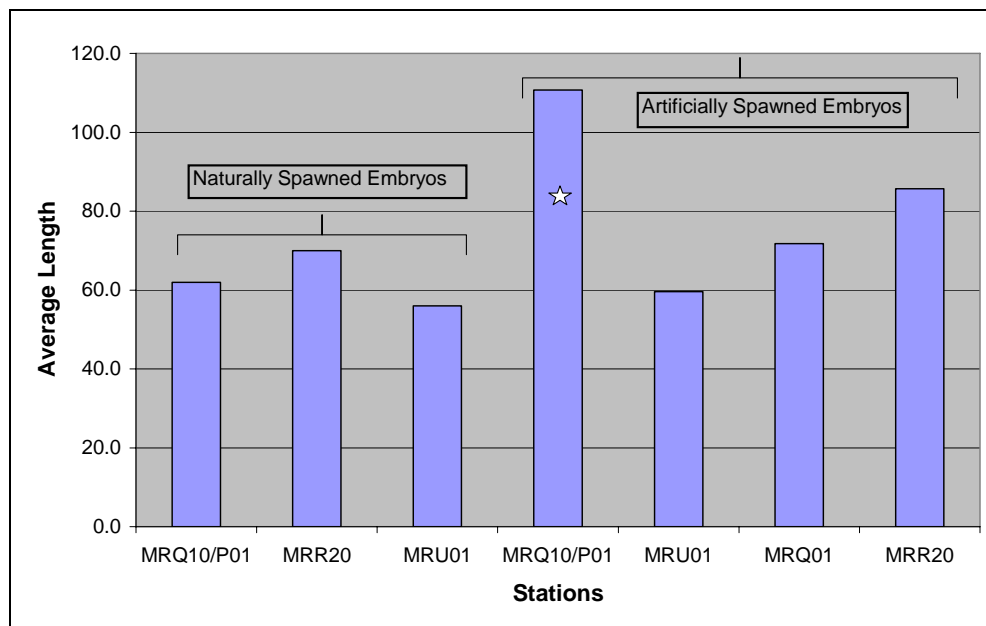
Measurement	Prob > F	Mean Measurement (Highest to Lowest)							
		Artificial MRQ10/P01	Artificial MRR20	Artificial MRQ01	Natural MRR20	Natural MRQ10/P01	Artificial MRU01	Natural MRU01	Natural MRQ01
AVPC Length (min and max)	<0.001	111	86	72	70	62	60	56	No Data
Heart Beat Rate	<0.001	Natural MRR20	Natural MRQ10/P01	Natural MRU01	Artificial MRR20	Artificial MRU01	Artificial MRQ01	Artificial MRQ10/P01	Natural MRQ01
		42	39	37	36	36	31	31	22
Heart Area (HA)	0.007	Natural MRU01	Artificial MRQ10/P01	Artificial MRU01	Natural MRQ10/P01	Artificial MRR20	Natural MRR20	Artificial MRQ01	Natural MRQ01
		11269	9040	8885	8642	8521	8518	8234	No Data
Pericardial Space (PS)	0.001	Artificial MRQ10/P01	Natural MRU01	Natural MRR20	Artificial MRQ01	Artificial MRU01	Natural MRQ10/P01	Artificial MRR20	Natural MRQ01
		35205	31325	30665	24205	23781	22677	20930	No Data
HA/PS	0.022	Artificial MRR20	Natural MRQ10/P01	Natural MRU01	Artificial MRU01	Artificial MRQ01	Natural MRR20	Artificial MRQ10/P01	Natural MRQ01
		44%	40%	38%	38%	37%	28%	27%	No Data
Yolk Flattening (%)	0.007	Natural MRR20	Natural MRQ10/P01	Natural MRU01	Artificial MRQ10/P01	Artificial MRU01	Artificial MRR20	Artificial MRQ01	Natural MRQ01
		56%	38%	37%	31%	28%	27%	18%	No Data

*Qualitative Description of Larval Stages in Natural and Artificial Spawnings.* The degree of embryo development in natural and artificially spawned samples generally represents transitional characteristics of stages  $p$  through pre-hatch ( $q$ ). The artificially spawned embryos represent the intermediate stage ( $q$ ; Figure 2b, Table 4) while the naturally spawned embryos represent more variation, ranging from younger ( $p$ ) to older stages ( $q$ ) (Figure 2a-c). Data from the natural spawned specimens from station MRQ01 represent earlier stages of development for all of the samples that were reviewed while the remaining stations with naturally spawned specimens represent more fully developed embryos (Table 4). Dechorionated embryos from naturally spawned eggs collected from station MRQ01 were the most problematic for scoring; most morphological characteristics were not visible for this group. Qualitatively, there is a difference between the larvae evaluated for the artificially and the naturally spawned specimens.

*Potential Differences in AVPC Length.* The video images provide a method of measuring the difference in AVPC length based on upbeat or downbeat of the heart within the pericardial space. This variation was calculated for each specimen examined using individual video clip images; thus providing an assessment of the natural variation associated with a given specimen based on the beat of the heart at the time of the measurement. The variation in the AVPC length for the

four study areas based on position of heart is 25% for natural and 36% for artificially spawned samples. The length of the beat distance is longer for the artificially spawned embryos compared to the naturally spawned embryos. This difference is associated with the observations from the artificially spawned larvae examined at station MRQ10/P01 (Appendix A). Results of ANOVA indicate that this station is statistically different from all others; MRR20 is statistically different from naturally spawned embryos from MRQ10 and MRU01 as well as the artificial spawned embryos from MRU01 as shown in Table 5. Anecdotal observations of the larval stage at MRQ10 indicate the artificially spawned embryos are probably at a later stage than most of the naturally spawned samples and consequently, the significant difference noted may be associated with the relative stage of development of the embryos.

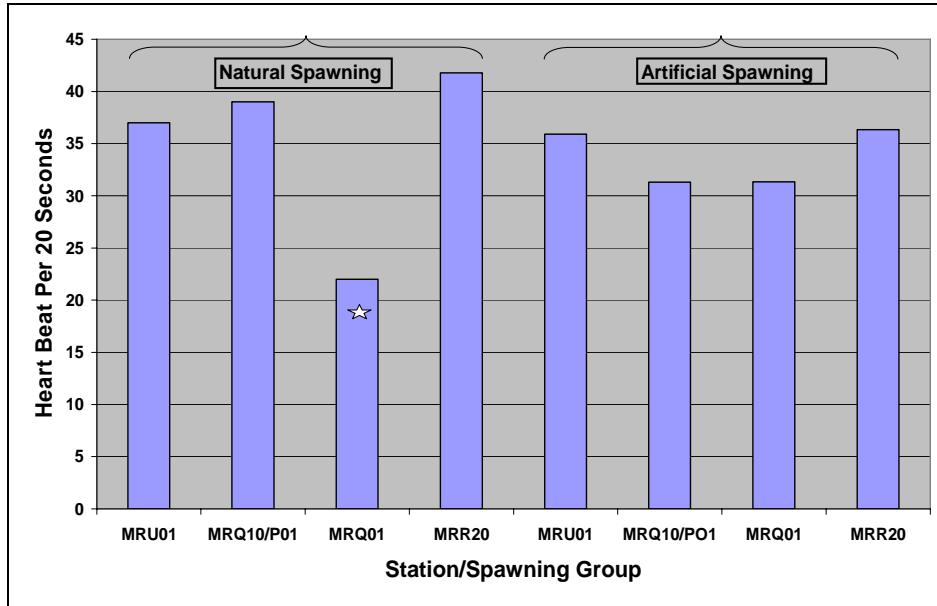
The differences between the lengths of an up- and downbeat are not statistically significantly different between those samples that were artificially or naturally spawned, nor are there statistically significant differences among the various stations (Appendix A). A biologically important difference is observed only if the length of AVPC distance among or between stations is greater than the variation within beat lengths for individual heart beats. This was examined by comparing variation within a subset of artificially and naturally spawned samples for each station. The number of specimens with longer beat lengths than expected within a station or compared by all stations was 2 or 3 instances per station; however, the incidence of longer beat lengths was consistently higher for artificially spawned samples from MRQ10/P01. These measurements were consistently larger and appear to represent an older larval stage than other location/spawning group assessments (Figure 10).



**FIGURE 10. ANOVA Results of Differences in AVPC Measurements**  
*[Star indicates statistically significant from all other stations]*

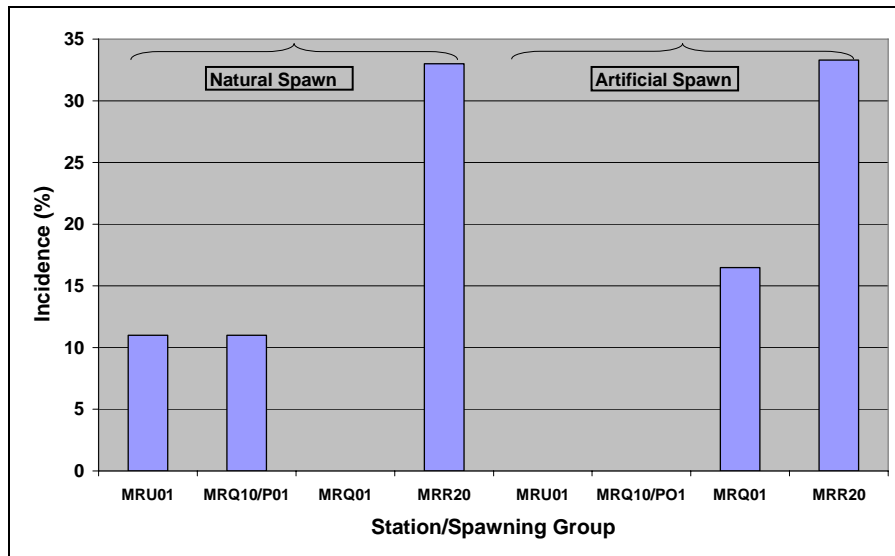
*Heart Rate and Arrhythmia.* The heart rate of the samples evaluated ranged from 11 to 50 beats per 20 second video clip. The heart rate for specimens collected from the natural spawn at station MRQ01 were significantly lower than those at all other stations. The variation of heart rates for naturally spawned samples from MRQ01 is consistent with the fact that all samples evaluated were early p-larval stage. The range of heart rate values observed with both the natural and artificial spawn at station MRR20 is larger than the other stations as measured by CV%; and these

samples also represent a larger percentage of p-larval stages (56 to 67%). Figure 11 shows the average heart rate per station/spawning group.



**FIGURE 11. Analysis of Heart Rate**  
*[Star indicates statistically significant from all other stations]*

The incidence of heart arrhythmia for the naturally spawned and artificially spawned samples from a subset of each of four stations was 0-33%. The incidence of arrhythmia in artificially spawned embryos was 0% at MRU01 and MRQ10/P01, 17% at MRQ01, and 33% at MRR20. The incidence of arrhythmia in naturally spawned embryos was 0% at MRQ01; 11% at MRU01 and MRQ10; and 33% MRR20. Figure 12 illustrates the incidence of arrhythmia by station.



**FIGURE 12. Incidence of Arrhythmia**

*Heart Area, Pericardial Region and Heart to Pericardial Region Ratio.* As the heart increases in size the pericardial space will either increase, maintaining a similar ratio; or will increase or decrease in size relative to the size of the heart. Pericardial edema is based on a swelling of the space within this region. Figures 13 and 14 present the average size of the heart, the average size of the pericardial space and the ratio of the heart to the pericardial space. The heart area for natural spawn MRU01 was significantly larger than artificial MRR20, natural MRR20, and artificial MRQ01 (Table 5). Pericardial space was significantly larger in artificial MRQ10/P01 than in artificial MRR20.

*Yolk Sac Flattening (% Incidence).* An apparent result of increasing heart or pericardial space area is a flattening of the yolk sac on the anterior face. The incidence of yolk sac flattening is relatively the same across natural and artificial spawning groups at all stations except MRR20 where the natural spawn flattening was significantly higher flattening incidence of artificially spawned samples from MRR20 (Figure 15).

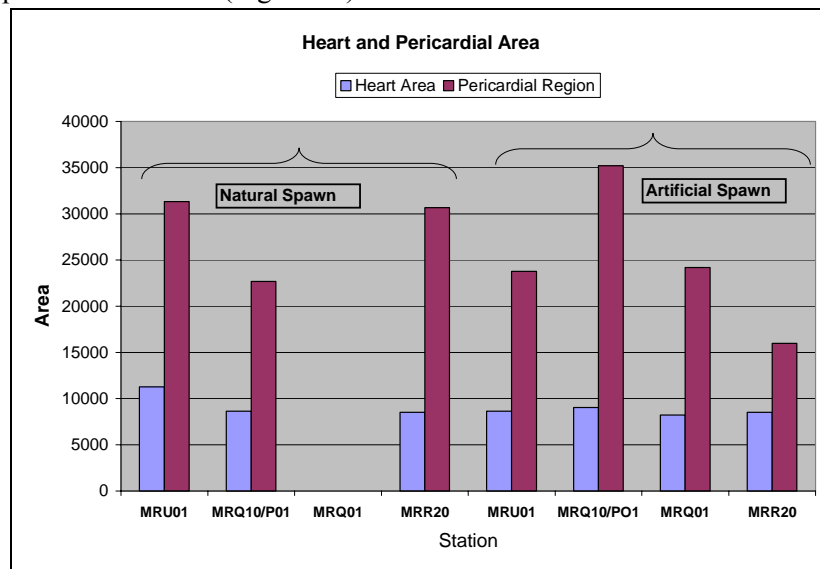


FIGURE 13. Comparison of Heart and Pericardial Areas (pixels)

