2020 CRFS SAMPLER MANUAL

CALIFORNIA RECREATIONAL FISHERIES SURVEY



A Cooperative Program of:

California Department of Fish and Wildlife Pacific States Marine Fisheries Commission NOAA Fisheries Sport Fish Restoration Act



California Department of Fish and Wildlife

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http://www.wildlife.ca.gov/marine/

TABLE OF CONTENTS

INTRODUCTION TO CRFS 1-1

CRFS-Definition and Goal 1-1 The Importance of Collecting Fishery Data 1-1 State and Federal Mandates 1-2 History 1-3

Marine Recreational Fisheries Statistics Survey (MRFSS) 1-3
Ocean Salmon Project (OSP) 1-3

Survey Design 1.4

Survey Design 1-4

CPFV Logbook Component 1-4 Telephone Survey Component 1-4 Field Sampling Component 1-5 Mode 1-5 Geographic Divisions 1-6

ROLES, RESPONSIBILITIES, GUIDELINES AND EXPECTATIONS 2-1

Roles 2-1

The Supervisor 2-1
The Lead Person (Lead or Field Lead) 2-1
The Fish and Wildlife Technician (Tech) 2-1
The Field Sampler (Sampler) 2-1
The Port Lead Sampler 2-2

Expectations 2-2

Safety 2-2

Data Quality and Due Dates 2-5

Timesheets and Travel Expense Claims 2-6

Professional Conduct 2-7

Attire 2-11

Vehicles 2-12

Equipment and Gear 2-13

CRFS Sampler Supply List 2-14

Region Specific Supplies 2-14

Printed Materials 2-15

Electronic Materials 2-16

CRFS ASSIGNMENTS, SITES AND SCHEDULES 3-1

CRFS Sites 3-1

Site Code 3-1

Site Name 3-1

Site List 3-1

Assignment ID 3-2

The Draw Program and Assignment Selection 3-2

Assigned Mode 3-2

Duration 3-2

Monthly Schedules 3-3

Assignment Spreadsheet Example 3-3

Managing Your Assignments 3-3

Assignment Goal 3-3

Rescheduling and Cancelation 3-4

Closed Sites 3-4

Refused Entry to a Site 3-4

Poor Weather 3-4

Fishing Tournaments 3-5

Opportunistic Sampling 3-5

Second Assignment 3-5

No Anglers at Site 3-6 Incorrect Assignment 3-6

GENERAL ONSITE PROCEDURES 4-1

Canvassing and Screening for Eligibility 4-1

Screening Divers 4-2

Kayak Anglers 4-2

CRFS Codes 4-2

Angler Residence Codes 4-2

Site Codes 4-2

Species Codes 4-2

Gear Type 4-3

Finfish Gear Codes 4-3

Salmon Gear Codes 4-3

Invertebrate Gear Codes 4-4

Areas Fished and Saltwater Cutoff Points 4-4

Saltwater Cutoffs 4-5

Catch Locations and Map Use 4-5

Latitude and Longitude and the One-Minute Grid 4-6

CRFS Block-Box Maps 4-6

Definition of Location 4-7

Accuracy of Location Grid 4-7

The Grid Size Item 4-8

Fishing Site Names 4-8

Fish Identification 4-8

Observed Catch (Sampler-Examined) 4-9

Unavailable Landed Catch (Angler-Reported) 4-9

Filleted Catch (Sampler-Examined or Angler Reported) 4-10

Unidentified Examined Catch 4-10

Released Catch (Angler-Reported) 4-10

Specimen Documentation: Rare or Large Fish 4-11

Sub-Sampling and CRFS Priority Species 4-11

CRFS Priority Species 4-12

CRFS Protocol for Dealing with Suspect Data 4-13

Biological Data Collection 4-14

Fork Length Measurement and Use of Measuring Board 4-15

Measuring Various Types of Fish 4-16

Weight Measurement and Scale Use 4-19

SPECIES SAMPLING PROCEDURES 5-1

Species Sampling Procedures 5-1

Salmon Sampling 5-1

Legal Authority 5-1

Salmon Equipment 5-2

Tagging the Head 5-2

Removing the Head 5-2

Non-Recovered Species (NRS) Protocol 5-3

Procedures for Tracking and Inventorying Salmon

Heads 5-4

Inventory Tags 5-4

Information Request Cards 5-5

Courtesy Head Tags 5-5

Important Salmon Goals to Remember 5-5

Salmon Head Drop Off Protocol 5-6

Yelloweye Rockfish Sampling 5-7

Collection Priorities 5-8

Equipment 5-8

Procedures for Collection 5-8

PR Form Example 5-10

White Seabass Sampling 5-10

White Seabass Equipment 5-11

Procedure for Sampling and Scanning 5-11

PR Form Example 5-12

Removing the Head 5-13

White Seabass Head Drop Off Protocol 5-13

Sexing Certain Species of Finfish 5-15

Elasmobranchs 5-16

Lingcod 5-16

Cabezon 5-17

California Sheephead 5-18

Surfperch (Amphistichinae) 5-18

California Halibut 5-20

```
Kelp Greenling 5-22
Handling Fish 5-23
        California Scorpionfish 5-23
        Rockfish 5-23
        White Seabass 5-24
        Lingcod 5-24
        Sharks 5-24
        Spiny Dogfish 5-24
        Ratfish 5-24
        Electric Rays 5-24
        Skates 5-25
        Stingrays and Bat Rays 5-25
Invertebrate Sampling 5-26
        Squid Sampling 5-27
        Abalone Sampling 5-27
        Lobster Sampling 5-28
        Crab Sampling 5-28
        Handling Crab 5-28
        Invertebrate Sampling Protocol FAQ 5-29
THE WEEKLY REPORT 6-1
Due Dates 6-1
Assignments Included in the Weekly Report 6-1
        PR1 6-2
        PR2 6-2
        PCO 6-2
        PCS 6-2
        PCD 6-2
        PCS/PCD Combination Trips 6-3
        PEC 6-3
        MM 6-3
        BB 6-3
General Instructions 6-3
        Excel Spreadsheet-Instructions for Completing the
        Weekly Report 6-3
        General Guidelines for the Spreadsheet 6-5
Weekly Report Item by Item Instructions 6-5
Weekly Report Coding Tips 6-11
Example of Weekly Report 6-12
ASSIGNMENT SUMMARY FORM (ASF) 7-1
```

Assignment Summary Form (ASF) Layout 7-1

Data Tracking 7-1

ASF Header 7-1

Hours 7-2

ASF Site Rows 7-2

ASF Footer-Boat Mode Totals 7-3

ASF Item by Item Instructions 7-3

ASF Coding Tips 7-15

Specific Editing Checks 7-15

Assignment Summary Form Examples 7-16

PR1-Salmon Season 7-16

BB 7-17

MM 7-18

PCO-Reassigned 7-19

PCO-Completed 7-20

PCD 7-21

PCD-Opportunistic 7-22

BEACH AND BANK (BB) MODE SAMPLING 8-1

BB Mode Definition 8-1

BB Survey Goal 8-1

BB Survey Methods 8-1

Example BB Clusters from Site List 8-2

BB Estimated Total Finfish Angler Counts 8-3

Canvassing 8-4

Incomplete BB Trips 8-4

Low Effort Protocol 8-4

No Anglers in BB Mode 8-4

Observed PR Accessed from BB 8-5

Anglers Fishing in Two Modes 8-5

Opportunistic PC Sampling 8-5

Screening Divers 8-5

Invertebrate Sampling in BB Mode 8-6

Two BB Assignments in One Day 8-6

Ways to Reduce Bias 8-6

2020 CRFS BB Mode Questionnaire 8-6

Shore Form Procedures for BB Mode 8-10

Numbering the Visits, Interviews and Shore Form

Pages 8-10

Shore Form Layout 8-10

Header Items 8-11

Fffort Items 8-11

Catch Items 8-11

Biological Data Collection 8-12

Shore Form Item by Item Instructions (BB Mode) 8-13

Shore Form Example (BB Mode) 8-22

MAN-MADE STRUCTURE (MM) MODE SAMPLING 9-1

MM Mode Definition 9-1

MM Survey Goals 9-1

MM Survey Methods 9-1

Example MM Clusters from Site List 9-2

MM Angler Counts 9-3

Canvassing 9-3

Anglers Fishing in Two Modes 9-3

Incomplete MM Trips 9-4

Low Effort Protocol 9-4

No Anglers in MM Mode 9-5

Opportunistic PC Sampling 9-5

Screening Divers 9-5

Invertebrate Sampling in MM Mode 9-5

2020 CRFS MM Mode Questionnaire 9-6

Shore Form Procedures for MM Mode 9-9

Numbering the Visits, Interviews and Shore Form

Pages 9-9

Shore Form Layout 9-10

Header Items 9-10

Effort Items 9-10

Catch Items 9-11

Biological Data Collection 9-11

Shore Form Item by Item Instructions (MM Mode) 9-11

Shore Form Example (MM Mode) 9-22

PRIMARY PRIVATE AND RENTAL BOAT (PR1) MODE SAMPLING 10-1

Introduction 10-1

PR1 Goals 10-2

Effort Data 10-2

Catch Data 10-2

Observe Kept Salmon 10-2

Location of Catch Data 10-2

Biological Data Collection 10-3

Sample Selection 10-3

Scheduling 10-3

PR1 Survey Procedures 10-3

Effort Data Collection 10-3

Arrival and Trailer Counts 10-4

Low Effort Protocol 10-4

No Boats in PR1 Mode 10-4

Sub-Sampling 10-5

Onsite Missed Boats 10-5

Offsite Missed Boats 10-5

Specific Off-Site Count Instructions 10-6

Catch Data Collection 10-7

Biological Data Collection 10-7

Sub-Sampling Lengths and Weights 10-8

Catch Location Average Depth Data Collection 10-8

Sub-Sampling Locations and Depths 10-8

Minimum PR Sample 10-8

Screening Divers 10-9

Fishing Tournaments 10-9

The PR Form (PR1 Mode) 10-10

Questionnaire Usage 10-10

Introduction to the PR1 Interview 10-10

Before the Assignment 10-11

Arrival on Site 10-11

Sampler Location Onsite 10-11

Multiple Samplers on One Assignment 10-11

Avoiding Duplication and Sharing Counts 10-12

Onsite Trailer Counts 10-12

Offsite Trailer Counts 10-12

Monitoring Boats 10-13

Multiple PR Trips on the Same Day 10-13

Determination of Boat Type 10-14

Non-Fishing (NF) Boat Types 10-14

CPFV and Commercial Boats 10-14

Opportunistic PC Sampling 10-14

Refused Boats 10-15

Language Barrier Boats 10-15

Anglers, Zip Code and Days Fished 10-16

Determination of Catch 10-16

Examining Catch 10-16

Salmon Head Recovery 10-17

Observed Catch 10-17

Unavailable Catch 10-18

Kept Unobserved Catch 10-18

Released Alive 10-18

Released Alive with Descending Device (DD) 10-18

Released Dead 10-19

Seal Take 10-19

Catch Location and Average Depth 10-19

Coding Location of Catch for Multiple Species on the

Same Trip 10-19

Measuring Catch 10-21

Interview Priorities 10-21

Required Fishing Effort Data 10-21

Required Catch Data 10-21

Sub-Sampled Data (Priority Order) 10-21

2020 CRFS PR Form Questionnaire 10-22

PR Form Layout 10-26

Header Row Items 10-26

Interview Effort Items 10-27

Individual Fish Data 10-27

Footer Totals 10-27

PR Form Item by Item Instructions 10-28

Specific Editing Checks 10-41

Summary of PR Counts 10-43

Form Examples 10-43

PR1 Form Example - Salmon 10-45

PR1 Form Example - Non Salmon 10-46

PR1 Form Example - Non Salmon 10-47

SECONDARY PRIVATE AND RENTAL BOAT (PR2) MODE SAMPLING 11-1

Introduction 11-1

PR2 Goals 11-1

Effort Data 11-2

Catch Data 11-2

Location of Catch Data 11-2

Biological Data Collection 11-2

Sample Selection 11-3

Scheduling 11-3

PR2 Survey Procedures 11-3

Effort Data Collection 11-3

PR2 Assignment Duration 11-3

Low Effort Protocol 11-4

No Boats in PR2 Mode 11-4

Arrival and Trailer Counts 11-4

Onsite Trailer Count 11-4

Onsite Missed Boats 11-5

Limited Activity at the Site 11-5

Offsite Trailer Counts 11-5

Launched Boats 11-6

Catch Data Collection 11-6

Biological Data Collection 11-6

Sub-sampling Lengths and Weights 11-6

Catch Location/Average Depth Data Collection 11-6

Sub-sampling Locations and Depths 11-6

Minimum PR Sample 11-7 Screening Divers 11-7 Fishing Tournaments 11-8 The PR Form (PR2 Mode) 11-8 Questionnaire Usage 11-8 Introduction to the PR2 Boat Interview 11-9 Before the Assignment 11-9 Arrival on Site 11-9 Sampler Location Onsite 11-9 Monitoring Boats 11-10 Multiple PR Trips on the Same Day 11-10 Determination of Boat Type 11-11 Non-Fishing (NF) Boat Types 11-11 CPFV and Commercial Boats 11-11 Opportunistic PC Sampling 11-11 Refused Boats 11-12 Language Barrier Boats 11-12 Anglers, Zip Code and Days Fished 11-12 Determination of Catch 11-13 Examining Catch 11-13 Observed Catch 11-13 Unavailable Catch 11-14 Kept Unobserved Catch 11-14 Released Alive 11-14 Released Alive with Descending Device 11-14 Released Dead 11-15 Seal Take 11-15 Catch Location and Average Depth 11-15 Coding Location of Catch for Multiple Species on the Same Trip 11-16 Measuring Catch 11-17 Interview Priorities 11-17

Required Counts 11-17

Required Boat Records 11-17 Required Catch Data 11-17

Sub-sampled CRFS Data (Priority Order) 11-18

2020 CRFS PR Form Questionnaire 11-18

PR Form Layout 11-22

Header Row Items 11-22 Individual Effort Items 11-23

Recording Launched Boats 11-23

Individual Fish Data: Catch and Biological Data 11-23

Footer Totals 11-23

PR Form Item by Item Instructions 11-24

Specific Editing Checks 11-37

Summary of PR Counts 11-38 PR2 Form Examples 11-40

PARTY/CHARTER BOAT ONBOARD (PCO) MODE SAMPLING 12-1

CPFV Survey Procedures 12-1

PC Assignments 12-1

PC Definitions 12-1

Party Boat 12-1

Chartered Boat 12-1

Six Pack 12-2

When to Ride or Sample Dockside 12-2

Opportunistic PC Sampling 12-2

Introduction to Onboard Sampling 12-2

Additional Data Collected Onboard 12-3

Unbiased Angler Sampling 12-3

Onboard Observer Protections 12-3

Title 14, California Code of Regulations Excerpts 12-3

Sampling Chartered Trips 12-5

CPFV Refusals 12-5

Chartered Trip Refusals 12-6

Alternate PC Trips 12-6

Onboard Fishing Locations 12-6

Onboard Catch by Location 12-7

PC Sample Scheduling 12-7

Scheduling PC Trips 12-8

Arrival at a PC Site 12-8

Onboard the Boat Trip 12-9

On the Way Out 12-9

During Fishing 12-10

On the Way Back 12-10

Boat Limits 12-11

NO CATCH Bags 12-12

Overnight Trips 12-12

Two PC Assignments in One Day 12-12

Special PC Assignment Summary and PC Effort Check

Instructions 12-12

No Anglers in PC Mode 12-13

Sampling Dungeness Crab On Board 12-13

Onboard Sampling Tips 12-13

PC Scheduling Questions and Answers 12-14

The CPFV Onboard Angler Form 12-16

CPFV Onboard Angler Form Layout 12-16

Boat Trip Data 12-16

Angler Data 12-16

CPFV Onboard Angler Form Item by Item Instructions 12-17

Specific Editing Checks 12-22

Example of Onboard Angler Form 12-23

CPFV Onboard Location Form 12-24

CPFV Onboard Location Form Layout 12-24

Boat Assignment Data 12-25

Fishing Location Data 12-25

Coordinate Data 12-25

Physical Data 12-25

Species Count Data 12-26

Recording Numbers Kept and Returned 12-26

Refused Items 12-26

CPFV Onboard Location Form Item by Item Instructions 12-27

CPFV Onboard Location Form Coding Tips 12-30

Trolling between Locations 12-30

Non-Stop Trolling 12-31

Specific Editing Checks 12-31

Example of Onboard Location Form 12-32

Onboard Location Form-Multiple Sheets 12-33

CPFV Onboard Catch and Discard Form 12-34

CPFV Onboard Catch and Discard Form Layout 12-34

Boat Assignment Data 12-34

Effort Data 12-34

Discard Fish Data 12-34

Handling Live Fish 12-35

Discard Data Coding Tips 12-35

Catch and Bio Data 12-36

Rockfish Combo Trips 12-36

Total Items 12-36

PC Onboard Catch and Discard Form Item by Item Instructions 12-37

Specific Editina Checks 12-42

Onboard Catch and Discard Form Example 12-44

Coding Boat Fish on the PC Onboard Catch and Discard Form 12-45

PARTY/CHARTER BOAT NON-SALMON DOCKSIDE (PCD) MODE SAMPLING 13-1

When to Sample Dockside 13-1 Scheduled vs. Opportunistic 13-1 Sampling Unit 13-1 PC Assignments 13-1 Data Collection 13-1 Combo Trips: PCD versus PCS Sampling 13-2
Sampling Chartered Trips 13-2
Sampling Guidelines and Procedures 13-2
CPFV Refusals 13-3
Interviewing Anglers 13-3
Boat Limits 13-4
NO CATCH Bags 13-4
No Anglers in PC Mode 13-4
Sampling Dungeness Crab 13-4
CRFS PC (CPFV) Dockside (PCD) Form Item by Item
Instructions 13-5

Specific Editing Checks 13-10
Example of PC Non-Salmon Dockside Form 13-12

PARTY/CHARTER BOAT SALMON DOCKSIDE (PCS) MODE SAMPLING 14-1

Sampling Unit 14-1
Data Collection 14-1
Sampling Guidelines and Procedures 14-1
CRFS-OSP Salmon CPFV Dockside Form Item by Item
Instructions 14-4
Specific Editing Checks 14-7
CRFS-OSP Salmon CPFV Dockside Form Example 14-8

PC EFFORT CHECKS 15-1

Need and Purpose 15-1
Methods 15-1
Types of Assignments 15-2
Additional PC Assignments 15-2
CRFS-OSP PC (CPFV) Effort Check Form Item by Item
Instructions 15-3
Specific Editing Checks 15-5
PEC Form Example – Salmon 15-7

SITE EFFORT CHECKS 16-1

PEC Form Example - Non Salmon 15-8

Need and Purpose 16-1 Scheduling 16-1 Methods 16-1 SEC Route 16-1 SEC Adjacent 16-2 Sampling SEC 16-2 Specific Editing Checks 16-3 SEC ASF Form Example – Routes 16-4 SEC ASF Form Example – Adjacent 16-6

SPECIES CODES 17-1

Sorted by Species Code 17-1 Sorted by Common Name 17-19 Sorted by AFS Common Name 17-37

OTHER CODES 18-1

PR Non-Fishing Codes 18-1
California Island Codes/Saltwater Cutoffs 18-1
CRFS Priority Species 18-1
PR1 Port Codes 18-2
PC Port Codes 18-3
California Saltwater Angling Records as of January 1, 2017
18-5
California Saltwater Diving Records as of January 1, 2017 18-9
Alpha Foreign Country Codes 18-12
Angler Slang Names 18-17
GLOSSARY 19-1

Appendix A. SAMPLER GUIDELINES

Appendix B. MARINE MAMMAL PROTECTION ACT 1972

INTRODUCTION TO CRES



The California Recreational Fisheries Survey (CRFS) Sampler Manual provides an explanation of the principles and goal of CRFS, detailed instructions regarding sampling procedures and protocols, and the proper coding of all forms. This manual describes the history of the survey, general information, methods, and the roles and responsibilities of Supervisors, Leads, Fish and Wildlife Technicians, and Samplers; it then follows with information specific to each fishing mode and the

corresponding form(s).

California's recreational fisheries are dynamic and diverse. Different sampling techniques may be employed based on area of the state, species or species group sought, time of year, and mode of fishing. Thorough reading of this manual coupled with on-going training will enable the Sampler to handle most situations, including unforeseen problems. Any situations, questions, or problems encountered that are not covered in this manual should be directed to the CRFS Field Lead or Supervisor.

To access the CRFS project page, please visit: https://www.wildlife.ca.gov/Conservation/Marine/CRFS

CRFS - Definition and Goal

The California Recreational Fisheries Survey is a multi-part survey implemented in 2004. The goal of CRFS is to produce, in a timely manner, marine recreational fishery data needed for sustainable management of California's marine resources. This goal is a high priority for the California Department of Fish and Wildlife (Department) as it provides the necessary information to make management decisions. The fishery data produced are catch and effort estimates for marine recreational finfish fisheries. Providing timely estimates is essential for the active management of marine species especially for species of concern. Funding for CRFS is provided through both state and federal sources.

CRFS is part of a larger network, the Pacific Coast Recreational Fisheries Information Network (RecFIN). RecFIN integrates state and federally funded marine recreational fisheries sampling programs from Washington, Oregon, and California by integrating estimates and data into a single database. The RecFIN database may be accessed by fishery managers, academia, constituents, and the public.

The Importance of Collecting Fishery Data

Understanding recreational fisheries and estimating total harvest is important, and legally mandated. Economically important marine fish species can be harvested by recreational anglers in bays and estuaries, nearshore areas, as well as the open ocean. Though these species are harvested by both recreational and commercial sectors, recreational catch

can be a significant portion of the total take of many marine fish species in California.

Catch and effort estimates are fundamental to assess the influence of fishing on a fish stock. The development of informed management plans, measures, and policies requires information about the species taken. fishing effort, and the inherent seasonal, temporal, and regional differences in those fisheries. Accurate and timely catch information on a wide range of marine fish species, coupled with their associated biological studies, allow management agencies to effectively protect, enhance, and maintain these living resources. These data are not only used by the Department, but also by state conservation agencies, recreational fishing industries, federal agencies, regional fishery management councils and commissions, academia and others interested in the productivity, management, conservation, and use of California's marine fisheries. Fishery stock assessments, the allocation of fishery resources between states, sectors, and regions, and the resulting management measures and regulations (both planned and emergency actions) depend on the results of these surveys.

State and Federal Mandates

The National Oceanic and Atmospheric Administration (NOAA) Fisheries and the Department have legal requirements for conducting surveys of marine recreational anglers to gather information on 1) catch, participation, and effort in marine recreational fishing; and 2) selected demographic characteristics.

Specifically, NOAA Fisheries is charged with administering a program of research and services relating to the ocean and inland waters of the United States (Title 16, Chapter 9, U.S. Code). Collecting statistics on marine recreational fisheries is authorized by:

- The <u>Fish and Wildlife Act of 1956</u>, Section 5(a)(4), which provides for the collection and dissemination of statistics on commercial and sport fishing;
- 2. <u>Migratory Game Fish Study Act of 1959</u> (Title 16, Chapter 9A, U.S. Code), which provides for continuing study of migratory marine fishes, including the effects of fishing on the species;
- 3. <u>Magnuson-Stevens Fishery Conservation and Management Reauthorization Act</u>, 2007 (Public Law 109-479), requires the collection of statistics for fishery conservation and management.

The Department must collect sport fishery catch information to meet the conservation and management policies for California's living marine resources. The authority to collect this information is specified in the California Fish and Game Code and the California Code of Regulations, Title 14.

History

Surveys of recreational fisheries tend to be more complex and expensive compared to data collection from the commercial sector. The recreational sector is quite diverse, involves more constituents, and is affected more by weather, the economy, and regional differences than the commercial sector. Recreational anglers can fish from boats, piers, jetties, docks, and beaches and can be widely and irregularly dispersed along the coast. Fishing habits and practices vary among fishing sites. Recreational anglers can be elusive; estimating recreational catch and effort can be difficult. Past surveys to assess the impact by recreational fisheries in California include:

Marine Recreational Fisheries Statistics Survey (MRFSS)



The Marine Recreational Fisheries Statistics Survey (MRFSS) was the main survey used by the Department to estimate catch and effort from marine fisheries in California from 1979 to 2003. MRFSS used complementary (dependent on each other) surveys for

catch and effort estimation. A random-digit-dialing household telephone survey was used to: 1) obtain participation and effort data; and 2) provide information on the proportion of fishing households in each county of the survey area. In addition to the telephone survey, MRFSS involved a field survey to intercept anglers to obtain information on catch (species, numbers of fish, lengths and weights) and area fished. The field intercept survey also collected information regarding: number of anglers, state or county of residence, length of trip, catch composition angler telephone numbers, and other items of interest to fishery managers.

In January 2004, CRFS was implemented and replaced MRFSS. The CRFS design was built off MRFSS but was adjusted to incorporate necessary changes to provide more reliable recreational fisheries data on a finer regional scale and quicker time frame.

Ocean Salmon Project (OSP)

The Department's Ocean Salmon Project (OSP) is in charge of recreational and commercial catch and effort estimates, utilizing coded wire tag (CWT) recoveries for California's ocean salmon fisheries. CWT estimates are a key component of salmon management because they identify the contribution of specific runs of salmon to the ocean fishery. OSP conducted a private bo at survey from 1962 to 2003.

Since 2004, CRFS collects recreational data from primary private boat surveys for OSP (the "PR1" mode). The CRFS "PR1" survey is designed to maintain the continuity of the historical OSP private boat estimates. CRFS also works with OSP to collect Commercial Passenger Fishing Vessel (CPFV) ocean salmon data, and to track the activities of the CPFVs during the season. During salmon season, the primary goals for CRFS include identifying adipose fin-clipped salmon for length measurements and head removal to recover the CWT. Samplers north of Point Conception will receive specialized training from OSP prior to the recreational salmon

opener. OSP processes the salmon sample data and salmon heads for CWT recovery. In addition, OSP produces biweekly catch and effort estimates and CWT contribution rates for salmon fishery management with the focus primarily on the major salmon ports. CRFS and OSP collaborate to implement effective CWT recovery and accurate salmon counts.

Survey Design

Catch estimates can most easily be understood by this simple model:

Estimated Total Angler Trips x Estimated Mean Catch Per Trip = Estimated Total Catch

Where the "Mean Catch Per Trip" is also known as catch per unit effort (CPUE). Since catch and effort are separate entities, the survey can be described as having separate collections for 1) effort, and 2) catch. The effort component is the estimation of "Total Angler Trips" and the catch component is the "Mean Catch per Trip" derived from the catch and effort on sampled trips. The estimated "Total Catch" is the product of the effort component and the catch component. CRFS estimates are produced on a monthly basis.

CRFS is similar to its predecessor, MRFSS, in that there are two main parts to the survey: a field survey component (on-site) and a telephone survey (off-site). These survey components also rely on data collected from mandatory CPFV logs and sport fishing license sales to estimate total catch and fishing effort of marine recreational anglers in California.

CPFV Log Component

CPFV operators are required to submit logs to the Department for each fishing trip. CPFV log data collection began in 1936. For each log entry, the vessel operator provides information on effort (number of anglers and number of hours fished) and take (species and number of fish caught). Logs are submitted monthly to CDFW and are mandatory. CRFS uses the CPFV effort data collected by Samplers and log records to estimate party/charter boat (PC) effort.

Telephone Survey Component

The Angler License Directory Telephone Survey (ALDTS) (conducted by a contractor) operates on a monthly basis. The data collected are used to estimate the total number of marine recreational fishing trips taken by license holders when field observations of effort are not feasible, such as night-time fishing and private-access sites. This off-site method uses the Department's Automated License Data System (ALDS) to select samples from a list of active fishing license holders. Data collected from licensed anglers is used to identify the number of anglers that completed saltwater sport fishing trips, and how many trips they completed, by fishing mode, over a specified period of time. ALDTS is used to collect effort data only for beach and bank fishing, night-time fishing, and private access fishing that might otherwise go unaccounted for.

Field Sampling Component

CRFS field sampling is conducted at over 500 publicly-accessible sites during daylight hours to gather catch and effort data. CRFS Samplers intercept recreational anglers at the completion of their fishing trips to collect on-site data by conducting the survey. On-site data is more reliable because it is not as susceptible to angler memory recall bias. On-site methods are used to collect all of the catch data and some of the effort data.

Mode

CRFS conducts four major angler surveys based on fishing mode, and each survey is different. A fishing mode is defined as the method of access to fisheries. The modes in CRFS are:

- MM Man-made structure fishing
- BB Beach and bank fishing
- PC Party and charter boat fishing
- PR Private and rental boat fishing

The following table shows the surveys that are used for each mode of fishing, access type (public or private), and time of day (day or night).

Surveys used in the California Recreational Fisheries Survey (CRFS) to collect data on fishing effort (Effort) and catch (fish caught and kept and fish caught and released) rates (catch per unit effort, CPUE).

and hish caught and released) rates (catch					
Mode	Estimate	Public Access (publicly-accessible sites covered by the field surveys)		Private Access (sites not accessible to the general public and not covered by the field surveys)	
		Day ♥	Night €	Day ♥	Night €
PR	Effort	Field Survey	Under- coverage adjustment ¹	Under- coverage adjustment ¹	Under- coverage adjustment ¹
	CPUE	Field Survey	Use estimate from day	Use estimate from day	Use estimate from day
PC	Effort	CPFV logs and Field Checks ²	CPFV logs and Field Checks ²	Not	Not Applicable
	CPUE	Field Survey (onboard & dockside)	Field Survey (onboard & dockside)	Applicable	
мм	Effort	Field Survey	NO	NO ESTIMATE	NO ESTIMATE
IVIIVI	CPUE	Field Survey	ESTIMATE		
ВВ	Effort	Telephone Survey ALDTS ³	Telephone Survey ALDTS ³	Telephone Survey ALDTS ³	Telephone Survey ALDTS ³
	CPUE	Field Survey	Use estimate from day	Use estimate from day	Use estimate from day

- 1. Under-coverage adjustment using estimates from the ALDTS and the field access point surveys.
- Operators of Commercial Passenger Fishing Vessels (CPFVs) are required as a condition of their license to submit logs for each fishing trip. The CPFV logs and a field survey to estimate compliance are used to estimate CPFV effort.
- 3. Angler License Directory Telephone Survey (ALDTS)

Geographic Divisions

Coastal California is divided into six CRFS Districts. District boundaries generally follow some county boundaries and/orimportant biogeographic lines. Also, Districts coincide with some important state and federal fishery management lines.

- 1. South District Los Angeles, Orange, and San Diego counties.
- 2. Channel District Santa Barbara and Ventura counties.
- 3. Central District Santa Cruz, Monterey, and San Luis Obispo counties.
- 4. San Francisco District Marin, San Francisco, San Mateo, and Sonoma counties on the Pacific coast and the eight counties surrounding San Francisco and San Pablo Bays: Alameda, Contra Costa, Solano, Sonoma, Marin, San Francisco, Santa Clara, and San Mateo counties.
- 5. Wine District Mendocino County and the Shelter Cove section of Humboldt County (to 40°10').
- Redwood District Humboldt County (from 40°10' north) and Del Norte County.

California Recreational Fisheries Survey (CRFS) Districts



ROLES, RESPONSIBILITIES, GUIDELINES AND EXPECTATIONS

The Supervisor oversees the field program and ensures the project goals and sampling needs are met, and works with other Supervisors to ensure proper and consistent sampling protocols are followed. Additional responsibilities include: personnel issues, timesheets and travel expense claims. The Supervisor has jurisdiction over hiring new field staff, and official employee action as needed.

The Lead Person (Lead or Field Lead) manages both field and officework in their respective District(s). The Lead's job is to make sure CRFS data is properly collected by Field Samplers, processed, and entered into the CRFS database. The Lead oversees the training and quality control of Field Samplers, including Fish and Wildlife Technicians and Scientific Aids, and monitors the data flow through the District office(s) including when data is received, edited, scanned, entered, uploaded, filed, and archived, Leads edit the data sheets and provide feedback to staff, maintain site lists and sample selections, and create the monthly Assignment Spreadsheet and sampling schedule in their respective District(s). Leads schedule staff meetings as needed, help review timesheets, recruit and interview new Samplers, and maintain/inventory all field equipment and forms. Leads review monthly CRFS estimates as they become available. They also work with the Department's Recreational Fishing Data Project and Ocean Salmon Project staff to ensure data and estimates are being properly reviewed, checked, and updated. Leads, along with Fish and Wildlife Technicians, will conduct quality control visits each month. During these "field checks" Sampler work performance will be reviewed and feedback will be provided at that time or later.

The District Lead Person is the Sampler's primary resource for training, procedural questions, and problem solving. The Lead is also the person whom anglers may contact concerning CRFS procedures and sampling issues. Media contacts should be directed to the Lead.

The Fish and Wildlife Technician (Tech) has many of the same responsibilities as the Lead and the Field Sampler. Techs may be sent into the field to sample or conduct field checks on other Samplers. They help train new Samplers, process data, maintain sampling gear, and complete general office tasks. Techs may help oversee data tracking and entry, enter data themselves, scan data sheets, help set up meetings and trainings, and provide direction for field staff when the Lead is unavailable.

The Field Sampler (Sampler) plays a vital role in the CRFS project since the key to accurate data collection is high-quality interviewing skills and attention to detail. The Field Sampler role is generally filled by Department Scientific Aids, but occasionally the Techs and even the Leads may act in this capacity. A good Sampler is one who can approach strangers with little reluctance, diplomatically handle sensitive situations, follow procedure and complete forms accurately and efficiently. Samplers are expected to act

professionally in both the field and office settings, accurately identify fish species, and communicate effectively with their Leads and coworkers. Sampler expectations and guidelines follow.

The Port Lead Sampler is an experienced Sampler assigned to track PC effort in a given port, and is usually responsible for ensuring sampling goals for CPFV salmon trips are met.

Expectations

Field Samplers are expected to:

- Be knowledgeable of CRFS goals and how the data is used
- Be knowledgeable of California's ocean sportfishing regulations
- Read a monthly schedule and complete assignments as scheduled
- Identify common marine fish species
- Use a dichotomous key to identify uncommon marine fish species
- Keep gear and equipment in proper working order
- Have all necessary forms and gear available
- Operate in a professional manner
- Wear appropriate field and office attire that is neat and clean
- Accurately complete and submit forms in a timely fashion
- Work at sea on commercial passenger fishing vessels
- Follow the procedures in this manual
- Lift up to 50 pounds
- Have a reliable personal transportation and a driver license
- Have good communication skills
- Be adaptable to changes in procedures, protocols, forms, and schedules

Safety

Sampler safety is more important than any CRFS task. Concerning personal safety there are two types of actions: 1) removing yourself from an immediate threat, 2) and taking care to avoid situations where your safety is at risk. If you find yourself in a situation of immediate danger, do whatever you can to make yourself safe. When you are safe, call the authorities or 9-1-1, and then call your Lead. Your Lead will provide you with a list of local emergency numbers.

If you are injured on the job, take care to memorize the chain of events that lead to the accident. Report your injury immediately and relay the detailed information so that proper action can be taken; a Worker's Compensation claim may be filed. Work with your Lead for proper documentation of work-related injuries and appropriate Personnel involvement, and doctor's visits.

The following sections describe ways to avoid accident or injury on the job with CRFS.

Driving in heavy traffic or in poor weather conditions is a part of this job. Take care when driving in parking lots; look for trailered boats, people, and animals. Winter conditions create additional potential dangers: rock-slides, icy patches on the road or poor visibility. Allowing extra space between your vehicle and the vehicle front of you can help, along with proper maintenance of the vehicle (whether personal or state-owned). If you are driving on state time, make sure you possess the proper vehicle accident report forms.