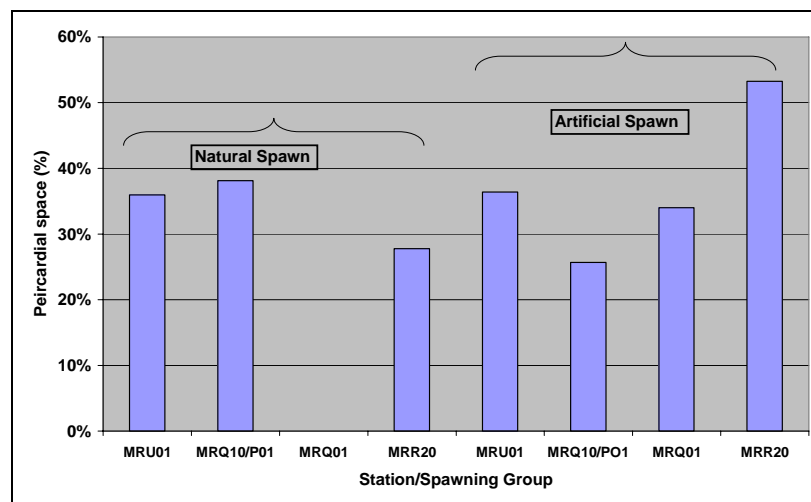


FIGURE 14. Ratio Heart: Pericardial Space



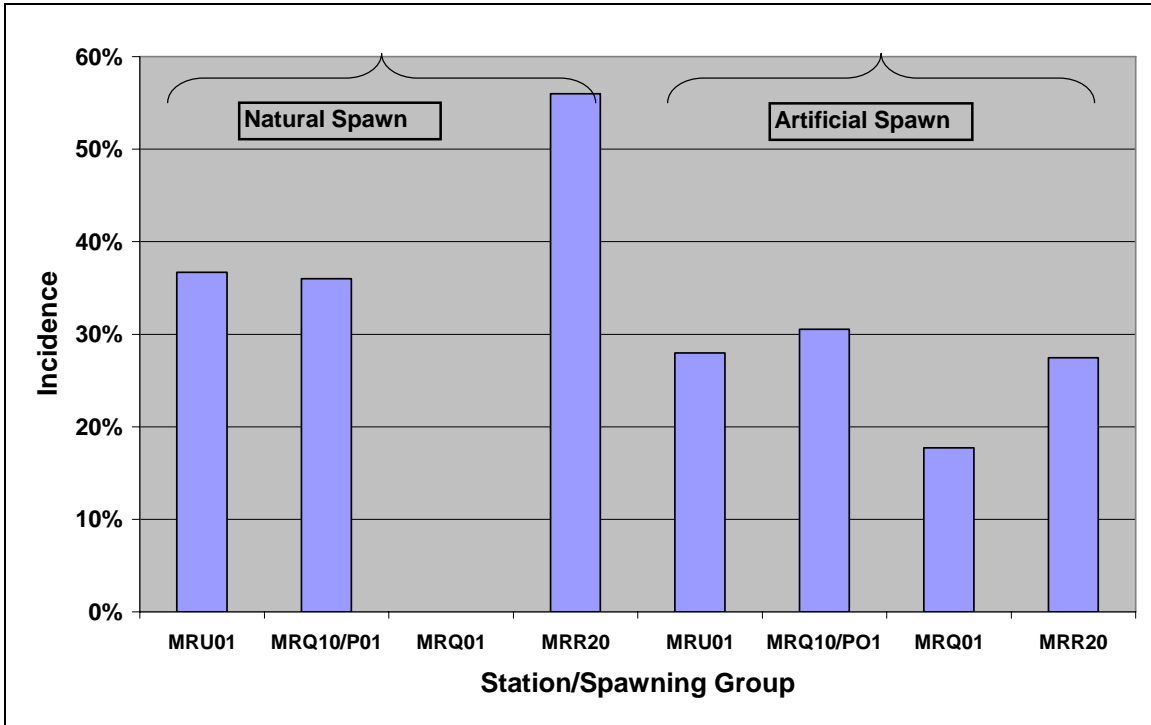


FIGURE 15. Observed Incidence of Flattened Yolk Sacs

*Background Illumination.* Data produced from measurement of mean gray values at several locations within individual video clips are presented in Table 6 and 7. The relationship between background illumination and illumination of various tissues (otic vesicle, eye, and yolk sac) are summarized in Figures 16 through 18. For features that are lighter (higher gray scale values) in color (otic vesicle and yolk sac) there are significant regressions ( $p < 0.05$ ) between the background illumination and the feature. There does not appear to be a difference in the relationship to background illumination for each feature between the natural and artificially spawned embryos.

**Table 6. Measured Mean Gray Values from Natural Spawn Video Recordings (first frame)**

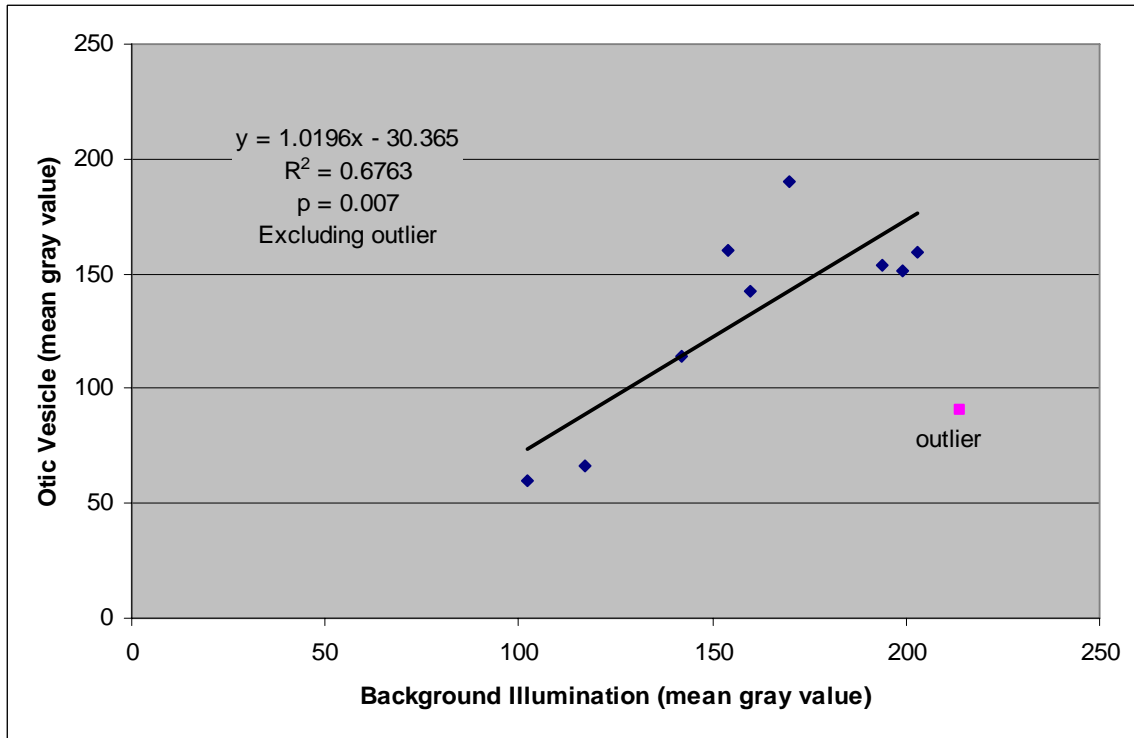
Natural Spawn										
Site	Video Clip #	Illumination (mean gray value*)								
		Background			Head			Eye		Yolk Sac
		Top Left	Bottom Rt	Bottom Left	Dorsal	Otic Vesicle	Dorsal, 2nd	Center	Outside	Not in Shadow
MRQ01-FEM1-022908-N4	11	194	210	206	118	154	146	27	37	159
MRQ01-FEM1-022908-N6	6	199	201	206	134	151	132	22	30	147
MRQ10/P01-FEM1-022708-N2	91	142	154	155	162	114	114	33	39	183
MRQ10/P01-FEM1-022708-N6	10	170	186	188	198	190	105	20	30	171
MRQ10/P01-FEM1-022708-N8	5	154	161	128	211	160	130	21	27	157
MRR20-FEM1-022808-N4	9	203	209	216	241	159	149	22	27	234
MRR20-FEM1-022808-N7	51	214	217	210	135	91	129	34	47	205
MRU01-FEM1-022608-N2	3	102	128	126	121	60	78	22	25	123
MRU01-FEM1-022608-N4	5	160	161	157	136	142	111	20	22	173
MRU01-FEM1-022608-N7	8	117	145	138	172	66	201	22	27	140
Mean		165.5	177.2	173.0	162.8	128.7	129.5	24.3	31.1	169.2
STD		37.7	31.3	36.0	41.9	43.8	32.7	5.2	7.6	32.4
CV		23%	18%	21%	26%	34%	25%	22%	25%	19%

\*Low numbers represent dark tones; high numbers represent light tones, gray values range from 0 to 255.

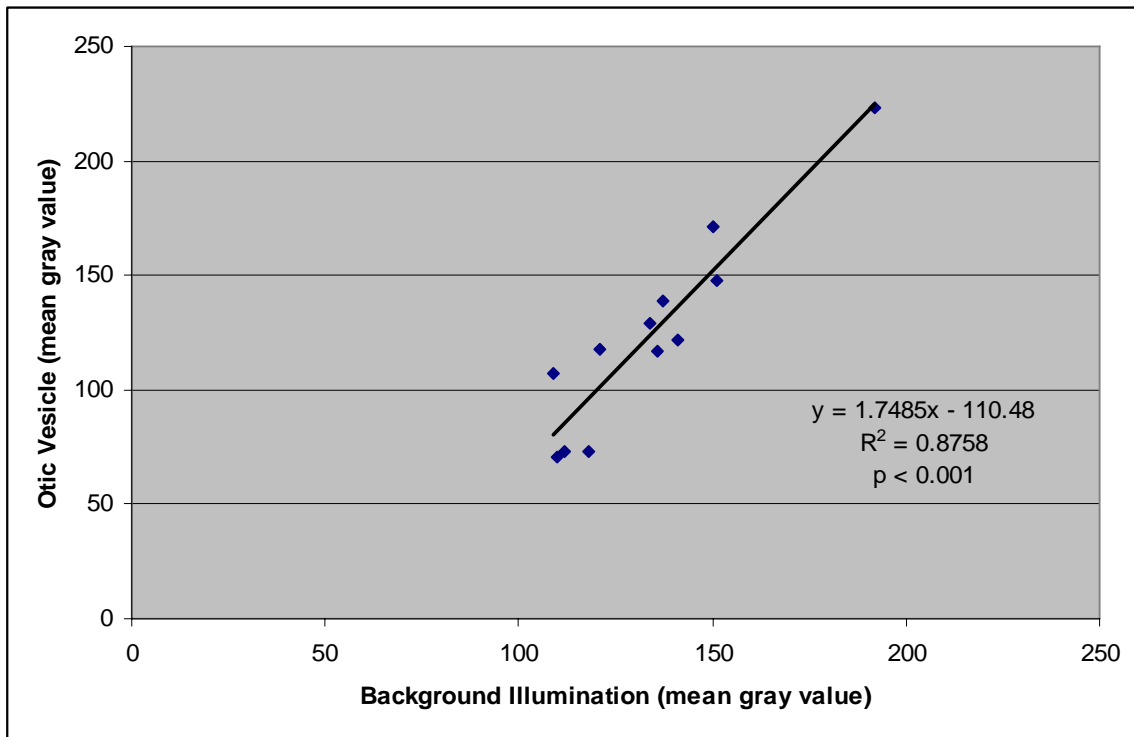
**Table 7. Measured Mean Gray Values from Artificial Spawn Video Recordings (first frame)**

Artificial Spawn										
Site	Video Clip #	Illumination (mean gray value*)								
		Background			Head			Eye		Yolk Sac
		Top Left	Bottom Right	Bottom Left	Dorsal	Otic Vesicle	Dorsal, 2nd	Center	Outside	Not in Shadow
MRQ01-FEM1-021808-A1	4	137	153	153	155	139	100	17	24	159
MRQ01-FEM1-021808-A3	8	136	129	135	116	117	105	17	25	141
MRQ10/P01-FEM1-021908-A1	1	151	174	174	131	148	85	20	22	172
MRQ10/P01-FEM1-021908-A2	3	110	140	137	149	71	107	17	29	139
MRQ10/P01-FEM1-021908-A3	29	118	141	145	154	73	82	17	26	156
MRQ10/P01-FEM1-021908-A5	1b	134	146	146	168	129	119	19	25	134
MRR20-FEM1-022008-A1	12	112	145	139	139	73	135	22	31	155
MRR20-FEM1-022008-A3	5	121	143	149	140	118	128	23	26	143
MRR20-FEM1-022008-A5	11	150	152	142	106	171	144	26	47	148
MRU01-FEM1-022208-A1	6	141	165	169	173	122	137	24	32	183
MRU01-FEM1-022208-A2	1c	192	191	191	167	223	183	21	26	205
MRU01-FEM1-022208-A3	1b	109	136	133	171	107	110	22	27	142
Mean		134.3	151.3	151.1	147.4	124.3	119.6	20.4	28.3	156.4
STD		23.5	17.5	17.9	21.8	43.9	28.1	3.1	6.5	20.9
CV		18%	12%	12%	15%	35%	23%	15%	23%	13%

\*Low numbers represent dark tones; high numbers represent light tones, gray values range from 0 to 255.

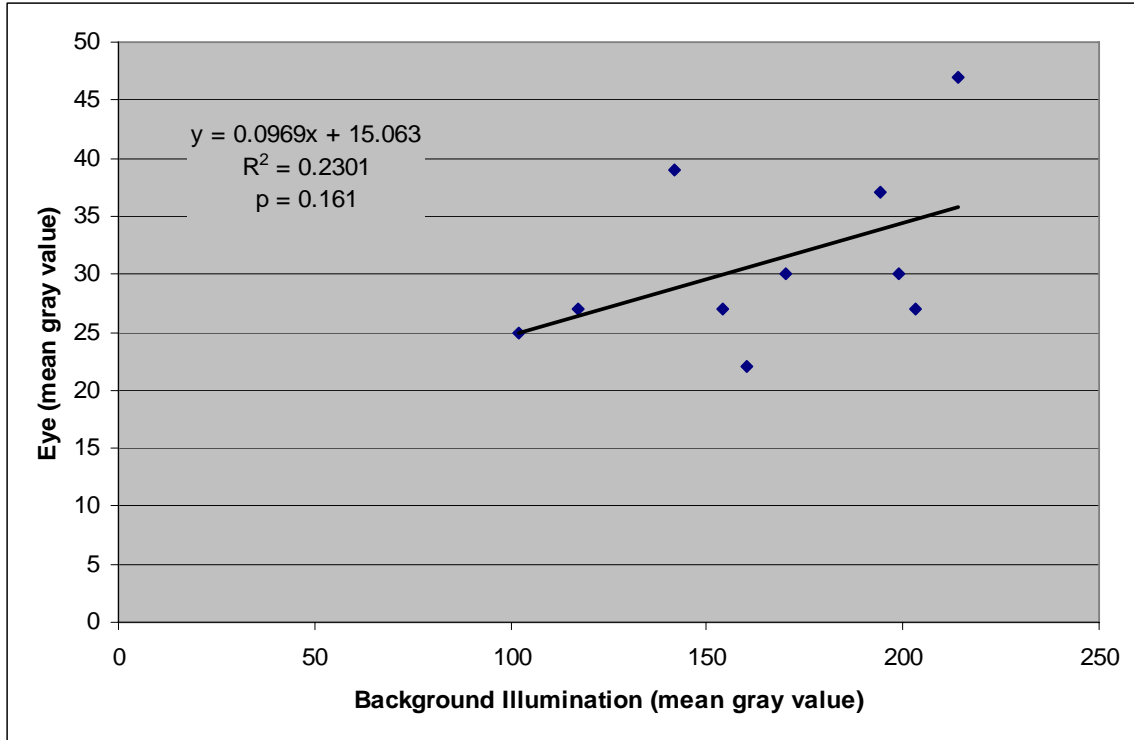


**A. NATURALLY SPAWNED EMBRYOS**

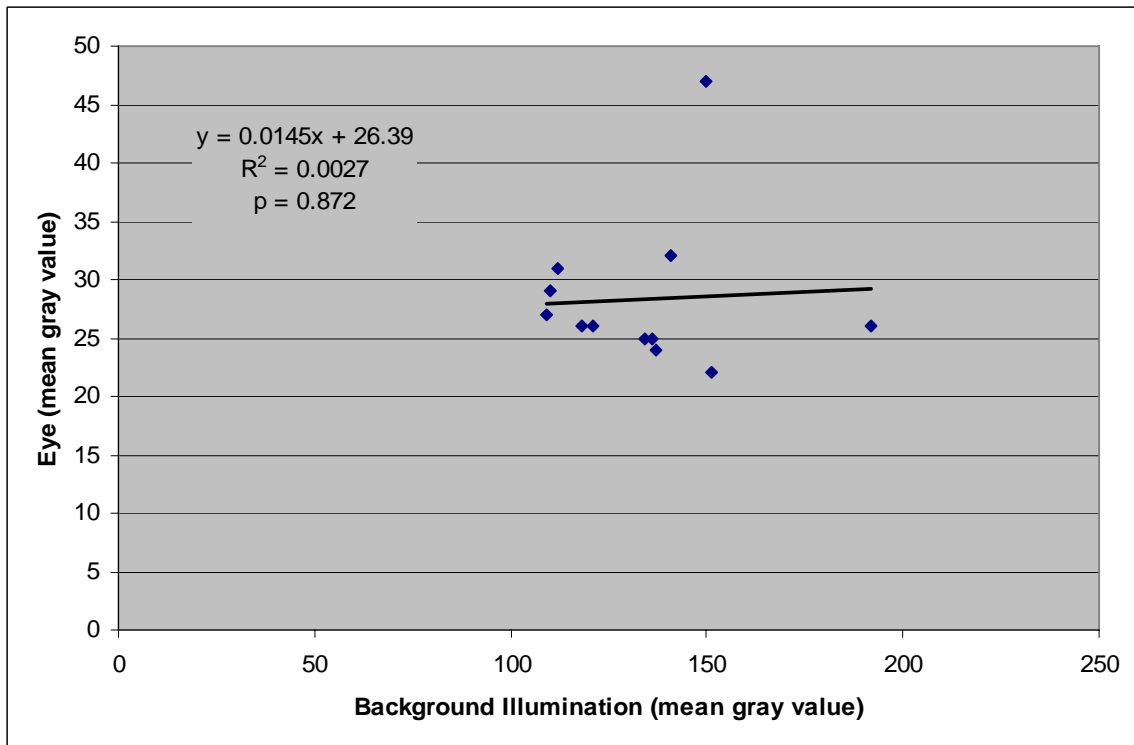


**B. ARTIFICIALLY SPAWNED EMBRYOS**

**FIGURE 16. Comparison of Background Illumination and Otic Vesicle**

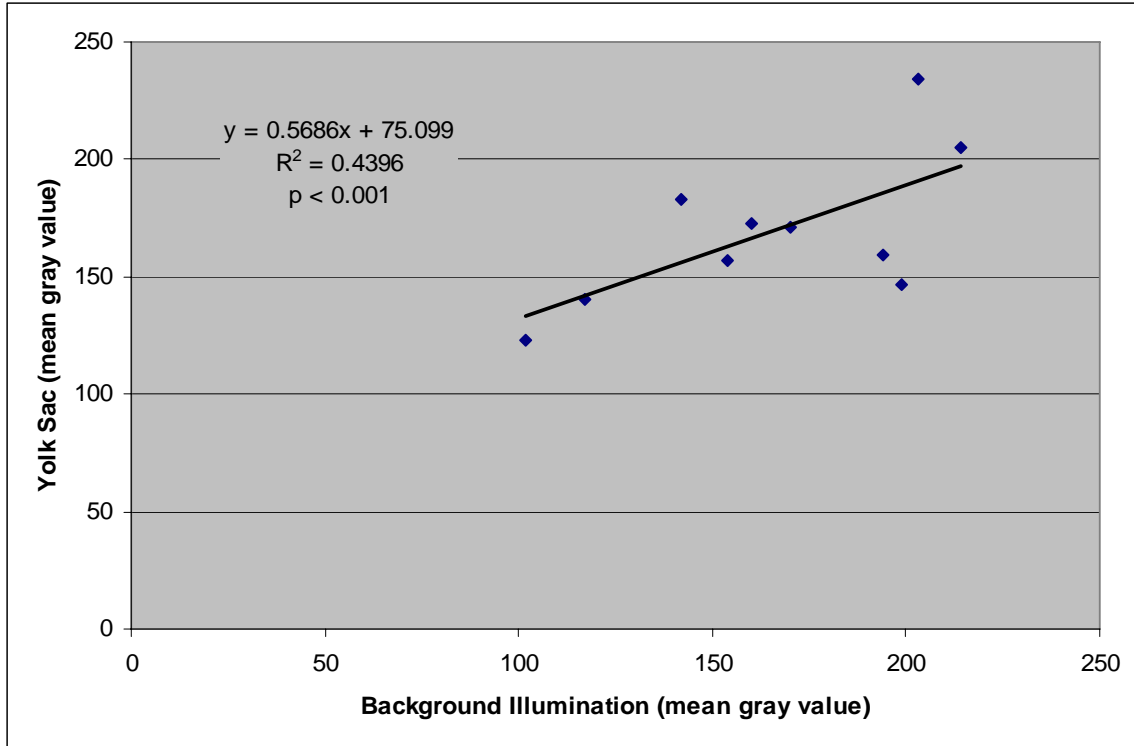


A. NATURALLY SPAWNED EMBRYOS

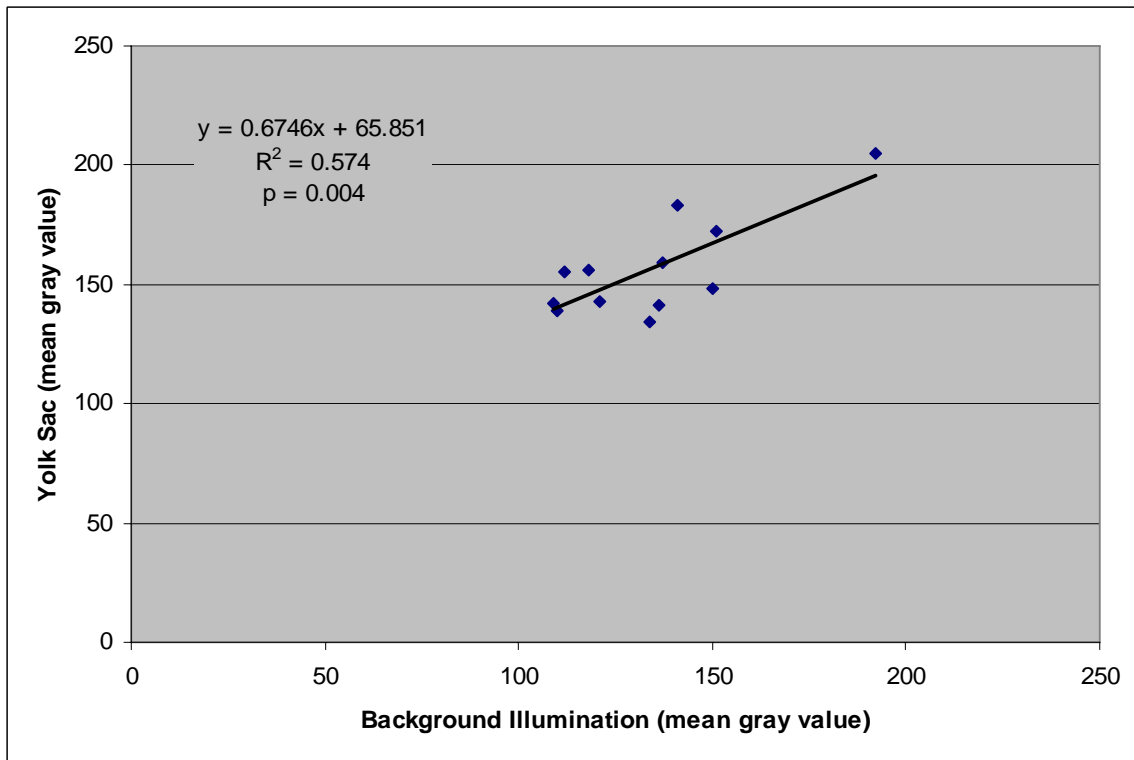


B. ARTIFICIALLY SPAWNED EMBRYOS

FIGURE 17. Comparison of Background Illumination and Eye Pigmentation



**A. NATURALLY SPAWNED EMBRYOS**



**B. ARTIFICIALLY SPAWNED EMBRYOS**

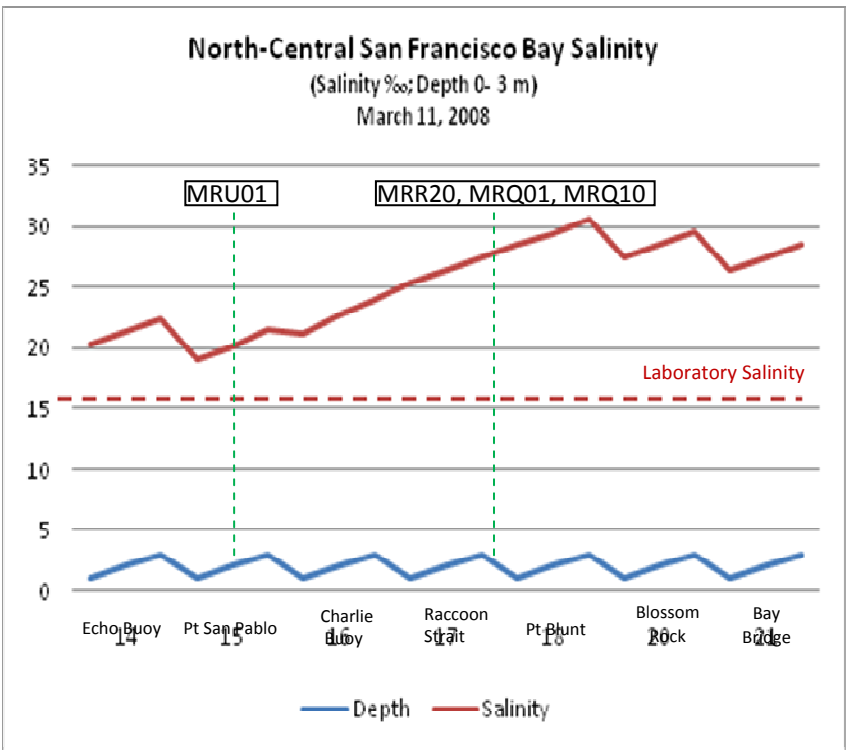
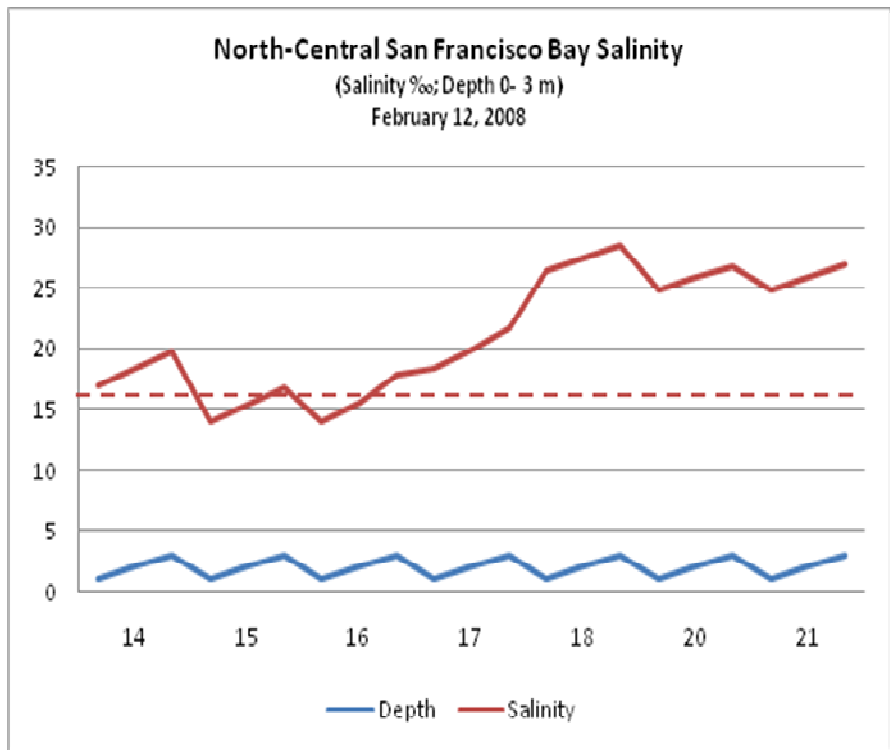
**FIGURE 18. Comparison of Background Illumination and Yolk Sac**

## Relationship of Qualitative Descriptions of Oiling, Local Salinity and Observations of Effects

Evaluations were made relative to the rank order of qualitative estimates of oiling provided (NOAA, BML, 2008). A summary of the relationships is presented in Table 8. Figure 19 portrays the salinities that were observed at nearby surface water stations during the spawning events. Figures 20 and 21 illustrate linear regression relationships among the various measurements with naturally and artificially spawned dechorionated embryos and degree of oiling present at the site locations. These regressions are built around average responses of the assessed embryos for each of the parameters based on 3 to 4 points of comparison. Caution is advised relative to the degree of confidence that can be placed in the observations. The majority of regression relationships have a low  $R^2$  and the slopes are insignificant (i.e., not different from a horizontal line) indicating a lack of linear correlation between the parameter and the rank order of estimated oiling.

**TABLE 8. Summary of Significance of Regression Relationships between Natural and Artificial Spawn Data**

Measurement	Natural (n=3)			Artificial (n=4)		
	Slope Direction	Prob > F	R <sup>2</sup>	Slope Direction	Prob > F	R <sup>2</sup>
AVPC Potential % Difference	Neg	<0.001	1.0	Neg	0.507	0.243
Yolk flattening (%)	Pos	0.160	0.938	Neg	0.670	0.109
Heart Rate	Neg	0.971	0.001	Pos	0.938	0.004
Heart Area	Neg	0.429	0.611	Neg	0.325	0.456
Pericardial Region	Pos	0.927	0.015	Neg	0.599	0.161



Data from <http://sfbay.wr.usgs.gov/access/wqdata>

**Figure 19. Salinity Profiles for Central San Francisco Bay, February and March 2008**

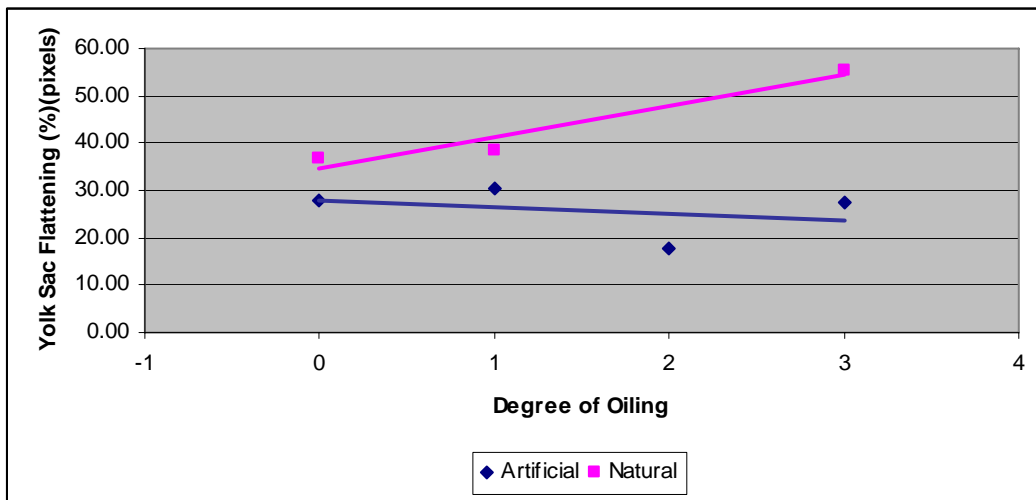
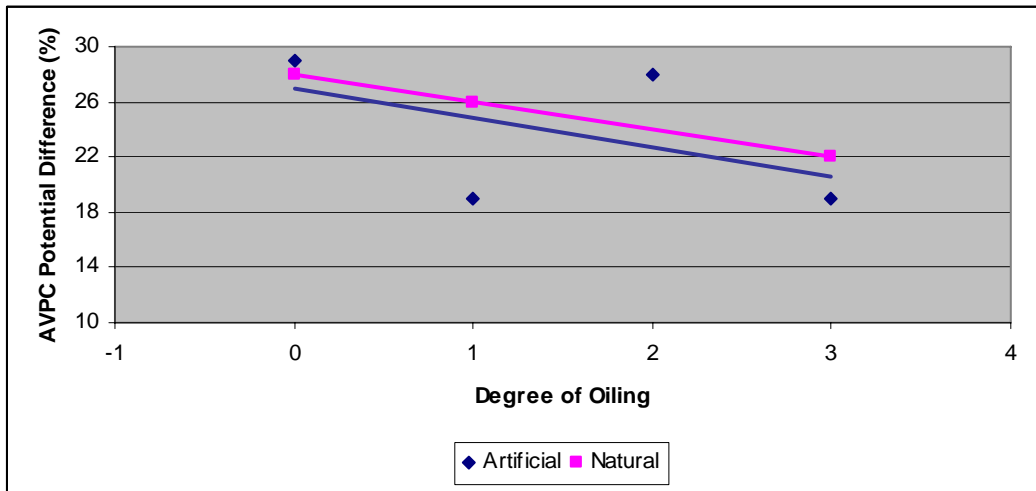