The job site can be dangerous. When you first arrive to the site, look around and make sure it is safe to work. At many sampling sites, you may wait for anglers in your vehicle. Leave valuables at home and lock your vehicle when you leave it. It is a good idea to carry your cell phone with you or know where local payphones are located. Make acquaintances with local Harbor Patrol, Police Departments, State Park Rangers, and Wildlife Officers. Local law enforcement officers tend to have certain areas they patrol, so you will tend to see the same people at the same sites.

Inclement weather or rough ocean conditions may render a site inaccessible and unsafe, for instance when waves are breaking over a jetty. Assess the ocean before walking too close. Keep an eye on the surf when working on a beach. Be prepared for bad weather, even if the weather at the dock is nice. Plan ahead and bring foul weather gear and/or dress in layers.

Use caution when walking on rough terrain such as jetty rocks, uneven banks, and steep beach cliffs. Slippery docks, uneven terrain, and unstable piers may all be sources of accidents. Slow down, pay attention, and move with purpose to avoid injury. Wear shoes with good traction.

Working around vehicles and boats requires a roving eye for vehicles, especially those towing a trailer. Use care when climbing onto a boat, since the boat or trailer may not be stable, or the foothold and/or handholds may be slippery. Samplers need to be especially safety-diligent when sampling party boats. Be aware of inexperienced anglers who do not pay attention when overhead casting. On rocking boats there is potential danger from swinging hooks and weights on fishing rods. Maintain a safe distance from fishing action whenever possible and be alert to situations where these problems may occur. Keep an eye on the swells so that you may prepare for unexpectedly large waves. Staying within grasp of something to hold on to or sitting down while observing are strategies that make PC sampling safer. Be cautious of leaning against deck railings, especially at the deck gate since the railings may be weak or the gate may not be secure.

Remote sites require you to be aware of your surroundings; approach public restrooms, foliage, and other "hiding spots" with caution. Pay attention to people in the vicinity and try to position the front of your body towards them.

Difficult people are occasionally encountered. Most anglers will cooperate with the CRFS interview, but some anglers will not want to be intercepted for whatever reason. Don't take it personally; it is their right to refuse. Be polite and try to get them to change their mind; you will be trained in gaining anglers cooperation, and your Lead will provide you with compelling arguments you can use to persuade anglers. Some anglers may be hostile toward you. Be aware of this potential when interviewing. If this should occur, stop the interview process and walk away. Even if you stay and engage the angry angler, it will rarely change their views towards you and/or the Department. If the situation escalates, or you witness criminal activity, call 9-1-1 immediately and be prepared to describe the person, place, or vehicle involved. Vehicle license plate or boat CF numbers are very helpful to authorities. If you are personally threatened either verbally or with physical harm, leave immediately. When you are safe, call the authorities and then call your Lead. If at any time you are unsure of how to handle a situation, contact your Lead. Though these situations are rare. knowing how to properly handle and report them is crucial.

Handling fish requires safety measures as well. Be aware that handling fish and invertebrates poses a hazard. For specific fish species to handle with care, please refer to the "Species Sampling" section of this manual.

Measuring fish on a deck or dock can make the area slippery. Kneepads, gloves, and shoes with grip will help while measuring fish. Please take care not to drop fish; use two hands to carry them. Remember that these fish are bound for someone's dinner table; treat them with respect. Don't measure them on a dirty measuring board. Do not lift the fish over open water.

Lifting large fish or bags of fish requires proper lifting procedures to avoid back injury. Lift fish in an area with enough space to work comfortably. Plan your lift, gauge how much strength to exert, and know where you are going with the fish before you lift. Tighten your stomach muscles while lifting and use your leg muscles to lift, not your back. Do not twist while you are lifting; try to maintain the natural curve of your spine. Store the load between knee and shoulder level, so lifting will be easier. Keep the load balanced evenly and close to your body. Maintain balance by keeping your feet shoulder width apart. Do not jerk the load up; move smoothly.

Wild bird and pinniped interactions are occasionally observed while working for CRFS. These animals may become beached, stranded, sick or otherwise injured near a site where you are working. Marine birds and pinnipeds (California sea lions, harbor seals, elephant seals, etc.) are wild animals and should be left alone by CRFS Samplers and only attended to by authorized personnel. Whether you notice a stranded or sick animal yourself or a concerned citizen is reporting it to you, under no circumstances are Samplers to attempt to rescue or assist a stranded marine bird or pinniped; this falls outside the scope of the Sampler's job duties. Please contact the proper local wildlife care center; your Lead should provide you with the appropriate contact information for your area.

As a side note, your Lead will give you contact(s) for reporting <u>dead</u> birds and mammals found onsite; certain research laboratories or veterinarian centers will perform necropsies on some or all of these species. On board CPFVs, let the crew handle situations where fishing line has become tangled around a bird or pinniped. Never try to assist a marine bird or pinniped while at sea. While pinnipeds are protected under the federal Marine Mammal Protection Act (MMPA), NOAA Fisheries has authorized several methods to deter pinnipeds from interfering with fishing operation s (see Appendix B).

Data Quality and Due Dates

You are responsible for the quality of your own data, from collection through submittal to your Lead. All CRFS forms should be completed onsite. Do not leave an assignment with the intention of filling them out later; complete the forms while your memory is fresh. All forms will be reviewed for quality by the Sampler before passing them on to the Lead. Always check the manual first for resolving form-coding issues; you are responsible for following the correct procedures form-by-form and item-by-item. Take the time to look up codes, protocols, etc. in this manual before contacting your Lead with questions. If you can't get a hold of your Lead or Fish and Wildlife Technician regarding a question, take detailed notes on your data she ets. Strive for clean, legible data, with as many good (complete) interviews as possible.

Editing should be done in the field during down time when possible. Write above, don't write over or erase; your changes should still be legible. If possible, editing should be spaced throughout the day, with minimal editing later on. Editing in this manner is cost effective and reduces the chance of errors, since you will be editing while the events of the day are still fresh in your mind. If you are unable to edit your paperwork the same day, you should take the forms with you the next sampling day, and edit during slack time. If it is necessary for you to finish your editing at home, your Lead expects you to make reasonable claims on your Assignment Summary Form and Weekly Report regarding your editing time. Under no circumstances should you "save up" all the editing until the last minute. Time spent editing is just as important as time spent collecting data. Errors or omissions found after forms are submitted require extra time to investigate and are often difficult to remedy. Errors create more work for the data editor and data entry staff, and may imply carelessness and lack of effort on the part of the Sampler. Timely, high quality, legible data is our primary goal and this depends on the punctuality of the Sampler.

Timely submittal of data sheets is extremely important. All forms for assignments scheduled the previous Monday through Sunday must be mailed to your Lead on Monday, or Tuesday if Monday is a holiday, OR hand-delivered by Wednesday. Your Lead may specify different deadlines. Understand that by the time your data sheets are ready to be submitted, they are of significant value to the Department. Take care in handling them, and delivering or mailing them to your Lead each week. Place your

Assignment Summary Form on top of the corresponding data sheets, and keep all the forms together, sorted by assignment date. The forms should all be in order by date, assignment number, and form number. Do not hold up forms for questions; instead, contact your Lead explaining the problems. Mail forms in a strong manila envelope. Do not fold forms. Tape the envelope. Mail forms by USPS first class post ensuring there is enough postage for delivery. Leads check the postmark to ensure data is mailed in a timely fashion. This is especially critical during the last week of the month.

In addition to weekly data, Samplers must meet other deadlines for Weekly Reports, OSP forms, timesheets, expense claims, fish quizzes, trainings, meetings, and/or conference calls. You are expected to be on time and participate in all of these job-related items as they are assigned by your Lead or Supervisor.

The Lead will review, edit, and scan the data sheets before passing them on to data entry. Your Lead or Tech can help you with your field, coding, or form questions. Return calls and emails inquiring about your data in a timely fashion, because some data questions require Sampler input before they can be entered and uploaded.

The data entry team will review the data sheets as they are entering them, and they may contact you with questions. Since each step takes time, it is very important that the forms keep moving through the system; therefore editing is an ongoing task (it doesn't end once the forms have been submitted). Occasionally, you may be contacted with questions about your data by another CRFS or OSP Lead, data manager, or data entry staff. Your quick response is essential to producing CRFS estimates on time. Poor quality editing will result in remedial action by your Lead.

Timesheets and Travel Expense Claims

Timesheets and expenses are due MONTHLY to your Lead around the same time each pay period. Your Lead will inform you of strict deadlines for these items. Timesheets have a specific template, in MS Excel, available for download from the intranet

(http://dfgintranet/portal/HumanResources/Personnel/InstructionsforCompletionofFG681/tabid/232/Default.aspx). Your Lead will direct you on where to find and how to use the current Department time keeping system. It is your responsibility to know the proper coding on the time sheet, from your position title and number, to the funding codes and how to claim absences like sick leave, holidays, etc.

Travel expense claims will be submitted through the Global CalATERS system. Work with your Lead and/or Supervisor to properly complete an expense claim. Expense claims are your responsibility; any errors or follow-ups to erroneous claims must be dealt with in a timely manner. The only expenses incurred during working hours that are typically reimbursable are: parking fees, tolls, mileage, meals and per diem (if applicable) and postage

for mailing data sheets to your Lead. All other expenses must be preapproved by the Supervisor.

Meals purchased while on assignment may be reimbursable; typically meals may be covered if the Sampler works over 24 hrs. As explained in your bargaining unit contract, the Department will only reimburse the Sampler for breakfast if the trip begins (departing headquarters) at or before 6 AM and the Sampler travels 100 miles or more that day. An example would be boat meal purchases while onboard a CPFV trip (which are encouraged as it is a friendly gesture and a way of supporting the galley). Boat meals (breakfast) can be reimbursed only for: 1) morning trips and 2) if the Sampler traveled a distance of 50 miles or greater one-way from headquarters to the landing. Lunch and daily incidentals are not reimbursed on trips that are less than 24 hours in duration. Dinner may be claimed if the Sampler travels 50 miles or greater one-way and returns to headquarters after 7 PM. For more information on per diem travel reimbursement, please visit http://www.calhr.ca.gov/. After selecting Bargaining Contracts, you will find your information under Memorandum of Understanding, Unit 11.

Parking should be exempt from charge with the parking placard provided to you. However, there might be times when a parking lot has automated gated access or the Sampler may be forced to feed a parking meter. If payment is required to park, retain the original receipt and it can be reimbursed.

Mileage reimbursement will be provided at the state government rate and is intended to cover the cost of fuel plus general wear and tear on the Sampler's personal vehicle. The use of a state vehicle is preferred, but if one is not available, a personal vehicle is allowed. Personal mileage reimbursement does not require documentation (e.g., gas receipts) but only includes that mileage generated while on assignment.

Postage for mailing original data sheets to the office is reimbursable but you must submit the original postage receipt. It is preferred to mail data from another Department office before using the post office or other mail service, to keep costs down and reduce the need for postage receipts. Only normal postage will be reimbursed; expedited mail services (e.g., express, overnight service) are not reimbursable unless pre-approved by the Supervisor.

Professional Conduct

As an employee working for the State of California, all CRFS staff are held to a high standard of professional conduct and work ethic. Understand that you are the face of the Department of Fish and Wildlife, and the public will turn to you for help and advice on many subjects, some of which have nothing to do with fish or wildlife. CRFS Samplers are to follow etiquette when working in both the field and office setting. Address your Lead, coworkers, and members of the public with courtesy and respect. Be polite and professional in all your endeavors. Your behavior serves to

substantiate the legitimacy of the survey and increase angler cooperation, builds positive workplace relationships, and helps you stand out as a responsible worker.

Etiquette can make or break a CRFS interview. Be aware of your body language and tone of voice. Before the CRFS interview, introduce yourself and the survey; do not assume anglers know who you are. Ask permission to board any boat or examine/handle any fish. Thank anglers after the interview is complete. Similarly for CPFVs or other privately owned/managed sites, always introduce yourself to the landing personnel and crew, and ask permission to conduct your business and board any vessel. Refrain from words that could be construed as inappropriate or vulgar. Be aware of diversity; CRFS Samplers interact with people from all walks of life. In the eyes of the public, CRFS Samplers are seen as the most visible and convenient way in which to contact fisheries regulators. You are a very visible person at any fishing site, especially while wearing CRFS attire and fully equipped. While you are observing the fisheries, you are being observed and judged by the public. Your actions and conversations reflect on the Department and state government in general. Do not do anything that could be viewed as a waste of time or state resources. Remember that you are a public employee. Working with others is part of the job. While sampling, you will use your interpersonal communication skills to gain access to sites, board and sample boats, and persuade anglers to cooperate with the survey. You will work with other CRFS samplers, other agency staff, and law enforcement, as well as with the public, including anglers and curious individuals. In the office setting. you will work with other CRFS staff, as well as Department staff who have no affiliations with CRFS. If you are working next to someone you don't know, introduce yourself and be friendly.

Stick to your job duties so the public understands your role. Do not engage in "deckhand" duties (helping anglers land fish, tying on hooks, etc.) while sampling onboard. Our insurance does not cover activities outside of your job description as a CRFS Sampler. Do not allow the public to think you are a Wildlife Officer, or any other role of authority. Your primary responsibility as a CRFS Sampler is to collect recreational fisheries data. If you do not know the answer to a question from the public, never guess; refer them to your Lead. If you are approached by a reporter in the field, you should provide your Lead's contact information, and inform your Lead; do not engage them in an interview of any kind. Most information sought by a reporter (survey design, catch success rate, cooperation, etc.) needs to be answered by either the Lead or someone else in the Department. Other projects may request your help in collecting field information. If you are approached in the field to do this, refer the person to your Lead.

Conflicts of interest and incompatible activities must be avoided while working for CRFS. When you are hired as a Scientific Aid you will be given a list of activities to avoid once you have signed your oath of allegiance to the State. Some political and employment activities should be avoided, for

example you may not work as a commercial fisherman while also working for CRFS. You may not use state resources, such as postage, office supplies, internet access, or state vehicles, for personal gain. You may not use your employment with the Department to promote your personal business or any other business. Gifts may not be accepted at any time, including free fishing trips on CPFVs, fish, clothing, or other goods and services. A primary reason for not accepting fish is due to the CPFV boat limit regulation. We do not want the Sampler to end up in a situation where enforcement finds the Sampler is either contributing to or helping the overlimit boat come down to the limit. This action may make it appear as if you are an agent of the vessel and that you "look the other way" when violations occur on the boat.

Tobacco use must be discreet. You may NOT smoke in view of the public. Never interview anglers while smoking or chewing tobacco. Do not throw cigarette butts on the ground or in the water. Do not spit chewing tobacco on the ground. If you do smoke, cover your CDFW patches and go away from the dock, anglers, and the public to a place where smoking is allowed; or do it in your personal car. It is against government policy to smoke in a state vehicle.

Working with Wildlife Officers can happen at any fishing site. Sometimes they will be undercover agents and you may interview a Wildlife Officer without your knowledge. If a Wildlife Officer asks you if you have seen any illegal activity, you should tell them generally what you know but ask them to be discreet with the information. If the officer asks to see your data, inform them that you have been instructed to refer such requests to your Lead. According to the Privacy Act Statement, the information we obtain through CRFS is confidential, and we do not want to jeopardize our presence at any site or CPFV operation. If a Wildlife Officer approaches while interviewing, let them proceed with their investigation first; stand back during their investigation. If possible, complete the interview with an investigated angler, and include confiscated catch, if any. Report the encounter on your forms.

Illegal fishing activity will probably occur at some point in your fishery sampling career. The purpose of the field sampling for CRFS is to collect an independent and unbiased sample of the fishing activity. Any behavior which would systematically exclude illegal take from the sample would create a bias in the sample.

Do not give the impression to anyone that you are a Wildlife Officer. Do not try to enforce fishing regulations in the field. However, you may educate the anglers as to fishing regulations. If you observe illegal fishing activities, pass the information along to your Lead or go somewhere private and call CalTIP. Distribute CalTIP business cards to concerned anglers who have witnessed illegal activity.

When you encounter an angler with a violation (e.g., a short fish, or over-limit), you should explain the violation and educate the angler <u>after</u> you have observed and measured the catch. The Sampler should inform the angler of violations if it appears the angler is unaware of the violation. Use a statement such as, "Did you know you have two undersized barracuda? The minimum size is 28 inches. I'm doing biological sampling, but if a Wildlife Officer were to come by, you might get a ticket." Usually, it is easier to obtain biological data on this catch if the angler is informed after you have concluded the interview.

Obvious violations of bag limit, size regulations, and other illegal activity should be reported to your Lead after your assignment. The Lead will take the appropriate action in regards to contacting enforcement. In this way, the Wildlife Officers can pay a visit to the site(s) where you saw violations occurring and issue citations when appropriate. This removes you from that process, as our function is biological sampling. With regard to illegal activity on party and charter boats, care should be taken not to disturb a good working relationship with captains and crew. Report any illegal activity to your Lead.

There may be occasions where an angler has kept a prohibited species, such as a giant sea bass, Yelloweye Rockfish, or Cowcod. Try to collect all retained Yelloweye, but do not collect any other prohibited species. A primary goal is to obtain length and weight data for prohibited species. If possible, take a photograph of the species. Under no circumstances should you engage in any sort of discussion or behavior that the angler may interpret as threatening enforcement action. If you are unsure of how to proceed in any situation, contact your Lead immediately.

Be sure to make a note next to sublegal fish, over limits and fish taken out of season that are recorded in your data. This helps data editors tremendously.

Education and outreach is the best approach when you encounter fishing violations or the general curious layperson. While improving public relations for CRFS and the Department is helpful, it is not your main responsibility. Do not let education and outreach get in the way of collecting high-quality data for the project. When you have time, distribute informational fliers and brochures, and answer questions from the public to the best of your abilities. Let your Lead know what questions you are getting, so they can share answers to those questions.

While in the field, you may have people ask you questions about fishing regulations. You are responsible for knowing the basics and how to look up the answers in the sport fishing regulation booklet. Samplers should be aware of the current fishing regulations in their District; however Samplers are not required to know the complex reasons why the regulations are as such. You should offer the person a copy of related outreach materials and show them where to find the answer, but never interpret regulations for the

angler. Suffice it to say that fishery managers are doing all they can to provide fishing opportunities while conserving fish populations for future generations. There is tremendous pressure on managers to allow angling and to justify every restriction. You may facilitate the outreach process by informing the angler that there is a process, explaining the limited role of the Sampler, providing contact information, explaining a regulation, or offering printed materials. If you do not know the answer to a regulation question and can't find it in your materials, never guess. Refer the angler to a CDFW office so they may get the answer directly. Often Samplers will not have time to get into a conversation and should politely explain that they are very busy with data collection at the moment.

Sometimes members of the public have stories of how they were mistreated by a Wildlife Officer, or other complaints regarding Department policies or regulations. You may listen but it is important to not take sides. Please explain that we are biologists, not policy-makers. You may suggest to the individual that they contact the Department with their concerns and/or write a letter to the California Fish and Game Commission.

Avid anglers may be approached several times per year by representatives conducting CRFS interviews. Angler cooperation is critical to the success of the survey. Samplers will encourage anglers to take the time to participate and thank them when they do. Every fishing trip may have different target species, locations, gear, etc. Therefore, it is necessary to have anglers provide data on each trip even if they have participated in the survey before. Anglers may also be telephoned to be asked about their trips in the past one to two months. Samplers should also encourage angler cooperation with telephone surveys of recreational fisheries.

Attire

Samplers are expected to look professional both in the field and the office setting. Samplers on assignment will wear their CRFS attire that is issued to them, including the polo shirt, hat and jacket. Do not wear attire with other logos (advertising logos, etc.), except on shoes it is okay. The public may be confused about who you are affiliated with if you display logos other than the Department shield while on assignment. Do not wear your CRFS attire when you are not working. No dangling jewelry. Samplers with long hair should tie it back.

Pants can be jeans, khakis, or corduroy. No sweats or tight-fitting pants (e.g. NO leggings, or jeggings, or "yoga pants"). Pants may not have holes, tears or obvious stains. You may wear long shorts (for warm weather), but they may not be too short and they must have a hem or be rolled (no cutoffs).

Shoes must be closed-toe. Do not wear 'flip-flops' or sandals. This is not only a safety issue, but also a matter of professionalism. Shoes should have rubber soles so you will have secure footing regardless of where you should find yourself sampling (e.g. on jetties, climbing into boats). You will

be provided rubber boots for onboard CPFV assignments, if you choose to wear them.

Hats will help prevent the sun from taking its toll and help identify you as a Department Sampler. Protect yourself against sunburn and heat exposure by wearing a hat. Hats help minimize glare from the sun which can tire you out and/or lead to a headache. It helps to keep your hair out of your face and therefore minimize the potential for accidents that can result from your inability to see clearly.

The California State ID Card is to be on your person the entire time you are on assignment. Do not use your ID card for any other purpose. Samplers will be viewed as representatives of the state of California conducting official business.

Sun screen is highly recommended to protect you from sunburn as well as reduce the risk of some skin cancers. You may be able to be reimbursed for sunscreen - contact your Supervisor prior to purchase.

Sunglasses will help protect your eyes from UV radiation. Sunglasses will not be provided to you.

Gloves will protect your hands while handling slippery wet fish. Gloves will be provided to you.

Vehicles

A valid California driver license and reliable personal transportation are required to work on the CRFS Project. Responsible driving may be checked with a Department driver's test before you may operate a state vehicle.

State vehicle use is preferred, but if one is not available, youwill use your own personal transportation. Work with your Lead to get a Voyager Fuel Card PIN assigned to you; this is required for fueling state vehicles. Your Lead will also provide you with information on how to reserve a state vehicle and how to fill out the state vehicle mileage log. Remember that only authorized persons may ride in a state vehicle and that state vehicles may only be used on official business.

Compensation for driving is defined here. Your Headquarters (HQ) address will be determined when you are hired. Once you leave your HQ while driving for CRFS, you are "on the clock" and will get paid your hourly rate to drive (i.e., travel time). Mileage from your home to HQ is part of your normal commute and that time will not be compensated. Mileage may only be reimbursed if you are using your personal vehicle. Please make notes on the data sheets if you used a personal vehicle or a state vehicle. Please make a note on your forms about unusual traffic conditions that would cause a long commute to your CRFS site.

Accidents happen. Make sure the proper state forms are with you when driving a state vehicle in case of a vehicular accident while on the job. Report all accidents, whether in a personal or state vehicle, to your Lead as soon as possible.

Citations are your responsibility. All California traffic laws apply when you are driving on state business. You are responsible for any traffic citations while driving on the job, including if you are in a state vehicle. Be a courteous and safe driver while driving on the job.

Parking can be difficult at some sites, especially during the busy summer months. Many access points to beach areas have pay parking lots. When the parking lot has an attendant, Samplers can almost always obtain free entrance, provided they are in uniform and have their ID card and parking placard with them. Please attempt to park legally. Use your good judgment about parking in a non-recognized parking space. Your Lead has included notes about special parking situations in your site descriptions. Your Lead will provide you with a parking placard to display on your dashboard to identify your vehicle. While you may sometimes need to park in a space reserved for boat trailers (as a last resort), never park in handicapped, fire hydrant, loading, tow-away or red zones. If you need to pay for parking, get a receipt and claim it on your monthly expense claim. When parking your vehicle at a launch ramp, be sure you give the anglers enough room to circle your vehicle with their boat trailers.

Parking citations should be brought to your Lead's attention right away. The placard may prevent ticketing; but it is not guaranteed that you will not get a parking citation while on assignment. If you do get a parking ticket, work with your Lead to attempt to dispute the ticket with city or harbor enforcement. This may involve your Lead writing a letter to the ticketing agency. Regardless of the outcome, the Department will not reimburse you for parking tickets or towing.

Equipment and Gear

At the beginning of employment, a vast array of gear will be issued to the Sampler. An itemized list of all gear will be signed by both the Lead and Sampler at the time of check-out and check-in. Since the value of this gear can exceed \$1000, it is important that all gear is returned upon the end of employment. Careful documentation will also ensure that Samplers aren't charged for gear that they were not issued in the first place.

Do not wait until the last minute to notify your Lead when you are short on forms or equipment. Give them a call or email as soon as you notice you need something. It may take time to get to you in the mail; they may need to make more copies or order replacements first.

Due to the rigorous nature of CRFS data collection, it is expected that normal wearing of gear will occur and may require periodic replacement (of scales, for example). Gear damaged during normal work duties will be

replaced free of charge to the Sampler, once the damaged unit is returned to the Lead. Gear that is damaged or lost outside of normal working duties, or due to Sampler negligence, may result in reduction in hours or termination. Examples of Sampler negligence include: leaving the measuring board on a pier, backing over it with your vehicle or leaving a scale on a CPEV.

All items must be returned clean and ready to be used again at the end of employment.

CRFS Sampler Supply List

- Site information: map or directions to the site, site codes and altern ate sites
- 2. Schedule of assignments and site clusters
- 3. Forms for assigned mode(s)
- 4. Assignment Summary Forms
- 5. Clipboard & Pencils
- 6. Measuring board & insert
- 7. Tape measure
- 8. 25, 12.5, 5, & 1 kg. scales
- 9. GPS receiver and extra batteries
- 10. CRFS Block/Box maps
- 11. Several copies of the Privacy Act Statement
- 12. CRFS ID Badge
- 13. Sampler Manual and interviewing reference materials
- 14. Field guide/keys appropriate to your area for fish ID.
- 15. Other administrative forms and supplies
- 16. Current fishing regulation booklets
- 17. Binoculars
- 18. CRFS/RecFIN brochure
- 19. Bucket and liner

Region-Specific Supplies

- 20. Salmon
 - a. Cutting board
 - b. Bags and collection tags for heads
 - c. Fillet knife
 - d. Rolling cooler
- 21. White seabass
 - a. Hand-held scanner with holster
 - b. Bags and collection tags for heads
 - c. Fillet knife

When sampling, you must have your fish ID books/sheets, measuring board, CRFS maps, and all scales with you at all times. You must also have your manual with you; it is acceptable to have it in your vehicle. Lacking these elements when a field check is performed may result in disciplinary action.

The Sampler should always plan ahead and have a sufficient number of forms on hand. Make arrangements to get more forms well in advance of getting low or running out.

Printed Materials

Listed below are a number of printed materials available to the Sampler to provide to anglers. Often a handout will be an incentive for anglers to participate. Be sure to have copies of the current regulations and handouts. You may be asked to supply businesses with printed information.

- CDFW Ocean Fishing Regulations Printed booklet, half letter size, double sided.
- 2. Bring That Rockfish Down Printed brochure through Sea Grant explains barotrauma and how to release rockfish properly.
- 3. Canary/Vermilion/Yelloweye Flier Printed color flier with illustrations of Canary, Vermilion, and Yelloweye Rockfishes. Also lists key characteristic for species identification.
- California Fishing Passport Printed booklet, half letter size, double sided.
- 5. CDFW Marine Region Card Business card with basic Marine Region contact information and web site address.
- 6. *CDFW CaITIP* Business card with contact information for CaITIP; telephone number for anonymous reporting of violations.
- RecFIN Card Business card with basic contact information and web site address.

Electronic Materials

Some printed materials are available in electronic form on the Department (http://www.wildlife.ca.gov/Fishing/) and RecFIN (www.recfin.org) websites on the internet.

A number of other interesting resources are available on these websites, including fish identification, access to the data, estimates, contact information, links to other agencies, and the ability to provide public feedback to the Councils.

Forms

Form	Survey Mode	Data
Assignment Summary	ALL, except PCS and	Effort & Form
Form	PEC	counts
Shore Form	BB and MM	Catch and Effort
Onboard Angler	PCO	Anglerinfo
Onboard Catch/Discard	PCO	Catch
Onboard Location	PCO	Catch and location
PC Dockside – Salmon	PCS	Catch and Effort
PC Dockside – Non-	PCD	Catch and Effort
Salmon		
PC Effort Check	ALL	Effort
PR Form	PR1 and PR2	Catch and Effort

CRFS ASSIGNMENTS, SITES, AND SCHEDULES

Site Code

The CRFS samples at coastal sites up and down the State of California. All fishing sites for the CRFS project are given a county and site code; and it is this combination of county and site codes together that make a unique way to represent each fishing site. County codes are represented first, with three numerical digits. The site codes come next and are also three numerical digits. So, together each individual site is represented by a six digit number. Most of the CRFS forms will require you to record the county and site codes for where you completed your assignment. After a while, you will become very familiar with these codes.

Site Name

CRFS sites are identified by site name in addition to the site code. The site names are very specific and should be followed exactly as they are presented on the site list. CRFS sites can be of varying sizes and may span different structures or shorelines. For instance, one site may be a launch ramp, a very specific point, whereas another site may include a pier or a stretch of beach. A site name can vary depending on what the assigned mode of sampling is at that site.

Site List

When you are hired, your Lead will give you a District site list which will have all the sites listed by county, corresponding maps, and driving directions to the sites. Leads may use a site description binder and/or a Wiki site to document the sites by District

(http://data.recfin.org/wiki/index.php/California Recreational Fisheries Survey). These descriptions not only give specific instructions on how to get to the site, but also include site boundaries (if any), fishing modes available at the site, Marine Protected Areas near the site, the facilities available at the site, and any phone numbers or web links you may need, such as party boat landing information. Notify your Lead if you discover information for a site is incorrect.

The District site list may change on a monthly basis. Some sites may be active one month and inactive the next. Your Lead and Supervisor will determine which sites stay active and which ones become inactive, based on time of year, budget, fishing seasons, effort, safety, etc. It is important that you pay attention to your Monthly Schedule and only go to sites you are instructed to visit. In addition, fishing sites may change over time. Launch ramps are constructed or torn down, or new CPFV landings open for business. CPFV boats or landings may move ports, change ownership, or go out of business from lack of customers. Regardless of whether these site changes are temporary or permanent, do not assume your Lead is aware of them. It is your responsibility to notify your Lead of these changes as soon as you are aware.

Assignment ID

Each assignment on the Monthly Schedule will have a unique six-digit assignment ID code (ASSN ID) for tracking purposes. The ASSN ID follows the assignment from the day it is populated from the Draw Program, to the sampling event, and through the data entry process. It is extremely important to use the correct ASSN ID on all your forms.

The Draw Program and Assignment Selection

Monthly assignments are determined by random selection through the Department's Data Portal CRFS Draw Program (called "Draw"). Stratified random sampling is used in all modes. Additionally, pressure-weighted sampling based on historical fishing patterns is used for PC, MM and PR2 modes. Each site has an estimate of past effort (fishing pressure) for each mode based on Sampler data collection of angler and boat counts. Use of average historical effort for future sampling can take into account the anticipated changes in fishing effort for each month based on regulations, etc. and kind of day (KOD) which is either weekends & holidays or weekdays (Monday-Friday). The Lead and Supervisor may utilize historical productivity data such as average interviews per site, MPA boundaries, safety at sites, or current budget status to adjust the number of active sites in the Draw. Fewer sites tend to give fewer assignments in the Draw.

Working with the Supervisor, the Lead generates the Draw each month, producing an Assignment Spreadsheet, Monthly Schedule, and Master Spreadsheet. The Assignment Spreadsheet is created in MS Excel and uploaded into the Data Portal where each assignment will be tracked by the Leads online. The Monthly Schedule is created in MS Excel and provided to the Samplers in a timely fashion so that they may plan their work month. The Master Spreadsheet is a printed spreadsheet that is used in the District office(s) as one way of tracking assignments through the data entry process.

Assigned Mode

The assigned mode (AMODE) for each assignment will dictate how the assignment is sampled, including the duration, forms to use, and site(s) to visit. Shore modes (MM and BB) are arranged into clusters of sites where the Sampler visits a group of sites in one assignment, whereas PC and PR mode assignments will require you to stay at one site, generally all day. See the specific Mode Sections in this manual for protocols by assigned mode.

Duration

The assignment duration will vary based on mode and the fishing effort that day. Most assignments require an 8-hour shift. Some assignments may be even longer work shifts, depending on the mode and time of year. CRFS is conducted during daylight hours only, so the summer months tend to have longer assignments. Including travel time, some PR assignments may last 16 hours. Sometimes riding onboard CPFVs will require up to a 15-hour day; overnight trips are even longer. In general, most assignments will require a minimum sampling time of two hours. Please see each Mode

Section for specific instructions regarding arrival and departure times, as well as low-or-no effort protocols. Please be aware that there are mode-specific instructions for each type of assignment, rescheduling, and on site procedures.

Monthly Schedules

CRFS Monthly Schedules are created in Excel on a monthly basis, around the 20th of the month. Requests for days off or changes to your availability must be submitted to your Lead no later than the 15th of the preceding month. It is important to let your Lead know your availability when it is due, so that the schedule can be finalized quickly. You will receive the schedule from your Lead approximately one week before the 1st of the following month. Monthly Schedules are produced in a tabular format, the Assignment Spreadsheet (i.e. DRAW tab), but may be copied into a calendar format, or a schedule by area or individual Sampler. The Monthly Schedule will tell you which day(s) you are to work, the mode, sites, start times, and coworkers. The columns in the assignment spreadsheet will be described to you by your Lead during training, so that you know how to interpret the Monthly Schedule. See below for an example of an Assignment Spreadsheet.

DISTRICT	YEAR	MONTH	KOD	DOW	DAY	CNTY	SITE	PORT	CLUSTER	NAME	ASSNID	AMODE	PC DURATION	ORDER	START CATEGORY
5	2014	AUG	WD	MON	4				WIN3		085202	MM		Α	early
6	2014	AUG	WD	MON	4				RED2		086201	MM		В	early
6	2014	AUG	WD	MON	4	15	301	CRD		Crescent City Docks	086712	PCD	dockside		
6	2014	AUG	WD	MON	4	15	301	CRD		Crescent City Docks	086523	PR1			
6	2014	AUG	WD	MON	4	23	103	FLD		Fields Landing	086509	PR1			
5	2014	AUG	WD	TUE	5	23	106	SHC		Shelter Cove	085702	PCD	dockside		
6	2014	AUG	WD	TUE	5	23	307	TRD		Trinidad Docks	086720	PCD	dockside		
5	2014	AUG	WD	TUE	5	23	106	SHC		Shelter Cove	085509	PR1			
6	2014	AUG	WD	TUE	5	23	307	TRD		Trinidad Docks	086531	PR1			
6	2014	AUG	WD	TUE	5	23	107			Eureka T-street launch	086301	PR2			late
6	2014	AUG	WD	WED	6	23	307	TRD		Trinidad Charters	086605	PCO	half day		
6	2014	AUG	WD	WED	6	15	400	CRL		Crescent City Harbor	086536	PR1			
6	2014	AUG	WD	WED	6	23	120	EUR		Eureka Marina	086515	PR1			
5	2014	AUG	WD	WED	6	45	100	FTB		Noyo River	085502	PR1			
5	2014	AUG	WD	THR	7				WIN3		085201	MM		В	early
6	2014	AUG	WD	THR	7	23	102	TRH		Trinidad Hoist	086501	PR1			
5	2014	AUG	WD	THR	7	45	103			Point Arena	085302	PR2			early

Managing Your Assignments

It is your responsibility to manage assignments scheduled to you. Assignments are required to be worked on the date they are assigned unless other arrangements have been made with your Lead. Office work and meetings, conference calls and trainings are just as important as field work. You are expected to be punctual to work, prepared and ready to go. Do not work on other Department projects without prior approval from the Lead and Supervisor.

Assignment Goal

The Sampler's daily goal is to obtain as many high-quality interviews as possible in a reasonable amount of time in the assigned mode.

Rescheduling and Cancelation

Do not move, reassign, or cancel an assignment without prior approval from your Lead. Do not swap assignments with another Sampler without prior approval from your Lead. There is more flexibility in the PC and BB assignments than in the PR1, PR2 and MM assignments, with PR1 being the most restrictive, especially during salmon season. If you cannot make an assignment, you are ill, or have an emergency, contact your Lead or Supervisor immediately. It is crucial for statistical validity that Samplers complete all assignments as scheduled.

If you miss an assignment and it can't be covered by another Sampler on that same day, it needs to be rescheduled to another day by the Lead. We cannot carry over missed assignments from one month to another. It is also important to understand that any assignment scheduled to you is not "your" assignment. The assignment needs to be completed on the date provided by the Draw; who completes it is not important. If you cannot work an assignment, the Lead will try to cover with another Sampler first and will only modify the assignment date as a last resort.

Closed Sites

If a site is closed upon your arrival, contact your Lead immediately. Your Lead needs to know the exact dates of closure so as to avoid extrapolating data to that site in estimates of fishing effort and catch. There is a difference between site and ocean conditions being unsuitable for fishing effort to develop and the site being closed by the city, county, harbor or police for reasons such as construction or maintenance. Examples of site closures include but are not limited to: crime scene investigations, boat hoist malfunction, oil spill or some other hazard, tsunami warning, fish kills, pier renovations, and closures due to an entertainment event like a concert or car show. Please notify your Lead if a site is closed while you are on assignment.

Refused Entry to a Site

In some cases you may be refused entry to a fishing site. If after explaining the project, admittance cannot be obtained, you should contact your Lead immediately. Working with your Lead, you should proceed with an alternative site for the assigned mode, move to a second assignment, or reassign the assignment. Your Lead must be notified about your refused entry/access, including refusals by CPFVs.

Poor Weather

Rain, wind, etc. might deter some anglers, but not all. In general, the rule to follow is that if people can fish, sampling should take place. If, on the day scheduled for sampling, the weather is obviously so bad that no one could be expected to fish, you should follow the instructions provided for such situations by your Lead and this manual. In some cases, lack of fishing effort at a particular site entails moving to the next site in a cluster of sites. In other cases, the assignment will be completed early if there is no effort, or you may have a second assignment that may be at a location where

effort is on-going. Your Lead may also provide you with another work activity. See each mode section in this manual for low-effort protocols.

Fishing Tournaments

For the purposes of CRFS sampling, a tournament is defined as a site and date specific fishing contest. Contestants usually have to return to tournament headquarters by a certain time for the "weigh in" which limits how far they can travel, and only specific species of fish may be taken on the date of the tournament event. Generally, tournaments are not sampled. Once the Sampler determines that a tournament is taking place, the Sampler should contact the Lead immediately to discuss rescheduling the assignment. Notify your Lead in advance when you learn the date and location of a tournament so that your Lead can make appropriate arrangements, if necessary. If a tournament is sampled, all sample numbers must be flagged with a "T". Informal 'pools', such as those arranged on CPFVs (jackpot contests), are not considered tournaments anglers participating in these types of contests should be sampled. Some fishing contests are regional (encompassing a large area) and span a long period of time (a week, a month, or an entire fishing season). Participants usually pre-register at a location such as a tackle or bait store, and may bring qualifying fish in during the entire time the contest is running. For the purposes of CRFS sampling, these types of contests are called derbies. Anglers participating in derbies are not in a tournament "T" and should be sampled as normal.

Opportunistic Sampling

In general, you must stick to sampling in the mode you are assigned for the day, even if you see recreational anglers in a different fishing mode at the site you are canvassing. The only exception is you may sample PC mode dockside while sampling in a different mode as long as you won't miss interviews in the assigned mode. Please see the PC section of this manual for further instructions on opportunistic PC sampling.

Opportunistic site effort checks (SEC) are mandatory at certain sites. Your Lead will provide you with directions on when and where to preform SECs. Site effort checks in BB and MM mode consist of counting the number of anglers fishing. SECs for PR modes are done by counting boat trailers. Sometimes SECs can be performed at nearby sites, or they may be completed at your current site for a mode other than your assigned mode.

Second Assignment

Multitasking is a handy skill to have as a CRFS Sampler. A successful Sampler is one who can take on many tasks, prioritize, and collect as much high-quality data as possible. Normally you will be scheduled for one CRFS assignment on any given day. However, budget or staffing issues may require you to work more than one assignment on the same day. If you have been given a second assignment by your Lead, generally you may begin working on it after completing your first assignment; both assignments would have a unique assignment ID. The second assignment would have a new primary site or cluster and possibly different mode

assigned. Sometimes you can complete one assignment during the down-time of another assignment, for instance, completing an opportunistic sample while sampling in another mode. Your Lead may give you side duties in conjunction with your primary assignment, such as pressure checks, helping another Sampler during peak hours or office work before or after your field shift. Work with your Lead to figure out a plan on these days with multiple tasks.

No Anglers at Site

After determining there are no anglers at your assigned site, please consult the mode sections of this manual for specific protocols. You may need to wait for effort to develop, or you may be able to move to another site, or the assignment might need to be rescheduled later in the month. If the weath er is bad and there are small craft advisories, the assignment might be terminated. If you have questions after reading the mode-specific instructions for no effort, please contact your Lead.

Incorrect Assignment

Occasionally a Sampler may accidently complete the wrong assignment. Examples could be: completing the wrong cluster assignment, sampling the wrong port, using the wrong form, or omitting a site in a cluster. In these cases, as soon as you discover the error, please inform your Lead immediately so a solution can be found. The assignment may need to be rescheduled or canceled. Do not discard any of the data sheets you may have completed during the incorrect assignment — submit them to your Lead along with the rest of your data. Be sure to include notes describing why the incorrect assignment was worked.

GENERAL ONSITE PROCEDURES

The onsite procedures differ somewhat by mode of fishing, geographic region, and the conditions at each site; however, the main goals are to collect effort information and to intercept and get complete interviews from as many eligible anglers as possible. Each CRFS interview may take anywhere from one to twenty minutes to complete, depending on the number of anglers contributing to that interview, how many fish they've retained, and the number of species present.

Surveys like this one require sampling of boats, anglers, and their catch in a random manner to produce a truly representative sample. Many systematic procedures have been developed that are intended to approximate a true random sample. Without any way to truly randomize effort you must use the methods described here to get a representative sample of anglers and fish. These methods described for anglers also apply to boats when the sampling unit is a boat (e.g. PR).

Overall, remember to reduce bias by not interviewing successful anglers only, or those at fillet stations, or by contacting just the friendly anglers. The sample of anglers should, without bias, accurately represent angler activity and catch rates of all species in the assigned mode on the date of your assignment.

Canvassing and Screening for Eligibility

Part of the CRFS Sampler's job is to build a rapport with anglers, make them feel comfortable, and determine if they are eligible for a CRFS interview. At shore sites you may canvass anglers to determine how long they have been fishing and how much longer they plan to fish at the site. You may also contact boat anglers who are launching and inform them you would like to speak with them when they come back to port. Anglers who have had the opportunity to meet the Sampler and discuss the survey will tend to be more cooperative when asked for an interview at the completion of their fishing trip. The canvassing can be informal; for example, the conversation might begin with "Catch anything?" or "How's the fishing?"

The purpose of canvassing is to screen anglers for eligibility for a CRFS interview. If they are NOT eligible, do not conduct a CRFS interview because the data may not be used. CRFS primarily samples recreational fin-fishing trips; however, CRFS interviews invertebrate only anglers but the emphasis will be restricted to collecting effort and reported catch information.

The definition of an eligible angler is one who has:

- been fishing recreationally in saltwater (seaward downstream of any saltwater cutoff)
- gear in the water or is part of a CPFV boat limit
- been targeting finfish or invertebrates
- completed their fishing trip in the assigned mode of fishing (exceptions for Shore modes, see MM and BB sections)

Examples of ineligible anglers would be: kelp harvesters, anglers setting crab pots, tide poolers, and anglers targeting sand crabs for bait.

Screening Divers

In addition to hook-and-line anglers, consider divers (spearfishers). If a diver uses a spear gun, they are to be considered "anglers" too. If the spearfisher targeted fish (speared a fish or intended to) they are eligible for a CRFS interview. Divers intending to take invertebrates by hand are also eligible anglers. Divers entering the water from the shore using a flotation device to kick out with fins to fish are considered MM or BB anglers (depending on the site and where they enter the ocean). Divers who access the water from a boat or other craft propelled by paddles/oars are considered PR anglers; this includes kayaks, stand up paddleboards, and pontoon boats with oars.

Kayak Anglers

Kayak anglers are of special interest to CRFS, as kayak fishing has become increasingly popular in recent years. Normally it is easy to tell a fishing kayak from a non-harvest kayak by the equipment onboard the kayak. However, you may need to speak to the kayaker directly to tell if they were fishing or not. Pay special attention to kayak fishers in the BB and PR modes.

CRFS Codes

CRFS codes are presented at the end of this manual. Your Lead may also provide you cheat sheets for common codes used in your District(s).

Angler Residence Codes: You will record the US zip codes for most of the anglers you interview. Use the foreign country codes for anglers who reside outside the US. If the angler does not know his zip code, you may record a city instead and try to look up the zip code later. Zip codes are not necessary for a valid interview so you may record a "don't know" or "didn't ask" as a last resort and proceed to the next part of the interview.

<u>Site Codes:</u> These are provided to you on your current site list, as part of your Monthly Schedule, and are also listed on the CRFS Wiki website. Site codes are numeric (NNN-NNN) and some sites can be represented using an OSP Port Code (XXX). OSP Port Codes are given to PR1 sites and CPFV landings. OSP Port Codes are listed at in the end of this Manual as well.

Species Codes: Five letter fish species codes have been provided to you in this manual and are sorted three ways: by code, common name, and by American Fisheries Society (AFS) common name. These lists include most finfish species (and some inverts) found on the Pacific coast. All codes should be listed, if not contact your Lead. These species codes are used for both the target(s) on the trip and for the catch records. You will become familiar with the species codes of fish targeted and caught in your District(s). When in doubt always look up the code in this manual; never guess or make up your own species code.

Gear Type

During a CRFS interview, the Sampler collects effort information including gear type used. The most common gear type encountered for anglers targeting finfish is hook-and-line (H). However, there are several other methods of take that anglers may use. The gear type should be recorded under GEAR on the following forms: PR, PCS, Shore, PCD, and PC Angler. A gear type is needed for both primary and secondary targets. If there is no secondary target, gear type should be left blank. Gear type is not a required data element for a valid CRFS interview, but it is important information to collect and it is mandatory for a salmon interview. See below for definitions of each finfish gear type.

The following gear codes apply to finfish targets:

 $H = \underline{\mathbf{H}}$ ook-and-line is a gear type used to take finfish. A hook or multiple hooks tied to a line that is attached to a reel mounted on a fishing rod, a handheld reel or the line can be tied directly to a rod without a reel. A poke pole, usually used in the intertidal, is another example of hook-and-line gear. Hook-and-line gear type is available for all finfish except salmon.

 $S = \underline{S}$ pear is a gear type used to take finfish; either fired from a gun-like launcher, powered by one or more elastic bands, or a pole launched by hand using a single elastic band (e.g. Hawaiian sling).

 $T = \underline{T}$ roll is angling from a boat or floating device that is making way by means of a source of power, other than drifting by means of the prevailing water current or weather conditions. Trolling is typically used to take salmon and large pelagic finfish like tunas.

 $N=Bait\ \underline{N}et$ is a category that includes cast and dip nets. Bait net is defined as any type of net actively propelled by hand through the water or thrown with the intent to capture fish. Examples include dip nets, Hawaiian throw nets, seine nets (also called beach nets), and A-frame nets (like those used in the night smelt fishery).

The following gear codes apply to salmon only:

M = Mooch is salmon fishing <u>with bait</u> from a boat or floating device that is making way by means of the prevailing water current or weather conditions only. Accurate recording salmon gear types is important for fisheries management. The mortality rate applied to released salmonids is determined by the gear type (i.e., 42.2% for mooching with bait, 14% for trolling).

 $B = \underline{B}$ oth (mooch & troll) is coded to indicate that salmon anglers used both mooch and troll gear types.

The following gear codes apply to invertebrates only:

P# = Pots, and the number of pots pulled (used on vessels). Pots are an enclosed trap with ports to allow entry to access bait and then prohibit legal sized invertebrates from escaping.

 \mathbf{F} # = $\underline{\mathbf{F}}$ lat ring/hoop nets, and the number used. Flat nets have collapsible sides that lie on the bottom when deployed. When retrieved, the sides of the net are raised which makes escape difficult.



 $R\# = \underline{R}$ igid hoop net, and the number used. Rigid nets have sides that are fixed in place with supports, making the net stand erect when deployed on the bottom. The diameter of the opening at the top is less than the diameter of the bottom making escape difficult.



 $\mathbf{E} = \operatorname{Snar} \mathbf{\underline{E}}$ (a rod and reel device). Snares are a small cage-like structure that holds bait, with up to six monofilament loops on the outside of the structure. It is attached to a rod and when reeled in the loops constrict, trapping the legs of the crab.



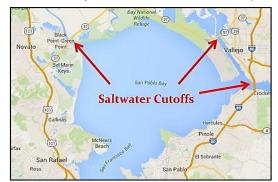
C = Hand while S**C**UBA diving with tanks

D = Hand while free **D**iving with no tanks

Areas Fished and Saltwater Cutoff Points

Samplers should pay attention to the area in which they are sampling. Open ocean typically refers to offshore areas, more than three miles out, in federal waters, indicated by a solid red line on your CRFS Block and Box maps.

Nearshore areas are within three miles of shore but outside an enclosed bay or estuary. Bay areas are inside enclosed bays, estuaries, or harbors.



River areas are typically not surveyed. CRFS is a marine survey and it is necessary to establish saltwater cutoff points at some locations. It is

possible to interview in the tidal portion of a river. It is mandatory to screen

anglers to see if their fishing was seaward of these saltwater cut off points. If any of their fishing was done seaward of these points, they are eligible anglers. If all their fishing was done above these points, they are ineligible for the CRFS interview. If you are recording catch, only record the catch caught seaward of these points. Some areas where anglers in freshwater need to be questioned regarding saltwater fishing are where US 101 or US 1 (Pacific Coast Highway) crosses estuaries and near rivers entering San Francisco Bay. Be sure to screen any boats that may have fished near these areas to see where their fishing was done. They may not be eligible for the survey.

Occasionally you might interview an angler who states they were fishing in "brackish" water. If the location cannot be determined from your laminated maps, ask, "If you had to pick either salt or fresh water, what would you pick for most of your fishing today?" If the angler chooses fresh water, you should stop the interview. Occasionally an angler will report saltwater fishing at a fresh water location, in this case complete the interview and write a comment on the form and follow up with your Lead.

Saltwater Cutoffs

County	River	Saltwater Cutoff Point
Del Norte	Smith River	1/4 way between mouth and Hwy 101
	Klamath River ¹	1/4 way between mouth and Hwy 101
Humboldt	Mad River	1/4 way between mouth and Hwy 101
	Eel River	Upper end of Cock Robin Island
	Redwood Creek	1/4 way between mouth and Hwy 101
Mendocino	Ten Mile River	Old dock,100 yards up from Hwy 1
	Noyo River	End of Dolphin Cove Marina
	Big River	Mid-2 nd turn upstream
	Albion River	Upperdock
	Navarro River	Highway 1 Bridge
Sonoma	Petaluma River	Highway 37 Bridge
	Coastal Rivers	Highway 1 Bridges
Napa	Napa River	Highway 37 Bridge
Solano	Sacramento River	Carquinez Bridge
Contra Costa	Sacramento River	Carquinez Bridge
San Mateo	Coastal Rivers	Highway 1 Bridges
Districts: 1-3	Coastal Rivers	PCH (Hwy 1) bridges (excludes
1	rding DD compling procedu	Elkhorn Slough)

¹Consult Lead regarding BB sampling procedures on the Klamath River.

Catch Locations and Map Use

Catch locations are important for boat modes. You will be given a set of laminated CDFW Block and Box maps. The location procedures gather information about the boat's location of catch, or effort if there is no catch. Catch location by species or species group is being used to study areas where species of interest are being caught (or not being caught) for purposes of protection/conservation through the use of conservation areas, depth restriction boundaries, and potential MPAs, or refugia. Rockfish depth-

dependent mortality rates applied on a depth-and-species basis may also differ by location.

The best person on the boat to contact dockside about catch locations will most often be the "pilot" of the vessel, also called the captain or skipper. We can think of every angler on a particular boat typically fishing and catching fish at the same locations.

Collecting location information may be one of the more difficult aspects of this survey. Anglers may not be able to provide their fishing location for a few reasons; anglers may be unaware of the location and unable to read or interpret your maps, they may not want to spend the time to provide this information, or they may be generally unwilling to divulge their favorite fishing spot. The Sampler will attempt to overcome these problems by being persistent, friendly, and helpful. Become familiar with local on-the-water and on-the-map landmarks and fishing site names to assist the angler in determining their fishing location. The Sampler must be convincing and credible while explaining the importance of gathering this data. While location is important, if an angler cannot or will not provide this information, continue with the CRFS interview. The absence of location data does not render the interview unusable: remember that the CRFS interview is voluntary.

Latitude and Longitude and the One-Minute Grid

The Equator has been designated as 0° (zero degrees) latitude and the north and south poles are 90°. Greenwich, England was designated as 0° (zero degrees) longitude. Any geographic location on Earth can be pinpointed on a map using the latitude-longitude grid system. The accuracy of the degree grid is increased using minute and second subdivisions of which there are 60 of each. A degree is about 60 nautical miles, a minute is about one nautical mile and a second is about 100 feet. Closer to the poles, longitude lines narrow and the grid is not as square. In California, we can assume square grids for this study. In this project, we will be working mainly at the minute level resolution (about a square nautical mile) for locations on maps. One-minute grid maps have been developed for this purpose. Latitude and longitude, common fishing sites and buoys, depth contour lines, and county lines are labeled on the maps for reference.

CDFW Block-Box Maps

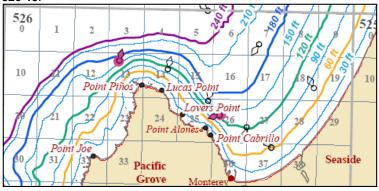
The CRFS format for mapping coordinates is the block-box system which uses pre-defined numbers to indicate a location within one nautical mile. Each block is 10 by 10 nautical miles with 100 boxes numbered 0-99 within each block. One box is approximately one square mile. Three digits are required for the block (BBB) and two digits for the box or microblock (bb). So, each pre-defined box can be expressed with the BBB-bb format. Multiple boxes in a block can be expressed by adding more box codes; BBB-bb-bb. Boxes inside enclosed bays and estuaries have boxes numbered with three digits (bbb) starting at 100. Each box can be converted to latitude and longitude coordinates (point data) if needed, using the center of the box. If all that is provided is the block (BBB) the coordinate will be the middle of the

block with a size of 100 square miles, which is not very precise. Try to get as fine scale catch locations as possible.

CDFW Fisheries Chart showing the 100 square mile CDFW blocks for the Monterey area.

524	523	522	521	520	519	518 517 516
		531	530	529	528	525 527 526 Monterey
546		537	536	535	534	533 532 Point Sur

Example Block-Box map; the box West of Pt. Piños is described as 526-13.



Definition of Location

A catch location can be described in several ways: a latitude/longitude, block-box, block and two boxes or area around a block-box. Location is always described to the nearest minute of latitude and longitude and is seen as one minute squares on a map with a point in the center where the east and west "minute" lines cross. A minute square is a large area of approximately a square mile.

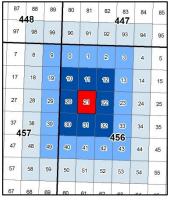
Accuracy of Location Grid

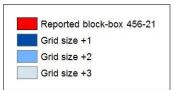
Recording a location can be seen as a tradeoff between getting an exact location for a fishing spot and including the majority of the catch. A less precise location covering a larger area may be used to encompass the majority of catch locations to form a "location cluster". The best location data captures catch location by species, to the box-level. Coding all catch on a boat to a broad area does not provide much for analysis. It's more important to try to identify catch locations based on species groups. For instance, anglers may target different species by fishing in a certain area, over a specific bottom-type or with specific gear; however, targets are one thing and catch is another. We are interested in where catch occurred.

The Grid Size Item

The grid size is used as a way to indicate the extent or size in minutes around

a block-box location. If an angler fishes over an area larger than a single 1minute block-box and doesn't specify exactly which additional boxes were visited, then a grid size should be used. A grid size of "1" increases the area fished to include the boxes immediately adjacent to the center block-box creating a total area fished encompassing 9 boxes equivalent to a 3-minute by 3minute area. Each grid size increment above 1 will increase the size of the area by adding the next row and column of adjacent boxes (see example). To include all the colored boxes in the example the location would be coded as 456-21 +3. The grid size can create a large area, so please record locations as precisely as possible and keep grid sizes to a minimum.





Fishing Site Names

You may use fishing site names to orient anglers when viewing the maps. Be aware that one site may have multiple local slang names. Please avoid prompting the angler when asking about catch locations, such as "Did you fish at Mulligan's Hill today?" Prompting with specific location names introduces bias into data collection. When receiving an unfamiliar location name, have the angler locate it on the block-box map. The intent of this survey is not to confirm pre-conceived fishing locations, but to collect unbiased catch locations. Many previously "known" fishing locations become "fished out" and effort may shift in location and extent along with fish availability in the wake of these events. Fishery managers need the ability to study these events when they occur.

Fish Identification

In addition to the comprehensive list at the end of this manual, your Lead will provide you with "cheat sheets" of local species by species groups. It is your responsibility to know and identify the more common species by sight. Studying the identification guides and training in the office and aquarium, combined with experience in the field, should make you knowledgeable in species identification in a short time. Learn the CRFS priority species. All fish that are presented to the Sampler should be identified to the species level. Fish should be recorded using American Fisheries Society Common Names and with the correct species code. Samplers should never code a fish to the species level when they are not certain of its identification. Be aware that anglers may use slang names; slang names are names other than American

Fisheries Society names. Samplers should not record a slang name in place of its AFS Common Name. The Other Codes section of this manual lists slang names.

You are issued two field guides: Miller and Lea's Guide to the Coastal Marine Fishes of California Fish Bulletin #157 and Peterson's Guide to Pacific Coast Fishes or its replacement, A Field Guide to Coastal Fishes from Alaska to California. Miller and Lea should be used as your first source of information and should be with you at all times in the field. You never know when you are going to run into a rare fish that will need to be keyed out. Other field guides or cheat sheets are available. Often your Lead will have additional identification books available in the office. Contact your Lead to inquire about using these books or making a photocopy. Your Lead should also have access to a fish photograph database; contact them to view these images. If you simply cannot identify a fish in the field, please take the time to key it out and/or take multiple photos of the fish (see Specimen Documentation below) and record any key features to help you and your Lead identify it later.

Observed Catch (Sampler-Examined)

The Sampler will strive to examine all landed catch. Examined/observed catch is the most robust catch category because the Sampler saw it, counted it, and identified it to species. All observed kept fish should be identified to the species level. Fish reported above the species level (i.e. genus, family, or group) must be recoded as "kept unobserved" fish (a type of angler-reported catch, see below) and not as observed catch. If the angler(s) refuses to have the landed catch examined, it must be coded as kept unobserved.

It is more important to count and identify all rockfish to the species level than to get bio data from those fish.

Unavailable Landed Catch (Angler-Reported)

The "Kept Unobserved" category includes catch that were kept by the angler, but for some reason the Sampler could not observe and identify to species. This type of catch category is angler-reported because fish were unavailable for positive identification by the Sampler (packed away, filleted, given away, fed to birds, or kept and used for bait).

For catch unavailable for identification (fish the Sampler cannot readily view), the Sampler will help the angler determine the catch to species level (ideally) or genus, family, or group. Hopefully, avid anglers can accurately identify catch, so you may be able to determine the unavailable catch to species level. To help the angler, you should be familiar with the fish caught in your area that are commonly released, used for bait, etc. Mark these species in your field guide, so if the angler doesn't know the species of unavailable catch, you can show them pictures. Never code the catch beyond a taxonomic level you feel confident with; however, keep in mind that fisheries managers rely on catch estimates by species. Try to determine unavailable catch to species level. If that cannot be done, code catch to genus or family, or an even more general code, like "bottomfish" (BOTOM).

Be persistent with anglers who have kept rockfish which are unavailable. The general 'RFGEN' code is not ideal for managing this fishery. Samplers should try to get visual observations to identify and enumerate rockfish to species whenever possible. Use your best effort to gain access to the catch for species identification.

Filleted Catch (Sampler-Examined or Angler-Reported)

Filleted catch are fish that have been somewhat processed before the sampler saw them. If the Sampler encounters filleted fish with attached skins, such as for rockfish or lingcod, the Sampler should try to identify those fillets to species. If the angler refuses or the Sampler is unable to identify the fillets, they should be recorded as "kept unobserved" fish. Count the fillets to get an accurate number of fish landed if the angler doesn't remember. Unidentified fillets may not be recorded as observed catch, even if you physically see a bag of fillets. This is especially important for filleted rockfish; never record unidentified rockfish fillets "RFGEN" as observed catch, even if you have enumerated them and identified the genus as rockfish. Only record fillets as "kept observed" when those fillets have been identified to the species level, by the sampler, based on the skins.

Oftentimes, a bag of fillets will contain some unidentified taxon such as rockfish genus, tuna, bottomfish, etc. and should be recorded as "kept unobserved". Try to have the angler provide the species of the fillets, if they are confident in their identification skills. Unfortunately, anglers will often refuse to let you open their bag of fillets. If this happens, make a note on the data sheets that you were dealing with fillets, as this explains why they were not "observed" and there is no bio data associated.

Unidentified Examined Catch

Samplers are expected to identify all fish to species level by recording the species name and/or code. You should be familiar with the species of fish caught in your area in the different fishing modes. It is important to know which species are commonly confused with each other. If you encounter a species you do not know, you should key out any marine sport-caught fish. Never code the catch beyond a taxonomic level you feel confident with. If you are not confident, take multiple photos, note key features, and share them with your Lead (see Specimen Documentation below) who will help you identify the fish. Take notes on what you think the species may be, and record the location of catch, depth, and any bio data which will help to identify the fish later.

Released Catch (Angler-Reported)

Released catch are fish that were intentionally released back into the water after being caught. In order to release the fish, the angler first must have had control and possession of it. Do not record fish the angler may have had on the line but didn't actually land. If the angler has available catch, you may be able to use it as a reference in determining what was discarded (e.g., ask,

"How many fish of this type were thrown back?"). Use fish identification charts and guides to help anglers determine the species of fish they released.

Fish that are released are further divided into two categories: released alive and released dead. The Sampler will ask the angler to determine if fish were released alive or dead. Fish that are not moving in the water are considered dead. Fish that are alive but are obviously not going to survive due to severe wounds or inability to swim down are to be coded as dead. Severe wounds include bleeding gills. For Sebastes spp., and other species with swim bladders brought up from deep water, there may be obvious signs of barotrauma (expanding gas) such as a protruding esophagus and eyes. Increased buoyancy may prevent the fish from swimming down from the surface. If the fish is unable to swim down, consider it dead. Fish returned to depth using descending devices are considered alive. The disposition of released catch is usually not something the Sampler can witness. The release event usually happened earlier in the day when the Sampler was not there. Record the disposition based on clarification with the angler if the released fish were returned alive or dead by asking, "Did the fish swim away?"

Specimen Documentation: Rare or Large Fish

If you encounter a rare species that you cannot positively identify, attempt to key it out. This may not be possible due to time. If the angler is in a hurry, make a note on the data form and take several photos with a camera or smartphone. Take a photo using these guidelines:

- Have the head of the fish pointing to the left
- Get as close as your camera will allow (try macro mode)
- Have something in the photo to provide scale or take the photo with the fish on your board
 - If on your board, move the fish to the center of the board do not have it pushed up against the stop as if you were measuring it. Maxillary extension is crucial so make sure the mouth is closed.
- Spread out the fins as much as possible
- Take the photograph in adequate light not in deep shade. Make sure the subject is completely, not partially, lit.
- Take two or three shots just in case

Be aware of the maximum lengths of species as listed in your copy of *Guide* to the Coastal Marine Fishes of California and take pictures of fish that exceed these lengths. Your Lead may also provide you with a "cheat sheet" of maximum reported lengths. Fish over the maximum size will be flagged in the database; without photographic proof, the record will be discarded from the database.

Sub-Sampling Biological Data and CRFS Priority Species

When sampling during busy periods, you may need to subsample weights and lengths from most species (excluding ad-clipped salmon and species of concern. This should be done in a random or systematic fashion. Subsampling fish should always be done at the species level. Never sub-sample within a species based on a particular length or weight (e.g. sampling big or

small fish). For a sample to be random every member of a given population must have an equal probability of being selected. Whenever the sample has more fish than will be measured, you should use one of the following procedures:

- (1) The Sampler should take out all the fish from the angler's 'bag' and line them up by species. Calculate the sampling fraction, *n* (e.g. every third fish), and weigh and measure every *n*th fish. Select the starting fish at random.
- (2) If there are too many fish to systematically sample in the given time frame, or if the surroundings make it very difficult to sample using the above method, you should randomly select 5 fish. At no time should you try to pick out the average or representative fish or the largest and smallest fish this is not a random sample.

The CRFS Priority Species table below shows the top priorities for fish species sampling. The most important fish to measure are salmon of all species with a clipped adipose fin, non-retention species (i.e. fish that are illegal to keep), species with special weekly tracking (species of concern), and species with harvest limits.

It is important to see all salmon catch. It is important to obtain lengths and weights of rockfish, especially rare and non-retention species. Priority species may be added as emerging fisheries develop on data poor species. Groundfish harvest limits are set in metric tons and to get the best estimates we need both lengths and weights. Salmon are managed using the number of fish, so fishery managers only need the lengths of salmon with clipped adipose fins. Paired length and weight measurements are preferred but length only measurements can still be used; we cannot have a weight without first having a length. However, both length and weight can help with data quality since most species have an established length-weight ratio that your data will be checked against. You should try to get as many lengths and weights as possible for all fish (except salmon), but when sub-sampling is required the following species categories should be measured first:

CRFS Priority Species

	ionity operate								
Highest P	riority:								
Ad-clipped	Ad-clipped salmon (both Chinook and Coho), length only thresher shark								
Higher Pri	ority: Species of Concer	n (in no par	ticular orde	r)					
yelloweye canary roo	, cowcod, bronzespotte ckfishes	d and	Pacific halibut		bluefin tuna				
High Prior	rity: Species with Harves	t Limits (in	no particula	r order)					
cabezon	California sheephead	greenlings		black, black-and-yellow, blue, bocaccio, brown, copper, calico, China, gopher, grass, kelp, olive,					
lingcod	ngcod California scorpionfish		(Hexagrammos spp.)		quillback, treefish, widow, and yellowtail rockfishes				

CRFS Protocol for Dealing with Suspect Data

In the field, if an angler reports unusual or suspect catch data to the CRFS Sampler, additional steps should be taken. Here are some clues to help the Sampler recognize suspect catch data:

- The reported species is out of documented range
- The reported species is unlikely to be taken in the fishing mode in which the angler is currently fishing
- The reported species is unlikely to have been caught in the area the angler claims (e.g. bay vs. open ocean)
- The reported species is unlikely to have been caught at the depth the angler is reporting
- The reported species is unlikely to be taken using the fishing gear the angler reported fishing with
- The angler incorrectly identifies the landed observed catch
- The novice angler admits that they may not be familiar with local species
- The angler reports a catch number that seems unrealistic

When the Sampler realizes that the data is suspect, the following steps should be taken:

- Identify the angler from the boat or bag that encountered the suspect fish
- 2. Use the available fish ID materials to confirm the species with the angler. Show the angler what characteristics are used to identify the species in question and point out other species that are commonly confused with the species in question. List the characteristics that may be used to distinguish similar species, and ask if the angler noted any of these key characteristics
- 3. Do your best to ascertain the angler's fish identification skills. Is the angler able to identify the kept observed catch? The avidity question will give a clue to how often the angler fishes. Does the angler frequently fish in this area or at this site?
- 4. For species reportedly taken at an unusual depth, ask the angler how confident they were in the accuracy of the depth reported. Did they have a depth finder on board? Was the suspect fish caught in the same location and depth as any of the kept species?
- 5. Ask the angler to quantify how certain they were in their identification of the suspect species (e.g. 100% certain, less than 50% certain). Record this information on the data sheet. If the angler is less than 50% certain, consider speaking with other anglers in the group who may be more knowledgeable, if possible.
- 6. Circle the suspect catch data on the data sheet
- Note on the ASF that the Sampler collected suspect data so that the data editor knows to look for and assess it
- 8. On a busy day, do not miss salmon boats to verify suspect data. When missing salmon catch is not a concern, Samplers should spend more time verifying suspect data (especially for Yelloweye

and Cowcod Rockfishes, Pacific Halibut, and other species of concern).

When the Sampler observes a species of concern, unusual or rare species that may be considered suspect, the Sampler should take steps to verify that what they observed was correct. Try to take a photo of all kept Yelloweye Rockfish, Cowcod, and any unusual species (such as out-of-range, oversize, or uncommon species) or fish that you are unable to identify (see Specimen Documentation above). Yelloweye Rockfish collection is covered in the Species Sampling section of this manual. Email the photo(s) to your Lead as soon as possible to document what you saw and validate the species identification. If you are unable to take a photo, please list the characteristics that you used to identify the species. Circle the suspect data that you observed on the data sheet and make a note on the ASF and in the Weekly Report. Follow step 8 above regarding missing boats when you observe what may be considered suspect data.

When the data editor comes across the data sheet with suspect data, the following steps should be taken:

- Review the steps the Sampler took to verify the accuracy of the data
- Determine if the Sampler missed any steps in the verification process listed above
- Contact the Sampler by phone as soon as possible to go over the situation, confirm all the details, and inquire if there is any additional information regarding the suspect data
- 4. The Sampler may be asked for a separate write-up to document the interaction with the angler claiming suspect data
- The data editor will take notes of the conversation with the Sampler and append them to the data sheet containing the suspect data
- 6. Using their fish ID skills and knowledge of the local fishery, the data editor will work with the Lead(s) to form an opinion as to the accuracy of the suspect data and provide a recommendation. Those notes will be appended to the data sheet
- 7. The Lead will notify the Supervisor and CRFS Coordinator about suspect data involving Yelloweye or Cowcod and provide the notes pertaining to the suspect data

Biological Data Collection

Species-level data collection is the most important. After determining catch species, the Sampler will measure as much of the finfish catch as possible (except for salmon, only adipose fin-clipped salmon are measured for length). Lengths and weights should still be taken from fish the Sampler is not able to identify, in hope of being able to use these data if the Lead is able to identify the fish based on photos/notes that the Sampler recorded.

Samplers should measure and weigh up to five (5) fish of each species in the bag or boat. If the bag or boat has more than 5 fish of one species, select no more than 5 for biological data collection. Missing biological data should be explained on the form. The goal is to get paired lengths and weights of 5 fish of each species. Weights may be missed if time does not allow. It is often difficult to obtain weights onboard PCs especially in less than ideal conditions. It is important to the CRFS program to collect biological data from fish that are under active management, also called "Species of Concem". Lengths can be used to predict weights and to examine length classes. For fish that are not weighed, weights will be calculated based on the length data. Weights are used to help with length-to-weight predictions, estimate mean weight and total metric tons harvested.

Fork Length Measurement and Use of the Measuring Board

The Sampler shall measure fish to the fork of the caudal fin for all species with such morphology. See instructions below for measuring species without a forked caudal fin. Fish fork lengths must be taken using the measuring board and recorded to the nearest millimeter. The measuring board is labeled in centimeters, but tick marked in millimeters. Remember to multiply the centimeter reading by 10 before adding the number of smaller markings past the label. For example, a fish that measures to the third line past 23 would be 233 millimeters. Samplers should never round lengths and weights. Rounding fish measurements will introduce a "digit bias" and will be seen in the data. Do not measure fillets. Fish must be laid flat with the mouth closed, pushed up against the stop. Keep head and tail in a straight line where possible. The tail fin may need to be spread flat to its natural position to allow for accurate identification of the fork or longest point.