Figure 20. Relationships between AVPC Length Data and Yolk Sac Flattening and Degree of Oiling



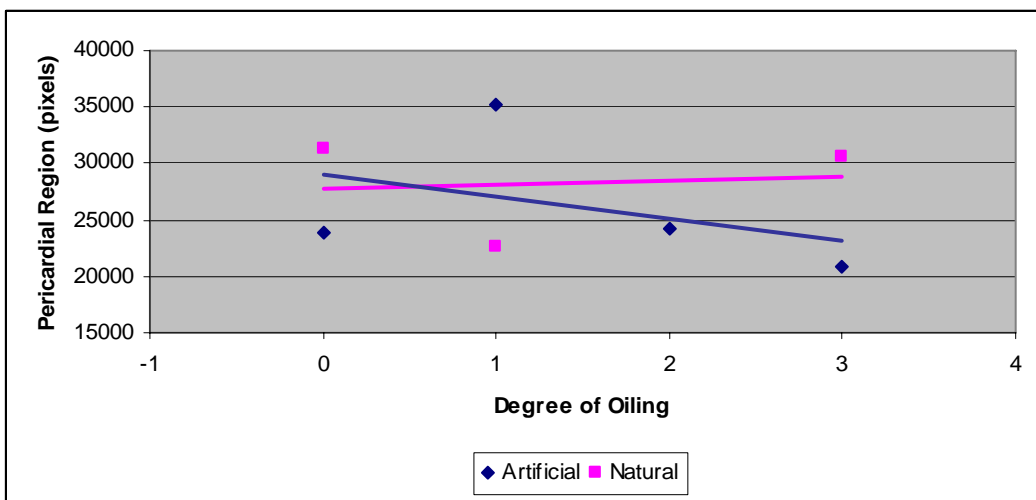
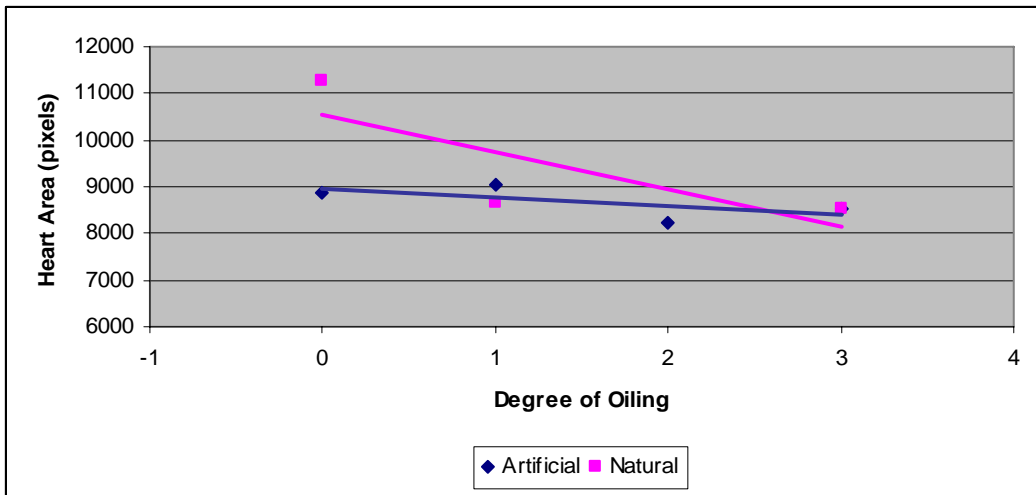
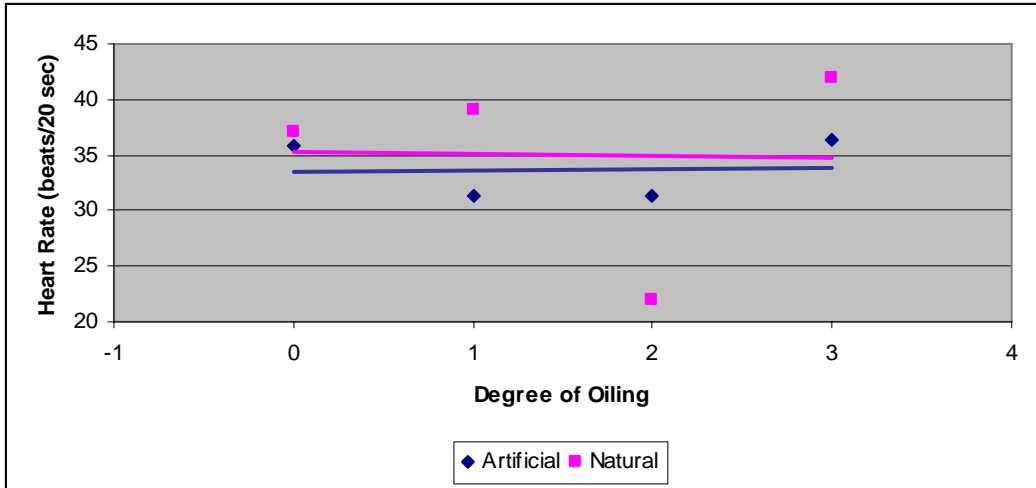


Figure 21. Relationship between Heart Rate, Heart Area, and Pericardial Area and Degree of Oiling

## Discussion

There is a distinct difference in the appearance of herring larvae from those organisms that were artificially spawned and developed at selected shallow subtidal locations and those that were spawned and developed naturally in nearshore, intertidal areas. The artificially spawned organisms are a known age and they have several differences between how they were handled compared to the naturally spawned embryos. The artificially spawned organisms were provided from harvest of adult herring selected for the larger sized individuals whereas the age structure of the adult herring comprising the Central San Francisco stock has not been evaluated at this time. The gonads used for the artificial spawning were harvested and fertilized at temperatures and salinities that were controlled (12°C and 16 ‰) and all organisms were spawned and collected on the artificial cages at the same time. Naturally spawned organisms were collected from areas with very different salinities ranging from salinities of ~16 - 28‰ (Figure 19). All organisms whether they were artificially or naturally spawned were exposed to seawater salinities of 16 ‰ during the dechoriation process. The artificially spawned embryos were subjected to scoring based upon video imaging of high quality while the naturally spawned embryos were primarily scored based on still images.

Several factors appear to be related to the observations collected on a subset of artificially and naturally spawned samples from four different locations. These factors probably had an impact on the assessment of oiling related impacts on Pacific herring in San Francisco Bay and need to be addressed in any future assessments. These factors include:

1. The location and environmental conditions of the water during spawning, development and hatching events.
2. The influence of environmental (salinity) and human factors (oiling) on the selected measurement end-points and how those influences effect assessments.
3. The stage of embryonic development when the fish were sampled and from which the measurements were made.
4. The consistency of the embryonic development stage upon which the measurements were made. The artificially spawned embryos are all one stage while the natural spawn are variable in developmental stage.
5. The illumination levels are variable and influence the density of pigmentation on various assessments of color and tissue density.
6. The age of the parental stocks and sizes of parents influence the size of eggs, development of embryos after fertilization and success of hatching. The age of parent stocks for naturally spawned embryos are unknown but the artificially spawned organisms were selected from larger individuals that were captured to provide the gametes used during this part of the project.

A key to understanding the effects of petroleum exposure on juvenile herring in San Francisco Bay requires a method to distinguish between the environmental and biological variables within the Bay that are also concordant with the rank order of oiling. Hershberger et al (2005) has demonstrated that younger adults have greater incidence of abnormal development in terms of notochord development, body shape and survival and successful development of the juveniles. The State of California's Herring Advisory Committee recognized various potential contributing

factors to the low population size of San Francisco Bay Herring stocks ranging from age structure (few 3-5 year fish), selective harvest of older fish, loss of habitat and the potential effects of contamination, including oil spills (California Fish & Game Commission 2008). The combination of many of these factors in addition to larger scale climate change has been attributed to be the cause(s) of the past decades historically low spawning biomass of herring in San Francisco Bay (SFEI Report). Since the characteristics being evaluated in this review are single measurements that may indicate responses to a variety of biological and chemical factors, it becomes important to evaluate factors which may be affecting the viability of natural larvae such as the age structure of the spawning populations. The winter of 2008 was different than many of the prior years with herring spawning in areas outside of their normal spawning sites. Cal Fish and Game regularly monitors the population structure of herring stocks in San Francisco Bay and these data should be available for examination to determine the size and age structure of the spawning stocks within the area of study as well as areas outside of the area of concern. The objective would be to compare the age structure of the stocks to data provided by Herschberger et al (2005) to determine if the age of the population of herring might have an influence on successful hatching.

The difference in handling of artificial and natural spawning events is an important element that needs to be addressed in any subsequent work. It is well known that herring can develop under a wide range of environmental salinities. Experimental laboratory protocols have demonstrated the wide range of tolerance to salinity and temperatures (Dinnel, et al. 2008). However, these protocols have been developed for determining success rates of fertilization, development, and hatching of larvae under consistent salinities throughout testing. The conditions associated with the natural and artificially spawned herring are different than these standardized protocols and would require appropriate acclimation to different salinities that occurred at points of capture and the subsequent laboratory trials for this study. Acclimation between the fertilization/development salinities and the water during hatching does not appear to have been addressed.

The potential and probable issue associated with this difference is built around the process of fertilization and early egg development. During the initial process of fertilization (after the sperm enters the egg) the eggs of fish imbibe the surrounding water and 'harden'. This captured water has its own characteristics and the egg is protected during development somewhat from the surrounding environment. Upon hatching, the larvae are then exposed to the surrounding environment. Under normal environmental conditions the ambient seawater at fertilization and at hatching should be relatively similar except in the intertidal under varying conditions of rainfall where local events alter this pattern and may explain some of the large variation in survival of intertidally spawned herring.

Standard technical procedures used in aquatic toxicology testing laboratories require slow acclimation of broodstock fishes over a period of at least two days to different salinities (ASTM E 1241). This slow acclimation over a period of at least two days and holding and developing the brood stock under these conditions during the collection and development of fertilized eggs assures minimum stress on the adults and developing eggs. The procedures outlined by Dinnel et al. (2008) follow this process as did the artificial spawning group during fertilization and dechoriation, although the development phase (post fertilization and hardening) was performed at environmental ambient conditions. The naturally spawned and developed eggs occurred in ambient water at the site but all of them were dechorionated in 16‰ water. It is anticipated that the larger the difference in the salinity of the imbibed water relative to the 'hatching' water used

during dechoriation the greater the stress to the newly 'hatched' individuals. The difference in salinity during spawning for the naturally spawned eggs is likely to result in osmotic gradients with water diffusing into tissues from the lower salinity environment during dechoriation. It is possible that the observations of extra membranes within the pericardial area, swelling of the pericardial area and flattening of the yolk sac during swelling could be related to the process just described. Edema created by this process or by petroleum exposures is unlikely to be distinguishable. The shapes and viability of the hatched/dechoriated larvae is also a characteristic that could result from being placed into different salinity environments. This is a key issue that needs to be addressed and incorporated into testing guidelines and protocols prior to doing future testing. An experiment to determine the influence of this process should be performed in order to eliminate this potential cause of any of the observations that were made in 2008 and that are proposed for 2009. This can be performed on a surrogate egg laying species (e.g., *Menidia beryllina*) prior to the herring spawning events in order to best establish a protocol for application to the herring study in 2009.

The last attribute that needs to be controlled is the amount of illumination transmitted to the viewing screens. An analysis of video images indicated that the density of pigmentation in the eye and otic vesicle were directly correlated to the amount of illumination measured by background grayness. Because of this factor the comments about pigmentation amounts is not supportable at this time. Future work will need to control this variable before significant conclusions can be made.

## References

ASTM. 2005. Annual Book of Standards: Standard Guide for Conducting Early Life-Stage Toxicity Tests with Fishes (ASTM E1241]. Vol.11.05 Biological Effects and Environmental Fate; Biotechnology; Pesticides. ASTM International, West Conshohocken, PA.

California Fish & Game Commission. 2008. Pre-publication Notice Statement on Proposed Hearing to amend Sections 163 and 164, Title 14, California Code of Regulations pertaining to Harvest of Herring and Harvest of Herring Eggs.

Dinnel P, R Hoover, L Lechuga, K Tobiason, and J Elphick. 2008. Development of Larval Pacific Herring, *Clupea pallasii*, Bioassay Protocols: Refinement, Validation, Refinery Effluent and Cherry Point Ambient Water Testing During 2007. Final Report for Washington Department of Ecology, Olympia WA.

Hill J and IA Johnston. 1997. Photomicrographic Atlas of Atlantic Herring Embryonic Development. Jnl Fish Biol. 51:960-977.

Hershberger PK, NE Elder, J Wittouck, K Stick, RM Kocan. 2005. Abnormalities in Larvae from the Once-Largest Pacific Herring Population in Washington State Result Primarily from Factors Independent of Spawning Location. Trans Am Fish Soc. 134:326-337.

NOAA/BML. 2008. The 2007 Cosco Busan Oil Spill: Assessing Toxic Injury to Pacific Herring Embryos and Larvae in the San Francisco Estuary. Draft Report. NOAA Fisheries, Northwest Fisheries Science Center, Seattle WA. 42 pp.

San Francisco Estuary Institute. 2008. The Pulse of the Estuary: Monitoring and Managing Water Quality in the San Francisco Estuary. Regional Monitoring Program for Water Quality in the San Francisco Estuary, SFEI. Oakland CA; 96 pp.