A measuring board used must be unless fish а exceeds the length of the board, then use a tape measure. Tο use the measuring board: Place measuring board on a hard, level surface

- 2. Straighten the fish as much as possible if rigor mortis has set in
- 3. Place the fish with the nose flush against the bracket end of the board and with the body centered over the measuring board
- 4. Close the fish's mouth

5. Keeping the nose of the fish against the bracket, press the tail down to the surface of the board. The fin may need to be spread flat to identify the fork. Read the length at the fork of the tail to the nearest millimeter.

Samplers will also carry a tape measure to be used only on specimens that exceed the length of the measuring board. To use a tape measure:

- 1. Pull some slack in the tape
- 2. Lay the tape on a hard surface
- 3. Place the fish on top of the tape (see example, right). The tape must not be on top of the fish as this will result in an exaggerated or inaccurate measurement as the tape bends to the contour of the fish's body
- 4. Pull the slack out of the tape make it tight under the fish's body
- 5. Read the length at the fork of the tail to the nearest millimeter
- 6. Clean the tape measure before it is used again

Alternate way to measure large fish:

- 1. Place the measuring board on a hard, level surface
- 2. Straighten the fish as much as possible
- Place the fish with the nose flush against the bracket end of the board and close the fish's mouth
- 4. Use the tape measure to measure the length of the fish that spills over the end of the board
- 5. Make sure to line up the tape's beginning with where the board ends

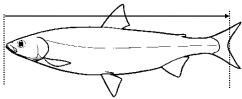




Measuring Various Types of Fish

Most species are measured from the most anterior tip of the longest jaw (mouth closed) or end of snout, whichever is terminal, to the posterior tip of the tail at its center line. This procedure is the same whether the tail forks in (e.g., mackerels) or protrudes out (e.g., flounders).

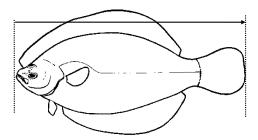
Salmonids - Salmonidae



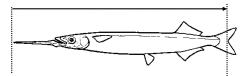
Eelpouts - Zoarcidae



Left eye flounders - Bothidae

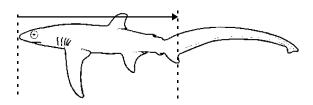


Halfbeaks - Hemiramphidae

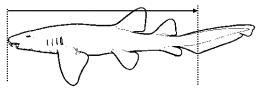


Sharks and sturgeons are measured from the tip of the snout to the center of the fork of the tail. For sharks without a fork, measure the shortest distance to the ventral lobe of the tail (See nurse shark below).

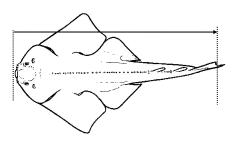
Thresher sharks – Alopiidae



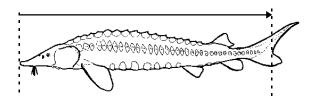
Nurse sharks - Ginglymostomatidae



Angel sharks - Squatinidae

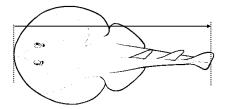


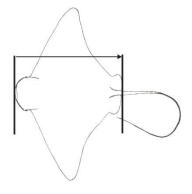
Sturgeons – Acipenseridae



Skates and rays are measured from the tip of the snout to posterior end of the pelvic fins. Do not include the claspers (if any). When a caudal fin is present, the fish is measured to the caudal fin.

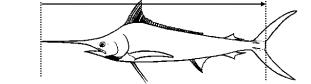
Electric rays - Torpedinidae





Bat rays - Myliobatididae

Billfish and swordfish are measured from the tip of the bill to the center of the fork of the tail.



Billfishes - Xiphiidae

Weight Measurement and Scale Use

Fish weights are to be recorded to the nearest hundredth of a kilogram (0.01 kg). The hundredths place may be a zero unless weighing small fish with the 1 kg hanging scale. Calibrate your scales weekly. Samplers should zero out all their scales at the start of each assignment. Four scales will be provided to each Sampler: One Pesola 1 kg scale, and three brass scales of 5 kg, 12.5 kg, and 25 kg capacity. The 25 kg scale is labeled in pounds and kilograms and displays measurements in 0.25 kg increments. The 12.5 kg scale is labeled in pounds and kilograms and is accurate to tenths of a kilogram. The smaller 5 kg scale is labeled in pounds and kilograms and is accurate to tenths of a kilogram (0.1 kg or 100 grams). The 1 kg scale is accurate to 1 hundredths of a kilogram (.01 kg or 10 grams).

You are expected to use the most accurate scale for each fish weighed. Do not record a fish weight that exceeds the capacity of the scale. Do not weigh a less-than one kilogram fish on a scale with a larger capacity than your one-kilogram scale. It is permissible to collect weights for bled fish. The weight of blood falls within the variability of stomach contents. With tunas record if the fish was bled next to the weight. Do not weigh gilled, gutted, or beheaded fish. Do not weigh salmonids. Do not weigh fish that are too lively to get an accurate reading from the scale.

After the scale has been exposed to saltwater and/or fish slime, rinse the scale in fresh water in the field if possible. At home wash the scale in hot soapy water. Rinse the scale in hot clean water to heat the metal to speed drying. Shake excess water from the scale. Place the scale in a dry warm place like in a sunny window, a warm oven, or under a hair dryer. When dry, spray with WD40.

Scales should be calibrated weekly or at least every month. Your Lead may require scale calibration documentation. Your Lead has calibration weights you may use to check your scales. Please calibrate outside in a well-ventilated area if you plan to use WD40. If the calibration knob seizes, notify your Lead for replacement. To adjust scales, here are a few items of known approximate weight you can use to check the accuracy of your scales:

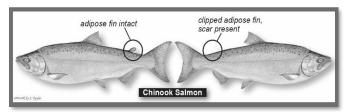
- 25 kg brass scale = 0.39 kg
- 1 liter of freshwater = 1.0 kg
- 1-gallon plastic jug of water = 3.9 kg

SPECIES SAMPLING PROCEDURES

Specific procedures have been developed for sampling salmon, Yelloweye Rockfish and White Seabass.

Salmon Sampling

All kept salmon must be examined for a clipped adipose fin (the small fleshy fin on the back of the fish between the dorsal and caudal fins). The adipose fin clip indicates the presence of a coded wire tag (CWT) in the salmon head. At least 25% of hatchery released salmon are tagged. Check to see if the salmon is missing its adipose fin. If so, explain to the angler that you need to collect the head for fishery management purposes. You have legal authority



to do so according to Section 1.73(b) of Title 14, California Code of

Regulations (see Section 'Legal Authority' below). Angler refusals are generally rare, but do occur (see Section 'Non-Recovered Species (NRS) Protocol' below). Attach the headtag to the salmon head, measure the fish, record the headtag number and fork length in millimeters on the data sheet and then remove the head. Place each tagged head in its own small clear zipper bag. It is important to follow this sequence. Store the head in a cool location untilyou can get the head into a freezer. Record the date, port, and sampling mode where each headtag was collected or issued on the Headtag Report Form. You will never need to weigh a salmon, even an adipose finclipped fish; sport salmon management is based on numbers of fish, not on weight.

Legal Authority

If an angler refuses to relinquish the head of a salmon inform them of the state law. Recovery of Coded-wire Tag from Salmon Head, Section 1.73(b) of Title 14, California Code of Regulations: Any person in possession of a recreationally taken salmon with a missing adipose fin (the small, fleshy fin on the back of the fish between the back fin and tail) shall immediately relinquish the head of the salmon, upon request by an authorized agent or employee of the



department, to facilitate the recovery of any coded-wire tag. The head may be removed by the fish owner or, if removed by the official department representative, the head shall be removed in a manner to minimize loss of salmon flesh and the salmon shall immediately be returned to the fish owner.

Salmon Equipment

- 1. Head removal equipment:
 - Knife and sheath
 - Cutting Board

2. Headtag Kit:

- Headtags
- Small clear zipper baggies (for each head/headtag)
- Headtag Report Form
- Large clear bags
- inventory tags

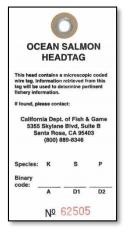
3. Courtesy Headtag kit

- Courtesy Tags
- Orange Information Request Cards
- Courtesy Headtag Report Form

Tagging the Head

A uniquely numbered headtag is issued for each adipose fin-clipped salmon observed while sampling. Place individual tagged heads in small clear zipper bags with the headtag number facing outward so it can be clearly seen from outside the bag. Place individually bagged heads into a large clear plastic bag. Attach an inventory tag to the outside of the large bag of heads. Using any type of non-clear bags will not be allowed as they can easily be confused with trash. See Section 'Non-Recovered Species (NRS) Protocol' below for instructions when the angler refuses to allow the tag to be applied to the salmon head.

Store the head in a clear zipper bag and freeze as soon as possible. If freezing is not immediately available keep the heads in a cool place to slow the



decomposition process. The zipper bag allows the lab to separate the frozen heads without damaging or tearing the headtag.

Removing the Head

- Using the metal wires of the headtag, securely attach a headtag to the lower jaw of an adipose fin-clipped salmon
- Lay the fish with the head on the cutting board portion of the measuring board and record the fork length and headtag number.
- Slide your knife under the gill plate and cut straight forward or at a 45 degree angle, until you are approximately 1 inch behind the eyes
- 4. Flip the fish over and repeat the cut until it meets the end of the first cut. You may have to angle the knife perpendicular to the ground to meet the other cut.
- 5. Once the two cuts have met, the head should come off cleanly





Make sure the cut exposes the least amount of meat possible and remove any gills or extra flesh attached to the head. Please keep your board and knife clean as you are dealing with fish that someone will be eating. If an angler prefers to cut the head off themselves, let them do it, however they must use their own knife.

Non-Recovered Species (NRS) Protocol

Non-recovered salmon Heads are rare. Most salmon anglers are aware of the CWT program and the legal requirement to relinquish the head of adipose fin clipped salmon. If you cannot remove a head for some reason, attempt to attach the headtag to the fish and record the species and fork length. Point out the toll-free number on the headtag to the angler; they may decide to relinquish the head later. Record this information on your data sheet (i.e. headtag number and fork length) and put NRS next to the headtag number. Record NRS and the species name on the back of the corresponding headtag and on the Headtag Report Form. If you are unable to attach the headtag to the head, record the headtag number and NRS on the data sheet and Headtag Report Form, place the headtag in its own small zipper bag and store it with the rest of your collected salmon heads. This information is important in tabulating the contribution rates of hatchery-origin fish to the year's catch.

Q. How do I persuade an angler to relinquish their salmon head?

A. If the angler refuses to relinquish their head, try these tactics:

- 1. Inform the angler about the importance of coded-wire tags to salmon management.
- 2. Offer to provide information to the angler about their fish through the information request card program.
- 3. Remind the angler that by law, he/she has to relinquish the head under Section 1.73(b), Title 14, CCR.
- 4. Try to attach the headtag to the fish if possible and point out the phone number on the tag. The angler may decide to relinquish the head later. If the angler still refuses, follow the NRS protocol and notify your lead promptly. Document the vessel's CF number and the license plate number of tow vehicle on your ASF.

Q. What if the salmon is confiscated by a Wildlife Officer?

A. Ad-clipped salmon that are confiscated should still have the headtag attached and fork length information collected. Record the head as an NRS. The attached tag will be a reminder that they are to be returned to the Ocean Salmon Project (OSP). Collect the name and contact information of the Wildlife Officer.

Enforcement personnel will be contacted to remind them that OSP needs the confiscated head.

Procedures for Tracking and Inventorying Salmon Heads

Each headtag is recorded on a Headtag Report Form. Fill out the form at the end of each sample day to accurately keep track of which headtags are used on each particular sample day. When inventorying your heads at the end of your sampling day, ensure each headtag number recorded on your data sheets match a salmon head in your possession. If there are discrepancies try to identify the source of the error. Each Monday a copy of the form will be sent to OSP. Once all tags in the series have been used, mail the original Headtag Report form with your weekly data.

NAME:_					ADTAG REPO s in NUMERICAL ord		SERIES#: 10000 - 10099				
Headtag#	MM / E	D/YY	Port	Sample Mode	Headtag#	MM / I	DD / YY	Port	Sample Mode		
10000				20	10050						
10001				20 10	10051				(A)		
10002			=		10052						
10003	/_	/			10053				2		
Port Codes: CRD = Crescent City Docks FLD = Fields CRL = Crescent City Launch SHC = Shelts CRC = Crescent City (PC, COM) FTB = Fort 8 TRD = Trinitad docks BOD = Bode		ter Cove Bragg	SAU = Sausalito RCH = Richmond BER = Berkeley EME = Emeryville	SCR = Santa C MOS = Moss Li MOH = Montere MON = Montere	anding by harbor (PR)		umeric County- for any port code				
TRH = Trinidad hoist DOR = Dorar EUR = Eureka LMD = Loch I					SNF = San Francisco PRI = Princeton	MOC = Montere MOR = Morro E AVI = Avila		Sample Mode Codes: PR1, PR2, PC, COM, MM, B			





Inventory Tags

Inventory tags are used for labeling bags of heads to allow for tracking as they make their way to the Santa Rosa Office. Each large bag of heads must be inventoried. Record your name, date and headtag series contained in the bag on the Inventory Tag and attach this tag to each large bag prior to storage and delivery to the head drop off location. Multiple

	4/17/16	
Name:	Series in Bag:	
Wong :	18702-78712	
Mattingly	83300-83304	

samplers can put their heads in one bag, but it needs to be clearly marked

which heads were collected by each sampler. Each bag's inventory will be confirmed by OSP staff and compared to headtag numbers recorded on sample forms. Using your headtags in consecutive order makes headtag tracking easier.

Information Request Cards

Information request cards are givento salmon anglers who are interested in learning about their fish. After the salmon heads are processed for the season, OSP will send anglers who requested it, information about their salmon such as: brood year, run, stock, hatchery where it was released, release date, and more. The cards are 3x5 cardstock and usually a bright color. The middle of



the card has a space where the sampler writes in the headtag number of the particular tagged fish the angler wants to know about. Multiple headtags listed for the same angler are okay. Information request cards can act as a positive outreach tool for salmon anglers, so Samplers are encouraged to hand out information request cards to anglers with tagged salmon.

Courtesy Headtags

If an angler approaches you with a tagged salmon from <u>outside your sample</u>, you may collect the head and assign it a courtesy headtag. Attach a courtesy headtag to the salmon and process the head as usual. Fill out an information request card and hand it to the angler and remind them to follow the instructions on the card so they can receive the information at the end of the year.

Important Salmon Goals to Remember

- Every boat needs to be checked for salmon effort, catch, and adipose finclipped fish.
- Each boat with salmon effort (or incidental salmon catch that they kept) should be noted as "a salmon boat". Determine if any
 - salmon were released and identify each salmon kept or released to species.
- All salmon must be counted and observed for the presence of an adipose fin. All heads from adipose fin-clipped fish must be retrieved.
- The heads should be frozen as soon as possible and delivered to the appropriate storage facility.



Salmon Head Drop Off Protocol

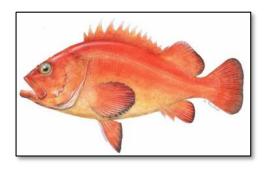
Salmon heads collected by field staff should be taken to one of the drop-off locations listed below (Listed north to south). Contact the office prior to head delivery to confirm office hours. For other arrangements, contact your Lead who will coordinate a meeting time and place to drop the heads off.

Ports	Salmon Head Drop Off Location	Contact Name and Phone
Crescent City, Trinidad, Eureka, Shelter Cove	CDFW – Eureka 619 2 nd Street Eureka, CA 95501	Ed Roberts (707) 441-5757
Fort Bragg	CDFW – Fort Bragg 32330 N. Harbor Way Fort Bragg, CA 95437	Ed Roberts (707) 441-5757
Bodega Bay, Sausalito	CDFW – Santa Rosa 5355 Skylane Blvd, Suite B Santa Rosa, CA 95403	James Phillips (707) 576-2899
Berkeley, Emeryville	Berkeley Marina 201 University Ave. Dock K-900, Men's restroom closet	James Phillips (707) 576-2899
San Francisco, Princeton	CDFW – Belmont 350 Harbor Blvd. Belmont, CA 94002	James Phillips (707) 576-2899
Santa Cruz	Long Marine Lab 1451 Shaffer Rd Santa Cruz, CA 95060	Jayna DaSilva (831) 649-7196
Moss Landing	Moss Landing Marine Labs, Corp Yard 8272 Moss Landing Rd. Moss Landing, CA 95039	Jayna DaSilva (831) 649-7196
Monterey	CDFW – Monterey 20 Lower Ragsdale Dr. Monterey, CA 93940	Jayna DaSilva (831) 649-7196
Morro Bay, Avila Harbor	CDFW – San Luis Obispo 3196 S. Higuera St., Suite A San Luis Obispo, CA 93401	Jayna DaSilva (831) 649-7196
Santa Barbara, Oxnard, Ventura	CDFW – Santa Barbara 1933 Cliff Dr. #9 Santa Barbara, CA 93109	Tamarind Harman (805) 564-1471

Yelloweye Rockfish Sampling

Yelloweye Rockfish have been a prohibited species in the recreational fishery since 2003. As a result, fewer Yelloweye Rockfish data have been available for stock assessments compared to other allowed species. With limited opportunities to encounter them, sampling priority is to collect the length,

weight, and catch location/depth data on all Yelloweye Rockfish (and other species of concem). Further, it is vital to get biological samples if the angler is agreeable to relinquishing the entire fish to you. Attempt to collect the whole carcass of all landed dead yelloweye rockfish to minimize the potential loss or cutting damage to



otoliths. The whole fish is preferred as it also provides sex information that is otherwise unknown. If the fish you encounter is still alive, encourage the angler to release it. If the angler is unwilling or uneasy with giving you the whole fish, ask to collect the head, or the filleted carcass. You do not have any legal authority to require anglers to provide you with Yelloweye Rockfish specimens or to allow you to cut the head off their fish; angler cooperation is strictly voluntary. Samplers must ask permission to first collect the carcass or as a last resort, the head. Do not collect heads or carcasses of Yelloweye Rockfish while on a CPFV.

The Groundfish Project removes otoliths for aging studies and collects sex information in the lab. The Yelloweye Rockfish stock assessment includes data from California, Oregon, and Washington. Yelloweye Rockfish growth rates may be higher in California waters than in cooler northern waters. It is important to capture both temporal and spatial changes in growth rates in order to effectively estimate the productivity of the stock. In prior years, biological samples of Yelloweye Rockfish from California have been limited but have recently increased, in part, because of CRFS collection efforts. The data and specimens CRFS Samplers are able to collect will improve the accuracy of growth curves and reduce uncertainty in future stock assessment modeling.

Be sensitive to the fact that retention of Yelloweye Rockfish is prohibited in California and the angler may be worried that you are collecting evidence against them. Let them know that while it is their responsibility to know the species of fish they are catching, identification of rockfish can be difficult. Stress that the Yelloweye Rockfish collection is for biological purposes only and CRFS is not affiliated with CDFW Wildlife Officers. The data you are collecting is confidential and protected under the Privacy Act. The angler cannot avoid a citation by allowing you to sample and/or collect the fish head.

If a Wildlife Officer is present and intends to confiscate the RFYEY from the angler, ask them to let you weigh, measure and tag the fish. After your interview with angler is complete, ask the Wildlife Officer to consider giving the fish to your Lead after the case against the angler is adjudicated. Explain why CRFS is collecting RFYEY and the importance of each fish to the officer, if necessary. Notify your Lead of the encounter and provide the name of the Wildlife Officer so that your Lead may follow up with him/her. If an enforcement officer is present, follow the guidelines in this manual for working with enforcement (Roles and Responsibilities Section).

Collection Priorities

During the salmon season do not miss boats to collect specimens, biological and location/depth data from Yelloweye Rockfish. Salmon are the priority. Remember that location and depth data are very important. Southem California Samplers should note fish caught in Mexican waters. Please note descending device usage for released Yelloweye.

Biological data priorities for yelloweye rockfish are as follows:

- 1. Length
- 2. Weight
- Whole fish (for otoliths and sex), or carcass or head. Only collect specimens that are landed dead, and NOT from onboard a CPFV.

Equipment

- 1. Knife and sheath
- 2. Cutting board
- Clear bag for storage (do not use opaque bags because they can easily be confused with trash)
- Rockfish headtags. Each Sampler will be provided with numbered headtags labeled "ROCKFISH HEAD TAG". Only use these headtags for Yelloweye Rockfish. Use the tags in order.

Procedures for Collection

- Measure the fork length and record on the data sheet and headtag.
- 2. Weigh the fish and record on the data sheet and headtag.
- 3. Inform the angler of their possession of an
 - illegal Yelloweye Rockfish and ask permission to collect the whole fish or to take the head if they refuse the whole fish.
- Inform the angler of the regulations and that it is





- illegal for them to retain yelloweye rockfish.
- 5. Show the angler how to identify a Yelloweye Rockfish
- 6. Explain to the angler that the Department can learn more about Yelloweye Rockfish populations if they would allow you to examine the fish and take the whole fish or head. Tell the angler that the head contains ear bones (otoliths) that we can use to determine the fish's age. Knowing the age of the fish will help us learn how fast Yelloweye Rockfish grow.
- 7. Fill out the headtag completely before attempting to attach it to the jaw of the yelloweye rockfish.
- 8. Tag the fish if at all possible. Even if they deny you the fish they may change their mind after it's photographed and/or filleted. Firmly attach the tag to the lower jaw of the fish.
- 9. Write the tag number on the data sheet to the right of the length measurement and circle the number.
- 10. If the angler will not give up the whole fish but will give you the carcass or the head after filleting, let them do so. They must use their own knife. Since it is somewhat difficult to fillet the fish without the head attached for leverage, the angler may wish to fillet the fish onsite and bring you the carcass or head. This is okay; tag the fish before the angler leaves to fillet the fish
- 11. Take the fish or remove the head. To remove the head:
 - Make sure to measure and tag the fish BEFORE removing the head.
 - Lay the fish with the head on the cutting board portion of the measuring board.
 - Slide your knife under the gill plate and cut straight down so that it clears about two inches behind the eyes.
 - Flip the fish over to the other side and repeat the cut until it
 - meets the end of the first cut. You may have to angle the knife perpendicular to the ground to meet the other cut.
 - Once the two cuts have met the head should come off. You
 may need to clean up around the gill arches to completely
 separate the head. Make sure the cut exposes the least amount
 of meat possible and remove any gills or extra flesh attached
 to the head. Keep your board and knife clean as you are dealing
 with fish that someone will be eating.
 - Place the head in a bag with the tag number visible from the outside for easy identification.
- Place the head or carcass in a clear bag. Store in a cool place and freeze as soon as





possible. Inform your Lead about collecting a Yelloweye Rockfish by writing a note on the ASF and in the Weekly Report.

- In the Weekly Report, enter the number of RFYEY kept and released for that particular assignment.
- In the far right of the Weekly Report, enter the species code for Yelloweye Rockfish (RFYEY), the number of heads collected, and the tag #s used (see example below).
 If no tags are used enter zero (0).
- Add a note on the Weekly Report email narrative whenyou deliver tagged Yelloweye Rockfish specimens to a "designated drop-off location".
- 13. Deliver the yelloweye rockfish specimen to the nearest designated CDFW office: Eureka, Fort Bragg, Santa Rosa, Belmont, Monterey, San Luis Obispo, Santa Barbara, Los Alamitos, or San Diego. Do not put salmon heads and Yelloweye Rockfish specimens in the same bag.

PR Form Example

		С	ATCH			BIO DATA						
	KEPT	RELS		SPECIES LOC	DEPTH	Fork length / carapace size (mm), sex (MF/T)						
SPECIES	obs *	alive *	(w/DD)	or effort loc if no catch	Average	Weight (decimal kg) or tag #						
CODE *	unobs *	dead * seal take		Block-box; Lat / Lon	Bottom (ft)	1	2	3	4	1 5		
RFYEY	obs 1	0	(0)	539-20	240	435	00055					
RETET	unobs O	dead 0	seal			1.90						

White Seabass Sampling

In Southern (and sometimes Central) California, CRFS samples White Seabass for the presence of a coded wire tag (CWT). The Ocean Resources Enhancement and Hatchery Program (OREHP), a Department sponsored program, raises White Seabass and releases juvenile fish into the wild. Prior

to release, each fish is tagged with a small (1.1 mm long by 0.25 mm diameter) CWT at the posterior edge of the left eye. The tag is not visible, and White Seabass do not possess an adipose fin that can be



removed like salmon to indicate the presence of the CWT. Instead, Samplers are to use a special metal-detecting scanner "wand" to scan each kept fish. Do not scan fish that the angler intends to release.

The purpose of the OREHP is to investigate the feasibility of enhancing marine fish species whose populations have substantially decreased over time through the introduction of hatchery produced fish. Legislation created

the Ocean Enhancement Stamp to fund this program in 1983. This stamp is required by all recreational anglers fishing south of Point Arguello. White Seabass was chosen because of the large decline in catch between the 1950s and 1980s, with annual sport fishing returns in California dropping from over 55,000 fish to less than 3,500 fish during this period. Since 2001, the OREHP has released over 100,000 juvenile White Seabass (8–12 in TL) annually into the waters off southern California. Prior to 2001, releases averaged 25,000 annually. Many of these fish have reached legal size (28 in. or 711 mm TL) and are now able to be caught by the recreational fishery. To assess the feasibility of using hatchery fish to enhance marine fish populations, it is critical to scan and recover tagged adult White Seabass.

Unlike salmon, you do not have legal authority to take the head. If an angler does not want to give up their fish head, inform them of the importance of the OREHP by giving them a flyer. If it's an issue of wanting the otoliths (ear bones), the Department can provide them with a replacement set. Be sure to write down the angler's name and address so that we can send them a replacement set of otoliths.

White Seabass Equipment

- Hand-held scanner with holster (To ensure that the hand-held scanner is not lost or stolen, we require that Samplers wear a belt with the hand-held scanner in its holster hanging from the belt)
- 2. Hand tally counter (The hand tally counter must be attached to the scanner's strap)
- 3. Knife and sheath
- 4. Large zipper bags to store heads
- 5. Ice chest with blue ice (when available)
- 6. CRFS White Seabass Head Collection Tags

Procedure for Sampling and Scanning

- 1. Measure the fork length and record.
- 2. Weigh the fish (be aware that some fish are quite large and may be too big for your scale or your physical ability to lift it)
- 3. Use the hand tally counter to keep track of the number of White Seabass scanned. This will give you the WSB Scan# during the assignment starting with '01'.
- 4. Before scanning, ask the angler if they left a hook in the mouth of the fish. The wand is very sensitive and the presence of a hook can cause a false positive reading. If there is a hook, try to remove it. If you cannot remove the hook, ask the angler if you may collect the head. The OREHP can remove the hook and rescan the head in their lab.
- 5. Turn on the hand-held scanner and check to see that it is working properly by passing it over the block of wood with metal in it which comes in the scanner carrier. If the wood block is unavailable, a piece of metal will work too. You should hear a beep to indicate it is working. If the scanner is not working, please notify your Lead

immediately to either replace the batteries or have the wand repaired.

- 6. Hold the fish up in front of you and away from any metal (e.g. jewelry, watch, measuring board, nails in the dock, coins).
- 7. Rub the scanner over the left side of the fish's head, focusing on the area under the eye and the cheek muscle.
- If no beep is emitted turn the fish over and scan the other side of the head. If no beep, code as a negative scan status on the data sheet
- If the scanner beeps, indicating the presence of a CWT, inform the angler that you would like to remove the head because it is a tagged hatchery White Seabass. Code as an H status if there is a positive scan and you collect the head. If you cannot get the head, code the status as P.
- 10. Record the scan number and status code on your datasheet. Use the area to the right of the weight field to code the scan number and status. You may need to skip a bio data column in order to allow room for lengths, weights, and scan codes for multiple fish from one boat.

The code is a three-digit sequence where the first 2 digits are the number of fish scanned on that assignment (01, 02, 03, etc.) and the third digit is a scan status alpha code (see below). If the fish is not scanned, omit the scan number and status code.

Scan Status Alpha Codes

H = positive scan, head taken by Sampler

N = negative scan

P = positive scan, no head taken

PR Form Example

1 10 1 011111									
		CATCH	ı					BIO DATA	
	KEPT	REI	_S	SPECIES LOC	DEPTH	Fork	length / car	apace size (n	nm), sex (M/F/T)
SPECIES	obs	alive total (w/DD)		or effort loc if no catch	Average			head was	tag#
code	unobs	dead	seal take	Block-box; Lat / Lon	Bottom (ft)	1	scan	ned, negative	4 5
SBWHT	^{вы} 2	0 ()		719-96-75	70	749	806		Fish head was not scanned
SBWHI	unabr doad soal		seal			01N K	02N	/	
SBWHT	_{вы}	alive O		h scan #03 scanned		926	1028	877/	Fish scan #04 scanned positive but the head was
SEWHI	0 unaps	0		sitive and the head v llected (H)	ras	03H	04P ←	K	not collected (P)

Q. What if the angler refuses to relinquish the head?

A. Inform the angler about the importance of coded-wire tags to White Seabass management. If they still refuse, record the length and note on the form that the head was scanned but not recovered using the count and the scan status code 'P'.

Removing the Head

- 1. Lay the fish on a flat surface
- 2. Slide the knife under the gill plate and cut forward or at a 45-degree angle until the cut is approximately one inch behind the eyes
- 3. Flip the fish over to the other side and repeat the cut until it meets the end of the first cut. You may have to angle the knife

perpendicular to the ground to meet the other cut

 Once the two cuts have met the head should come off cleanly. Make sure the cut exposes the least amount of meat possible



and remove any gills or extra flesh attached to the head. Please keep your knife and board clean as you are dealing with fish that someone will be eating. If an angler prefers to cut the head off themselves, let them do it with their own knife

- Fill out a White Seabass headtag form. Write the assignment ID in the ASSN ID fields and write the 2-digit numerical fish number of the scanned fish
- Place the completed headtag form in a clear zipper bag with the head
- 7. Keep the head in a cool place and freeze it as soon as possible

White Seabass Head Drop Off Protocol

If at all possible, you should drop off the White Seabass head(s) at the end of each day at one of the locations listed below. You can also call (877) 728-3972 to find the nearest location to drop off a White Seabass head. If you cannot drop off the head that day, freeze the head until you can drop it off. Contact the business prior to delivery to confirm office hours.

Ports	White Seabass Head Drop Off Location	Phone
Monterey, Moss Landing, and Santa Cruz	CDFW – Monterey 20 Lower Ragsdale Drive, Suite 100 Monterey CA 93940	(831) 649-7196
Morro Bay, Avila Harbor	CDFW – San Luis Obispo 3196 S. Higuera St., Suite A San Luis Obispo, CA 93401	(831) 649-7196
Santa Barbara	CDFW – Santa Barbara 1233 Cliff Drive, Suite 9	(805) 568-1221
	Sea Landing	(805) 963-3564

Ports	White Seabass Head Drop Off Location	Phone
	301 W. Cabrillo Blvd.	
Ventura	Eric's Tackle 2127 E. Thompson, Ventura	(805) 648-5665
Oxnard	Channel Islands Sportfishing Center 3900 Pelican Way, Oxnard	(805) 985-8511
Marina Del Rey	Marina Del Rey Sportfishing 13795 Fiji Way	(310) 822-3625
Redondo Beach	Redondo Beach Boat Hoist 181 N. Harbor Drive	(310) 374-3481
San Pedro	22 nd Street Landing 141 W 22 nd Street	(310) 832-8304
Sair Feuro	LA Harbor Sportfishing Ports 'O Call Village	(310) 547-9916
Long Beach	Long Beach Sportfishing 555 Pico Ave., Berth 55	(562) 432-8993
Long Beach	Pierpoint Landing 200 Aquarium Way	(562) 983-9300
	Avalon Seafood At the end of the green pier	(310) 510-0197
Catalina Island	Two Harbors Harbor Patrol Office On the Pier	(310) 510-4211
Los Alamitos	CDFW – Los Alamitos 4665 Lampson Ave., Suite C	(562) 342-7111
LOS AIAMITOS	Huntington Harbor Fuel Dock Mariner's Point	(562) 592-4975
Huntington Beach	Pacific Edge Bait and Tackle 5042 Edinger Ave.	(714) 840-4262
	Mako Matt's Marine 6411 Edinger Ave.	(714) 893-7743
	Angler's Center 419 Old Newport Rd.	(949) 642-6662
	Balboa Angling Club	(949) 673-6316
Newport Beach/Irvine	Davey's Locker 400 Main Street	(949) 673-1434
	Newport Landing Sportfishing 309 Palm Street	(949) 675-0550
Dana Point	Dana Wharf Sportfishing 34675 Golden Lantern Street	(949) 496-5794

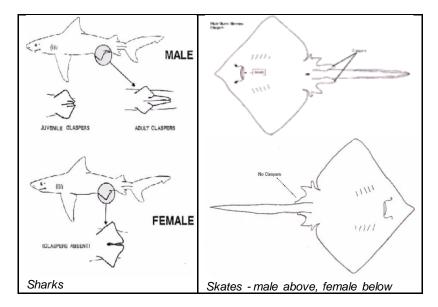
Ports	White Seabass Head Drop Off Location	Phone
	Hogan's Bait and Tackle 34320 Pacific Coast Hwy #G	(949) 493-3528
	Jig Stop Tackle and Tours Dana Point	(949) 496-3555
	Helgren's Sportfishing 315 Harbor Drive South	(760) 722-2133
Oceanside/Carlsbad	Leon Raymond Hubbard Jr. Hatchery 4200 Garfield Street	(760) 434-9501
Solana Beach	Blue Water Tackle 124 Lomas Fe Drive #207	(858) 350-8505
	Dana Landing Mission Bay	(619) 226-2929
San Diego	Hubbs-Sea World Research Institute 2595 Ingraham Street	(619) 227-3870
	CDFW – San Diego 3883 Ruffin Road	(858) 467-4201

Sexing Certain Species of Finfish

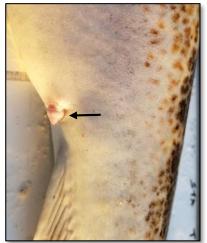
The sex of fishes should be recorded on the data form whenever possible. This information should be considered a bonus and should in no way interfere with your ability to get length and weight data. The codes for fish sex are: M=Male, F=Female, T=Transitional. Transitional California Sheephead may be coded with 'T'.

Some species of fish can be sexed using external characteristics; for other species, sex may be determined when the fish is being filleted (for party or charter boat mode which requires dissection of the gut), or by using season-specific external characteristics. If a fish is releasing live young or eggs, it's a female; the presence of white milt indicates that it's a male.

The sex of **elasmobranchs** can always be determined from external characters because male fish have a pair of mixopterygia (intromittent organs, claspers) which are visible from an early stage of development on the inside edge of the pelvic fins (see below). The females do not have mixopterygia.



Adult **Lingcod** can be sexed externally. View the ventral side of the fish near the posterior end. Males have a distinct papilla next to the anus. Females do not. If necessary, press around to reveal the papilla. Sometimes the male papilla does not protrude from the abdomen and will initially appear flush. See below, male Lingcod (pictured left) with papilla protrusion and female Lingcod (pictured right) without a visible papilla. Note, color is not indicative of sex.





Lingcod sexing; female

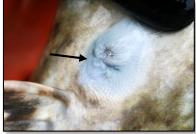
Cabezon may also be sexed in a similar manner. Unlike Lingcod, male and female Cabezon both have papillae. The papillae on male and female Cabezon differ in shape. It is necessary to press around the papilla with your thumb to reveal features of its shape. The male papilla has ridges on its edges and protrudes slightly from the abdomen, resembling a cruller donut. The female papilla is conical in shape and has smooth edges and is surrounded by folds of skin, resembling a cinder cone (see below). Note, color is not indicative of sex.