## APPENDIX A

- A-1 Summary of Available Video and Still Images
- A-2 Summary of NewFields Review
- A-3 Artificial Spawn: Commentary from Video Analyses
- A-4 Artificial Spawn: Results from Video Analysis
- A-5 Natural Spawn: Commentary from Video Analyses
- A-6 Natural Spawn: Results from Video Analyses
- A-7 Natural Spawn: Analysis of Still Images for Occurrence of Body Axis Irregularities
- A-8 Illumination Assessment of Video Files (Artificial & Natural)

### A-1 SUMMARY OF AVAILABLE VIDEO AND STILL IMAGES

File Descriptor	# Still Images	# Video Clips	Quick Assessment
MRP04-FEM1-021708-A1	32	36	Usable
MRP04-FEM1-021708-A2	30	34	Usable
MRP04-FEM1-021708-A3	30	37	Usable
MRP04-FEM1-021708-A4	30	34	Usable
MRP04-FEM1-021708-A5	30	34	Usable
MRQ01-FEM1-021808-A1	31	31	Usable
MRQ01-FEM1-021808-A2	31	31	Usable
MRQ01-FEM1-021808-A3	30	31	Usable
MRQ01-FEM1-021808-A4	30	30	Poor, very pixilated screen captures, no video
MRQ01-FEM1-022908-N1	20	20	Immature animals
MRQ01-FEM1-022908-N2	20	21	Poor quality; immature animals, various planar views
MRQ01-FEM1-022908-N3	23	21	Poor quality; immature animals, various planar views
MRQ01-FEM1-022908-N4	20	20	Poor quality; immature animals, various planar views
MRQ01-FEM1-022908-N5	20	20	Poor quality; immature animals, various planar views
MRQ01-FEM1-022908-N6	20	21	Poor quality; immature animals; Clip 22 good image
MRQ01-FEM1-022908-N7	20	22	Poor quality; immature animals, various planar views
MRQ01-FEM1-022908-N8	20	20	Poor quality; immature animals, various planar views
MRQ10/P01-FEM1-021908-A1	31	31	Usable
MRQ10/P01-FEM1-021908-A2	30	32	Usable
MRQ10/P01-FEM1-021908-A3	31	33	Usable
MRQ10 P01-FEM1-021808-A4	31	31	Poor, very pixilated screen captures, no video
MRQ10/P01-FEM1-021908-A5	30	32	Usable
MRQ10 P01-FEM1-022708-N1	27	21	Poor quality; immature animals, various planar views
MRQ10 P01-FEM1-022708-N2	21	21	Poor quality; immature animals, various planar views
MRQ10 P01-FEM1-022708-N3	22	21	Poor quality; immature animals, various planar views
MRQ10 P01-FEM1-022708-N4	19	20	Poor quality; immature animals, various planar views
MRQ10 P01-FEM1-022708-N5	27	20	Poor quality; immature animals, various planar views
MRQ10 P01-FEM1-022708-N6	20	20	Poor quality; immature animals, various planar views; Clips 6 and 20 possibly good video
MRQ10 P01-FEM1-022708-N7	19	21	Usable
MRQ10 P01-FEM1-022708-N8	21	22	Usable
MRR20-FEM1-022008-A1	31	33	Usable
MRR20-FEM1-022008-A2	29	33	Usable
MRR20-FEM1-022008-A3	30	31	Usable
MRR20-FEM1-022008-A4	28	30	Usable
MRR20-FEM1-022008-A5	30	30	Usable
MRR20-FEM1-022808-N1	22	20	Poor quality; immature animals, various planar views
MRR20-FEM1-022808-N2	26	22	Poor quality; immature animals, various planar views

**SUMMARY OF AVAILABLE VIDEO AND STILL IMAGES (CONTINUED)**

MRR20-FEM1-022808-N3	19	22	Poor quality; immature animals, various planar views
MRR20-FEM1-022808-N4	26	23	Poor quality; immature animals, various planar views; there are a few that may be usable
MRR20-FEM1-022808-N5	20	19	Videos do not appear to be usable; have indistinct heart imaging
MRR20-FEM1-022808-N6	19	21	Videos do not appear to be usable; have indistinct heart imaging
MRR20-FEM1-022808-N7	21	20	Videos do not appear to be usable; have indistinct heart imaging
MRR20-FEM1-022808-N8	22	19	Videos do not appear to be usable; have indistinct heart imaging
MRT04-FEM1-022508-A1	30	31	Usable
MRT04-FEM1-022508-A2	30	35	Usable; clip 30 is a good heart image
MRT04-FEM1-022508-A3	30	32	Usable; clip 11 shows truncated pericardial region
MRT04-FEM1-022508-A4	32	35	Usable; clip 17 irregular shaped yolk sac
MRT04-FEM1-022508-A5	30	30	Usable
MRU01-FEM1-022208-A1	30	30	Usable
MRU01-FEM1-022208-A3	30	30	Usable
MRU01-FEM1-022208-A2	30	33	Usable
MRU01-FEM1-022208-A4	29	32	Usable; note may need to increase contrast
MRU01-FEM1-022608-N1	29	21	Usable; clip 20 may have been over exposed?
MRU01-FEM1-022608-N2	26	20	Usable
MRU01-FEM1-022608-N3	30	21	Usable
MRU01-FEM1-022608-N4	26	20	Some are usable but most do not clearly show heart; several images (clip 16, 21 show possible differences in pericardial region
MRU01-FEM1-022608-N5	24	20	Usable
MRU01-FEM1-022608-N6	22	24	Usable; CLIP 23 undeveloped jaw
MRU01-FEM1-022608-N7	30	20	Usable; Clip 14: elongated pericardial region
MRU01-FEM1-022608-N8	26	20	Usable; Clip 7, 4 compared to Clip 14: variation, elongation of pericardial region (how much variation is natural? Is this a developmental artifact?
whleggMRQ01-FEM1-022908-N3	6		images show process of dechoriation
Total Number	1579	1565	
# Stills for Natural Spawn (excluding MRP and MRT)	727		
# Stills for Artificial Spawn (excluding MRP and MRT)	542		
# Usable video		1109	



## A-2 SUMMARY OF NEWFIELDS REVIEW

Parameters Assessed	Artificial Spawn (#Assessed)		Natural Spawn (#Assessed)	
	MRQ10/P01 FEM1 021908 A1 (3), A2 (3), A3 (1), A5 (3)		MRQ01 FEM1 022908 N3 (3), N6 (3);	
	MRU01 FEM1 022208 A1 (3), A2 (4), A3 (3)		MRQ10 P01 FEM1 022708 N2 (3), N6 (3), N8 (3);	
	MRQ01 FEM1 021808 A1 (3), A3 (3);		MRR20 FEM1 022808 N4 (3), N7 (3)	
	MRR20 FEM1 022008 A1 (3), A 3 (3), A 4 (3)		MRU01 FEM1 022608 N2 (3), N4(3), N7(3)	
	Still Images	Video Clips	Still Images	Video Clips
Number Assessed (Status)	Previewed only	35/243	Preview only; detailed analysis of 10 samples	30/208
Development (qualitative)	x		x	
Body Axis Alignment		x	x *	x
Appearance		x	x	x
Degree of Coiling:			x	
Eye Pigmentation		x		x
Development of Jaw		x		x
Yolk Sac Descriptors		x		x
Notochord		x		x
Body Pigmentation		x		x
Heart Structure		x		x
Heart Performance		x		x
Pericardial Region/Area		x		
AV Measurement		x		
Development of Otic Vesicle		x		x
Background Illumination (qualitative)			x	x
Background Illumination (quantitative)		x		x
Other Comments		x		x

x\* = all still images (727)

**A-3 COMMENTARY FROM VIDEO ANALYSES: ARTIFICIAL SPAWN  
 MRQ01**

Site	Video Clip #	Morphology of Specimen								Comments
		Heart Structure	Heart Rate	Pericardial Region	Cranial Region	Eye Pigmentation	Jaw	Yolk Sac	Arrhythmia	
MRQ01- A1	4	Appears to have different orientation, atrium appears rounded (top view); ventricle appears enlarged	32 in 20 sec	Heart occupies 17% of the pericardial region	Otic vesicle has differentiation with two otoliths present; notochord has faint segmentation	85%	Jaw notch present, jaw not well developed	Yolk sac is flattened, rounded not pigmented	6/31, Clip 4 is flattened	
	15	Normal, backward L	31 beats per 20 sec	Heart occupies 40% of the pericardial region	Otic vesicle has differentiation with two otoliths present; notochord is not segmented; darkened	85%	Jaw notch present, jaw not well developed	Rounded, pigmented		
	28	Normal, backward L	28 beats in 20 second period	Heart occupies 32% of the pericardial region	Otic vesicle has differentiation with two otoliths present, but very heavily pigmented; segmentation of notochord not discernible	85%	Jaw notch present, jaw developing	Rounded		
MRQ01--A3	8	Normal, backward L	33 beats in 20 sec	Appears to be membrane developed which extends to heart; heart occupies 37% of pericardial region	Otic vesicle has differentiation with two otoliths present; segmentation of notochord	85%	Jaw notch present, jaw developing noticeably			
	13	Normal, backward L, anterior/posterior extension	34 beats in 20 sec	Extended posterior/anterior; heart occupies 51% of pericardial region	Head is not bent, linear; well defined otic vesicle; very faint segmentation of notochord	85%	Very developed	Elliptical		
	30	Normal, backward L	30 beats in 20 sec	Heart occupies 44% of the pericardial region	Otic vesicle has differentiation with two otoliths present; notochord is not segmented	85%	Very developed	Rounded, flattened on anterior side	Yes	

**COMMENTARY FROM VIDEO ANALYSES: ARTIFICIAL SPAWN  
 MRQ10**

Site	Video Clip #	Morphology of Specimen								Comments
		Heart Structure	Heart Rate	Pericardial Region	Cranial Region	Eye Pigmentation	Jaw	Yolk Sac	Arrhythmia	
MRQ10-A1	1	Normal in shape backward L shape with atrium most dominant, ventricle elongated	36 beats in a 20 second period; pattern was consistent	Heart occupies approximately 18% of pericardial region	Less highly differentiated	Eye appears to be colored	Jaw is not fully developed; no protrusion			Pericardium occupies a larger space than clip 5. This may be related to developmental age or possibly edema ; sample appears to be highly pigmented and granular in appearance
	5	Normal in shape backward L shape with atrium most dominant, ventricle elongated	33 beats in a 20 second period; steady beat	Heart occupies 39% of pericardial region	Less highly differentiated; notochord not clearly segmented.	Eye appears to be colored	Jaw is not fully developed; notch developing on anterior side	Round in nature and heart is overlapped by yolk sac		Pericardium may be more developed than the organism shown in Clip 1 based on the heart filling more of the pericardial space and the eye size tends to be smaller than the one in clip one. Image is darker making differentiation more difficult. Dorsum above notochord looks granulated.
	19	Normal in shape backward L shape with atrium most dominant, ventricle elongated ; ventral triangle anterior of yolk sac with septum	32 beats in 20 second period	Heart occupies 36% of pericardial region; a more defined membrane bridging the ventral pericardial area to the yolk sac	Highly differentiated	Eye appears to be colored	Jaw is starting to become more pronounced	Normal in appearance		Granulated yolk sac
MRQ10-A2	3		28 beats in 20 second period	Heart occupies 19% of the pericardial region	Well defined; notochord not segmented	>85%; appears colored	Jaw is not fully developed; no protrusion	Yolk sac appears rounded; resembles stage q development		
	18		30 beats in 20 second period	Heart occupies 29% of the pericardial region	Well defined; notochord not segmented	>85%; appears colored	Jaw not fully developed; no protrusion- anterior notch	Yolk sac appears rounded; resembles stage q development		
	26		30 beats in 20 second period	Heart occupies 29% of the pericardial region	Well defined; notochord not segmented	>85%; appears colored	Jaw not fully developed; no protrusion	Yolk sac appears rounded; resembles stage q development		

**COMMENTARY FROM VIDEO ANALYSES: ARTIFICIAL SPAWN**  
**MRQ10 - Continued**

Site	Video Clip #	Morphology of Specimen								Comments
		Heart Structure	Heart Rate	Pericardial Region	Cranial Region	Eye Pigmentation	Jaw	Yolk Sac	Arrhythmia	
MRQ10-A3	29		34 beats in 20 seconds	Heart occupies 28% of the pericardial region	Obvious segmentation in the notochord	85% pigmentation	Jaw notch present, jaw not well developed	Rounded		
MRQ10-A5	1	Heart has a pinch on the ventral side when it is at the bottom of the down beat otherwise backward L; goes into the yolk sac on anterior side	30 beats in 20 second period	Heart occupies 27% of the pericardial region	Otic vesicle has differentiation with two otoliths present; notochord is not segmented	85% eye pigmentation	Jaw notch present, jaw not well developed	Yolk sac is intermediate between round and elliptical and appears to be granulated		
	8		30 beats in 20 second period	Heart occupies 24% of the pericardial region	Otic vesicle has differentiation with two otoliths present; notochord has faint segmentation	85% eye pigmentation	Jaw notch present, jaw not well developed	Yolk sac is intermediate between round and elliptical and appears to be granulated		
	20	Appears to have a different orientation , atrial side is bulbous and extends beneath the yolk sac	30 beats in 20 second period	Heart occupies 28% of the pericardial region which appears to be compressed than other images	Otic vesicle has differentiation with two otoliths present; notochord is not segmented	85% eye pigmentation	Jaw is not developed but a notch is forming	Yolk sac is rounded and pushes into the pericardial region definitive and yolk sac is granulated		Pigmentation on dorsal side of head

**COMMENTARY FROM VIDEO ANALYSES: ARTIFICIAL SPAWN  
 MRR20**

Site	Video Clip #	Morphology of Specimen								Comments
		Heart Structure	Heart Rate	Pericardial Region	Cranial Region	Eye Pigmentation	Jaw	Yolk Sac	Arrhythmia	
MRR20-FEM1-022008-A1	12	Not normal, extremely compressed pericardial region	24 beats per 20 sec	Completely abnormal, heart intruding into yolk sac or vice versa; Not measured	Not well defined; pigmentation	<50% eye pigmentation	Evident, but not protruded, and seems out of normal position	Flattened		
	25	Y- shaped; atrium is on top of yolk sac	29 beats per 20 sec	Heart occupies 29% of the pericardial region	Otic vesicle has differentiation with two otoliths present; notochord has faint segmentation	60% eye pigmentation	Jaw notch present	Rounded, pigmented	No	
	32	Y-shaped; atrium is on top of yolk sac	33 beats per 20 sec	Heart occupies 43% of the pericardial region	Otic vesicle has differentiation with two otoliths present; segmentation of notochord	60% eye pigmentation	Jaw notch present	Rounded; slight flattening	No	
MRR20-FEM1-022008-A3	5	Backwards L, with atrial intrusion into yolk sac area	39 beats per 19 sec; major shutter at end of video (animal moved out of frame)	Pigmented; secondary pericardial membrane evident; heart occupies 38% of pericardial region	Otic vesicle has differentiation with two otoliths and semi-circular canals present dorsally and ventrally; notochord has faint segmentation; pigmentation	<85%	Developing jaw notch, intermediate protrusion	Rounded, flattened anterior side		
	19	Normal backwards L	42 beats per 20 sec	Heart occupies 37% of pericardial region	Differentiated, otoliths developed, branching semi-circular canals; notochord faintly visible	60%	Developing jaw notch, protrusion	Elliptical		
	26	Tipped backwards L; heart and yolk region overlaps	37 beats per 20 sec	Heart occupies 37% of pericardial region	Heavy pigmentation; indistinct features in otic vesicle, notochord faintly segmented	<50 %; Eyes appear large compared to head region	Rudimentary notch present	Elliptical		