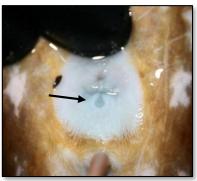
Cabezon sexing; male papilla resembles a cruller donut





Cabezon sexing; female papilla resembles a cinder cone





Cabezon sexing; male

Cabezon sexing; female

California Sheephead can be sexed externally by color. Sheephead are protogynous hermaphrodites, meaning they are born female and become male later in their development. They change color as they age and change from female to male. There are four life stages: juvenile, female, transitional and male. <u>Juveniles</u> are bright orange-red with black spots on the fins and caudal peduncle. They frequently have a white strip along their sides from head to tail.



Figure 12 - Juvenile Sheephead



Figure 13 - Female Sheephead



Figure 14 – Transitional Sheephead

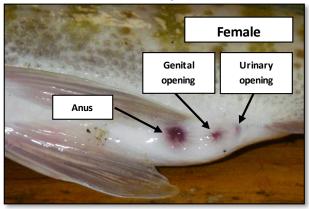


Figure 15 - Male Sheephead

We code the females, transitionals and males. <u>Females</u> are a faded rose to brownish red with a white chin. <u>Transitionals</u> are a dusky rose to a deeper reddish-orange with darkening of the anterior and posterior thirds of the body. Those areas may appear light brownish or grayish in color. The chin remains white. <u>Male</u> fish are dark brown or black on the first and last third. The central third is a deep orange to red. The chin is white.

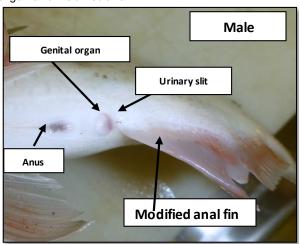
Surfperch of the subfamily Amphistichinae which includes species such as barred, Redtail, Silver, Walleye, and Calico Surfperch, can be sexed externally by noting the number of openings between the anal and pelvic fins: males have two while females have three. To clarify, males have three openings; however, the genital and urinary openings appear as slits and the genital opening is obscured. Only two openings are visible on males. Note: other species of surfperch may be too difficult to sex.

Ventral view: female Barred Surfperch.

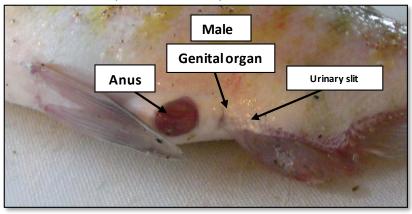


Pictured above and below are ventral views of a female Barred, male Redtail, and male Barred Surfperch, respectively. The anal, genital, and urinary openings appear as purplish "spots" in the female Barred Surfperch shown above. Male surfperch possess a genital organ and modified anal fin ray as shown below.

Ventral view: ripening male Redtail Surfperch, exhibiting bulbous genital organ and modified anal fin.

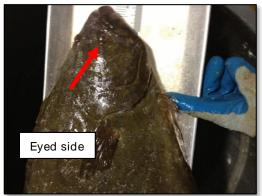


Ventral view: a non-ripe male Barred Surfperch

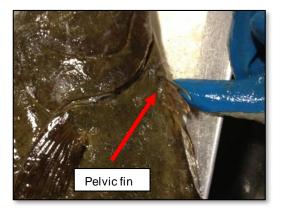


Note: male Barred Surfperch taken during the non-mating season may have genital organs that are not bulbous as shown above.

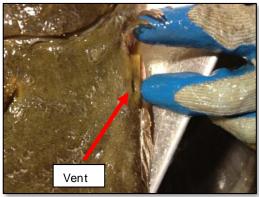
California Halibut can be sexed by squeezing the abdominal cavity to extrude sex products. Males will release milt 100% of the time; if no milt is released, the halibut is female.



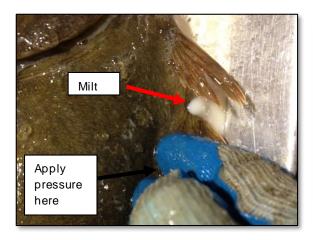
Step 1: Position the halibut so that the eyed side or pigmented side is facing upwards



Step 2: Locate the pelvic fin, which is just posterior to the gill cover and below the pectoral fin



Step 3: Flip the pelvic fin back towards the head and locate the vent underneath where sexual products are extruded



Step 4: Apply pressure to the organ cavity just posterior to the vent. If the halibut is male, milt will be released, as shown left. If no milt is extruded the halibut is female

Step 5: Both sexes may release a clear fluid prior to sexual products. It is important to note that both sexes may also release contents from other organs prior to sexual products. These contents may appear off-white in color and chunky (see photo). This halibut was not male. Milt is pearly white, opaque, and never chunky. If you see this, keep applying pressure. If milt is released after this then the halibut is male: if no milt then female.



Kelp Greenling are sexually dimorphic – the basic male pattern is brownorange and red-purple with bright blue spots. These blue spots are often surrounded by smaller black spots. Colors can change during the reproductive season – courting males are often uniform gray with blue spots. The rule of thumb is if it has blue spots it is male. Females tend to be gray or brown with brown or yellow spots.

Male Kelp Greenling



Female Kelp Greenling



Handling Fish

California Scorpionfish

This pretty fish has a serious toxin in its dorsal, anal and pelvic fin spines. At

the minimum, a poke from a spine is very painful, but it can also be life-threatening for some people. These fish should only be handled with the utmost care. Pliers are good to use rather than hands so that there is minimal chance of being stuck by one of the spines. On many PC boats, the deckhand will break off the spines with pliers while holding the fish



over the side before bringing it aboard. Do not be deceived; small specimens can be just as dangerous.

Remedies:

- 1) For a serious situation, get to the nearest emergency room because anaphylactic shock can occur from the toxin.
- 2) For a minor situation, soak the injured body part in water that is as hot as can be tolerated (the hotter, the better) or apply meat tenderizer (not "Accent," which is only a flavoring). Tenderizers that contain papaya enzyme are good because the toxin is a protein, and papaya enzymes (and other tenderizers) break down protein.

Rockfish (Sebastes spp.)

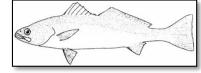
Most, if not all, 67 species of rockfish have some toxin in their spines, so use care when handling them. If a spine breaks your skin and the pain is more than minor, damp heat or meat tenderizer will usually take care of the problem. While rockfish are not nearly as dangerous as California scorpionfish, you should watch for reactions, especially if there



are subsequent injuries because people can develop a reaction to the rockfish toxin if they are injured a number of times.

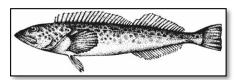
White Seabass

This fish has many sharp teeth, so steer clear of the mouth when handling.



Lingcod

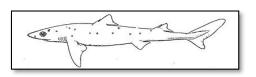
Lingcod have large, sharp teeth and sharp gill rakers. NEVER pick up this fish by inserting your hand under the gill cover. Instead, pick up the fish by inserting the thumb and forefinger of one hand into the eye sockets and use the other hand to lift the fish by the tail.



Sharks can be dangerous, even when they appear to be dead. There are many reports of anglers being bitten by a shark lying on the deck that was thought to have been dead for hours. Use caution when measuring these fish.

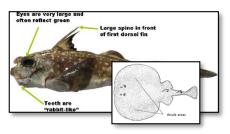
Spiny Dogfish

Dogfish should be handled with care, as the large spines at the leading edge of each dorsal fin are venomous and can inflict painful wounds.



Ratfish

Ratfish are rarely seen by Samplers because they are caught in deep water and most people who catch them throw them back. If you should need to handle a specimen, use care to avoid the very large, venomous spine in front of the dorsal fin. The



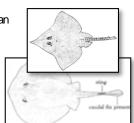
toxin is a protein, so heat or meat tenderizer can probably be used to relieve the pain.

Skates and Rays

Electric rays can be dangerous. Do NOT touch the disk part of this fish! The name is self-explanatory. While you won't suffer permanent damage, the shock can be very strong and painful.

Skates have sharp scapular and tail spines that can be painful.

Stingrays and Bat Rays have a venomous stinger at the base of the tail which can inflict a painful wound. Again, heat or meat tenderizer may minimize the pain.



Invertebrate Sampling

The primary goal of CRFS is to collect data on finfish trips. In general, finfish sampling has a higher priority than invertebrate sampling. A Sampler should never miss finfish boats or anglers to obtain more than the minimum data needed for a complete interview from invertebrate-only anglers. Observation of invertebrate catch for species composition and enumeration is not necessary. Angler reported kept and released is all that is required for invertebrate catch and effort.

Invertebrate anglers should be screened for finfish catch. If the angling party has only targeted invertebrates but incidentally caught finfish you will code the second target as UNIFH along with recording catch.

BB invertebrate-only anglers should not be interviewed along with MM invertebrate-only anglers after the stop count. Invertebrate-only anglers intercepted during the PR surveys should be interviewed to obtain the minimum data needed for a complete CRFS sample. Observation of invertebrate catch is not necessary. Eligible anglers/boats targeting invertebrates will get their own CRFS sample number. Invertebrate-only anglers will not be included in any start, stop, or instantaneous angler counts while sampling in shore modes; invertebrate-only boats/trailers will be included in all counts while sampling in PR modes – make notes regarding known invertebrate-only boats included in counts. Code boats that are only setting invertebrate gear, not pulling gear/checking pots, as NFOTH when they return to the ramp.

In order to avoid missing finfish effort, Samplers may save time while sampling invertebrate-only boats/anglers by collecting only the minimum data elements required for a complete CRFS interview.

Note: conduct complete interviews whenever possible; this interview of minimum data elements should be completed only when pressed for time to avoid missing finfish effort.

The target boxes on all CRFS forms are primarily for finfish. If the targets are invertebrates and finfish, record the targets in the order that they are given in the CRFS interview (e.g., if an invertebrate is the primary target, record the appropriate invertebrate code in the primary target space and the finfish target in the secondary space). If an angler only targeted invertebrates then, as with finfish, only the primary space would need to be coded.

The "Area fished" for invertebrates is the same as for finfish. Use the proper codes for nearshore, bay, offshore, etc. Gear for invertebrates is somewhat different though. In addition to Hook-and-line (H) gear that may be used to take invertebrates, there are special invertebrate-only gears (see Gen. Onsite Procedures). The number of pots/nets employed to catch invertebrates must be recorded. Be aware that anglers may employ two different kinds of gear at the same time, so both the primary and secondary

targets will be filled out and the gear codes will reflect the two separate gears used.

Even if invertebrate pots were left out to soak overnight, or for many days, anglers may only harvest one day's worth of limits on any one day that they check their gear. In most cases the "Days Fished" should always be "1" except in the case of boat anglers who have completed and filed a Declaration for a Multi-Day Fishing Trip (see Section 27.15 in the Ocean Sport Fishing Regulations booklet) with the Department. Although this occurrence is rare, multi-day fishing trips do occur in southern California, typically targeting lobster.

The avidity question should be delivered the same way, no matter what the anglers were targeting. Even if the angler you are interviewing was targeting invertebrates only, you would still ask for how many days they fished for finfish in a 12-month period in California marine waters. Make sure they are not counting invertebrate-only trips in their estimate of avidity.

Squid Sampling



CRFS Samplers should collect catch and effort data for market squid (SQDMK), or Humboldt/jumbo squid (SQDJU). With market squid, make sure to ask the angler if their squid bait was caught on the trip or bought at the store (do not include store-bought squid in the catch data).

Collect the minimum data elements required for a good interview from a squid-only boat. Code squid-only boats that you are not able to sample at all, or are

unable to collect the minimum data elements from, as missed boats, just like finfish boats. Make a note on your form as to the activity of the boat.

Abalone Sampling

The vast majority of abalone effort is contained in the BB mode. Do not collect catch and effort data from abalone-only anglers in BB mode. Abalone anglers should be screened for effort as it is common for them to have a secondary target of finfish using the spear gear type. For those dual target trips, the goal would be to observe finfish catch and collect all catch and effort data for finfish prior to collecting abalone



catch data. Angler reported catch for abalone meets CRFS needs and therefore it is not necessary to observe. Only catch that has been incidentally observed while collecting finfish data should be recorded as kept observed.

Lobster Sampling

The code for California Spiny Lobster is LOBSP. The following target codes apply to the take of spiny lobster:

F# = flat hoop net,

R# = rigid, or "modified" hoop net

= number of hoop nets used

C = hand while S**C**UBA diving with tanks

D = free **D**iving with no use of air tanks.



If hoop nets are used (gear codes F or R) then the <u>number of nets</u> employed will be recorded as well. The number of nets used follows the gear code (e.g., 3 flat hoop nets = F3).

Crab Sampling

This sampling includes Dungeness Crab (*Metacarcinus magister*) Red Rock Crab (CRBRR), Brown Rock Crab (CRBBR), Yellow Rock Crab (CRBYR), Slender or Graceful Rock Crab (CRBGR), and the general Cancer genus (CRBGN). Sheep Crabs or Kelp Crabs may be coded as true crabs (CRABS). For crab identification, please see:

https://www.wildlife.ca.gov/Fishing/Ocean/Dungeness-Crab http://www.dfw.state.or.us/mrp/shellfish/crab/Crab ID.asp

Handling Crab

In most cases you will not have to handle crabs except to remove finfish from a mixed catch within the same cooler or bag. Always wear gloves when handling crabs. Keep your fingers away from the chelipeds (claws). Crabs are usually not landed dead and can be quite lively. Take care to not have



any part of your hand near the claws of any crabs in proximity, especially those beneath the crab you're grabbing! The crab will pinch you if possible. The best way to handle a crab is to grab the last (posterior most) set of legs at the point nearest to carapace and squeeze them together. This will give you a 'lever' to hold the crab. Do not squeeze too tightly or the legs

may detach. If you do get pinched, break the claw off from the body first to release the claw. Do not pull your finger (or other body part) out of the claw because this will result in lacerations.

Invertebrate Sampling Protocol FAQ

General Protocol

1. How do I word the avidity question for invertebrate-only anglers?

Word it the same as usual, "Not counting today, how many days have you gone saltwater sport fin fish fishing in California in the last 12 months". Be clear with invertebrate-only anglers that you are just asking about fin fish trips.

2. How do I show that the angler or boat fished for invertebrates AND finfish?

Record what the angler says was their primary and secondary target (so they can have a primary target of crab and secondary target of finfish). Each target will have its own area and gear specified.

3. If a boat or angler reports only invertebrate target(s), but they kept or released finfish, should finfish be added as the secondary target?

Yes. If finfish are caught, there needs to be a finfish target recorded. Edit your data as follows:

Primary target = invertebrate Secondary target=

(Write UNIFH as the secondary target or a more specific finfish target the angler gives when probed further)

Primary target = invertebrate Secondary target = invertebrate

(Change secondary target to UNIFH or a more specific finfish target the angler gives when probed further)

4. Should I identify and count invertebrates that an angler or boat has kept?

No. Ask the angler(s) for the number of invertebrates they kept and released by species. Record angler reported kept invertebrate species as kept unobserved. If the invertebrate catch is easily observed while sampling finfish you should record those observed invertebrate as Kept/obs.

5. How do I code gear used for an invertebrate target that does not have a CRFS gear code?

Anglers targeting invertebrates may often use gears for which there is no applicable CRFS code – gears such as slurp guns, clam guns, shovels, rakes, pitchforks or other manual gears, including by hand. This is seen most often with clammers. In these instances, the Sampler will leave the gear field on the data sheet blank, but include a note on the data sheet indicating what the clammers were using to take clams (or other invertebrates).

6. Should I record biological data for invertebrates?

No. Do not measure or determine the sex of any invertebrate catch.

MM and BB Modes

7. Should I count invertebrate-only anglers during a shore assignment?

No. Invertebrate-only anglers are NOT included in start, stop or instantaneous counts for MM mode. Invertebrate-only anglers are NOT included in the BB total estimated finfish anglers counts.

8. Should I interview invertebrate-only anglers during a shore assignment?

MM: Yes, but do not interview incomplete or partial trip invertebrateonly anglers after the stop count and only if invertebrate sampling does not deter from finfish sampling.

BB: No, do not interview invertebrate-only anglers.

PR1 and PR2 Modes

9. If a crab/lobster boat returned from JUST placing or setting pots/hoops, do I interview them?

No, boats that are only placing pots/hoops without finfish effort are not eligible for a CRFS interview.

Code them as a NFOTH boat. You will record the time of the sample, the sample number, and "0" total anglers for this NFOTH boat.

10. If an invertebrate boat just returned from harvesting but has no finfish target, do I interview them?

Yes. Invertebrate-only boats are interviewed for CRFS. Conduct the interview as normal except, we would not observe the catch and instead have the angler report the number of invertebrates kept and released by species. Do not collect biological data (sex & lengths).

11. How do I code days fished for a crab boat that soaked pots for 72 hours?

For crab-only boats' days fished, we just ask for days the boat was out picking up pots. The boat fished for 1 day. Even if the pot(s) had a limit of crab for every day they soaked, the angler would only be able to bring in 1 limit of crab that day. For crab-only boats, code 1 day fishing regardless of how long the pots soaked. Even though the pots soaked overnight, do not check the Night Fishing box.

12. The last finfish boat just returned to the ramp. There are two trailers left in the lot, but I know they were fishing for invertebrates only – no finfish targets – because I spoke with them before they launched. Should I stay on site waiting for them to return?

Do not stay on site waiting for known invert-only boats to return. Do include them in your PR stop count, with a note regarding activity. Do your best to canvass boats as they launch to determine activity/target — this will help you to determine whether you should stay on site. Stay onsite for the prescribed period of time (until sunset for PR1, up to 8 hours for the assignment for PR2) for trailers with unknown activity (trailers that were present on your arrival or trailers for boats that you were not able to canvass).

13. How do I record location and depth for invertebrate boats?

We want location and bottom depth to reflect where the species were caught. Only code location and depth for effort, if the boat had no catch at all.

If the boat caught baitfish and crab, for example, it should have two sets of locations and depths listed, one for the baitfish and one for the crab pots (lined up by catch species, not targets). Remember to re-record the location for finfish if there were crab measured and recorded in between the finfish species.

14. If the anglers had placed 5 pots/hoops but could only find 3 pots/hoops, how many total pots/hoops do I code?

Three pots/hoops (P₃/H₃)

15. If an invertebrate-only boat is launching, how do I code that?

PR1: Launched boats are not tracked in PR1 mode. However, notes on launching boats that are determined to be invertebrate-only during canvassing should be recorded elsewhere. This documentation will clarify to the Lead that the Sampler does not have to remain on site for known invertebrate-only boats at the end of the day.

PR2: We would code invertebrate-only boats that launched under the PR2 launched category. Please make a note that it was invertebrate only. You would not need to wait all day at the PR2 for an invertebrate-only boat.

16. A boat has an ice chest with limits of crab and rockfish for five anglers. I'm pressed for time. What do I do?

CRFS prioritizes finfish data over invertebrate data and we don't collect biological data from invertebrates, so ignore the crab. Be careful sorting the fish from the crab. Once you're done with the rockfish, ask the angler(s) for counts of kept and released crab by species.

17. A boat of four crabbers tells you that in addition to the four limits of Dungeness crab on their boat, they left another 15 legal-size crab in their pots, and they plan on going back out in a day or two to pull them again. How do I code these 15 crabs that are not on the boat?

These crabs will not show up anywhere on your data sheets for this assignment. If you were to count them, say as kept/unobserved, and another randomly-assigned CRFS sampling assignment was conducted when and where these crabbers came back with these crabs, they then would be double-counted.

PCO and PCD Modes

18. Should I record invertebrate catch and bio data for a PC trip?

No, do not record any invertebrate catch or bio data for a PCO or PCD sample. This includes unobserved and observed kept catch, released catch, measurements, and sex. You should only record an invertebrate as a target if it is reported as being a primary or secondary target, along with the water area and gear. Do not sample invertebrate-only PC trips.

THE WEEKLY REPORT

Weekly Reports that summarize catch and effort data for species that require close monitoring are provided to fishery managers to ensure harvest guidelines are not exceeded. District Leads also use Weekly Reports to track Sampler activity and make sure assignments are completed during the sample week. Leads use the Weekly Report to track Sampler hours and check that Samplers' timesheets match what is reported, while the CRFS executive team uses the reports for budgeting purposes. The Weekly Report consists of a spreadsheet with a list of every assignment the Sampler worked or was scheduled to work, quantitative catch and effort data, and a qualitative description of weather, catch and effort for each assignment, and any other work done for CRFS or another CDFW project.

Due Dates

All CRFS Samplers are to submit a Weekly Report to their District Lead and OSP by 8:00 AM each Monday throughout the year. Weekly Reports from Samplers in Districts 1 and 2 may have different deadlines and content requirements given by their Leads. The report covers ALL assignments worked during the previous Monday through Sunday sampling week. Nonsample time for data entry, office work, training, meetings, and travel should also be included in the Weekly Report. The report also includes assignments that were scheduled, but not completed. Sick, vacation, and holiday hours to be claimed on Sampler's timesheets should also be reported in the Weekly Report.

Assignments included in the Weekly Report:

MODE code	Mode Description
PR1	primary private rentals (skiffs)
PR2	secondary private rentals (skiffs)
PCO	PC-CPFV onboard sampling
PCS	PC-CPFV salmon dockside sampling
PCD	PC-CPFV non-salmon dockside sampling
PEC	PC-CPFV effort checks
MM	man-made structures
BB	beach-bank
Non-Sampling	Mode Codes
DAT	data-entry work in CDFW office
SEC	site effort checks
SK1	SEC data-entry work in CDFW office
OFC	all other work in CDFW office
TRV	travel time to and from sample site
SIC	sick hours claimed on timesheet
HOL	holiday hours claimed on timesheet
VAC	vacation hours claimed on timesheet
OPW	work for and paid by another CDFW project
TRN	specified training
PDD	professional development day
MTG	staff meeting

Sampling Assignment Summary in the Weekly Report

PR1

For the PR1 mode, the Weekly Report data is transcribed from the footer of the Assignment Summary Form (ASF). The Sampler sums the page totals from all of his/her PR1 forms onto the footer of the ASF. The ASF footer totals align with the fields of the Excel Weekly Report. Remember to include the onsite and/or offsite start count if you are the first Sampler, onsite and/or offsite stop count if you are the last Sampler and the names and numbers of other Samplers you worked with and whether or not they have data. Do not include known non-fishing boats in the stop count.

PR2

For the PR2 mode, the Weekly Report data is transcribed from the footer of the Assignment Summary Form (ASF). The Sampler sums the page totals from all of his/her PR2 forms onto the footer of the ASF. The ASF footer totals align with the fields of the Excel Weekly Report. Remember to include the onsite and/or offsite start count and onsite and/or offsite stop count. Do not include known non-fishing boats in the stop count.

PCO

PCO is for onboard CPFV sampling. These assignments have an ASSN ID. Include the boat name, trip type and target(s) in the notes field; include your plans for rescheduling if you didn't make it out. Fill out the number of headtags used, HALPA, RFYEY, RFCOW, RFCAN and RFBLK kept and released, and other species headtags field (yellow groundfish items). Remember to only include HALPA, RFYEY, RFCOW, RFCAN and RFBLK counts captured on the PC Onboard catch and discard form, transcribed from the "totals boxes" on the footer of the form.

PCS

Salmon dockside assignments do not have an ASSN ID. Fill out the OSP port code and orange salmon items transcribed from the footer of the form. Samplers are to record only data he/she collected; when assisting another Sampler with a PCS sample, record hours worked on the Weekly Report without completing the orange salmon items. A new/separate sheet does not need to be created. Include the names and numbers of other Samplers you worked with and whether or not they have data in the notes field of the spreadsheet.

PCD

Non-salmon dockside assignments have an ASSN ID or are conducted opportunistically. All of these PC assignments should be included in the Weekly Report. Fill out the number of headtags used and yellow groundfish items, transcribed from the "totals boxes' on the footer of the form. Remember to include a row on the weekly even if the PCD was opportunistic.

PCS/PCD Combination Trips

Boats with both salmon and non-salmon targets are frequently sampled concurrently during a sampling assignment. Data from these two different targets are recorded on separate rows on the Weekly Report – one row for the PCS data, and one row for the PCD data.

PEC

Party/Charter effort checks (PECs) do not have an ASSN ID. PECs are important for validating CPFV logbooks and should be reported on the Weekly Report. This includes time spent checking in with the landings in person or by phone, emailing the Port Lead to report CPFV activity, filling out the PEC forms, etc. The mode should be listed as PEC and remember to include the OSP port code.

MM

These are clusters that include man-made structure sites. Sample data fields are for boat modes only – do not fill out any fields between mileage and notes when sampling in shore modes, except for Headtags.

BB

These are clusters that include beaches and bank sites. Sample data fields are for boat modes only – do not fill out any fields between mileage and notes when sampling in shore modes, except for Headtags.

General Instructions

Excel Spreadsheet – Instructions for Completing the Weekly Report
Samplers are to report their activities, and catch and effort data using the
MS Excel spreadsheet template provided by their Lead. At the beginning of
the season, your District Lead will provide you with the MS Excel
spreadsheet template that should not be altered. It is important that the
order of the MS Excel columns be retained so that the data will align and
merge among all Sampler assignments.

Even if you did not work any assignments or claim any hours during the week you are still required to send an email stating, "I did not work and have nothing to report", or something to that effect. You are also required to send any other items that Leads are expecting, for example, if you did not collect any salmon heads that week you are still required to submit an OSP headtag report.

Leads will compare your time sheets with the work reported in your weekly reports. All time claimed on your timesheet must be reflected on a weekly report; total hours claimed on your time sheet must equal total hours claimed in your weekly reports.

Samplers must complete and send the Weekly Report to their Lead and OSP via email every Monday morning no later than 8:00 AM (Weekly

Reports from Samplers in Districts 1 and 2 may have different deadlines and content requirements given by their Leads):

- Save the Excel file to your computer
- 2. Locate the Template tab (orange tab)
- 3. At the top of the Template fill out the green fields with your information.
- 4. Your last name, Sampler number, Monday's date, Lead's last name, and District will autofill in the gray columns.
- 5. Fill out the rest of the spreadsheet with all of your activities for the week (Mon-Sun), including field and office work, training, meeting, and "absent" hours such as vacation, sick, and holiday. Use the SamplerExample (blue tab), Field Name Definitions (green tab), and Mode & Port Codes (yellow tab) as guidance for filling out the report correctly. Non-applicable fields should be left blank. For days with multiple modes sampled, use multiple rows and split up the time accordingly.

Once you have entered all your information for that week:

- 1. Right click on the Tab name (Template)
- 2. Select "Move or Copy"
- 3. Under the "To Book" heading, choose "new book" from the drop down menu
- 4. Check the box that says "Create a Copy"
- 5. Click "OK"
- 6. Save this new Excel file in the appropriate format: "mmddyy WeeklyReport D# LastName"
 - The "mmddyy is the Monday date that began the sampling week
 - b. D# = CRFS District where MOST sampling occurred
 - c. LastName = Sampler's last name
- Email this Excel file with a brief description of the weather, catch and effort for the week in the body of the email to your Lead and OSP.

For example; "Weather was great all week but salmon fishing was poor. A lot of anglers switched to bottomfishing and caught mostly RFBLK but one RFYEY was released. A few small salmon were kept. Worked with James Phillips (302) who also has data – we each used our own headtags". Also include the status of any remaining PR1 trailers.

Some items on the Weekly Report are only applicable to salmon trips; those

column labels on the Template tab are color-coded in orange. These fields include (leave these fields blank for assignments and/or regions that do not apply):

Salmon boats



- Salmon anglers
- Kings kept
- Kings released
- Coho kept
- Coho released
- Number of headtags used
- Commercial pounds sampled (OSP only)

Some items on the Weekly Report are only applicable to groundfish; those column labels on the Template tab are color-coded in yellow. These fields include:

- Pacific halibut kept
- Pacific halibut released
- Yelloweye rockfish kept
- Yelloweye rockfish released
- Cowcod kept
- Cowcod released
- Canary rockfish kept
- Canary rockfish released
- Black rockfish kept
- Black rockfish released
- Species of head collected
- Number of rockfish headtags used
- Rockfish head tag series number(s)
- Other pertinent notes: Comments about descending device usage, depth the fish was taken, associated catch species.



General Guidelines for the Spreadsheet:

- 1. Do not insert or reorder the columns
- 2. Do not leave blank rows between data rows and column headers
- 3. Non-applicable items are left blank
- 4. Applicable counts of zero are not left blank
- Only report data you collected. If another Sampler worked with you and also collected data, he/she should report that data on their own Weekly Report

Weekly Report Item by Item Instructions

FIELD	INSTRUCTIONS	CODES AND FORMATS
Header		
Week	Fill out in header only:	Gray column will autofill when
beginning	Monday's date in which	mode data is added
Monday	sample week began.	



FIELD	INSTRUCTIONS	CODES AND FORMATS
		Example: 7/10/2017
Sampler Last Name	Fill out in header only: Enter your last name.	Gray column will autofill when mode data is added. Example: TROXEL
Sampler Number	Fill out in header only: assigned CRFS or OSP Sampler 3-digit number.	Gray column will autofill when mode data is added) Example: 312
District Number	Enter the CRFS District where MOST of the sampling occurred.	1 = South 2 = Channel 3 = Central 4 = San Francisco 5 = Wine 6 = Redwood
Lead Last Name	Fill out in header only: last name of CRFS or OSP Lead.	Gray column will autofill when mode data is added. Example: ROBERTS
Daily Data		
Assn ID num	Assignment ID number (include leading zero) The assignment ID number should be reported on each line relevant to the assignment, including travel rows.	Leave blank if not applicable.
Date mm/dd	Enter the date that sampling/work occurred. Do not alter the date format. REQUIRED FOR EACH ROW OF DATA.	Example: 7/10
OSP Port, Cluster or Cnty Site	Enter the 3-letter alpha OSP port code for PR1, PCO, PCD, PCS, PEC, and COM assignments. Enter the cluster designation for BB and MM assignments. Enter the county and site codes for PR2 assignments (include leading zeros and NO dash). The sample site OSP port code, cluster, or Cnty Site should be reported on each line relevant to the assignment, including travel rows.	Example: MOS for Moss Landing PR1 site Example: SFO10 for San Francisco MM cluster Example: 023107 for Samoa Bridge Boat Ramp PR2 site
MODE	Enter the appropriate sampling mode for the sampled assignment or code	PR1 = primary private and rental boats

FIELD	INSTRUCTIONS	CODES AND FORMATS
	for other work. Do not leave blank; REQUIRED FOR EACH ROW OF DATA.	PR2 = secondary private and rental boats PC0 = CPFV onboard
	OR	PCS = CPFV salmon dockside PCD = CPFV non-salmon
	Enter the appropriate non- sampling mode code for travel, data entry, office work, trainings, meetings, and other non-field assignments or "absent" hours such as sick, vacation, and holiday. REQUIRED FOR EACH ROW OF DATA.	dockside PCO = PC-CPFV onboard PEC = PC-CPFV effort check MM = man-made structures BB = beach and bank SEC = site effort check routes MTG = staff meeting TRN = specified training OFC = all other office work TRV = travel hours OPW = other project hours SK1 = SEC data entry SK# = SK (non-SEC) data entry DAT = data entry SIC = sick HOL = holiday VAC =
Sample -Time	Enter the 4-digit 24 hour time	vacation ALL Modes.
Start	When sampling started; REQUIRED FOR EACH ROW OF DATA.	Use military time (0000-2400) without colon (:) Use the leading zero for times before
End	When sampling ended; REQUIRED FOR EACH ROW OF DATA.	1000. Example: 0800
Decimal Hours	Do NOT enter in this field. Based on the Start and End Sample Times the total Decimal Hours is automatically calculated for	Decimal Hours column will autofill when Sample Time – Start and End data is added. Example: 7.75
Mileon	that row.	
Mileage	Enter the nearest whole number of miles to/from headquarters to/from sampling site and mileage accrued while traveling between sites in a personal vehicle.	Example: 25 Enter zero "0" if a state car was used; only enter miles that will be claimed for reimbursement. Be sure to deduct commute miles before reporting "net" miles. Miles are reported only on TRV rows

FIELD	INSTRUCTIONS	CODES AND FORMATS
Refu +	Enter the total number of	PR only
Barrier	boats where angler(s)	<black> if non-applicable</black>
	refused to be sampled or a	
	language barrier occurred.	
Total Boats	Enter total number of	PR and PC modes only
	sampled boats (not counting	<black> if non-applicable</black>
	refusals, language barriers	
	or missed boats.	
Salmon	Enter the total number of	PCS, PR and COM only
Boats	salmon boats that were	<black> if non-applicable</black>
	targeting and/or kept	
	salmon.	
Salmon	Enter the total number of	PCS and PR only
Angls	anglers who targeted and/or	<black> if non-applicable</black>
	keptsalmon.	
Kings Kept	Enter the sum of king	PCS, PR and COM only
	salmon kept.	0 = No kings kept
	•	# = Number of kings kept
		<black> if non-applicable</black>
Kings Rels	Enter the sum of king	PCS, PR and COM only
3	salmon released.	0 = No kings released
		# = Number of kings released
		<black> if non-applicable</black>
Coho Kept	Enter the sum of Coho	PCS, PR and COM only
	salmon kept.	0 = No coho kept
		# = Number of coho kept
		<black> if non-applicable</black>
Coho Rels	Enter the sum of Coho	PCS, PR and COM only
	salmon released.	0 = No coho released
		# = Number of coho released
		<black> if non-applicable</black>
Head Tags	Enter the total number of	ALL Sampling Modes
	salmon head tags used	0 = No head tags used
	(includes tags collected in	# = Number of head tags
	shore modes).	used
HALPA	Enter the total number of	PR and PC only
Kept	Pacific Halibut kept.	0 = No HALPA kept
	. asmorianoa Ropu	# = Number kept
HALPA	Enter the total number of	PR and PC only
Rels	Pacific Halibut released.	0 = No HALPA released
1	. asino Hansac Toroacoa.	# = Number released
		<pre> <</pre>
RFYEY	Enter the total number of	PR and PC only
Kept	Yelloweye Rockfish kept.	0 = No RFYEY kept
vehr	renoweye Nockiisii kept.	0 - NO IN TET KEPT

FIELD	INSTRUCTIONS	CODES AND FORMATS
		# = Number kept
RFYEY	Enter the total number of	PR and PC only
Rels	Yelloweye Rockfish	0 = No RFYEY released
	released.	# = Number released
DECOM	Enter the tetal accept on et	
RFCOW	Enter the total number of	PR and PC only
Kept	Cowcod kept.	0 = No RFCOW kept # = Number kept
		<pre># = Number kept <black> if non-applicable</black></pre>
RFCOW	Enter the total number of	PR and PC only
Rels	Cowcod released.	0 = No RFCOW released
17612	Cowcod released.	# = Number released
		<pre> <</br></pre>
RFCAN	Enter the total number of	
Kept	Canary Rockfish kept.	PR and PC only 0 = No RFCAN kept
Kept	Canary Rockiish kept.	# = Number kept
		<pre></pre>
RFCAN	Enter the total number of	PR and PC only
Rels	Canary Rockfish released.	0 = No RFCAN released
11010	Carrary Recikinshi released.	# = Number released
RFBLK	Enter the total number of	PR and PC only
Kept	Black Rockfish kept.	0 = No RFBLK kept
· '	•	# = Number kept
		 diank> if non-applicable
RFBLK	Enter the total number of	PR and PC only
Rels	Black Rockfish released.	0 = No RFBLK released
		# = Number released
		<black> if non-applicable</black>
Missed	Enter the total number of	PR only
Boats	onsite missed boats.	0 = No missed onsite boats
Onsite		# = Number of missed boats
		onsite
		<black> if non-applicable</black>
Missed	Enter the total number of	PR only
Boats	offsite missed boats.	0 = No missed offsite boats
Offsite		# = Number of missed boats
		offsite
		<black> if non-applicable</black>
Trailer	Enter the onsite trailer start	PR only
Counts:	count (should only be on first	# = number of trailers in the
Onsite Start	Sampler's form).	onsite count area at start
		count
		0 = no trailers in onsite count
		area at start count

FIELD	INSTRUCTIONS	CODES AND FORMATS
		 dank> if non-applicable
Trailer Counts: Offsite Start	Enter the offsite trailer start count (should only be on the first Sampler's form).	PR only # = number of trailers in the offsite count area at start count 0 = no trailers in offsite count area at start count
Trailer Counts: Onsite Stop	Enter the onsite trailer stop count (should only be on last Sampler's form).	PR only # = number of trailers in the onsite count area at stop count 0 = no trailers in the onsite count area at stop count
Trailer Counts: Offsite Stop	Enter the offsite trailer stop count (should only be on last Sampler's form).	PR only # = number of trailers in the offsite count area at stop count 0 = no trailers in the offsite count area at stop count
Other SPP Headtags: SPP	Enter the species code for heads taken from non-salmon species.	PR and PC only Example: RFYEY <blank> if non-applicable</blank>
Other SPP Headtags: #used	Enter the total number of non-salmon head tags used.	PR and PC only # = Number of other spp. head tag used 0 = No head tags used from other spp. <black>if non-applicable</black>
Other SPP Headtags: HT#	Enter the head tag series number(s) used for non-salmon species.	PR and PC only ##### = the head tag number used for other spp. 0 = No other spp. head tags used <black> if non-applicable</black>
Weather and other pertinent notes	Enter notes about weather, other Samplers you worked with, training, CPFV activity, other Sampler headtags on your data sheets, driving conditions, etc. REQUIRED FOR EACH ROW.	ALL MODES Please be concise - max 75 characters allowed in database

Weekly Report Coding Tips

The following coding tips and examples address the most common types of errors that occur on the Weekly Report. The most common errors fall into the following categories; 1) fields inappropriately left blank or not blank, 2) transcription errors between the ASF and the Weekly Report and, 3) incorrect coding.

- Make sure you do not include data from other Samplers on your Weekly Report.
- 2. Do not fill in trailer count items if you were not in charge of tallying those for the PR assignment.
- Fill out Pacific halibut, yelloweye rockfish, cowcod, canary and black rockfish counts for PR and PC modes (excluding PCO onboard catch location).
- Salmon items are only needed for PR, PCS, and COM assignments except for Head Tags, which are reported for all sampling modes.
- 5. EVERY row on the Weekly Report needs: Sampler name and number, MODE, date, sample times, and comments filled in.
- 6. Use the "Example" tab in the Excel file for information on how to code each assignment mode on the Weekly Report.
- 7. If both PCS and PCD data are recorded on the same date a line will be necessary for each mode.
- 8. Use the "Mode and Port Code" tab in the Excel file for a list of Mode codes, PR1 sites, and PC sites.
- The assignment ID and sample site (port code, cluster or cnty/site) should be reported on each line relevant to the assignment, including the travel rows.

Example of Weekly Report - 1

Z			,		Rels			0	0							0								
-			RFCOW	_							_					H							H	
-		ROBERTS	æ		Kept			0	0							0							Ц	
×			RFYEY		Rels			2	0							က								
>		NAME:	RF		Kept			-	0							0								
>		Lead LAST NAME:	PA		Rels			0	0							0								
0		Lea	HALPA		Kept			-	0							0								
တ		9		Head	Tags			30	0	2						5				0				
œ		District #:	910		Rels			3		0						3								
o		Ö	Coho		Kept			0		0						-								
۵.		312	gs		Rels			7		3						2								
0		Sampler# 312	Kings		Kept			119		50						40								+
z		San	non					63		9						36								
Σ			Salmon		Boats Angls			30		-						9								
_			Γ	Total	Boats			47	2	-						24								
¥		TROXE		Reiu +	_			-								0								
_		NAME:		Mileage Barrie	1		0					0			0		0		23		23			
_		Sampler LAST NAME: TROXEL		Decima	Hours		2.00	10.03	0.33	0.33	0.00	2.00	2.00	2.00	2.50	7.08	2.58	0.00	0.53	7.53	0.43	1.00	000	
I		Samp	Time		E G		_	1832	1335	1335	1200	2230	1200	1700			2210			1535		1800		
Ö			Sample Time	_	Start			0830	1315	1315	1200	2030	1000	1500	1000	1230	1935			_	1536	1700	H	
L.	50		ľ	MODE			TRV [PR1	PCD	SS	PEC	TRV	DAT	MAO	TRV	PR1	TRV	PR4	TRV	MM	TRV	OFC		
ш	dsheet 20.	05/04/20			_	Cnty Site	SHC	SHC	SHC	SHC	SHC	SHC			CRL	CRL	CRL	EUR	RED6	RED6	RED6			
٥	ry Spread	0/90		Date	mm/dd/yy		5/4	5/4	5/4	5/4	5/4	5/4	2/2	2/2	9/9	9/9	9/9	9/9	6/9	6/9	2/6	6/9		(P)
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A	4 CRFS-OSP Weekly Summary Spreadsheet 2020	Week Beginning Monday:	Sampler		LAST NAME		TROXEL		TROXEL	TROXEL	TROXEL		<u></u>											
7	4	2	9			_	œ	6	9	F	12	13	4	5	16	17	9	19	20	21	52		24	

duster assignment on 5/9. Later that day, he concluded his workweek with office hours (OFC) spentediting data, drafting and submitting Weekly Report Example 1 (Columns A-Z) – Sampler Troxel (312) worked three different modes (PR1, PCD, & PCD) on 5/4. While on site at Shelter Cove (OSP port code "SHC"), he collected PEC information for three boats. On 5/5, Troxel worked non-samplinghours consisting of CRFS data entry (DAT) and other CDFW project work (OPW) approved by his Lead. On 5/6, he was originally scheduled his Weekly Report. ***Note**: Sample Time is blank for ASSN 056533 since the Sampler was scheduled but did not work. ****Note**: Each mode requires differentassignment summary totals. Reference the Weekly Report template workbook "Field Definitions" tab for full to work an EUR PR1 (ASSN 056533). He was reassigned to a CRL PR1 (ASSN 056509) instead. Troxel then worked a RED6MM specifics

AQ	V7. 2020		District	9	9	9	9	9	9	9	9	9	9	9	9	9	9	9	9	Î
AP		CDFW	Lead	ROBERTS	ROBERTS	ROBERTS	ROBERTS	ROBERTS	ROBERTS	ROBERTS	ROBERTS	ROBERTS	ROBERTS	ROBERTS	ROBERTS	ROBERTS	ROBERTS	ROBERTS	ROBERTS	
A0		Monday	Date	5/4/2020	5/4/2020	5/4/2020	5/4/2020	5/4/2020	5/4/2020	5/4/2020	5/4/2020	5/4/2020	5/4/2020	5/4/2020	5/4/2020	5/4/2020	5/4/2020	5/4/2020	5/4/2020	
AN		Notes	(e.g., marine conditions, CPFV activity, trailer status)	Travel from HQ to Shelter Cove in	Walkenhauer (303) has data also.	Sampled Squirrel and C'mon (RL)	Sampled Squirrel (ST/R) trip. Exce	Checkmate (2) non-fishing trip. Se	Travel from SHC to HQ. Overtime	CRFS data entry in Eureka office.	Other Project Work: Video data an	Travel from HQ to Fort Bragg in st	Worked for Bondoux (325) was sid	Travel from Fort Bragg to HQ in sta	I was asked to work in EUR for Bo	Weekly reporting, emails, mailing	Travel from HQ to Crescent City N	Early Shift, Order ADCB. Moderat	Weekly reporting, emails, mailing	
AM		dtags	# H		00186	0	0						0							
₹		Other SPP Headtags	# Used		-	0	0						0							
¥		Other S	Spp		RFYEY	0	0						0							
3			OffSite Stop										-							
₹		Frailer Counts	OnSite OffSite OnSite OffSite Start Start Stop Stop		0								0							
₩		Trailer (OffSite Start																	
AG					33															
ΑF		Boats	Onsite Offsite										4							
Æ		Missed Boats			0								0							
P		RFBLK	Rels		83	22							47							Ţ
AC		Æ	Kept		96	30							106							
B		RFCAN	Rels		9	0							12							
*		Æ.	Kept		00	0							9							

Weekly Report Example 1, Continued, (Columns AA-AQ) – Each mode requires different assignment summary totals (or data totals) totals. Whereas, PCD mode assignments require ColumnL, S-AD, and AK-AM. PR1 or PR2 mode assignments require assignment Columns K-AM). Reference the bottom of the ASF (for PR totals) and the Weekly Report template workbook, "Field Definitions" tab. for full details on all mode totals. For instance, PCS mode assignments only require Columns L-S and Columns AK-AMassignment Column E, cells in that row will highlight in green or blue. Green cells indicate a summary total is required. Blue œlls on PR1 or PR2 summary totals for Columns K- AM; Offsite Missed Boats and Office Site Trailer Counts do not apply to all sites. Reference the PR sampling sections for details. BB or MM mode assignments only require Column S to be totaled. *Note: Once a mode is filled into rows indicate a summary total may or may not be required. Reference the PR sampling sections for details on which sites require

Offsite Missed Boats and Office Site Trailer Counts. All grey columns will autofill based on the header fields of the report. **Note: all

rows listed in the Weekly Report requires the Notes (Cdumn AN) to be filled in.

Example of Weekly Report - 2

	ка	m	p	le	of	W	е	eł	(l)	y	R	ep	oc	rt	t -	- 2	2			
7			MO		Kels									0						
\		RTS	RFCOV	2	Kept									0						
×		ROBE	EY	-	Kels									0						
W		VAME:	HALPA RFYEY		Kept									0						
^		Lead LAST NAME: ROBERTS		-	Kels									0						
Π		Lead		2	Kept									0						
S		9		Head	lags			0			0			2						
æ		District #:	Coho		Kels									0						
ø		Ö	ပိ		Kept									0						
۵		425	Kings		Kels									-						
0		Sampler # 425	Ϋ́		Kept									12						
z		Sa	Salmon	_	Boats Angls									10						
Σ			Sal		Boats									4						
٦		NS		Refu + Total	Barrier Boats									7						
¥		COLLI		Refu +	Darrier									0						
٦		I NAME		Mileage	•		0		0	22		22	9		9	19		20		
-		Sampler LAST NAME: COLLINS		Decimal	Hours	0.50	1.72	1.03	2.00	0.33	5.33	0.33	0.23	8.00	0.25	0.25	7.35	0.83	1.00	
Ξ		Sam	Sample Time	ī	End	0815	0958	1100	1300	0820	1340	1400	0859	1700	1716	0815	1536	1626	1730	
g			Samp	č	Start	0745	0815	0958	1100	0800	0820	1340	0845	0060	1701	0800	0815	1536	1630	
ш	020			MODE		OFC	TRV	88	TRV	TRV	Σ	TRV	TRV	PR2	TRV	TRV	BEC	TRV	OFC	
В	adsheet 2	05/04/20		OSP Port,	mm/dd/yy Cluster, or Cnty Site		RED9	RED9	RED9	RED4	RED4	RED4	023107	023107	023107	BEC1	BEC1	BEC1		
Q	ary Sprea	0/90		Date	mm/dd/yy	2/9	2/9	2/9	2/9	8/9	8/9	8/9	6/9	6/9	6/9	9/10	9/10	9/10	9/10	
၁	Summe	Monday:		Assn ID	num		056101	056101	056101	056206	056206	056206	056309	056309	056309					+
В	Weekly	Week Beginning Monday:	Je.		Number	425	425	425	425	425	425	425	425	425	425	425	425	425	425	050420
A	4 CRFS-OSP Weekly Summary Spreadsheet 2020	5 Week	6 Sampler		LASI NAME	8 COLLINS	6 COLLINS	10 COLLINS	11 COLLINS	12 COLLINS	13 COLLINS	14 COLLINS	15 COLLINS	16 COLLINS	17 COLLINS	18 COLLINS	19 COLLINS	20 COLLINS	21 COLLINS	4 4

Since she drove a state vehide, her mileage was listed as "0". On 5/8 Collins sampled the RED4 MM duster. Her PR2 assignme**r**t on Fags" in Colum S only. Samplers should also note the start time and order the cluster was sampled in Column AN. **Note: PR2 rows 5/9 was completed at County 23, Site 107. Her final workday for the week, on 5/10, included samping a BEC route and office hours (OFC) spent completing her Weekly Report and data edits. *Note: BB and MM rows have an assignment summary total for "Head vehide at the office. This time was coded to the non-sampling mode code OFC. Directly after, she sampled the RED9 BB duster. Weekly Report Example 2 (Columns A-Z) – Sampler Collins (425) started her workweek on 5/7 by picking up forms and a state have the CNTY SITE code listed in Column E as a six-digit number with no spaces or special characters.

AQ	V7. 2020		District	9	9	9	9	9	9	9	9	9	9	9	9	9	9	•
ΑP		CDFW	Lead	ROBERTS	ROBERTS	ROBERTS	ROBERTS	ROBERTS	ROBERTS	ROBERTS	ROBERTS	ROBERTS	ROBERTS	ROBERTS	ROBERTS	ROBERTS	ROBERTS	
AO		Monday	Date	5/4/2020	5/4/2020	5/4/2020	5/4/2020	5/4/2020	5/4/2020	5/4/2020	5/4/2020	5/4/2020	5/4/2020	5/4/2020	5/4/2020	5/4/2020	5/4/2020	
AN		Notes	(e.g., marine conditions, CPFV activity, trailer status)	Picking up forms, talking with Lea	Travel from HQ to Oregon border in	Some BB effort at Pelican Beach;	Travel from Crescent City to HQ	Travel from home to Trinidad in per	No effort at Trinidad pier, some effi	Travel from North Jetty to home.	Travel from HQ to Samoa Bridge T	Boats able to make it out of the ba	Travel from Samoa Bridge T. Stree	Travel from HQ to Eureka Slough	Moderate BB and MM effort inside	Travel from South Spit to HQ	Completing and submitting weekly	
AM		dtags	# 1H									0						
٩٢		Other SPP Headtags	# Used									0						
AK		Other 5	Spp									0						
A			OffSite Stop															
A		railer Counts	OnSite OffSite OnSite OffSite Start Start Stop									0						
ΑH		Trailer (OffSite Start															
AG												9						
ΑF		Missed Boats	Onsite Offsite															
AE		Missec	Onsite									0						
AD		RFBLK	Rels									0						¥
AC		RF.	Kept									6						
AB		RFCAN	Rels									0						
¥		FF.	Kept									0						

Weekly Report Example 2, Continued, (Columns AA-AQ) – Each mode requires different assignment summary totals (or data totals) Columns K-AM). Reference the bottom of the ASF (for PR totals) and the Weekly Report template workbook, "Field Definitions" tab. totals. Whereas, PCD mode assignments require ColumnL, S-AD, and AK-AM. PR1 or PR2 mode assignments require assignment Offsite Missed Boats and Office Site Trailer Counts. All grey columns will autofill based on the header fields of the report. **Note; all Column E, œlls in that row will highlight in green or blue. Green œlls indicate a summary total is required. Blue cells on PR1 or PR2 for full details on all mode totals. For instance, PCS mode assignments only require Columns L-S and Columns AK-AMassignment summary totals for Columns K- AM; Offsite Missed Boats and Office Site Trailer Counts do not apply to all sites. Reference the PR sampling sections for details. BB or MM mode assignments only require Column S to be totaled. *Note: Once a mode is filled into rows indicate a summary total may or may not be required. Reference the PR sampling sections for details on which sites require rows listed in the Weekly Report requires the Notes (Cdumn AN) to be filled in.

THE ASSIGNMENT SUMMARY FORM (ASF)

The ASF is a cover sheet used to track CRFS assignments as well as a summary page of all data sheets used on the assignment. The ASF moves with the data sheets through delivery, editing, scanning, and data entry. ASFs are used by the Lead to monitor time on site, travel time, assignment disposition, effort levels, and to make sure Samplers conducted the correct assignment, cluster and order.

An ASF must be submitted for each CRFS assignment that has an ASSN ID. regardless of the assignment's disposition (including reassigned and canceled), even if the Sampler never went out to sample. The ASF is mandatory and will serve as a record of what happened to every issued assignment. Every site visited is logged on the ASF, even if no anglers are interviewed. The ASF is also used to record PR trailer counts and to summarize data for boat mode assignments. You may also be scheduled to conduct Site Effort Checks (SEC) at certain sites and these counts would be recorded on the ASF as well. Of note, ASFs are not needed for dockside salmon PC samples during salmon season (these assignments do NOT have ASSN IDs).

Assignment Summary Form (ASF) Layout

The ASF is structured into three general areas: header, site rows, and footer. The header is for recording information about the CRFS assignment as a whole, including the number of hours the Sampler worked the assignment. The majority of the form is structured into site rows, where specific information is recorded about each site. The footer is used to record a daily summary of data in boat modes but excludes PCO-Onboard Location Form data.

Data Tracking

The top margin of each ASF has six fields (RCVD ON/BY, EDIT ON/BY, SCAN ON/BY, ENTER ON/BY, UPLD ON/BY and FILE ON/BY) used by your Lead and other data editors/data entry personnel to track the progress of the assignment through the editing and entry process. Leave these fields blank – your Lead will complete them.

RCVD ON/BY:	EDIT ON/BY:	SCAN ON/BY:
ENTR ON/BY:	UPLD ON/BY:	FILE ON/BY:

ASF Header

The header section is required to track the Assignment ID, who worked, and the fishing mode and cluster (if appropriate). Each assignment record is identified in the database with the assignment number, Sampler ID, date and Assignment ID. The header section is also used to report the Sampler's hours and mileage, assignment disposition, other Samplers on the assignment and general comments about marine conditions, effort and catch, PC activity and/or any other pertinent information that may have influenced fishing or sampling.

RCV	/D ON/BY: E	:DIT ON/BY:		50	CAN ON/BY	r:		
ENT	'R ON/BY: L	IPLD ON/BY: _		FI	LE ON/BY:			
Asan #	V16 3/4/19	CRFS	ASSIGNMENT	SUMMARY F	ORM			
_	SAMPLER NAME:	SAMPLER #	DATE (MM/DD/YY)	ASSN ID	MODE	PORT/ CLUS	HOURS	
1	Joe Sampler	187	07/04/19	075503	PR1	PRI	10	TOTAL
	OTHER SAMPLER(S): NAME & # (w/data)	1	PLER(5): NAME & # (w/data)	1	ASSN DISP*	8.5	SAMPLING
	Tom Johnson 156 (Y/N) circle one (Y/N) circle one					State Car or Pers Vehicle	1	TOT TRV
	MARINE CONDITIONS: 3-4 foot swell, light winds, patchy fog					ODO START	0.5	TRV TIME HQ to site 1
	EFFORT: Fairly high. 35 trailers to start					ODO STOP	0.5	TRV TIME
SINE	CATCH: Mix of RFGEN and Salmon. Many anglers had limits 50 MILEAGE						0.5	-
COMMENTS	PC ACTIVITY: One 6 pack (Sara Bella RFGEN charter) was sampled opportunistically Round all hours to							i all hours to
ö	OTHER PERTINENT INFORMATION:							st 0.25 hour
	The Princeton Launch Ramp is scheduled to close 08/05/19 for repairs. Johnson (156) has stop count.						0.00hr = 53-07 mins 0.25hr = 08-22 mins	
	No offsite trailer count						0.50hr = 23-37 mins 0.75hr = 38-52 mins	
	HEAD TAGS USED (PR/PC): 12001-12036	EAD TAGS USED (PRIPC): 12001-12036 HEAD TAGS USED (MIMBB):						

Hours

For each CRFS assignment, record sampling, travel, and edit hours to the nearest quarter (0.25) hour, rounding up or down as necessary. Travel time is the time spent driving from headquarters to the first site plus the time spent driving from the last site to headquarters at the end of the day. Sampling time is the time you arrived at your first site until the time you leave your last site, and includes time spent driving between sites. Edit time is time spent editing data outside of the assignment. Normally, CRFS Samplers would edit data during slow periods onsite, but if this is not possible, at-home editing time should be reported here. Time spent at a site conducting a PEC or SEC is considered sample time.

ASF Site Rows

This area is used to report and describe each site visit during an assignment ASFs for MM and BB cluster assignments and SEC assignments will often have quite a few site rows filled out, since the Sampler is roving among multiple sites. Generally, for PR and PC modes, only one site will be listed because the Sampler will be at that same site for the entire working day. In addition, SECs conducted at sites adjacent to sites in the assigned mode may be added and recorded on the same ASF.

	*Assignment dispositions: 1=Complete, 2=Reassigned, 6=Canceled Check box if continued on 2nd page HO Departure Time: 1700 **ROW#* SITE NAME / SITE COMMENT** TIME						ite Effort Check applicable)	Onsite Start	Offsite Start	Onsite Stop	Offsite Stop
	Princeton PC	CNTY	81	ARRV	0800	вв	ANGS COST				Г
4	Lots of late launches due to nice weather. One	SITE	100	START	0805	мм		35			
Ι'	angler had a cowcod. Informed him the fish was no	DISPO**	1	STOP	1625	ММ		35			
	take and gave him a reg book.	HR5	8.5	DEPR	1630	PR					

This section is also used to record how much of the Sampler's time was spent at each site, the reason for leaving a site (disposition), arrival/departure time, time active sampling started/stopped, and PR trailer counts. Make sure the site name matches exactly what is on the current monthly Site List. For assignments which are reassigned or canceled, record the site name, county

and site code, and disposition (the reason why it was reassigned or canceled). Under certain circumstances you may be assigned to conduct site effort checks at various sites. Your Lead will give you more information on SEC assignments.

ASF Footer - Boat Mode Totals

The footer of the ASF consists of totals for boat modes and is used to monitor sampling and catch for weekly tracking. The footer is to be filled out for boat mode assignments only, with the grand sum of the page totals from each form. No zeros are needed in this section for MM and BB modes. You are required to report these PR totals to your Lead on your Excel Weekly Report every Monday by 8:00 AM. Weekly Reports from Samplers in Districts 1 and 2 may have due dates and content requirements given by their Leads.

ASF Item by Item Instructions

Field Name	Instructions	Coding_
	l	Examples and Formats
	HEADER	
Assn#	The assignment number will	1=first assignment
	be "1" unless you are issued	2=second assignment
	more than one assignment in	
0	a day.	La a Oarran Lan
Sampler	Print your full name. Do not	Joe Sampler
Name	sign.	100 1 000
Sampler #	Enter your personal 3-digit	100 to 399
	CRFS Sampler ID code.	Example: Joe Sampler = 150
Date	Enter the assignment date.	MM/DD/YY
		Example: 08/05/14 for
		Aug, 5 2014
ASSN ID	Enter the 6 digit assignment ID. Each CRFS assignment is given a unique identification number. The number should be used on its issued date and every time the assignment is attempted or if it is canceled.	011001 to 126999. The first two fields are the month (e.g. 08= August). The third field is the CRFS District (i.e. 4 = SF). The fourth field identifies the mode and the last two fields from 01-99 are generated by the schedule draw program

Field Name	Instructions	Coding Examples and Formats
Comment	Summarize and describe your day's activities in the space provided. Include marine conditions that may have affected catch or effort. Report on unusual events, angler activities, and species targets. Describe rescheduled assignments, missed and canceled assignments, and alternate site, if completed at a different site than assigned. Include descriptions of effort levels and catch, especially rare species. Report the names and sampler numbers of additional Samplers whom you worked with. Circle appropriate letter indicating whether or not additional Samplers collected data. List names of CDFW staff who conducted field checks. List any PC activity, other pertinent information, and headtags used.	"The beaches were empty due to NW gale force winds." "Waves washing over jetty made it unsafe to conduct counts." "Effort was high but catch avg. 4 RF each; a few ling also landed" "Worked with Jane Doe (217) and had a field check by John Doe" "Used headtag series 50000-06" "Unable to obtain weights due to rough conditions" "Worked with Sampler Joe Smith today. We both have data" "No PC effort at assigned site Emeryville, assignment completed at Berkeley instead".

Field Name	Instructions	Coding Examples and Formats
ASSN MODE	Enter the assigned mode. The assigned mode will appear on your Sampler Schedule.	PR1 and 2 = Primary and secondary private / rental boat sites. MM= Manmade structures BB= Beaches and banks. PCO= Party and charter boats sampled on- board PCD= Non-Salmon party and charter boats sampled dockside PEC= Party/Charter effort check SEC= Site Effort Check
CLUSTER	For MM, BB, and SEC assignments record the cluster site code. The cluster code is the county or District alpha-code with a number suffix. For PR and PC modes use the three letter OSP port code.	San Diego 1 = SDG1 Central 1 = CEN1 Bodega PR1=BOD

Field Name	Instructions	Coding Examples and Formats
ASSN DISP	Report the number code for the assignment disposition. The codes are defined under the "COMMENT" box. The assignment dispositions are 1, 2 or 6. For Dispositions other than "1" the Sampler must record the conditions or reasons in the comments section. Note: There is another disposition below for each site (site disposition). Do not get the two disposition types confused. For assignments which are reassigned or canceled, record the site name, and county and site codes on the first ASF Site Row.	1=Complete: When you "complete" the assignment and it is done. 2= Reassigned: When the assignment needs to be moved or rescheduled. Examples: You missed the PC boat and there are no eligible alternates; personal reasons approved by your Lead; the site is closed to fishing; no boats going out due to weather or low effort; you get sick or injured during the assignment; the situation is unsafe or unhealthy 6= Cancelled: When your Lead notifies you that the assignment cannot be rescheduled before the end of the month
State car or pers vehicle	Record "SC" for state car or "PV" for personal vehicle.	"SC" or "PV". Make notes in comments if used some other transportation
Odo Start	Record your odometer mileage at start of driving (when you leave home/HQ).	Example:10001
Odo Stop	Record your odometer mileage at end of driving (when you arrive at home/HQ).	Example: 10044

Field Name	Instructions	Coding Examples and Formats
MILEAGE	Compute the total miles you drove to the nearest whole mile for the day. This will help the Sampler fill out a CalATERS claim for the	Miles, to the nearest whole number Example: 43
	month. For Samplers using a State car, do not record mileage (leave blank).	
HOURS TOTAL	The total hours for the day. Incudes: sampling, travel and edit	Round to nearest 0.25 hour.
	A conversion chart has been provided in the COMMENTS section to help with rounding hours to the nearest 0.25 hours.	Example: 8 hours and 20 minutes would be rounded to 8.25
HOURS SAMPLING	The time you spent from arrival at your first site to the time you leave your last site.	Round to nearest 0.25 hour. Example: 8 hours and
		40 minutes worked would round to 8.75
HOURS TOT Travel	The time you spent travelling from your HQ to your first site, plus the time you spent travelling from your last site to HQ.	Travel between sites is counted in your HOURS SAMPLING.
Field Name	Instructions	Coding Examples and Formats
HOURS TRV time HQ to first site	The time you left your HQ to the time you arrive at your first site.	Round to nearest 0.25 hour. Example: 45 minutes
HOURS TRV time last site to HQ	The time from when you left your last site to when you arrived at HQ.	rounds to 0.75 hours Round to nearest 0.25 hour. Example: 55 minutes rounds to 1.0 hours

Field Name	Instructions	Coding Examples and Formats
HOURS EDIT	Extra time spent editing forms at home or office. You are expected to edit your forms during slack time between interviews; however, occasions may arise when you require more time to edit forms.	Round to nearest 0.25 hour. Example: 25 minutes rounds to 0.5 hours
	SITE ROWS	
SITE NAME	Name of this site as listed on your current Site List. The Site Name and Site Code must match.	"Santa Cruz Marina Launch Ramp"
SITE COMMENT	Record comments regarding: unusual circumstances at this site, effort levels, missed anglers or boats, language barriers, catch.	"Boat angler was in a kayak" "Missed one BB angler"
CNTY = County	Record the 3-digit county code.	1=Alameda 111=Ventura

Field Name	Instructions	Coding Examples and Formats
SITE	Record the 3-digit site code corresponding to the site name.	"104" = Moss Landing Launch Ramp
DISPO = Site Disposition	The site disposition is recorded for each site sampled and indicates the status of the effort there and the reason for leaving the site. The lowest valid disposition code should be used. Site disposition is recorded just prior to departure from the site. The site disposition code needs to be 0, 1, 4, 5, or 7.	Use the lowest valid code: 0= Site Effort Check: You have performed a trailer/angler count only (i.e., drive-bys) 1= Complete/Done 4= Low Effort (PC only): There are no anglers. The assn will need to be rescheduled; work with your Lead on this. 5= Other: Examples: Time spent at site (outside of assigned cluster) interviewing anglers; you can't ride the boat for whatever reason (not allowed by captain, PC had motor problems, poor weather). 7= Roving (Clusters): you are sampling a cluster of sites and you are moving between sites as scheduled. Your last site visited will get a DISPO=1

Field Name	Instructions	Coding Examples and Formats
HRS =Hours	Enter the total amount of time spent at the site (time between arrival and departure times). Do not include time traveling to or from the site. Include time spent driving between access points at that site.	Round to nearest 0.25 hours. Example: 55 minutes rounds to 1.0 hours
ARRV =Arrival	Time in 24 hour format when you physically arrived at the site.	24 hour format: "0701" = 7:01am
START	This is the time when you physically start sampling, usually after doing an arrival count of trailers or anglers.	24 hour format: Must be at least one minute later than ARRV time
STOP	This is the time you physically stopped sampling.	24 hour format: For MM assignments, you will do your end count in between STOP and DEPR times
DEPR =Departure	This is the time when you physically departed the site.	24 hour format: "2359" = 11:59pm

Field Name	Instructions	Coding Examples and Formats
Site Effort Check (if applicable)	You will only use this if you are assigned to specifically go check fishing effort for a certain site or group of sites. Write in the number of anglers/boats/trailers in the appropriate cell for the mode(s) you are assigned to check (MM, BB, PC, PR).	Example: You are assigned to go check a series of sites in a specified route. You would record the number of anglers or trailers seen at each site. Example: While traveling
		to or from a CRFS assignment you might be instructed to conduct SEC counts at adjacent sites.
PR Trailer Counts	Record the number of fishing trailers Onsite, Offsite or both, if applicable.	See Summary of PR Counts table in PR1 and PR2 sections
	FOOTER – BOAT MODE T	
Refu + Barrier	Total number of PR fishing boats that refused the CRFS survey or were language barriers on all PR/PCO forms.	Sum of Refu + Barrier from each PR/PCO/PCD page
Total Boats	Total number of boats on all PR pages; includes fishing and NF boats (But not missed boats, refusals, or language barriers).	Sum of 'Total Boats' from each PR/PCO/PCD page

Field Name	Instructions	Coding Examples and Formats
Boats Salmon	Total number of boats sampled that were targeting and/or kept salmon on all PR forms. Include "Salmon Refusals" as salmon boats.	Sum of 'Salmon Boats' from each PR page 0= no salmon boats
Angs Salmon	Total number of anglers sampled from boats targeting and/or keeping salmon on all PR forms. Include "Salmon Refusal" anglers as salmon anglers.	Sum of 'Salmon Angs' from each PR form
Kept Kings	Total number of Chinook salmon kept observed + kept unobserved on all PR/PCO forms.	Sum of 'Kept Kings' from each PR/PCO/PCD Angler form
Rels Kings	Total number of Chinook salmon released alive + released dead on all PR/PCO forms.	Sum of 'Rels Kings' from each PR/PCO/PCD form

Field Name	Instructions	Coding Examples and Formats
Kept Coho	Total number of Coho salmon kept observed + kept unobserved on all PR/PCO forms.	Sum of 'Kept Coho' from each PR/PCO/PCD form
Rels Coho	Total number of Coho salmon released alive + released dead on all PR/PCO forms.	Sum of 'Rels Coho' from each PR/PCO/PCD form
# Head Tags	Total number of salmon head tags issued (including NRS tags) on all PR/PCO forms.	Sum of '# Head tags' from each PR/PCO/PCD form
Kept Pac. halibut	Total number of Pacific halibut 'observed kept' and 'unobserved kept' from all PR/PCO forms.	Sum of 'Kept Pacific halibut' from each PR/PCD/PCO/PCD Angler Form
Rels Pac. halibut	Total number of Pacific halibut 'released dead + released alive' from all PR/PCO forms.	Sum of 'Rels Pacific halibut" from each PR/PCO/PCD form
Kept Yelloweye	Total number of Yelloweye Rockfish 'observed kept' and 'unobserved kept' from all PR/PCO forms.	Sum of 'Kept Yelloweye' from each PR/PCO/PCD form
Rels Yelloweye	Total number of Yelloweye Rockfish 'released dead + alive' from all PR/PCO forms.	Sum of 'Rels Yelloweye' from each PR/PCO/PCD form

Field Name	Instructions	Coding Examples and Formats
Kept Cowcod	Total number of Cowcod 'observed kept' and 'unobserved kept' from all PR/PCO forms.	Sum of 'Kept Cowcod' from each PR/PCO/PCD form
Rels Cowcod	Total number of Cowcod 'released dead + alive' from all PR/PCO forms.	Sum of 'Rels Cowcod' from each PR/PCO/PCD form
Kept Canary	Total number of Canary RF 'observed kept' and 'unobserved kept' from all PR/PCO forms.	Sum of 'Kept Canary RF' from each PR/PCO/PCD form
Rels Canary	Total number of Canary RF 'released dead + alive' from all PR/PCO forms.	Sum of 'Rels Canary RF' from each PR/PCO/PCD form
Kept Black RF	Total number of Black RF 'observed kept' and 'unobserved kept' from all PR/PCO forms.	Sum of 'Kept Black RF' from each PR/PCO/PCD form
Rels Black RF	Total number of Black RF 'released dead + alive' from all PR/PCO forms.	Sum of 'Rels Black RF' from each PR/PCO/PCD form
On Missed	The total number of onsite missed boats on all PR forms.	Sum of 'On Missed' from each PR form
Off Missed	The total number 'offsite missed boats' returning to the PR1's offsite area (usually a marina or private slip) on all PR forms.	Sum of 'Off Missed' from each PR form
		vp

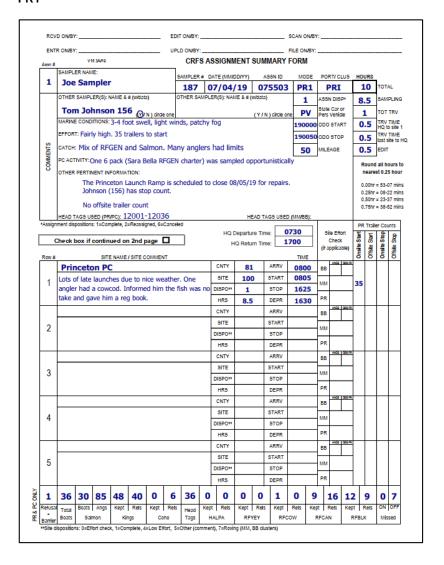
ASF Coding Tips

The following coding tips and examples address the most common types of errors on the Assignment Summary Form. The most common errors fall into the following three categories: 1) items left blank or not blank in appropriately, 2) mathematical errors and 3) incorrect assignment procedures followed.