**COMMENTARY FROM VIDEO ANALYSES: ARTIFICIAL SPAWN  
 MRR20 - Continued**

Site	Video Clip #	Morphology of Specimen								Comments
		Heart Structure	Heart Rate	Pericardial Region	Cranial Region	Eye Pigmentation	Jaw	Yolk Sac	Arrhythmia	
MRR20-FEM1-022008-A5	11	Backward L, with atrial intrusion into yolk sac area	41 beats per 20 sec		Rudimentary development with faint segmentation of notochord; organism more transparent than most	ND	Rudimentary notch present	Elliptical; 6/30	Yes	
	17	Very compressed; shape not clearly visible, very elongated	45 beats per 20 sec	Very Compressed;	Rudimentary development, w/ 2 otoliths; faint segmentation of notochord; organism more transparent than most	20%	Transparent, rudimentary, slight notch	Elliptical, flattened	Yes	
	24	Very compressed; shape visible; Backward L; atrium overlapping yolk sac	37 beats in 20 sec	Very Compressed;	Rudimentary development with hardly any segmentation or the notochord	30%	Early stage of notch formation	Rounded, flattened	yes	

**COMMENTARY FROM VIDEO ANALYSES: ARTIFICIAL SPAWN  
 MRU01**

Site	Video Clip #	Morphology of Specimen								Comments
		Heart Structure	Heart Rate	Pericardial Region	Cranial Region	Eye Pigmentation	Jaw	Yolk Sac	Arrhythmia	
MRU01-FEM1-022208-A1	6	Normal in shape backward L shape with atrium most dominant, ventricle elongated	36 beats in a 20 second period; pattern was consistent	Heart occupies approximately 37 % of pericardial region	Highly differentiated	85% pigmented	Jaw is starting to develop with a notch	Yolk sac is elliptical		Specimen has a pigmented granular appearance
	18	Abnormal in shape; V-shape, horizontal plane and extends into the yolk sac	33 beats in a 20 second period; steady beat	Heart occupies approximately 46% of pericardial region;	Highly differentiated; notochord segmentation is not obvious	>85% pigmented	Jaw is not fully developed; notch starting to developing on anterior side			Specimen has a pigmented granular appearance
	22	Normal in shape backward L shape with atrium bulbous appearance, ventricle elongated	32 beats in 20 second period	Heart occupies approximately 38% of pericardial region;	Highly differentiated; some segmentation of the notochord	Colored eye	Jaw well developed and starting to protrude	Elliptical shape		Specimen has a pigmented granular appearance
MRU01-FEM1-022208-A3	1		42 beats in 20 second period	Heart occupies 41% (measured) of the pericardial region	Well defined	~85% pigmentation	Jaw is starting to become more pronounced	Yolk sac is elliptical (checked with elliptical tool)		
	14		38 beats in 20 second period	Heart occupies approximately 33% (measured) of the pericardial region	Well defined	> 85% colored	Jaw very pronounced			
	21		39 beats in 20 second period	Heart occupies 35% (measured) of the pericardial region; heart has v-shape aspect rather than backwards L	Well defined	~85% pigmentation	Jaw very pronounced			Ventral edge of pericardial region not smooth arc with jaw protrusion; some pigmentation

**COMMENTARY FROM VIDEO ANALYSES: ARTIFICIAL SPAWN  
 MRU01 - Continued**

Site	Video Clip #	Morphology of Specimen								Comments
		Heart Structure	Heart Rate	Pericardial Region	Cranial Region	Eye Pigmentation	Jaw	Yolk Sac	Arrhythmia	
MRU01-FEM1-022208-A2	1		31 beats in 20 second period	Heart occupies 33% (measured) of the pericardial region	Difficult to evaluate due to lighting; notochord segmentation evident	~85% pigmentation	Jaw is starting to protrude	Yolk sac is elliptical		Very light specimen, difficult to perceive depth
	8		35 beats in 20 second period	Heart occupies 34% (measured) of the pericardial region; heart protrudes into yolk region	Difficult to evaluate due to lighting; notochord segmentation evident	> 85% colored	Jaw is starting to protrude	Yolk sac is intermediate; more flattened appearance on pericardial side		
	10		34 beats in 20 second period	Heart occupies 46% (measured) of the pericardial region;	Difficult to evaluate due to lighting; notochord segmentation not as developed	> 85% colored	Jaw notch is forming	Yolk sac is rounded		
	17		39 beats in 20 second period	Heart occupies 33% (measured) of the pericardial region; atrial side is bulbous in shape	Well defined; segmentation of notochord very well defined (more than clip 10)	> 85% colored	Jaw notch is well formed	Yolk sac is elliptical		



**A-4 RESULTS FROM ARTIFICIAL SPAWN VIDEO ANALYSIS  
 MRQ01**

Site	Video Clip #	Frame	Position of AV Canal during Heartbeat	Record	Angle	Length (px)	Potential % Length Difference	Heart Rate / 20sec	Heart Area	Pericardial Region	%Heart: Pericardium	Yolk Sac Flattening (Group Video Images)			Arrhythmia (Individual Video Images)		
												Yes	# in Sample	Freq	Yes/No	# in Sample	Freq
												<b>MRQ01-FEM1-021808-A1</b>					
	4		Minimum	Clip 4 Downbeat jpg	-119.8	134.8	9.9%	32	5973	35196	17%	yes			no		
	4		Maximum	Clip 4 Downbeat jpg	110.1	148.1	9.0%										
	15		Minimum	Clip 15 Upbeat jpg	125.5	51.6	38.8%	31	7867	19627	40%	no			no		
	15		Maximum	Clip 15 Upbeat jpg	-125.9	71.6	27.9%										
	28	27/618	Minimum	Clip 28 Downbeat jpg	108.7	59.1	35.4%	28	9125	28743	32%	no			no		
	28		Maximum	Clip 28 Downbeat jpg	108.2	80	26.1%										
<b>Mean</b>							<b>24.5%</b>	<b>30.33</b>	<b>7,655.0</b>	<b>27,855.3</b>	<b>30%</b>						
<b>MRQ01-FEM1-021808-A3</b>												5	31		1	3	
	8	48/622	Minimum	Clip 8 Upbeat jpg	-103.2	48.3	36.6%	33	10752	28734	37%	no			no		
	8		Maximum	Clip 8 Upbeat jpg	-103	66	26.8%										
	13	44/623	Minimum	Clip 13 Downbeat jpg	105.3	34.2	42.7%	34	8167	15977	51%	no			no		
	13		Maximum	Clip 13 Downbeat jpg	-100.6	48.8	29.9%										
	30	135/619	Minimum	Clip 30 Upbeat jpg	109.9	50	37.2%	30	7522	16954	44%	yes			yes		
	30		Maximum	Clip 30 Upbeat jpg	108.7	68.6	27.1%										
<b>Mean</b>							<b>33.4%</b>	<b>32.33</b>	<b>8,813.7</b>	<b>20,555.0</b>	<b>44%</b>	11	62	18%	1	6	17%

**RESULTS FROM ARTIFICIAL SPAWN VIDEO ANALYSES  
MRQ10**

Site	Clip #	Frame	Position of AV Canal during Heartbeat	Record	Angle	Length (px)	Potential % Length Difference	Heart Rate / 20sec	Heart Area	Pericardial Region	%Heart: Pericardium	Yolk Sac Flattening (Group Video Images)			Arrhythmia (Individual Video Images)		
												Yes	# in Sample	Freq	Yes	# in Sample	Freq
<b>MRQ10/P01-FEM1-021908-A1</b>												11	31		0	3	
	1		Minimum	Clip 1 Upbeat SC	69.3	113.3	29.6%	36	9966	54305	18%						
	1		Maximum	Clip 1 Upbeat SC	109.9	146.8	22.8%										
	5		Minimum	Clip 5 Downbeat SC	104.9	93.1	15.1%	33	8635	21986	39%						
	5		Maximum	Clip 5 Downbeat SC	104	107.2	13.2%										
	19		Minimum	Clip 19 Downbeat SC	72.5	79.7	32.1%	32	9350	25879	36%						
	19		Maximum	Clip 19 Downbeat SC	104.3	105.3	24.3%										
	Mean						22.9%	33.7	9,317	34,057	31%						
<b>MRQ10/P01-FEM1-021908-A2</b>												10	32		0	3	
	3		Minimum	Clip 3 Upbeat SC	74.5	134.9	22.6%	28	8842	46591	19%						
	3		Maximum	Clip 3 Upbeat SC	104.7	165.4	18.4%										
	18		Minimum	Clip 18 Upbeat SC	99.7	95.4	18.9%	30	11108	38878	29%						
	18		Maximum	Clip 18 Upbeat SC	99.1	113.4	15.9%										
	26		Minimum	Clip 26 Upbeat SC	104.3	97	12.5%	30	8896	38894	23%						
	26		Maximum	Clip 26 Upbeat SC	103.8	109.1	11.1%										
	Mean						16.6%	29.3	9,615.3	41,454.3	23%						
<b>MRQ10/P01-FEM1-021908-A3</b>												9	33		0	1	
	29		Minimum	Clip 29 Downbeat SC	105.4	105.8	16.6%	34	9943	35114	28%						
	29		Maximum	Clip 29 Downbeat SC	107	123.4	14.3%										
	Mean						15.4%										

**RESULTS FROM ARTIFICIAL SPAWN VIDEO ANALYSES**  
**MRQ10 - Continued**

Site	Video Clip #	Frame	Position of AV Canal during Heartbeat	Record	Angle	Length (px)	Potential % Length difference	Heart Rate / 20sec	Heart Area	Pericardial Region	%Heart: Pericardium	Yolk Sac Flattening (Group Video Images)			Arrhythmia (Individual Video Images)		
												Yes	# in Sample	Freq	Yes	# in Sample	Freq
MRQ10/P01-FEM1-021908-A5												9	32		0	3	
	1b		Minimum	Clip 1 Upbeat SC	106.5	112.6	24.2%	30	8570	31534	27%						
	1b		Maximum	Clip 1 Upbeat SC	106.6	139.8	19.5%										
	8		Minimum	Clip 8 Upbeat SC	102.7	86.1	21.7%	30	8490	35228	24%						
	8		Maximum	Clip 8 Upbeat SC	104.4	104.8	17.8%										
	20		Minimum	Clip 20 Upbeat SC	105.3	80.2	25.7%	30	6603	23643	28%						
	20		Maximum	Clip 20 Upbeat SC	110.2	100.8	20.4%										
<b>Mean</b>							<b>21.5%</b>	<b>30.00</b>	<b>7,887.7</b>	<b>30,135.0</b>	<b>26%</b>	39	128	30%	<b>0</b>	<b>10</b>	<b>0%</b>

**RESULTS FROM ARTIFICIAL SPAWN VIDEO ANALYSIS  
MRR20**

Site	Video Clip #	Frame	Position of AV Canal during Heartbeat	Record	Angle	Length (px)	Potential % Length Difference	Heart Rate / 20sec	Heart Area	Pericardial Region	%Heart: Pericardium	Yolk Sac Flattening (Group Video Images)			Arrhythmia (Individual Video Images)		
												Yes	# in Sample	Freq	Yes/No	# in Sample	Freq
<b>MRR20-FEM1-022008-A1</b>												11	33		0	3	
	12	ND	Minimum	Not Measured	ND	ND	ND	24	ND	ND	ND	yes			no		
	25	26/616	Minimum	Clip 25 Upbeat jpg	117.6	77.8	24.2%	29	6378	22227	29%	no			no		
	25		Maximum	Clip 25 Upbeat jpg	117.1	96.6	19.5%										
	32		Minimum	Clip 32 Upbeat jpg	131.1	62.4	29.5%	33	8292	19217	43%	yes			no		
	32		Maximum	Clip 32 Upbeat jpg	131	80.8	22.8%										
Mean							24.0%	28.67	7,335.0	20,722.0	36%						
<b>MRR20-FEM1-022008-A3</b>												9	31		0	3	
	5		Minimum	Clip 5 Downbeat jpg	70.6	90.1	4.2%	39	8930	23503	38%	yes					
	5		Maximum	Clip 5 Downbeat jpg	71.4	93.9	4.0%										
	19	58/620	Minimum	Clip 19 Upbeat jpg	108.7	59.1	39.1%	42	11145	30132	37%	no					
	19		Maximum	Clip 19 Upbeat jpg	108.4	82.2	28.1%										
	26	182/614	Minimum	Clip 26 Upbeat jpg	90.7	77	24.7%	37	10788	29014	37%	no					
	26		Maximum	Clip 26 Upbeat jpg	90	96	19.8%										
Mean							20.0%	39.33	10,288	27,549.7	37%						

**RESULTS FROM ARTIFICIAL SPAWN VIDEO ANALYSIS**  
**MRR20 - Continued**

Site	Video Clip #	Frame	Position of AV Canal during Heartbeat	Record	Angle	Length (px)	Potential % Length Difference	Heart Rate / 20sec	Heart Area	Pericardial Region	%Heart: Pericardium	Yolk Sac Flattening (Group Video Images)			Arrhythmia (Individual Video Images)		
												Yes	# in Sample	Freq	Yes/No	# in Sample	Freq
<b>MRR20-FEM1-022008-A5</b>												6	30		3	3	
	11	137/609	Minimum	Clip 11 Upbeat jpg	48.2	51	39.0%	41	9284	21837	43%	no			yes		
	11		Maximum	Clip 11 Upbeat jpg	49.6	70.9	28.1%										
	17	223/611	Minimum	Clip 17	-123.1	109.8	0.0%	45	5606	10861	52%	yes			yes		
	17		Maximum	Clip 17	123.1	109.8	0.0%										
	24	300/599	Minimum	Clip 24 Upbeat jpg	131.7	97.7	18.3%	37	7748	10652	73%	yes			yes		
	24		Maximum	Clip 24 Upbeat jpg	-130.4	115.6	15.5%										
<b>Mean</b>				-			<b>16.8%</b>	<b>41.00</b>	<b>7,546.0</b>	<b>14,450.0</b>	<b>56%</b>	<b>26</b>	<b>94</b>	<b>28%</b>	<b>3</b>	<b>9</b>	<b>33%</b>

**RESULTS FROM ARTIFICIAL SPAWN VIDEO ANALYSIS**

**MRU01**

Site	Video Clip #	Frame	Position of AV Canal during Heartbeat	Record	Angle	Length (px)	Potential % Length difference	Heart Rate / 20sec	Heart Area	Pericardial Region	%Heart: Pericardium	Yolk Sac Flattening (Group Video Images)			Arrhythmia (Individual Video Images)		
												Yes	# in Sample	Freq	Yes/No	# in Sample	Freq
<b>MRU01-FEM1-022208-A1</b>												11	30		0	3	
	6		Minimum	Clip 6 Upbeat SC	106.2	57.3	28.8%	36	8653	23198	37%						
	6		Maximum	Clip 6 Upbeat SC	105.7	73.8	22.4%										
	18		Minimum	Clip 18 Upbeat SC	117.3	34.9	49.9%	33	9298	20303	46%						
	18		Maximum	Clip 18 Upbeat SC	116.7	52.3	33.3%										
	22		Minimum	Clip 22 Upbeat SC	112.1	39.9	56.9%	32	9666	25138	38%						
	22		Maximum	Clip 22 Upbeat SC	109.6	62.6	36.3%										
Mean				-			37.9%	33.67	9,205.7	22,879.7	41%						
<b>MRU01-FEM1-022208-A2</b>												9	33		0	4	
	1c		Minimum	Clip 1c Upbeat SC	114.7	40.7	35.6%	31	5514	16737	33%						
	1c		Maximum	Clip 1c Upbeat SC	67.6	55.2	26.3%										
	8		Minimum	Clip 8 Downbeat SC	107.9	64.5	18.3%	35	4914	14576	34%						
	8		Maximum	Clip 8 Downbeat SC	109.1	76.3	15.5%										
	10		Minimum	Clip 10 Downbeat SC	121.1	67.6	35.7%	34	10898	23493	46%						
	10		Maximum	Clip 10 Downbeat SC	122.1	91.7	26.3%										
	17		Minimum	Clip 17 Upbeat SC	111.4	50.3	36.4%	39	9764	29864	33%						
	17		Maximum	Clip 17 Upbeat SC	114	68.6	26.7%										
Mean				-			26.5%	36.00	8,525.3	22,644.3	38%						

**RESULTS FROM ARTIFICIAL SPAWN VIDEO ANALYSIS**  
**MRU01 - Continued**

Site	Video Clip #	Frame	Position of AV Canal during Heartbeat	Record	Angle	Length (px)	Potential % Length difference	Heart Rate / 20sec	Heart Area	Pericardial Region	%Heart: Pericardium	Yolk Sac Flattening (Group Video Images)			Arrhythmia (Individual Video Images)		
												Yes	# in Sample	Freq	Yes/No	# in Sample	Freq
												<b>MRU01-FEM1-022208-A3</b>					
	1b		Minimum	Clip 1 Upbeat SC	-108.8	49.3	28.2%	42	9672	23590	41%						
	1b		Maximum	Clip 1 Upbeat SC	-108.4	63.2	22.0%										
	14		Minimum	Clip 14 Downbeat SC	-111.8	53.9	39.1%	38	10283	31419	33%						
	14		Maximum	Clip 14 Downbeat SC	-111.1	75	28.1%										
	21		Minimum	Clip 21 Upbeat SC	120.3	55.6	12.6%	39	10192	29489	35%						
	21		Maximum	Clip 21 Upbeat SC	-118.6	62.6	11.2%										
<b>Mean</b>							<b>23.5%</b>	<b>39.67</b>	<b>10,049</b>	<b>28,166.0</b>	<b>36%</b>	<b>26</b>	<b>93</b>	<b>28%</b>	<b>0</b>	<b>10</b>	<b>0%</b>

**A-5 COMMENTARY ON NATURAL SPAWN VIDEO ANALYSES  
 MRQ01**

Site	Video Clip #	Morphology										Comments
		Planar View	Rotation from Lateral Plane	Heart Structure	Heart Rate	Pericardial Region	Development of Otic Vesicle	Eye Pigmentation	Jaw	Yolk Sac	Arrhythmia	
MRQ01-N3	11	Left side		Abnormal	11 beats/20 sec	Compressed	Rudimentary	<50%	NM	Rounded, heavily pigmented		
	12	Left side	Yes	Not discernible	29/27 sec ~22/20	Compressed	Not discernible	<50%	Not present	Rounded, flattening, secondary membrane		
	17	Left side ventral	Yes	Not discernible	27/20 sec	Not discernible	No view	<50%	Not present	Rounded, flattening, granular appearance		
MRQ01-N6	9	Left side	Yes	Not discernible	31/20 sec	Compressed	No view	<50%	Not present	Rounded, heavily pigmented, secondary membrane		
	10	Left side	Yes	Not discernible	28/20 sec	Compressed	No view	NM	Not present	Rounded, heavily pigmented, granular appearance		
	22	Left side ventral	Yes	Not discernible	13/20 sec	Not discernible	No view	NM	Not present	Rounded, flattening of anterior side, secondary membrane		



**COMMENTARY ON NATURAL SPAWN VIDEO ANALYSES  
MRQ10**

Site	Video Clip #	Morphology										Comments
		Planar View	Rotation from Lateral Plane	Heart Structure	Heart Rate	Pericardial Region	Otic Vesicle	Eye Pigmentation	Jaw	Yolk Sac	Arrhythmia	
MRQ10-N2	91	Left side	No	Backward L	35/20 sec	Compressed, overlaps with yolk sac	Yes with some definition	NM	Rudimentary	Rounded, flattening, granular appearance		
	100	Left side	No	Backward L	38/21 sec	Appears normal may be some slight swelling need to make measurements	Yes with definition	~50%	Yes-slight protrusion	Intermediate between rounded and elliptical, somewhat granular in nature		
	101	Left side	No	Backward L	36/20 sec	Appears normal need to make measurements	Yes with definition	<50%	Yes with protrusion	Elliptical with granulation		
MRQ10-N6	10	Left side	Yes	Backward L	38/20 sec	Appears normal need to make measurements	Yes with definition	NM	Rudimentary	Rounded, flattening on anterior side		
	20	Left side	No	Backward L	48/20 sec	Appears normal may be some slight swelling need to make measurements	Yes with some definition	~50%	Yes-slight protrusion	Rounded, some granulation, secondary measurement		Segmented notochord
	22	Left side	No	Backward L	44/20 sec	Compressed, overlaps with yolk sac	Yes with definition	NM	Yes-slight protrusion	Intermediate between rounded and elliptical, somewhat granular in nature		Segmented notochord
MRQ10-N8	5	Left side	No	Backward L	37/20 sec	Appears normal, measurements possible	Yes with definition	>50%	Yes-slight protrusion	Rounded		Segmented notochord
	15	Left side	No	Backward L	34/20 sec	Compressed, overlaps with yolk sac, measurements possible	Yes with definition	<50%	Yes with protrusion	Rounded		
	22	Left side	No	Backward L	40/20 sec	Appears normal need to take measurements	Rudimentary	<50%	Rudimentary	Rounded		Segmented notochord, sample is washed out

**COMMENTARY ON NATURAL SPAWN VIDEO ANALYSES  
 MRR20**

Site	Video Clip #	Morphology										Comments
		Planar View	Rotation from Lateral Plane	Heart Structure	Heart Rate	Pericardial Region	Otic Vesicle	Eye Pigmentation	Jaw	Yolk Sac	Arrhythmia	
MRR20- N4	9	Left side	No	Backward L	39/20 sec	Appears normal need to take measurements	Yes with definition; notochord very faint	~50%	Yes-slight protrusion	Rounded		
	20	Left side	Yes	Not discernible	42/20 sec	Compressed; some overlap with yolk sac, may be a secondary membrane anterior of yolk sac in lower corner	Yes with some definition; notochord not discernible	NM	Yes-slight protrusion	Rounded, with some flattening anterior side of yolk sac		
	23	Left side	No	Backward L	50/21 sec	Appear elongated from head to yolk sac; ventricle bulbous; heart occupies left side of pericardial region, measurements possible	Rudimentary, some definition of notochord	50%	Rudimentary	Only partially viewed in image		Segmented notochord
MRR20-N7	51	Ventral	Yes	Not discernible	38/20 sec	Not discernible	Rudimentary	<50%	Not present	Rounded with flattening of anterior edge, pigmented		Specimen is very granular
	57	Left side	No	Not discernible	50/20 sec	Elongated appearance, may have swollen pericardial, measurement not possible; secondary membrane is distinct and appears granulated	Yes with some definition	<50%	Yes-slight protrusion	Elliptical with a pinch on anterior side, double membrane		Notochord with some segmentation, specimen has ghost-like appearance
	66	right side	Yes	Not discernible	34/21 sec	Not discernible	Not discernible	Not discernible	Not discernible	Rounded with flattening of anterior edge, pigmented		Pectoral fin is present on right side of pericardial region

**COMMENTARY ON NATURAL SPAWN VIDEO ANALYSES**  
**MRU01**

Site	Video Clip #	Morphology										Comments
		Planar View	Rotation from Lateral Plane	Heart Structure	Heart Rate	Pericardial Region	Otic Vesicle	Eye Pigmentation	Jaw	Yolk Sac	Arrhythmia	
MRU01-FEM1-022608-N2	3	Left side	No	Backward L-attached to jaw region on the ventricle side	42/20 sec	Appears normal, measurements possible	Yes with definition	Not discernible	Yes-intermediate protrusion	Rounded		Notochord with segments, still images look good most are linear
	14	Left side	No	Backward L-attached to jaw region on the ventricle side	35/20 sec	Appears normal, measurements possible, secondary membrane anterior of yolk sac	Yes with definition	Not discernible	Yes, intermediate protrusion	Rounded		
	19	Left side	No	Backward L-attached to jaw region on the ventricle side	34/20 sec	Appears normal, measurements possible	Yes with definition	Not discernible	Yes-slight protrusion	Rounded		Image somewhat grainy and dark
MRU01-FEM1-022608-N4	5	Left side	No	Backward L-attached to jaw region on the ventricle side	36/20 sec	Appears normal, measurements possible	Yes with some definition	Not discernible	Yes-slight protrusion	Rounded, with some flattening anterior side of yolk sac		Notochord with segments, still images look good most are linear
	11	Left side	No	Y - laying on its side	37/20 sec	Appears normal, measurements possible	Yes with definition	85%	Yes with protrusion	Rounded		Notochord with segments
	14	Left side	No	Backward L	36/20 sec	Elongated appearance, may have swollen pericardial with extended secondary pericardial membrane evident, measurement possible	Yes with some definition	Not discernible	Yes-very protruded	Rounded with double membrane on ventral edge		Notochord with some segmentation

**COMMENTARY ON NATURAL SPAWN VIDEO ANALYSES  
 MRU01- CONTINUED**

Site	Video Clip #	Morphology										Comments
		Planar View	Rotation from Lateral Plane	Heart Structure	Heart Rate	Pericardial Region	Otic Vesicle	Eye Pigmentation	Jaw	Yolk Sac	Arrhythmia	
MRU01-FEM1-022608-N7	8	Left side	No	Not discernible	37/20 sec	Elongated appearance almost looks like two distinct pericardial regions, may have swollen pericardial, measurement possible	Yes with definition	<50%	Yes-very protruded, extends under eye	Appears rounded, only partially in view		Notochord with some segmentation
	16	Left side	No	Backward L-attached to jaw region on the ventricle side	37/20 sec	Heart occupies left side of pericardial region, measurements possible	Yes with definition	<50%	Yes-very protruded, extends under eye	Appears rounded, only partially in view		Notochord with some segmentation, image has some granulation
	21	Left side	No	Backward L-attached to jaw region on the ventricle side	40/20 sec	Elongated appearance almost looks like two distinct pericardial regions, secondary pericardial membrane evident, measurement possible	Yes with definition	~50%	Yes-very protruded, extends under eye	Appears rounded with double membrane, only partially in view		

**A-6 RESULTS FROM NATURAL SPAWN VIDEO ANALYSES**

**MRQ01**

Site	Clip #	Frame	Position of AV Canal during Heartbeat	Record	Angle	Length (px)	Difference Length (%)	Heart Rate / 20sec	Heart Area	Area Pericardial Region	%Heart: Pericardium	Yolk Sac Flattening (Group Video Images)			Arrhythmia (Individual Video Images)		
												Yes	# in Sample	Freq	Yes	# in Sample	Freq
MRQ01-FEM1-022908-N3												NM*	NM	NM	0	3	0
	11		NM	NM	NM	NM	NM	11			NM	no					
	12		NM	NM	NM	NM	NM	22			NM	yes					
	17		NM	NM	NM	NM	NM	27			NM	yes					
MRQ01-FEM1-022908-N6																	
	9		NM	NM	NM	NM	NM	31			NM	no			0	3	0
	10		NM	NM	NM	NM	NM	28			NM	no					
	22		NM	NM	NM	NM	NM	13			NM	yes					
<b>MRQ01 Mean</b>								<b>22</b>				<b>NM</b>	<b>NM</b>	<b>NM</b>	<b>0</b>	<b>3</b>	<b>0</b>

**RESULTS FROM NATURAL SPAWN VIDEO ANALYSES  
 MRQ10**

Site	Clip #	Frame	Position of AV Canal during Heartbeat	Record	Angle	Length (px)	Difference Length (%)	Heart Rate / 20sec	Heart Area	Area Pericardial Region	%Heart: Pericardium	Yolk Sac Flattening (Group Video Images)			Arrhythmia (Individual Video Images)		
												Yes	# in Sample	Freq	Yes	# in Sample	Freq
MRQ10/P01-FEM1-022708-N2												9	21	43%	1	3	
	91		NM	NM	NM	NM	NM	35			NM	yes					
	100		Minimum	Clip 100 Upbeat.jpg	66.6	47.9	57%	38	9811	24398	40%	no					
	100		Maximum	Clip 100 Upbeat.jpg	109.9	146.8	67.4%										
	101	62/620	Minimum	Clip 101 Upbeat.jpg	55.3	47.4	36.9%	36	6464	20752	31%	yes					
	101		Maximum	Clip 101 Upbeat.jpg	56.3	64.9	27.0%										
MRQ10/P01-FEM1-022708-N6												9	20	45%	0	3	
	10		Minimum	Clip 10 Downbeat.jpg	-78.3	78.6	19.7%	38	7480	19338	39%						
	10		Maximum	Clip 10 Downbeat.jpg	-77.7	94.1	16.5%										
	20	54/614	Minimum	Clip 20 Upbeat.jpg	63	55	34.2%	48	10342	31904	32%						
	20		Maximum	Clip 20 Upbeat.jpg	63.4	73.8	25.5%										
	22		Minimum	Clip 20 Upbeat.jpg	69.4	59.8	12.7%	44	8947	18607	48%						
	22		Maximum	Clip 20 Upbeat.jpg	69.1	67.4	11.3%										

**RESULTS FROM NATURAL SPAWN VIDEO ANALYSES**  
**MRQ10 – Continued**

Site	Clip #	Frame	Position of AV Canal during Heartbeat	Record	Angle	Length (px)	Difference Length (%)	Heart Rate / 20sec	Heart Area	Area Pericardial Region	%Heart: Pericardium	Yolk Sac Flattening (Group Video Images)			Arrhythmia (Individual Video Images)		
												Yes	# in Sample	Freq	Yes	# in Sample	Freq
MRQ10/P01-FEM1-022708-N8												6	22	27%			
	5		Minimum	Clip 5 Upbeat	95.1	45.2	40.0%	37	10713	27950	38%	no			0	3	0
	5	26/612	Maximum	Clip 5 Upbeat	95.4	63.3	28.6%										
	15		Minimum	Clip 15 - Upbeat	112	50.7	30.8%	34	8113	13612	60%	no					
	15		Maximum	Clip 15 - Upbeat	113.1	66.3	23.5%										
	22	23/621	Minimum	Clip 22 - Downbeat	100.8	47.9	11.5%	40	7269	24852	29%	no					
	22		Maximum	Clip 22 - Downbeat	77	53.4	10.3%										
<b>MRQ10 Mean</b>						<b>26.4%</b>	<b>37.6%</b>	<b>39</b>	<b>,642</b>	<b>22,677</b>	<b>40%</b>	<b>24</b>	<b>63</b>	<b>36%</b>	<b>1</b>	<b>9</b>	<b>11 %</b>