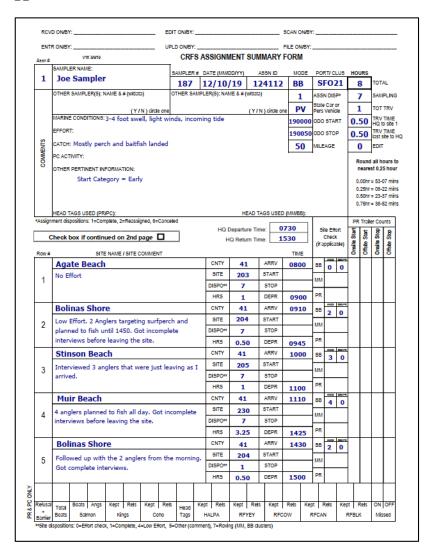
Specific Editing Checks

- The date should be recorded in MM/DD/YY format, like the other CRFS forms
- 2. BB and MM assignments are disposition 7 = 'roving' until the last site visit, which is 1 = 'complete'
- 3. Round all recorded time to nearest 0.25 hour. The table in the COMMENTS area is provided to help with rounding
- If a field is not applicable, such as SEC or PR Trailer Counts, leave it blank

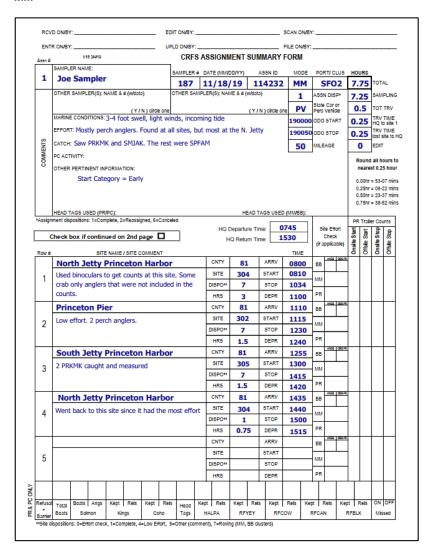
Assignment Summary Form Examples PR1



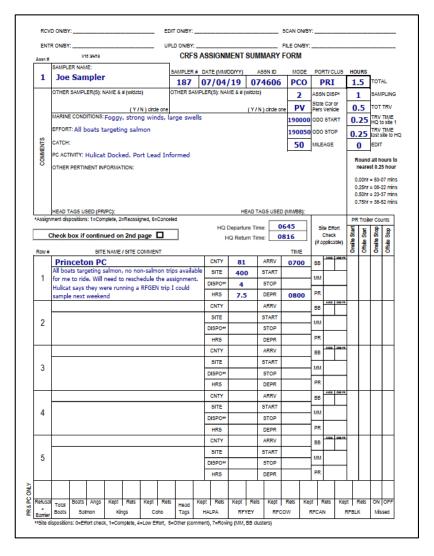
Assignment Summary Form Examples BB



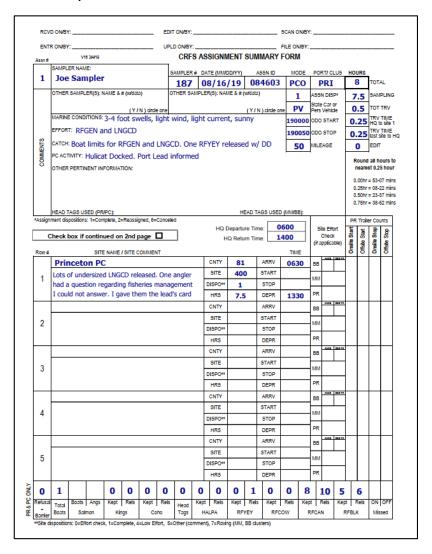
Assignment Summary Form Examples MM



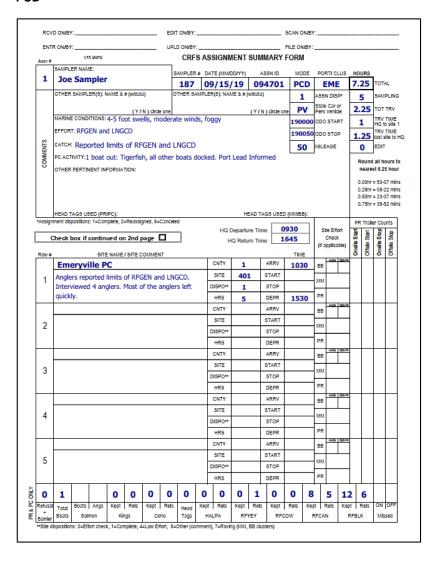
Assignment Summary Form Examples PCO – Reassigned



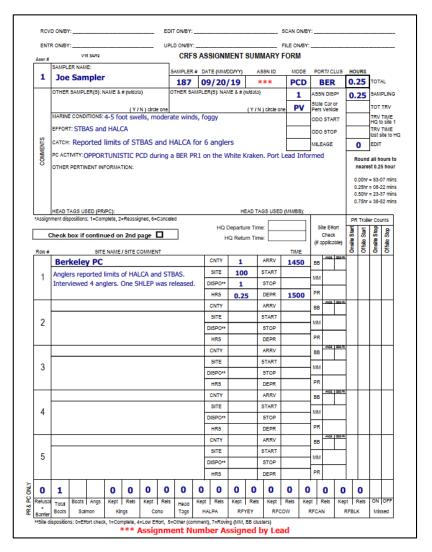
Assignment Summary Form Examples PCO – Completed



Assignment Summary Form Examples PCD



Assignment Summary Form Examples PCD – Opportunistic



Beach and Bank (BB) Mode Sampling

BB Mode Definition

Beach and Bank mode (BB) is defined as a shore mode where recreational fishing occurs on beaches and/or banks. Beach is defined as the ocean shore made up of sand or pebbles, usually washed by high tide waters. Bank is defined as the slope of elevated land adjoining with the ocean or bay, can be rock or an overhanging cliff, and may be reinforced with materials placed there by humans. A beach or bank may be part of a nearshore area or a bay or estuary.

BB Survey Goal

The primary goal for BB sampling is to estimate catch per unit of effort (CPUE) as catch per angler trip in this mode. CPUE is calculated by multiplying the catch per minute fished by the average trip length. Fishing effort is provided by the telephone survey. Other relevant data collected by the BB survey include area fished, gear type, target(s), and fish biological data. The Sampler's goal while on a BB assignment is to obtain as many high-quality interviews from as many BB anglers as possible.

BB Survey Methods

In comparison to other modes of fishing sampled by CRFS, BB mode is sampled at a lower rate. BB sites are grouped into clusters. The Sampler will rove and sample all sites in a cluster assignment. The sites within a cluster are defined by a site list which will be provided by the Lead via the Monthly Schedule. The number of sites or if they are active/inactive may depend on the season and/orthe geographic proximity among sites. The cluster/site list changes and is unique by month. It is important for Samplers to use the cluster/site list that matches the month of the sample selection.

The BB sample draw selects one assignment per BB cluster per month. Randomly, 2/3 of all the District's BB clusters are assigned to weekend days and 1/3 of the clusters are assigned to weekdays. BB effort is expected to be different for these separate kinds of days (KODs). Therefore, expect more BB assignments to be scheduled on weekends and holidays than on weekdays. The Sampler is to begin a BB sample day by consulting the Monthly Schedule, which will list the date of the assignment and BB cluster to visit. Leads may set the start times for the assignment and/or direct the order in which you will visit the sites in the cluster. Without direction from the Lead, the start time and the first site visited are determined by the Sampler. If start location is at the Sampler's discretion, they will move through the sites in a geographically efficient order, and randomize start times and starting sites. All of the BB sites are publicly accessible which allow Samplers to access them without problems.

Example BB Clusters from Site List

DISTRICT	MONTH	CNTY	NAME	SITE	AMODE	CLUS
1	NOV	37	Alamitos Bay	214	ВВ	LOS1
1	NOV	37	Marine Stadium	215	ВВ	LOS1
1	NOV	37	Pier J	201	вв	LOS1
1	NOV	37	Shoreline Village	202	ВВ	LOS1
1	NOV	37	Cabrillo Beach	110	BB	LOS2
1	NOV	37	Cherry Beach	402	ВВ	LOS2
1	NOV	37	King Harbor	303	вв	LOS2
1	NOV	37	Abalone Cove	205	ВВ	LOS3
1	NOV	37	Palos Verdes	211	ВВ	LOS3

The Sampler is to contact the Lead immediately if they cannot complete an assignment due to illness or emergency. For the proper implementation of statistical methods it is crucial that Samplers try to complete all assignments as scheduled. As with any mode, rescheduling of BB assignments is not desirable to the survey. If necessary, the Lead can reschedule a BB assignment. Leads will attempt to conserve the original KOD when rescheduling a BB assignment; however, the KOD may be changed if necessary.

Sampling will normally take place within an eight hour work day during daylight hours. Samplers will strive for six hours of sampling time and all ow up to two hours for travel time while on assignment. Samplers are to avoid working over an eight hour day for BB assignments. BB angler (angler parties) interviews are completed on the CRFS Shore Form. The Sampler will obtain an estimate of total BB anglers for each site visit and record any fishing kayak or fishing personal water craft activity at those sites. In addition, the Sampler may perform SECs at adjacent sites using the Assignment Summary Form and may also perform CPFV checks at adjacent PC sites using the PC Effort Check Form (PEC).

When a BB cluster is assigned, the Sampler will typically have to cover an extensive stretch of coast. The Sampler will move through all the access points defined in the cluster, counting and/or canvassing any anglers they encounter. An access point is a pre-defined location within a BB site where the Sampler may access a beach/bank in an attempt to intercept anglers. Some BB sites have one access point while others have many access points. The Sampler may use a site map binder or the CRFS Wiki site for driving directions, site boundaries, and a list of access points. After visiting the first site in a cluster, the Sampler should move through the other sites looking for angler activity and keeping detailed data records. It is important for the Sampler to keep in mind that there may only be one reasonable order of sites to visit to minimize driving time (i.e. starting at the northern or southernmost site) and keep project costs down. Unlike MM mode, BB clusters do not have a predefined or scheduled order. After the Sampler has visited all sites within the assigned cluster they may return to previously

visited sites where they expect to obtain interviews. All of the sites within the assigned cluster must be visited in order for the assignment to be considered complete. The Sampler is to notify their Lead immediately if they are not able to complete a BB assignment by visiting all sites within your assigned BB cluster.

The general rule is for the Sampler to stay at a site where they expect to get one interview per hour. If they do not expect to obtain at least one interview per hour, the Sampler should move to the next site in the cluster. In an effort to obtain as many valid and high-quality BB interviews as possible at a site, it may be necessary to stay at certain access points where there is high angler activity. The Sampler should take up a strategic position so they can intercept a majority of the anglers. If no such point exists, the Sampler should be posited where the majority of the anglers are within sight and easily accessible. At crowded beaches, close observation of the fishing activity is required since the Sampler must be alert to those anglers leaving the site. In other instances, roving through access points by vehicle is the easiest way to spot and count anglers at a site. Each BB cluster is unique and the Sampler will be trained on the best way to sample any specific BB site.

BB Estimated Total Finfish Angler Counts

An important aspect of BB sampling is obtaining estimated total finfish angler counts by site. The BB survey does <u>not</u> collect start, stop and instantaneous counts like for the MM survey. Since BB sites are vast and angler effort tends to be low, CRFS only needs an overall estimate of the number of finfish anglers participating in BB mode while you are there sampling a site. Since BB sites are divided up into access points, the Sampler may not know the estimated total of finfish anglers until the last access point has been checked. A good way for the Sampler to perform this estimated count is to keep a running tally of BB anglers while moving through all access points. The Sampler should also keep note of any arriving BB anglers.

For BB estimated total angler counts the Sampler is to count finfish anglers only. A finfish angler is defined as an angler that has wet gear hours and has or is targeting finfish during the survey day or has the immediate intent to finfish. This includes anglers taking a break, re-baiting or moving between locations within the site. Invertebrate-only anglers are NOT included in BB angler counts. If an angler is moving back and forth between a BB and a MM, they should be included in the BB Estimated Total Finfish Anglers Count. The Sampler should try not to double count or miss anglers behind bluffs or obstructions. Oftentimes it is difficult to determine the number of anglers on a long open beach so the Sampler should use binoculars and their best judgment. If it is difficult or dangerous to walk on a beach or bank, it is recommended to count finfish anglers using binoculars. The Sampler is to pay attention to site boundaries and only count BB anglers at that specific BB site. It is important for the Sampler to remember this is just an estimate. Times are not associated with any BB counts.

Canvassing

A useful tactic for sampling in BB mode is to complete a preliminary canvass to determine the number and location of anglers at a site and a rough approximation of the duration of their trips. With this information the Sampler is able to maximize intercept coverage by planning their movements around those of the anglers. It may give the Sampler a good indication of when to stay onsite and when to move to the next site in the cluster.

Incomplete BB trips

While complete-trip interviews are preferred, BB mode sampling protocol allows Samplers to interview anglers who have not yet completed their fishing trips. Anglers must have fished for at least 30 minutes to be eliqible for a CRFS interview in BB mode. Unlike MM mode, anglers in BB mode do not have to be more than halfway done with their fishing trip to be interviewed. Incomplete-trip interviews are allowed in an effort to get as much BB data as possible; normally this mode has lower effort than other modes and it has historically been a challenge to get an adequate number of BB interviews per assignment. The Sampler may get incomplete-trip interviews at any time while working a BB assignment.

Before getting an incomplete-trip interview, the Sampler is to canvass the angler(s) to determine if they should stay on site to get the interview or return to the site later. After visiting all sites in a cluster, the Sampler may encounter the same angler(s) again upon return to that site. When this happens, the Sampler is to attempt to update the interview. The Sampler should update the interview by copying the relevant information onto the Shore Form for the most recent visit to the site and deleting the interview from the Shore Form for the previous visit to the site.

Do not conduct incomplete trip interviews of anglers targeting invertebrates only without any incidental finfish by catch.

Low Effort Protocol

The general sampling guideline for clusters is to strive to obtain at least one interview per hour. If the Sampler cannot do such, they are to move to the next site in the cluster, return to a previous site in the cluster if they have gone through all sites, or terminate the assignment. If there is low effort at a BB site, the Sampler is to canvass the angler(s). The Sampler may decide to wait for anglers to complete their trip. The Sampler should continue to rove from site to site in the cluster until the day's fishing activity has ceased or the Sampler has worked to the limit of six sample hours. Other reasons to leave the assignment early would be if the site is unsafe, darkness, or extreme weather conditions.

No Anglers in BB Mode

The ASF and Shore Form header information, estimated BB angler counts and times, must be completed for each visit to a site even if there are no

finfish anglers present. After determining there are no BB anglers at the first site, the Sampler will go immediately to the next nearest site in the cluster. If no anglers are present at the next site, the Sampler will go immediately to the next nearest site. The Sampler is to keep searching for anglers by roving through sites and access points for up to two hours. If the Sampler does not find any BB anglers after roving through all sites in the cluster and two hours of sampling time has passed and no effort seems likely to develop, the Sampler may terminate the assignment; the assignment is complete. While waiting for effort to develop or anglers to complete their trip at an assigned mode and site, the Sampler may sample in PC mode opportunistically if this mode exists at the site.

Observed PR Accessed from BB

Fishing kayaks, and personal water crafts (PWC) are considered PR mode and may not be interviewed while you are sampling a BB. However, CRFS is interested in documenting where kayak and PWC-based angling activity takes place. Determining kayak angler launch sites can be difficult. In an effort to gather as much data on kayak fishing activity as possible, the Sampler will keep track of the number of fishing kayaks launched from the BB shore. These data will be used to determine if the creation of additional PR2 sites is warranted. The Sampler is to use discretion on the intended target based on visual observations and make the determination to include kayak and PWC anglers in the count only if it believed finfish is either the primary or secondary target.

Anglers Fishing in Two Modes

When interviewing an angler who has been fishing in two different modes (i.e. BB mode and MM mode), ask the angler where they have spent most of their time fishing. The angler is eligible for an interview if they have spent more than half of their fishing trip in the BB mode. The Sampler will collect only the information which pertains to the angler's time on the BB, i.e. catch, target(s), gear, area. The angler is ineligible for an interview if they have spent less than half of their time fishing in the BB mode.

To accurately portray the angler's time fishing in the BB mode, the Sampler will need to adjust the angler's arrival time. The Sampler will ask the angler how long they spent fishing at the BB and will calculate the angler's arrival time by taking the interview time and subtracting the angler's total time fishing in BB mode to create the estimated arrival time.

Opportunistic PC Sampling

It is possible to sample PC boats during BB sampling. For instance, the Sampler may encounter PC boats at beaches next to a PR site or PC landing. However, the Sampler may not leave the site or miss any BB interviews to complete an opportunistic PC sample.

Screening Divers

The Sampler is to be aware that divers at a beach or bank may be targeting finfish. If a diver used or intended to use a spear gun, they can be interviewed as eligible anglers (gear = S). Divers entering the water from

the BB shore using fins and a flotation device (such as a dive tube) to fish are considered BB anglers. Spearfishers using kayaks or personal watercraft are PR anglers and may not be interviewed while the Sampler is sampling a BB assignment.

Invertebrate Sampling in BB Mode

The goal of BB sampling is to collect as much information on finf is h catch and an estimate of the total number of anglers fishing at each site. However, on beach and bank assignments it is possible for the Sampler to find anglers targeting invertebrates. The Sampler must be aware of invertebrate-only anglers to avoid including these anglers in the BB estimated total finf ish angler counts and the observed PR accessed from BB counts.

Sometimes the Sampler may find an angler targeting both finfish and invertebrates. These anglers, of course, qualify for a CRFS interview and are included in the effort count. For the invertebrate component of the catch the Sampler will record all retained invertebrates as kept unobserved – do not count or collect biological data from retained invertebrates. Anglers targeting invertebrates only could have incidental catch of finfish; in this case record UNIFH as the secondary target. Beach and bank anglers targeting invertebrates only without incidental finfish bycatch are not included in the survey and will not be interviewed.

Two BB Assignments in One Day

Rarely a Sampler will be given two BB assignments on the same day. The Lead should specify which assignment to work first. The Sampler must work that assignment before the second assignment is attempted. In other words, before beginning the second assignment, the Sampler must visit all of the sites in the first cluster assigned before starting the second assignment.

Ways to Reduce Bias

Some ways to reduce potential bias in the BB sample include:

- BB anglers may fish during incoming tides, however the Sampler should not introduce bias into the survey by only interviewing anglers during that time period.
- Samplers should be sure to check all access points, not just the most frequently used, or the most easily accessible.

2017 CRFS BB Mode Questionnaire

The wording of the questions (i.e. script) has been structured to capture the required information for this survey in an efficient and thorough manner. A laminated copy of the BB questionnaire will be provided. It is important that the Sampler use the wording of questions as stated in the BB script since slight changes in wording can result in different responses. The Sampler will be canvassing, screening, introducing the survey, and providing the Privacy Act information. After screening for angler-eligibility, the Sampler will introduce the survey to the angler(s) to be sampled by saying:

SCREENING: Have you fished from a beach or bank in saltwater today?

Yes: Go to the next question.

No: If not complete but fished for at least 30 minutes, go to next

No: If not complete and have not fished for at least 30 minutes = Ineligible;

stop interview.

Refused: Code Sample # as R, record the number of anglers in the group,

terminate interview.

"Hello, my name is _____ and I represent CDFW. We are interviewing marine recreational anglers for the California Recreational Fisheries Survey. May I ask you a few questions about your fishing trip?"

The Sampler will state the Privacy Act by saying, "This study is being conducted in accordance with the Privacy Act of 1974. You are not required to answer any question that you consider to be an invasion of your privacy."

ANGS Total: How many of you had gear in the water?

Enter the total number of anglers that fished together. Go to next.

Refused: Code Sample # as R, terminate interview.

Unlicensed: What type of sport fishing license does each of you have? Enter the number of the ANGS (above) who fished without a current California sport fishing license. Go to next.

Refused: Code R in (unlic) and continue the interview.

TRIP LENGTH: At what time did you arrive at the fishing site today? Enter the time in 24 hr format when the angler reported arriving at the site. Go to next.

Refused: Code Sample # as R, terminate interview.

If incomplete trip but fished for AT LEAST 30 minutes: How many additional hours and minutes do you plan to fish here today?

Enter the number of additional hours and minutes the angler intends to fish. Go to next.

Refused: Code Sample # as R, terminate interview.

DAYS FISHED 12 months: Ask a random angler in the group. Not counting today, within the past 12 months, how many days have you gone saltwater sport fin-fishing in this state or from a boat launched in this state?

Refused: Code R, Don't know: Code DK

Sampler didn't ask: Code DA

ZIP CODE: Ask a random angler in the group. What is the ZIP code of your residence? If ZIP unknown, ask: What city or town do you live in?

Refused: Code R, Don't know: Code DK

Sampler didn't ask: Code DA

Non U.S. resident: Code Foreign Country

PRIMARY TARGET: What type of fish were you primarily trying to

catch? Code the taxon of the angler's primary target.

Anything: Code UNIFH

Refused: Code Sample # as R, terminate interview.

SECONDARY TARGET: What type of fish were you secondarily trying

to catch? Code the taxon of the angler's secondary target.

Anything: Code UNIFH

No secondary target: Leave blank

EFFORT AREA: Was your rimary target> fishing in the ocean or enclosed bay/estuary/harbor?

Nearshore (< 3 miles): Code N

Bay/Estuary/Harbor: Code B. Be aware of freshwater cutoffs. Refused: Sampler will determine and code appropriately

EFFORT AREA: Was your <secondary target> fishing in the ocean or enclosed bay/estuary/harbor?

Nearshore (< 3 miles): Code N

Bay/Estuary/Harbor: Code B. Be aware of freshwater cutoffs. Refused: Sampler will determine and code appropriately

GEAR: What gear type did you use for <pri>primary target>?

Finfish

Hook & Line: Code H Spear: Code S Bait Net: Code N

Shellfish

Pot #: Code Pn

Flat Hoop Net #: Code Fn Rigid Hoop Net #: Code Rn

Snare: Code E SCUBA: Code C Free Diving: Code D

GEAR: What gear did you use for <secondary target>?

Finfish

Hook & Line: Code H Spear: Code S Bait Net: Code N

Shellfish

Pot #: Code Pn Flat Hoop Net #: Code Fn

Rigid Hoop Net #: Code Rn

Snare: Code E

SCUBA: Code C Free Diving: Code D

SPECIES CODE: Did you catch any fish today?

Yes: Record Species Code and go to next.

No: Record "No Catch" in Species Code box and zeros in KEPT obs. KEPT

unobs. RELS alive total and RELS dead.

Refused: Code Sample # as R, terminate interview.

KEPT OBSERVED: May I see the catch?

Yes: Sampler will identify and count all fish by species.

No: Enter zero and code numbers of all fish as Kept Unobserved.

Fillets: If fillets can't be ID'ed by skin, enter zero and code numbers of Kept

Unobserved.

Refused to let you see fish or tell you the types and numbers of fish kept (i.e., kept un observed): Code Sample # as R, terminate interview.

KEPT UNOBSERVED: Did you retain any other fish? Probe for any fish given away, filleted, used for bait or thrown away.

Yes: Record species and number of fish.

No: Enter zeroes in Kept Unobserved boxes for all Kept Observed species recorded.

Refused: Code Sample # as R, terminate interview.

Don't Know: Code Sample # as R, terminate interview.

RELEASED ALIVE: Were any fish released alive? Probe for any fish that were purposely released alive (swam away).

Yes: Record species and number of fish.

No: Enter zeroes in Released Alive Total boxes for all Kept Observed or

Unobserved species recorded.

Refused: Code Sample # as R, terminate interview.

Don't Know: Code Sample # as R. terminate interview.

RELEASED DEAD: Were any fish released dead? Probe for any fish that were thrown back dead (did not swim away).

Yes: Record species and number of fish.

No: Enter zeroes in Released Dead boxes for species recorded Kept Observed or Unobserved.

Refused: Code Sample # as R, terminate interview. Don't Know: Code Sample # as R, terminate interview.

BIO DATA: May I measure your kept fish today?

FORK LENGTH (MM) SEX (M/F/T): Enter the fork length in millimeters of each fish measured above the dotted line. Add a suffix of M (male), F (female) or T (transitional) for each sexed fish.

WEIGHT (DECIMAL KG) OR HEAD TAG # (CIRCLE TAG #): Enter the weight in kilograms of the fish below the length. Do not record a weight

without a length. Do not weigh salmon. Enter the head tag number for an adipose fin-clipped salmon below the length in place of the weight. If the salmon head is lost or refused write NRS after the tag number. For collected yelloweye rockfish and white seabass, enter a length and weight and put the head tag number or scan code to the right of the measurements. Circle salmon and yelloweye head tag numbers.

Shore Form Procedures for BB Mode

This section describes specific procedures for the BB survey using the Shore Form. The Shore Form is used for shore modes only (MM and BB).

Numbering the Visits, Interviews and Shore Form Pages

Each BB site visited should be recorded on the ASF in the order visited. The Sampler must start a new Shore Form page when they arrive at a site. The first interview at a site will be "1" and all other interviews at that site, during that visit will be numbered sequentially. At the next site, the Sampler must start a new Shore Form page and begin renumbering interviews with "1" again. If the same site is visited more than one time in a day, a new page will be used for each visit, and the Sampler must start with interview "1" at each visit. An example is shown below.

First site in the cluster

- Site 1 on ASF
- Start a new Shore Form page
- Number interviews sequentially starting with 1

Second site in the cluster

- Site 2 on ASF
- Start a new Shore Form page
- Number interviews sequentially starting with 1

Third and last site in the cluster

- Site 3 on ASF
- Start a new Shore Form page
- Number interviews sequentially starting with 1

Return to site 2

- Site 4 on ASF
- Start a new Shore Form page
- Number interviews sequentially starting with 1

Shore Form page numbers will start at "1" and be numbered sequentially for the entire assignment.

Shore Form Lavout

The Shore Form is used for both BB and MM cluster assignments. The Shore Form allows for one angler or a group of anglers (angler-group) to be interviewed at one time. It is preferred to interview individual anglers, unless their catch is grouped together and cannot be separated. Angler-group data

are recorded in rows (angler-rows) with specific data items arranged by columns. The Shore Form divides each angler-row into three main sections: Effort, Catch, and Bio Data. These sections, along with the header and footer are explained below. Each angler-row has two sub-rows to record two observations for each item in some columns. Catch and Bio Data may span multiple rows and sub-rows as needed to code additional species, fish counts, and measurements. An angler-group may be continued on the next page.

Header Items

The Shore Form header identifies the type of shore mode, assignment, date of the assignment, and the site, cluster and sampler completing the assignment. These items are required for each page header, on all sheets used in the assignment.

The second part of the header is divided into an MM section and a BB section. For BB mode, the MM items are left blank. The BB items include estimated total finfish angler count and observed PR accessed from BB count. This count includes any fin-fishing kayak or personal water craft (PWC) that launched from the beach and bank site.

CRFS SHORE FORM			Імм		вв	(V8 12/2	22/2016)				Fo	r Assignm	ent,	Page	of
ASSN ID	Date (MM/DD/YY)	CNTY SITE			SITE NAME				CLUSTER Sampler#		Sampler Last Name		Name		
MM COUNTS (inst. at least every 1.	5 hr)	Start	Inst#1	Inst #2	Inst #3	Inst #4	Inst #5	Stop		BB Estimate	ed		OBS PR	
Time Count Began		jan									Total Finfis	h		Accessed	
Finfish Anglers		lers									Anglers			From BB	

Effort Items

Individual (or group) angler data fields include sample number, time, total anglers, unlicensed anglers, arrival time, additional time, 12 month avidity, zip code, primary and secondary target species, and water area and gear (for each target).

EFFORT

SAMPLE #	ANGS	TRIP LENGTH	DAYS FISHED	TARGET		
[or R or B]	Total	Arrival Time	12 months	1st	REA	GEAR
Time	(unlic)	Additional Time hrs min	Zip Code	2nd	₹	<u>5</u>
		time	12 mo			
A	()	ADD-hrs ADD-min	zip			

Catch Items

Catch data recorded include any species caught, number of fish by species examined (kept obs), number of fish by species landed unavailable (kept unobs), number of fish by species released alive, and number of fish by species released dead.

Biological Data Collection

For each fish examined, lengths, weights and sex when appropriate are recorded in the bio data section. Also head tag numbers for specimens collected (salmon, White Seabass, Yelloweye Rockfish) are recorded here. There are no required items in the Bio Data section. However, it is important to gather as much finfish biological data (length, weight and sex) as possible.

CATCH			BIO DATA						
	KEPT	RELS	Fork length/Carapace size (mm), Sex (M/F/T)						
SPECIES	obs	alive	Weight (decimal kg) or (tag #						
CODE	unobs	dead	1	2	3	4	5		
	obs	alive							
	unobs	dead							

Shore Form Item by Item Instructions (BB Mode)

HEADER □MM □BB Check the box for the assigned survey mode listed in the column AMODE of the Monthly Schedule. Only required to check the box on the first page of the assignment. Page of Enter in sequence the form and the total number of pages for the assignment. Enter page info on all pages. ASSN ID Enter the six digit assignment ID number on all pages. The ASSN ID is listed on the Monthly Schedule and is unique to each CRFS assignment. CRFS assignment. CRFS assignment. Example: Page 2 of 1		Shore Form Item by Item Instructions (BB Mode)									
HEADER □MM □BB Check the box for the assigned survey mode listed in the column AMODE of the Monthly Schedule. Only required to check the box on the first page of the assignment. Page of Enter in sequence the page number of the form and the total number of pages for the assignment. Enter page info on all pages. ASSN ID Enter the six digit assignment ID number on all pages. The ASSN ID is listed on the Monthly Schedule and is unique to each CRFS assignment. CRFS assignment. ASSN ID is isted on the Monthly Schedule and is unique to each CRFS assignment. CRFS assignment. District from 1-6; NN the sequence number generated by the draw 5=PR1 generated by the draw 5=PR1 generated by the draw 5=PR1 generated by CSP 6=PCD 9=Opportunistic PCE Example: 076103 is from July, District 6, BB assn, third assn randomly chosen by draw Date Enter the date the Format is MM/DD/YY	Field Name	Instructions	Coding Examples and								
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	ASSN ID	assignment ID number on all pages. The ASSN ID is listed on the Monthly Schedule and is unique to each	format MMDNNN where MM is the calendar month ranging from 01-12; D is the CRFS District from 1-6; NNN is the sequence numbers generated by the draw from 101 to 999 where the first N digit is the mode: 1=BB 2=MM 3=PR2 4=PR1 generated by the draw 5=PR1 generated by OSP 6=PCO 7=PCD 9=Opportunistic PCD Example: 076103 is from July, District 6, a BB assn, third assn randomly chosen by the draw								
assignment was Example: 07/01/17 = completed on all July 1, 2017	Date	assignment was	Format is MM/DD/YY Example: 07/01/17 =								

Field Name	Instructions	Coding Examples and Formats
	pages.	
CNTY	Enter the 3 digit numeric county code. Must be entered on all pages. County codes are listed on the Site List.	Example: 053 = Monterey County
SITE	Enter the 3 digit numeric site code. Must be entered on all pages. Site codes are listed on the Site List.	Example: 205 = Zmudowski State Beach
SITE NAME	Enter the correct name of the BB site which can be found on the Site List. Must be entered on all pages.	Example: Zmudowski State Beach
CLUSTER	Enter the Cluster code/name which can be found on the Site List. Must be entered on all pages.	Example: CEN5 This is the fifth BB cluster in Central District (District 3)
Sampler #	Enter your 3 digit numeric Sampler identification number on all pages.	Example: 207 = Jayna Da Silva
Sampler Last Name	Write your last name completely and legibly on all pages.	Example: Da Silva
	MM COUNTS	
Time Count Began – Start	Leave blank for BB.	
Finfish Anglers – Start	Leave blank for BB.	
Time Count Began – Inst#1-5	Leave blank for BB.	
Finfish Anglers – Inst #1-5	Leave blank for BB.	
Time Count Began – Stop	Leave blank for BB.	
Finfish Anglers – Stop	Leave blank for BB.	
BB Estimated Total Finfish Anglers	Enter the total estimated finfish anglers present at the site while you are there. Tally and sum all the anglers at each access point and add	Example: 3 = three BB finfish anglers

Field Name	Instructions	Coding Examples and Formats
	any additional anglers who start fishing.	
OBS PR Accessed From BB	Enter the total number of fin fishing kayaks or personal water craft (PWC) that accessed the ocean from the BB site you are sampling. Kayak or PWC anglers may not be interviewed as they are PR mode.	Example: 3 = three fin fishing kayaks were observed on site
	EFFORT	
SAMPLE # [orR orB]	Record BB sample numbers in consecutive order starting with 1 for every angler or anglergroups interviewed during a visit to a site.	Sample # = 1, meaning the first interview collected at that site during that visit Refusal = R Language Barrier = B
	Refusals: anglers who refuse to participate, record an R without a sample number. Barriers: anglers who cannot participate due to a language barrier, record a B without a sample number.	Sample flags: T = tournament Record a T after the sample number if the anglers are part of a fishing tournament Example: 3T = the third interview at the site and the angler(s) participated in a fishing
Time	Enter the time for each interview attempted. Refusals and barriers should have a time recored.	tournament Format is military time (24-hr format) Example: 1730 = 5:30 pm
ANGS Total	Enter the total number of anglers who contributed to the bag of the interview you are conducting. Refusals and Barriers: Count the number of angler(s) in the group	Example: 3 = three anglers contributed to the bag Refused or Barrier: Code an R or B in the Sample # field, respectively, and terminate the interview;

Field Name	Instructions	Coding Examples and Formats
	and report that number. Terminate the interview.	include the total number of anglers
ANGS (unlic)	Enter the number of anglers out of the total contributors to the bag who do NOT have a	Example: 0 = All anglers in the bag were licensed
	current CA fishing license.	Refused or Barrier: Code an R or B in the Sample # field,
	Note that "unlicensed anglers" field is a subset of the total	respectively, and terminate the interview
	anglers, therefore unlicensed ≤ total anglers.	This is not a required field for MM sampling; it is required for BB sampling
Arrival Time	Enter the time that the angler(s) in the bag arrived at the site.	Format is military time (24-hr format). Example: 1730 = 5:30 pm
		Refused or Barrier: Code an R or B in the Sample # field, respectively, and terminate the interview
Additional Time – ADD-hrs	Enter the number of whole hours that the angler(s) in the bag plan to continue to fish.	Complete-trip: 0, meaning they have no additional hours and are done fishing for the day
	Note: for an incomplete BB interview to be considered valid, the	Incomplete trip example: 2, means they intend to fish for 2 more hours
	angler(s) must have been fishing for at least 30 minutes.	Refused or Barrier: Code an R or B in the Sample # field, respectively, and terminate the interview
Additional Time – ADD-min	Enter the number of whole minutes that the angler(s) in the bag plan to continue to fish.	Complete-trip: 0, meaning they have no additional minutes and are done fishing for the day

Field Name	Instructions	Coding Examples and Formats
		Incomplete trip example: 30, means they intend to fish for 30 more minutes Refused or Barrier:
		Code an R or B in the Sample # field, respectively, and terminate the interview.
DAYS FISHED 12 months	Ask the angler, or a random angler contributing to the bag, about their recreational fishing avidity for the past 12 months. Record whole number of days fished.	Example: 52 = not counting today, the angler went saltwater sport fin-fishing in CA or from a boat leaving from CA 52 days in the past 12 months, or about once per week Refused = R Don't know = DK Sampler didn't ask = DA
Zip Code	Record the five digit numeric zip code residence of the angler, or a random angler contributing to the bag. You may select the same angler who answered the avidity question.	Example: 90210 = Beverly Hills Refused = R Don't know = DK Sampler didn't ask = DA Foreign Country = use the 3 letter country code; Example: Ireland = FIE
TARGET – 1 st	Record the five letter species code of the primary target sought for the angler, or group of anglers. Anglers who do not have a specific target may be coded to unidentified fish.	Example: LNGCD = targeting lingcod UNIFH = "Anything" target or unspecified target. Refused or Barrier: Code an R or B in the Sample # field, respectively, and terminate the interview.
TARGET – 2 nd	Record the five letter species code of the secondary target sought for the angler, or group of anglers.	Example: ABALO = targeting abalone Blank = no secondary target

Field Name	Instructions	Coding Examples and Formats
AREA	You may need to probe for secondary targets. However, a secondary target is not required, may leave blank. Record the water area	N = Nearshore (Ocean
	where the majority of fishing effort occurred by primary and secondary target.	< 3 miles out) B = Enclosed bay, estuary, or harbor
GEAR	Record the fishing gear type used by primary and secondary target.	H = Hook and line S = Spear N = Bait net Invert gear only: Pn = Pot and number of pots used Fn = Flat hoop net and number of nets used Rn = Rigid hoop net and number of nets used E = Snare C = SCUBA diving (by hand) D = Free diving (by hand)
	CATCH	
SPECIES	Record the five letter species code for each species or taxon caught. Use additional rows for angler-bags with multiple catch species.	Example: HALCA = California halibut NOCATCH = nothing was caught Refused or Barrier: Code an R or B in the Sample # field, respectively, and terminate the interview
KEPT obs	Enter the whole number of fish by species that were retained in the bag by the angler(s) and	Example: 5 = five specimens of this species were examined and counted

Field Name	Instructions	Coding Examples and Formats
	examined by the Sampler. Only fish/inverts that the Sampler is able to see and count are recorded here. May include fillets that can be counted and identified to species. Fillets that can't be identified to species should be recorded in kept unobs. Make a note on the form confirming when overlimits are recorded.	0 = no catch of any species examined for this bag Refused: Try to get angler-reported catch if no fish are allowed to be examined Barrier: Record a B in the Sample # field and terminate the interview
KEPT unobs	Enter the number of fish by species that were retained in the bag by the angler(s) but not examined by the Sampler. This includes any fish/inverts that the Sampler is not able to see, identify, or count. This includes fish given away, packed away on ice, thrown away, fillets that are not identifiable or countable, or used for bait. Probe for catch that may not be remembered such as bait species. Make a note on the form confirming when overlimits are recorded.	Example: 5 = five specimens of this species were reported by the angler as kept 0 = no unavailable catch of any species for this bag Refused or Barrier: Code an R or B in the Sample # field, respectively, and terminate the interview
RELS alive	Enter the whole number of fish by species reported as released alive by the angler(s) contributing to this bag. This	Example: 3 = three specimens of this species were reported by the angler as released with no mortal injuries

Field Name	Instructions	Coding Examples and Formats
	includes fish released with and without a descending device. Fish must have been landed and intentionally released. Probe for catch that may not be remembered. "Alive" means the fish was not dead upon release, did not have significant wounds, and swam away after release.	0 = no fish released alive Refused or Barrier: Code an R or B in the Sample # field, respectively, and terminate the interview
RELS dead	Enter the whole number of fish by species reported as released dead by the angler(s) contributing to this bag. Fish must have been landed and intentionally released. Probe for catch that may not be remembered. "Dead" means the fish was not moving upon release, had significant wounds, and could not swim	Example: 2 = two specimens of this species were reported by the angler as released with mortal injuries, were floating, and/or unable to swim away 0 = no fish released dead Refused or Barrier: Code an R or B in the Sample # field, respectively, and
	away after release. BIO DATA	terminate the interview
Fork length (mm), Sex (M/F/T)	In the top box, enter the catch species' fork length.	Example: 321 = 321 mm fork length. If no length can be obtained, leave blank
GEX (IVI/F/1)	Record M, F, or T after the length for sexed species. Do not collect lengths from salmonids with an intact adipose fin. Make a note on the	F = female M = male T = transitional If no sex can be obtained, omit sex Example: 443F = the fish was 443 mm and a female

Field Name	Instructions	Coding Examples and Formats
	form confirming when sublegal fish are recorded.	
Weight (decimal kg) or Tag#	In the box below the length, enter the catch species' weight in kg. Do not weigh filleted, beheaded, or gutted fish. You may weigh bled fish For salmon, enter the 5 digit numeric headtag number below the length in place of the weight and circle the tag number. Do not weigh the salmon	Example: 5.3 = 5.3 kg weight If no weight can be obtained, leave blank Example: (12345) = tagged salmon where head was collected

Shore Form Example (BB Mode)

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Man-Made Structure (MM) Mode Sampling

MM Mode Definition

Man-made mode is defined as a human-made structure where recreational fishing occurs. Man-made (MM) structures include piers, jetties, bridges, docks or other similar structures.