**RESULTS FROM NATURAL SPAWN VIDEO ANALYSES  
 MRR20**

Site	Clip #	Frame	Position of AV Canal during Heartbeat	Record	Angle	Length (px)	Difference Length (%)	Heart Rate / 20sec	Heart Area	Area Pericardial Region	%Heart: Pericardium	Yolk Sac Flattening (Group Video Images)			Arrhythmia (Individual Video Images)		
												Yes	# in Sample	Freq	Yes	# in Sample	Freq
MRR20-FEM1-022808-N4												NM	NM	NM	0	3	
	79	100/678	Minimum	Clip 9 Downbeat	90	91	9.9%	39	8194	30013	27%	no					
	9		Maximum	Clip 9 Downbeat	90	100	9.0%										
	20	161/622	Minimum	Clip 20 Downbeat	90	49	40.8%	42	8913	26509	34%	yes					
	20		Maximum	Clip 20 Downbeat	90	69	29.0%										
	23	18/634	Minimum	Clip 23 Upbeat	58.2	49.4	24.7%	48	8446	35472	24%	no					
	23		Maximum	Clip 23 Upbeat	57.6	61.6	19.8%										
MRR20-FEM1-022808-N7												10	18	56%	2	3	
	51		Minimum	Clip 51	NM	NM	NM	38	NM	NM	NM						
	51		Maximum	Clip 51	NM	NM	NM		NM	NM							
	57		Minimum	Clip 57	NM	NM	NM	50	NM	NM	NM						
	57		Maximum	Clip 57	NM	NM	NM		NM	NM							
	66		Minimum	Clip 66	NM	NM	NM	34	NM	NM	NM						
	66		Maximum	Clip 66	NM	NM	NM		NM	NM							
<b>MRR20 Mean</b>							<b>22.2%</b>	<b>41.8</b>	<b>8518</b>	<b>30665</b>	<b>28%</b>	<b>10</b>	<b>18</b>	<b>56%</b>	<b>0</b>	<b>6</b>	<b>33%</b>



**RESULTS FROM NATURAL SPAWN VIDEO ANALYSES  
 MRU01**

Site	Clip #	Frame	Position of AV Canal during Heartbeat	Record	Angle	Length (px)	Difference Length (%)	Heart Rate / 20sec	Heart Area	Area Pericardial Region	%Heart: Pericardium	Yolk Sac Flattening (Group Video Images)			Arrhythmia (Individual Video Images)		
												Yes	# in Sample	Freq	Yes	# in Sample	Freq
MRU01-FEM1-022608-N2												8	20	40%	0	3	
	3	9/616	Minimum	Clip 3 Upbeat	112	50.7	32.9%	42	12744	29101	44%	no					
	3		Maximum	Clip 3 Upbeat	110.9	67.4	24.8%										
	14	57/604	Minimum	Clip 14 Downbeat	114.9	45.2	38.7%	35	11691	19242	61%	no					
	14		Maximum	Clip 14 Downbeat	66.3	62.7	27.9%										
	19	111/607	Minimum	Clip 19 Downbeat	90	41	39.0%	34	11950	21157	56%	no					
	19		Maximum	Clip 19 Downbeat	90	57	28.1%										
MRU01-FEM1-022608-N4												7	20	35%	0	3	
	5	17/621	Minimum	Clip 5 Upbeat	92.7	63.1	28.4%	36	11395	42087	27%	yes					
	5		Maximum	Clip 5 Upbeat	90	81	22.1%										
	11	38/614	Minimum	Clip 11 Upbeat	98.1	49.5	36.8%	37	11378	34116	33%						
	11		Maximum	Clip 11 Upbeat	102.8	67.7	26.9%										
	14	45/610	Minimum	Clip 14 Upbeat	90	45	15.6%	36	11457	37783	30%						
	14		Maximum	Clip 14 Upbeat	88.9	52	13.5%										
MRU01-FEM1-022608-N7												7	20	35%	1	3	
	8	31/608	Minimum	Clip 8 Upbeat	90	44	29.5%	37	10692	35770	30%						
	8		Maximum	Clip 8 Upbeat	90	57	22.8%										
	16	23/612	Minimum	Clip 16 Upbeat	90	36	44.4%	37	8063	30769	26%						
	16		Maximum	Clip 16 Upbeat	90	52	30.8%										
	21	12/615	Minimum	Clip 21 Upbeat	90	59	30.5%	40	12048	31898	38%						
	21		Maximum	Clip 21 Upbeat	90	77	23.4%										
<b>MRU01 Mean</b>							<b>28.7%</b>	<b>37</b>	<b>11,269</b>	<b>31,325</b>	<b>38.4%</b>	<b>22</b>	<b>60</b>	<b>36.7%</b>	<b>1</b>	<b>9</b>	<b>11%</b>

**A-7 ILLUMINATION ASSESSMENT - ARTIFICIAL SPAWN (SUBSET OF VIDEO FILES)**

Artificial Spawn										
Site	Clip #	Illumination (mean gray value*)								
		Background			Head			Eye		Yolk Sac
		Top Left	Bottom Right	Bottom Left	Dorsal	Otic Vesicle	Dorsal, 2nd	Center	Outside	Not in Shadow
MRQ01-FEM1-021808-A1	4	137	153	153	155	139	100	17	24	159
MRQ01-FEM1-021808-A3	8	136	129	135	116	117	105	17	25	141
MRQ10/P01-FEM1-021908-A1	1	151	174	174	131	148	85	20	22	172
MRQ10/P01-FEM1-021908-A2	3	110	140	137	149	71	107	17	29	139
MRQ10/P01-FEM1-021908-A3	29	118	141	145	154	73	82	17	26	156
MRQ10/P01-FEM1-021908-A5	1b	134	146	146	168	129	119	19	25	134
MRR20-FEM1-022008-A1	12	112	145	139	139	73	135	22	31	155
MRR20-FEM1-022008-A3	5	121	143	149	140	118	128	23	26	143
MRR20-FEM1-022008-A5	11	150	152	142	106	171	144	26	47	148
MRU01-FEM1-022208-A1	6	141	165	169	173	122	137	24	32	183
MRU01-FEM1-022208-A2	1c	192	191	191	167	223	183	21	26	205
MRU01-FEM1-022208-A3	1b	109	136	133	171	107	110	22	27	142

\*low numbers = dark tones (dark orange color); high numbers = light tones (light orange color)

**ILLUMINATION ASSESSMENT - NATURAL SPAWN (SUBSET OF VIDEO FILES)**

Natural Spawn										
Site	Clip #	Illumination (mean gray value*)								
		Background			Head			Eye		Yolk Sac
		Top Left	Bottom Rt	Bottom Left	Dorsal	Otic Vesicle	Dorsal, 2nd	Center	Outside	Not in Shadow
MRQ01-FEM1-022908-N4	11	194	210	206	118	154	146	27	37	159
MRQ01-FEM1-022908-N6	6	199	201	206	134	151	132	22	30	147
MRQ10/P01-FEM1-022708-N2	91	142	154	155	162	114	114	33	39	183
MRQ10/P01-FEM1-022708-N6	10	170	186	188	198	190	105	20	30	171
MRQ10/P01-FEM1-022708-N8	5	154	161	128	211	160	130	21	27	157
MRR20-FEM1-022808-N4	9	203	209	216	241	159	149	22	27	234
MRR20-FEM1-022808-N7	51	214	217	210	135	91	129	34	47	205
MRU01-FEM1-022608-N2	3	102	128	126	121	60	78	22	25	123
MRU01-FEM1-022608-N4	5	160	161	157	136	142	111	20	22	173
MRU01-FEM1-022608-N7	8	117	145	138	172	66	201	22	27	140

\*low numbers = dark tones (dark orange color); high numbers = light tones (light orange color)

**A-8 ANALYSIS OF STILL IMAGES OF NATURALLY SPAWNED EMBRYOS FOR OCCURRENCE OF BODY AXIS IRREGULARITIES**

**MRQ01**

Site	# Still Images	Quick Assessment	Appearance	Body Axis Defect	C	G	O	S	NM	Linear	Total #	Comments
MRQ01-FEM1-022908-N1	20	Immature animals	Gold tone present (19/20)	9	1	1	3	2	3	1	20	Linear specimens appear to have less gold than others
MRQ01-FEM1-022908-N2	20	Poor quality; immature animals, various planar views	Gold tone present	10			5	3	1	1	20	Linear specimens appear to have less gold than others
MRQ01-FEM1-022908-N3	22	Poor quality; immature animals, various planar views	Gold tone present	13		1	5	3			22	Jpeg 9 &10 same image; 18 & 19 are same image
MRQ01-FEM1-022908-N4	20	Poor quality; immature animals, various planar views	Gold tone present	12			1	3	4		20	Background illumination is very dark and textured
MRQ01-FEM1-022908-N5	20	Poor quality; immature animals, various planar views	Gold tone present	9		1	4	4	2		20	
MRQ01-FEM1-022908-N6	19	Poor quality; immature animals; Clip 22 good image	Gold tone present	9	1	1	2	6			19	Background very white, washed out, textured
MRQ01-FEM1-022908-N7	20	Poor quality; immature animals, various planar views	Gold tone present	11				1	8		20	Background illumination is very dark and textured
MRQ01-FEM1-022908-N8	20	Poor quality; immature animals, various planar views	Gold tone present	11				1	8		20	
			<b>Total Number</b>	<b>84</b>	<b>2</b>	<b>4</b>	<b>20</b>	<b>23</b>	<b>26</b>	<b>2</b>	<b>161</b>	

Codes: C = c-shaped; G = g-shaped; O = o-shaped; S = s-shaped; NM = Not measurable; Linear = Normal horizontal orientation

**ANALYSIS OF STILL IMAGES OF NATURALLY SPAWNED EMBRYOS FOR OCCURRENCE OF BODY AXIS IRREGULARITIES  
 MRQ10**

Site	# Still Images	Quick Assessment	Appearance	Body Axis Defect	C	G	O	S	NM	Linear	Total #	Comments
MRQ10 P01-FEM1-022708-N1	25	Poor quality; immature animals, various planar views	Gold tone present	18		2		1	3	1	25	Repeated image 16 &17; linear specimen have less gold tone
MRQ10 P01-FEM1-022708-N2	21	Poor quality; immature animals, various planar views	Gold tone present	13	1		2		1	4	21	Background illumination is very dark and textured
MRQ10 P01-FEM1-022708-N3	21	Poor quality; immature animals, various planar views	Gold tone present	13			2			6	21	Repeat image 3&4, 18&19; background very dark
MRQ10 P01-FEM1-022708-N4	19	Poor quality; immature animals, various planar views	Gold tone present	19							19	Background illumination is very dark and textured
MRQ10 P01-FEM1-022708-N5	21	Poor quality; immature animals, various planar views	Gold tone present	14	3		1		2	1	21	At image 18, background illumination changes to lighter (very textured)
MRQ10 P01-FEM1-022708-N6	20	Poor quality; immature animals, various planar views; Clips 6 and 20 possibly good video	Gold tone present	16	1				1	2	20	Background illumination changes
MRQ10 P01-FEM1-022708-N7	18	Usable	Gold tones not noticeable	6					2	10	18	Image 17 good of linear development; 1 image not usable
MRQ10 P01-FEM1-022708-N8	21	Usable	Gold tones not noticeable	8				1		12	21	Washed out, white background
			<b>Total Number</b>	<b>107</b>	<b>5</b>	<b>2</b>	<b>5</b>	<b>2</b>	<b>9</b>	<b>36</b>	<b>166</b>	

Codes: C = c-shaped; G = g-shaped; O = o-shaped; S = s-shaped; NM = Not measurable; Linear = Normal horizontal orientation

**ANALYSIS OF STILL IMAGES OF NATURALLY SPAWNED EMBRYOS FOR OCCURRENCE OF BODY AXIS IRREGULARITIES  
 MRR20**

Site	# Still Images	Quick Assessment	Appearance	Body Axis Defect	C	G	O	S	NM	Linear	Total #	Comments
MRR20-FEM1-022808-N1	22	Poor quality; immature animals, various planar views	White background	16	1				2	3	22	Various tones in background
MRR20-FEM1-022808-N2	21	Poor quality; immature animals, various planar views	Gold tones, Varied dark to light	19					1	1	21	
MRR20-FEM1-022808-N3	19	Poor quality; immature animals, various planar views	Gold tones, Varied dark to light	19							19	
MRR20-FEM1-022808-N4	21	Poor quality; immature animals, various planar views; there are a few that may be usable	Washed out white; some gold tones when non-linear	14					1	6	21	Image 6 & 7 repeated; 8&9 repeated; 10&11 repeated; 23 &24 repeated; 25&26 repeated
MRR20-FEM1-022808-N5	20	Videos do not appear to be usable; have indistinct heart imaging	Washed out white; gold tones	15			3	1	1		20	
MRR20-FEM1-022808-N6	18	Videos do not appear to be usable; have indistinct heart imaging	Washed out white; gold tones	16			2				18	18 & 19 repeated
MRR20-FEM1-022808-N7	21	Videos do not appear to be usable; have indistinct heart imaging	Washed out white; gold tones	10		1	6	1	1	1	20	12&13 repeat
MRR20-FEM1-022808-N8	22	Videos do not appear to be usable; have indistinct heart imaging	Washed out white; gold tones	13		1	4	3	1		22	
			<b>Total Number</b>	<b>122</b>	<b>1</b>	<b>2</b>	<b>15</b>	<b>5</b>	<b>7</b>	<b>11</b>	<b>163</b>	

Codes: C = c-shaped; G = g-shaped; O = o-shaped; S = s-shaped; NM = Not measurable; Linear = Normal horizontal orientation

**ANALYSIS OF STILL IMAGES OF NATURALLY SPAWNED EMBRYOS FOR OCCURRENCE OF BODY AXIS IRREGULARITIES  
 MRU01**

Site	# Still Images	Quick Assessment	Appearance	Body Axis Defect	C	G	O	S	NM	Linear	Total #	Comments
MRU01-FEM1-022608-N1	21	Usable; clip 20 may have been over exposed?	Washed out white to gray; no gold tones							21	21	Image 2,3,4 are repeats; 17&18 same; 19&20 same image; 21&22 same; 23 &24 same; 26,27, 28 same image
MRU01-FEM1-022608-N2	20	Usable	Mixed; Transparent white to gray (washed out white at bottom)							20	20	2,3 same; 4,5 same; 6,7 same; 8,9 same; 10,11 same; 17,18 same
MRU01-FEM1-022608-N3	17	Usable	White to gray background; no gold tone							17	17	2,3 same; 4,5 same; 6,7,8 same; 9,10,11,12 same; 15,16, 17 same; 18,19 same; 21,22 same; 23,24 same; 28,29 same
MRU01-FEM1-022608-N4	19	Some are usable but most do not clearly show heart; several images (clip 16, 21 show possible differences in pericardial region)	washed out white; no gold tones	5						14	19	5,6 same; 7,8 same; 9,10 same; 11,12 same; 17,18 same; 20,21 same; 25,26 same
MRU01-FEM1-022608-N5	19	Usable	Mixed; no gold tones		1					18	19	2,3 same; 11,12 same; 21,22 same; 23,24, 25 same
MRU01-FEM1-022608-N6	21	Usable; CLIP 23 undeveloped jaw	Mixed; no gold tones	4	3					14	21	12,13 same; 15,16 same; 17,18 same; 24, 25 same
MRU01-FEM1-022608-N7	21	Usable; Clip 14: elongated pericardial region	Translucent, shades of gray							21	21	6,7 same; 10,11 same; 12,13 same; 14,15 same; 16, 17 same; 19, 20 same; 22,23 same; 25, 26 same; 30,31 same
MRU01-FEM1-022608-N8	20	Usable; Clip 7, 4 compared to Clip 14: variation, elongation of pericardial region (how much variation is natural? Is this a developmental artifact?)	Translucent, shades of gray	6	1					13	20	4,5 same; 11,12 same; 13,14 same; 15, 16 same; 20,21 same; 24, 25 same
			<b>Total Number</b>	<b>15</b>	<b>5</b>	<b>0</b>	<b>0</b>	<b>0</b>	<b>0</b>	<b>138</b>	<b>158</b>	

Codes: C = c-shaped; G = g-shaped; O = o-shaped; S = s-shaped; NM = Not measurable; Linear = Normal horizontal orientation