MM Survey Goals

The primary goals for MM sampling are to estimate catch per unit of effort (CPUE) as catch per angler trip and fishing effort as the number of angler trips. CPUE is derived from counts of fish by species and catch type (sampler observed kept, and angler reported kept, released alive and released dead) and from the number of anglers in the interviewed fishing parties. Fishing effort is based on the counts of anglers while on site and the duration of the fishing trip for each angling party. Other relevant data collected by the MM survey include area fished, gear type, target(s), fish length and weight measurements. The goal of the Sampler while on an MM assignment is to obtain as many high-quality interviews from as many MM anglers as possible and to obtain accurate counts of anglers fishing for finfish.

MM Survey Methods

MM sites are grouped into clusters. The number of sites in a cluster will vary but the Sampler must visit all sites within the cluster for the assignment to be considered complete. The sites within a cluster are defined by a site list which will be provided by the Lead via the Monthly Schedule. The number of sites or if they are active/inactive may depend on the season and/or the geographic proximity among sites. The cluster/site list changes and is unique by month. The Sampler is to use the cluster/site list that matches the month of the sample selection included in the monthly schedule.

The MM sample draw takes into consideration the overall effort of the MM cluster. Clusters with high effort have a higher probability of being drawn for sampling than those with lower effort. Not all MM clusters will be sampled every month. The site list will designate the site labels (e.g. A, B, C...) for sites within a cluster. Sites within a cluster will be sampled in a predetermined order. The starting site will be randomly selected by the draw program. A Sampler will begin their MM sample day by consulting the Monthly Schedule, which will provide the start time (early or late) and the first site to visit. Leads will set the times for early and late start times each month based on knowledge of the fisheries and the daylight hours available. The Sampler is to rove through the sites in alphabetical order or in a manner designated by the Lead. All of the MM sites are public access sites so you should be able to access them without problems.

Example MM Clusters from Site List

DISTRICT	AMODE	CLUS	SUBSITE	TMODE	CNTY	SITE	MONTH	YEAR	NAME
5	MM	WIN3	A	MM	45	204	3	2014	Fort Bragg
5	MM	WIN3	В	MM	45	103	3	2014	Point Arena
6	MM	RED2	A	MM	23	211	3	2014	South Spit
6	MM	RED2	В	MM	23	105	3	2014	King Salmon: Rock Fingers
6	MM	RED2	С	MM	23	305	3	2014	Del Norte St Pier
6	MM	RED4	A	MM	23	307	3	2014	Trinidad Pier
6	MM	RED4	В	MM	23	210	3	2014	North Jetty Eureka
6	MM	RED6	A	MM	15	300	3	2014	North Jetty Crescent City
6	MM	RED6	В	MM	15	314	3	2014	B Street Pier
6	MM	RED6	C	MM	15	302	3	2014	Citizen's Dock
6	MM	RED6	D	MM	15	303	3	2014	South Jetty Crescent City

The Sampler is to contact the Lead immediately if they cannot complete an assignment due to illness or an emergency. For the proper implementation of statistical methods, it is crucial for statistical methods that Samplers try to complete all assignments as scheduled. As with any mode, rescheduling of MM assignments is not desirable to the survey. If necessary, the Lead can reschedule an MM assignment. Leads must conserve the effort category, kind of day (KOD), start time and cluster order when rescheduling MM assignments – if there are no available KODs during the remainder of the month the Lead will cancel the assignment.

Sampling will normally take place within an eight hour work day during daylight hours. The Sampler is to strive for six hours of sampling time and allow up to two hours for travel time while on assignment. A Sampler is to try and avoid working over an eight hour day for MM assignments. Interviews of MM anglers (or angler parties) are recorded on the CRFS Shore Form. The Sampler will obtain accurate start, stop and instantaneous counts of finfish anglers while on site. In addition, the Sampler may perform pressure checks at adjacent sites using the Assignment Summary Form and may also perform CPFV checks at adjacent PC sites using the PC Effort Check Form (PEC).

The Sampler may use a site map binder or the CRFS Wiki site for driving directions and site boundaries. After visiting the first site in the cluster, the Sampler should move through the assigned order of sites looking for angler activity and keeping detailed data records. After visiting all sites in the prescribed order for the MM cluster, the Sampler may return to any previous sites where they expect to obtain interviews. All of the sites within a cluster must be visited in order for the assignment to be considered complete. A Sampler is to notify the Lead immediately if they are not able to complete an MM assignment by visiting all sites within the assigned MM cluster.

The general rule is to stay at a site where it is expected to get one interview per hour. If the Sampler does not expect to obtain at least one interview per hour, they should move to the next site in the cluster. In an effort to obtain as many valid and high-quality MM interviews as possible, it may be necessary to stay at certain sites where there is high angler activity. The Sampler should take up a strategic position so they can intercept a majority of the anglers. Samplers are to avoid surveying only at cleaning stations on wharfs or piers

as this will bias data towards successful anglers. Close observation of fishing activity is required, especially at crowed piers where the Sampler must be alert to any anglers leaving the site. Each MM cluster is unique and new Samplers will be trained on the best way to sample any specific MM site.

MM Angler Counts

An important aspect of MM sampling is obtaining accurate counts of finfish anglers by site. The survey uses start, stop and instantaneous counts to collect effort data. A start count is performed upon your initial arrival to the site. Instantaneous counts are done while on site; the Sampler will stop actively sampling and conduct a count. These angler counts may not happen instantly; they may take from a few minutes to half an hour to complete (depending on the size of the site). Instantaneous counts must be completed every 1.5 hours or less while onsite. Finally, a stop count is conducted when sampling ceases but before incomplete-trip interviews are attempted.

For MM angler counts the Sampler is to count finfish anglers only. A finfish angler is defined as an angler that has wet gear hours and has or is targeting finfish during the survey day or has the immediate intent to finfish. This includes anglers taking a break, re-baiting or moving between locations within the site. Invertebrate-only anglers are NOT included in MM angler counts. The Sampler should begin the count at the far end of the MM structure and count as they return to the origin. The origin is where a Sampler can see all people leaving the structure or a constriction point all anglers must pass. It is important for the Sampler to try not to double count or miss anglers behind obstructions. Oftentimes it is difficult to determine the number of anglers when there are multiple fishing rods so the Sampler must use their best judgment. If it is too difficult or dangerous to walk on a jetty or other structure, it is recommended to count finfish anglers using binoculars. The Sampler will use local knowledge and their discretion to determine the activity of those inaccessible anglers and only count anglers believed to be targeting finfish. The Sampler is to write a note on the ASF with the count of anglers and the proportion that you used. The time the angler count began is always recorded on the ASE and in the header of the Shore Form.

Canvassing

A useful tactic for sampling in MM mode is to complete a preliminary canvass to determine the number and location of anglers at a site and a rough approximation of the duration of their trips. With this information the Sampler is able to maximize intercept coverage by planning their movements around those of the anglers. It may give the Sampler a good indication of when to stay onsite and when to move to the next site in the cluster.

Anglers Fishing in Two Modes

When interviewing an angler who has been fishing in two different modes (i.e. BB mode and MM mode), ask the angler where they have spent most of their time fishing. The angler is eligible for an interview if they have spent more than half of their fishing trip in the MM mode. The Sampler will collect only the information which pertains to the angler's time on the MM, i.e. catch,

target(s), gear, area. The angler is ineligible for an interview if they have spent less than half of their time fishing in the MM mode.

To accurately portray the angler's time fishing in the MM mode, the Sampler will need to adjust the angler's arrival time. The Sampler will ask the angler how long they spent fishing on the MM and will calculate the angler's arrival time by taking the interview time and subtracting the angler's total time fishing in MM mode to create the estimated arrival time.

When completing instantaneous start or stop counts, if the angler is fishing on the MM during the time of your count, include them in the count. If they are not fishing on the MM during the instantaneous count they are NOT included. Do not adjust counts if the angler reports they have spent most of their time fishing in another mode.

Incomplete MM trips

While complete-trip interviews are preferred, MM mode sampling protocol allows the Sampler to interview anglers who have not yet completed their fishing trips. Incomplete trips should only be conducted for anglers targeting finfish. Anglers in MM mode <u>must</u> be at least halfway done with their fishing trip to be interviewed. Incomplete-trips are allowed in an effort to get as much MM data as possible. Incomplete-trips are adjusted based on the catch rates for the time fished to account for additional fishing time. The Sampler may ONLY get incomplete-trip interviews after the site's stop count.

Before getting an incomplete-trip interview, canvass the angler(s) to determine if the Sampler should stay on site to get the interview or return to the site later. The Sampler may encounter the same angler(s) again if you return to the site after visiting all of the other sites in the cluster. When this happens, the Sampler is to attempt to update the interview. The Sampler should update the interview by copying the relevant information onto the Shore Form for the current visit to the site and deleting the interview from the Shore Form for the previous visit to the site.

Do not conduct incomplete-trip interviews of invertebrate-only anglers without incidental finfish by catch.

Low Effort Protocol

The general sampling guideline for clusters is to strive to obtain at least one interview per hour. If the Sampler cannot do such, they are to move to the next site in the cluster, then return to a previous site in the cluster if they have gone through all sites, or terminate the assignment. If there is low effort at a MM site, the Sampler is to canvass the angler(s) and determine the duration of their trip. The Sampler may decide to wait for them to complete their trip. The Sampler should continue to rove from site to site in the cluster in order until the day's fishing activity has ceased or the Sampler has worked to the limit of six sample hours. Other reasons to leave the assignment early would be if the site is unsafe, darkness, or extreme weather conditions.

No Anglers in MM Mode

The ASF and Shore Form header information, start/stop counts, and times must be completed for each visit to a site, even if there are no finfish anglers present. After determining there are no MM anglers at the first site, the Sampler should go immediately to the next site in the order. If no anglers are present at the next site, the Sampler should go immediately to the next site in the order. The Sampler is to keep searching for anglers by roving through sites for up to two hours. If the Sampler does not find any MM anglers after roving through all sites in order and two hours of sampling time has passed and effort does not seem likely to develop, they may terminate the assignment; the assignment is complete. Knowing that the cluster had zero effort for that day is important.

Opportunistic PC Sampling

It is possible to sample PC boats during MM sampling. For instance, the Sampler may encounter PC boats at large piers. However, they may not leave the site or miss any MM interviews to complete an opportunistic PC sample.

Screening Divers

The Sampler is to be aware that divers at a dock, pier or jetty may be fishing If a diver used or intended to use a spear gun, they can be interviewed as eligible anglers (gear = S). Divers entering the water from the MM structure using fins and a flotation device (such as a dive tube) to fish are considered MM anglers. Spearfishers using kayaks or personal water craft are PR anglers and may not be interviewed while the Sampler is sampling a MM assignment.

Invertebrate Sampling in MM Mode

The goal of MM sampling is to collect information on finfish effort and catch. However, man-made structures are often found to be popular places for anglers to target invertebrates. The Sampler must be aware of invertebrate-only anglers to avoid including them in their MM angler counts. Especially on man-made structures it can be difficult to determine fishing targets since at first glance it can appear the anglers are using the hook and line gear type. It is common for invertebrate gear to be fitted on a standard fishing pole, which can make it easy for the Sampler to miscount the angler as targeting finfish. Therefore, the Sampler should pay attention to what anglers are reeling in or ask their intended target.

Sometimes the Sampler may find an angler targeting both finfish and invertebrates. These anglers, of course, qualify for a CRFS interview and are included in the effort count. For the invertebrate component of the catch the Sampler will record all retained invertebrates as kept unobserved – do not count or collect biological data from retained invertebrates. Anglers targeting invertebrates only could have incidental catch of finfish; in this case record UNIFH as the secondary target. The Sampler should not miss interviews of finfish anglers while sampling anglers with invertebrate only

targets. Do not interview invertebrate only anglers after the stop count (incomplete trip interviews).

2017 CRFS MM Mode Questionnaire

The wording of the questions (i.e. script) has been structured to capture the required information for this survey in an efficient and thorough manner. You will be provided with a laminated copy of the MM questionnaire. It is important that you use the wording of questions as stated in the MM script since slight changes in wording can result in different responses. You will be canvassing, screening, introducing the survey, and providing the Privacy Act information. After screening for angler-eligibility you will introduce the survey to the angler(s) to be sampled by saying:

SCREENING: Have you completed a saltwater sport fin-fishing trip today?

Yes: Go to next.

No: If after the stop count and has completed at least 50% of the

anticipated fishing trip, go to next

No: If not complete and not at least 50% done with the trip after the stop

count = Ineligible; stop interview.

Refused: Code Sample # as R, record the number of anglers in the group,

terminate interview.

"Hello, my name is _____ and I represent CDFW. We are interviewing marine recreational anglers for the California Recreational Fisheries Survey. May I ask you a few questions about your fishing trip?"

The Sampler will state the Privacy Act by saying, "This study is being conducted in accordance with the Privacy Act of 1974. You are not required to answer any question that you consider to be an invasion of your privacy."

ANGS Total: How many of you had gear in the water?

Enter the total number of anglers that fished together. Go to next.

Refused: Code Sample # as R, terminate interview.

Unlicensed: What type of sport fishing license does each of you have? Enter the number of the ANGS (above) who fished without a current California sport fishing license. Go to next.

Refused: Code R in (unlic) and continue interview

TRIP LENGTH: At what time did you arrive at the fishing site today? Enter the time in 24 hr format when the angler reported arriving at the site. Go to next.

Refused: Code Sample # as R, terminate interview.

If incomplete-trip but fished for at least 50%: How many additional hours and minutes do you plan to fish here today?

Enter the number of additional hours and minutes the angler intends to fish. Go to next.

Refused: Code Sample # as R, terminate interview.

DAYS FISHED 12 months: Ask a random angler in the group. **Not** counting today, within the past 12 months, how many days have you gone saltwater sport finfishing in this state or from a boat launched in this state?

Refused: Code R, Don't know: Code DK

Sampler didn't ask: Code DA

ZIP CODE: Ask a randomangler in the group. What is the ZIP code of your residence? If ZIP unknown, ask What city or town do you live in?

Refused: Code R, Don't know: Code DK

Sampler didn't ask: Code DA

Non-U.S. resident: Code Foreign Country

PRIMARY TARGET: What type of fish were you primarily trying to

catch? Code the taxon of the angler's primary target.

Anything: Code UNIFH

Refused: Code Sample # as R, terminate interview.

SECONDARY TARGET: What type of fish were you secondarily trying

to catch? Code the taxon of the angler's secondary target.

Anything: Code UNIFH

No secondary target: Leave blank

EFFORT AREA: Was your cprimary target> fishing in the ocean or enclosed bay/estuary/harbor?

Nearshore (< 3 miles): Code N

Bay/Estuary/Harbor: Code B. Be aware of freshwater cutoffs.

Refused: Code Sample # as R, terminate interview

EFFORT AREA: Was your <secondary target> fishing in the ocean or enclosed bay/estuary/harbor?

Nearshore (< 3 miles): Code N

Bay/Estuary/Harbor: Code B. Be aware of freshwater cutoffs.

Refused: Code Sample # as R, terminate interview

GEAR: What gear type did you use for <pri>primary target>?

Finfish

Hook & Line: Code H Spear: Code S Bait Net: Code N

Shellfish Pot #: Code Pn

Flat Hoop Net #: Code Fn Rigid Hoop Net #: Code Rn

Snare: Code F

SCUBA: Code C Free Diving: Code D

GEAR: What gear did you use for <secondary target>?

Finfish

Hook & Line: Code H Spear: Code S Bait Net: Code N

Shellfish

Pot #: Code Pn

Flat Hoop Net #: Code Fn Rigid Hoop Net #: Code Rn

Snare: Code E SCUBA: Code C Free Diving: Code D

SPECIES CODE: Did you catch any fish today?

Yes: Record Species Code and go to next.

No: Record "No Catch" in Species Code box and zeros in KEPT obs, KEPT

unobs, RELS alive total and RELS dead.

Refused: Code Sample # as R, terminate interview.

KEPT OBSERVED: May I see the catch?

Yes: Sampler will identify and count all fish by species.

No: Enter zero and code numbers of all fish as Kept Unobserved.

Fillets: If fillets can't be IDed by skin, enter zero and code numbers of Kept

Unobserved.

Refused to let you see fish or tell you the types and numbers of fish kept (i.e., kept unobserved): Code Sample # as R, terminate interview.

KEPT UNOBSERVED: **Did you retain any other fish?** Probe for any fish given away, filleted, used for bait or thrown away.

Yes: Record species and number of fish.

No: Enter zeroes in Kept Unobserved boxes for all Kept Observed species recorded.

Refused: Code Sample # as R, terminate interview. Don't Know: Code Sample # as R, terminate interview.

RELEASED ALIVE: **Were any fish released alive?** Probe for any fish that were purposely released alive (swam away).

Yes: Record species and number of fish.

No: Enter zeroes in Released Alive Total boxes for all Kept Observed or

Unobserved species recorded.

Refused: Code Sample # as R, terminate interview. Don't Know: Code Sample # as R, terminate interview.

RELEASED DEAD: **Were any fish released dead?** Probe for any fish that were thrown back dead (did not swim away).

Yes: Record species and number of fish.

No: Enter zeroes in Released Dead boxes for species recorded Kept

Observed or Unobserved.

Refused: Code Sample # as R, terminate interview. Don't Know: Code Sample # as R, terminate interview.

BIO DATA: May I measure your kept fish today?

FORK LENGTH (MM) SEX (M/F/T): Enter the fork length in millimeters of each fish measured above the dotted line. Add a suffix of M (male), F (female) or T (transitional) for each sexed fish.

WEIGHT (DECIMAL KG) OR HEAD TAG # (CIRCLE TAG #): Enter the weight in kilograms of the fish below the length. Do not record a weight without a length. Do not weigh salmon. Enter the head tag number for an adipose fin-clipped salmon below the length in place of the weight. If the salmon head is lost or refused write NRS after the tag number. For collected Yelloweye Rockfish and White Seabass, enter a length and weight and put the head tag number or scan code to the right of the measurements. Circle salmon and yelloweye head tag numbers.

Shore Form Procedures for MM Mode

This section describes specific procedures for MM surveys using the Shore Form. The Shore Form is used for shore modes only (MM and BB).

Numbering the Visits, Interviews and Shore Form Pages

Each MM site visited should be recorded on the ASF in the order visited. The Sampler must start a new Shore Form page when they arrive at a site. The first interview at a site will be "1" and all other interviews at that site will be numbered sequentially. At the next site, the Sampler must start a new Shore Form page and begin renumbering interviews with "1" again. If the Sampler visits the same site more than one time in a day, a new page will still be used for each visit, and the interview numbering will start with "1" at each visit. An example is shown below.

First site in the cluster

- Site 1 on ASF
- Start a new Shore Form page
- Number interviews sequentially starting with 1

Second site in the cluster

- Site 2 on ASF
- Start a new Shore Form page
- Number interviews sequentially starting with 1

Third and last site in the cluster

- Site 3 on ASF
- Start a new Shore Form page
- Number interviews sequentially starting with 1

Return to site 2

- Site 4 on ASF
- Start a new Shore Form page
- Number interviews sequentially starting with 1

Shore Form page numbers will start at "1" and be numbered sequentially for the entire assignment.

Shore Form Layout

The Shore Form is used for both BB and MM cluster assignments. The Shore Form allows for one angler or a group of anglers (angler-group) to be interviewed at one time. It is preferred to interview individual anglers, unless their catch is grouped together and cannot be separated. Angler-group data are recorded in rows (angler-rows) with specific data items arranged by columns. The Shore Form divides each angler-row into three main sections: Effort, Catch, and Bio Data. These sections, along with the header and footer are explained below. Each angler-row has two sub-rows to record two observations for each item in some columns. Catch and Bio Data may span multiple rows and sub-rows as needed to code additional species, fish counts, and measurements. An angler-group may be continued on the next page.

Header Items

The Shore Form header identifies the type of shore mode, assignment ID, number, number of sites visited, date of the assignment, county, site, cluster, and Sampler completing the assignment. These items are required for each page header, on all sheets used in the assignment.

The second part of the header is divided into an MM Counts section and a BB Counts section. For MM mode, the BB items are left blank. The MM items include angler counts and the times each count was conducted.



Effort Items

Individual (or group) angler data fields include sample number, time, total anglers, unlicensed anglers, arrival time, additional time, 12 month avidity, zip code, primary and secondary target species, and water area and gear (for each target).

EFFORT

SAMPLE #	ANGS	TRIP LI	ENGTH	DAYS FISHED	TARGET		
[or R or B]	Total		I Time	12 months	1st	REA	GEAR
Time	(unlic)	Addition hrs	nal Time min	Zip Code	2nd	₹	<u>5</u>
			time	12 mo			
А	()	ADD-hrs	ADD-min	zip			

Catch Items

Catch data recorded include any species caught, number of fish by species examined (kept obs), number of fish by species landed unavailable (kept unobs), number of fish by species released alive, number of fish by species released dead.

Biological Data Collection

For each fish examined, lengths, weights and sex when appropriate are recorded in the bio data section. Also head tag numbers for specimens collected (salmon, White Seabass, Yelloweye Rockfish) are recorded here. There are no required items in the Bio Data section. However, it is important to gather as much biological data (length, weight and sex) as possible.

CATCH BIO DATA

	KEPT	RELS	Fo	rk length/Car	rapace size (m	nm), Sex (M/F	/T)
SPECIES	obs	alive		Weight	(decimal kg)	or (tag#)	
CODE	unobs	dead	1	2	3	4	5
	obs	alive					
	unobs	dead					

Shore Form Item by Item Instructions (MM Mode)

Field Name	Instructions	Coding Examples and Formats
	HEADER	
□мм □вв	Check the box for the assigned survey mode listed in the column AMODE of the Monthly Schedule.	⊠мм
Page of	Enter in sequence the page number of the form and the total number of pages for the assignment. Enter page info on all pages.	Example: Page 2 of 7

Field Name	Instructions	Coding Examples and Formats
ASSN ID	Enter the six digit assignment ID number on all pages. The ASSN ID is listed on the Monthly Schedule and is unique to each CRFS assignment.	ASSN ID is in the format MMDNNN where MM is the calendar month ranging from 01-12; D is the CRFS District from 1-6; NNN is the sequence numbers generated by the draw from 101 to 999 where the first N digit is the mode: 1=BB 2=MM 3=PR2 4=PR1 generated by the draw 5=PR1 generated by OSP 6=PCO 7=PCD 9=Opportunistic PCD Example: 076203 is from July, District 6, a MM assn, third assn randomly chosen by the draw
Site Visit of	Each site sampled in the cluster requires a new Shore Form and a unique Site Visit number, starting with 1. Enter in chronological order the Site Visit number and the total number of site visits for the cluster. Enter Site Visit info on all pages.	(Visit#) of (Total Site Visits in cluster assn) Site Visit 3 of 6
Date	Enter the date the assignment was completed on all pages.	Format is MM/DD/YY Example: 07/01/15 = July 1, 2015

Field Name	Instructions	Coding Examples and Formats
CNTY	Enter the 3 digit numeric county code. Must be entered on all pages. County codes are listed on the Site List.	Example: 023 = Humboldt County
SITE	Enter the 3 digit numeric site code. Must be entered on all pages. Site codes are listed on the Site List.	Example: 211 = South Spit
SITE NAME	Enter the correct name of the MM site which can be found on the Site List. Must be entered on all pages.	Example: South Spit
CLUSTER	Enter the Cluster code/name which can be found on the Site List. Must be entered on all pages.	Example: RED3 This is the third MM cluster in Redwood District (District 6)
Sampler #	Enter your 3 digit numeric Sampler identification number on all pages.	Example: 301 = Marc Heisdorf
Sampler Last Name	Write your last name completely and legibly on all pages.	Example: Heisdorf
	MM COUNTS	
Time Count Began – Start	Enter the time in which you begin the MM start count.	Format is military time (24-hr format). Example: 1300 = 1:00 pm
Finfish Anglers – Start	Enter the number of finfish anglers at the site. Omit invert-only anglers from the count.	Enter positive whole number of anglers. Example: 3 = three MM finfish anglers
Time Count Began – Inst #1	Enter the time in which you begin the first instantaneous count. This should be done within 1.5 hrs of the start count.	Format is military time (24-hr format). Example: 1430 = 2:30 pm
Finfish Anglers – Inst #1	Enter the number of finfish anglers at the	Enter positive whole number of anglers.

Field Name	Instructions	Coding Examples and Formats
	site. Omit invert-only anglers from the count. Instantaneous counts are not required if you leave the site 1.5 hours or less after arriving; in this case there would only be a start and stop count.	Example: 6 = six MM finfish anglers
Time Count Began – Inst#2	Enter the time in which you begin the second instantaneous count. This should be done within 1.5 hrs of the previous count.	Format is military time (24-hr format) Example: 1600 = 4:00 pm
Finfish Anglers – Inst #2	Enter the number of finfish anglers at the site. Omit invert-only anglers from the count. Instantaneous counts are not required if you leave the site 1.5 hours after arriving; in this case there would only be a start and stop count.	Enter positive whole number of anglers Example: 8 = eight MM finfish anglers
Time Count Began – Inst#3, Time Count Began – Inst#4, Time Count Began – Inst#5	Continue entering times for each instantaneous count until you are ready to leave the site by doing a stop count (see below).	Format is military time (24-hr format) Example: 1730 = 5:30 pm
Finfish Anglers – Inst #3, Finfish Anglers – Inst #4, Finfish Anglers – Inst #5	Continue entering the number of finfish anglers for each instantaneous count until you are ready to leave the site by doing a stop count (see below).	Enter positive whole number of anglers Example: 0 = zero MM finfish anglers
Time Count Began – Stop	Enter the time in which you begin the MM stop count.	Format is military time (24-hr format). Example: 1800 = 6:00 pm

Instructions	Coding Examples and Formats
Enter the number of finfish anglers at the site. Omit invert-only anglers from the count.	Enter positive whole number of anglers Example: 0 = zero MM finfish anglers
Leave blank for MM sampling.	
Leave blank for MM sampling.	
EFFORT	
Record MM sample numbers in consecutive order starting with 1 for every angler or angler-groups interviewed during a visit to a site.	Sample # = 1, meaning the first interview collected at that site during that visit Refusal = R Language Barrier = B
Refusals: anglers who refuse to participate, record an R without a sample number. Barriers: anglers who cannot participate due to a language barrier, record a B without a sample number.	Sample flags: T = tournament Record a T after the sample number if the anglers are part of a fishing tournament Example: 3T = the third interview at the site and the angler(s) participated in a fishing tournament
Enter the time stamp for each interview attempted. Refusals and barriers should get a time.	Format is military time (24-hr format) Example: 1730 = 5:30 pm
Enter the total number of anglers who contributed to the bag of the interview you are conducting. Refusals and Barriers: Count the number of	Example: 3 = three anglers contributed to the bag Refused or Barrier: Code an R or B in the Sample # field, respectively, and terminate the interview
	Enter the number of finfish anglers at the site. Omit invert-only anglers from the count. Leave blank for MM sampling. Leave blank for MM sampling. EFFORT Record MM sample numbers in consecutive order starting with 1 for every angler or angler-groups interviewed during a visit to a site. Refusals: anglers who refuse to participate, record an R without a sample number. Barriers: anglers who cannot participate due to a language barrier, record a B without a sample number. Enter the time stamp for each interview attempted. Refusals and barriers should get a time. Enter the total number of anglers who contributed to the bag of the interview you are conducting. Refusals and Barriers:

Field Name	Instructions	Coding Examples and Formats
	and report that number. Terminate the interview.	
ANGS (unlic)	Enter the number of anglers out of the total contributors to the bag that do NOT have a current CA fishing license. Note that "unlicensed anglers" field is a subset of the total anglers, therefore unlicensed ≤ total anglers.	Example: 0 = All anglers in the bag were licensed Refused or Barrier: Code an R or B in the Sample # field, respectively, and terminate the interview This is not a required field for MM sampling; it is required for BB sampling
Arrival Time	Enter the time that the angler(s) in the bag arrived at the site.	Format is military time (24-hr format). Example: 1730 = 5:30 pm Refused or Barrier: Code an R or B in the Sample # field, respectively, and terminate the interview
Additional Time – ADD-hrs	Enter the number of whole hours that the angler(s) in the bag plan to continue to fish. Note: for an incomplete MM interview to be considered valid, the angler(s) must have completed at least	Complete-trip: 0, meaning they have no additional hours and are done fishing for the day Incomplete trip example: 2, means they intend to fish for 2 more hours Refused or Barrier:
	50% of the fishing trip at the time of the interview	Code an R or B in the Sample # field, respectively, and terminate the interview

Field Name	Instructions	Coding Examples and Formats
Additional Time – ADD-min	Enter the number of whole minutes that the angler(s) in the bag plan to continue to fish.	Complete-trip: 0, meaning they have no additional minutes and are done fishing for the day
		Incomplete trip example: 30, means they intend to fish for 30 more minutes
		Refused or Barrier: Code an R or B in the Sample # field, respectively, and terminate the interview.
DAYS FISHED 12 months	Ask the angler, or a random angler contributing to the bag, about their recreational fishing avidity for the past 12 months. Record whole number of days fished.	Example: 52 = not counting today, the angler went saltwater sport finfishing in CA or from a boat leaving from CA 52 days in the past 12 months, or about once per week. Refused = R Don't know = DK Sampler didn't ask = DA
Zip Code	Record the five digit numeric zip code residence of the angler, or a random angler contributing to the bag. You may select the same angler who answered the avidity question.	Example: 90210 = Beverly Hills Refused = R Don't know = DK Sampler didn't ask = DA Foreign Country = use the 3 letter country code; Example: Ireland = FIE
TARGET – 1 st	Record the five letter species code of the primary target sought for the angler, or group of anglers. Anglers who do not have a specific target	Example: LNGCD = targeting lingcod UNIFH = "Anything" target or unspecified target

Field Name	Instructions	Coding Examples and Formats
	may be coded to unidentified fish.	Refused or Barrier: Code an R or B in the Sample # field, respectively, and terminate the interview
TARGET – 2 nd	Record the five letter species code of the secondary target sought for the angler, or group of anglers. You may need to probe for secondary targets. However, a secondary targetis not required, may leave blank.	Example: ABALO = targeting abalone Blank = no secondary target
AREA	Record the water area where the majority of fishing effort occurred by primary and secondary target. Note that the AREA of fishing effort and the species catch location may differ.	N = Nearshore (Ocean < 3 miles out) B = Enclosed bay, estuary, or harbor Refused or Barrier: Code an R or B in the Sample # field, respectively, and terminate the interview
GEAR	Record the fishing gear type used by primary and secondary target.	H = Hook and line S = Spear N = Bait net Invert gear only: Pn = Pot and number of pots used Fn = Flat hoop net and number of nets used Rn = Rigid hoop net and number of nets used E = Snare C = SCUBA diving (by hand) D = Free diving (by hand)
	CATCH	
SPECIES	Record the five letter species code for each	Example: HALCA = California Halibut

Field Name	Instructions	Coding Examples and Formats
	species or taxon caught. Use additional rows for angler-bags with multiple catch species.	NOCATCH = nothing was caught Refused or Barrier: Code an R or B in the Sample # field, respectively, and terminate the interview
KEPT obs	Enter the whole number of fish by species that were retained in the bag by the angler(s) and examined by the Sampler. Only fish/inverts that the Sampler is able to see and count are recorded here. May include fillets that can be counted and identified to species. Fillets that can't be identified to species should be recorded in kept unobs. Make a note on the form confirming when overlimits are recorded	Example: 5 = five specimens of this species were examined and counted 0 = no catch of any species examined for this bag Refused: Try to get angler-reported catch if no fish are allowed to be examined Barrier: Record a B in the Sample # field and terminate the interview
KEPT unobs	Enter the whole number of fish by species that were retained in the bag by the angler(s) but not examined by the Sampler. This includes and fish/inverts that the Sampler is not able to see, identify, or count. This includes fish given away, packed away on ice, thrown away, fillets that are not identifiable or	Example: 5 = five specimens of this species were reported by the angler as kept 0 = no unavailable catch of any species for this bag Refused or Barrier: Code an R or B in the Sample # field, respectively, and terminate the intervie

Field Name	Instructions	Coding Examples and Formats
	countable, or used for bait. Probe for catch that may not be remembered such as bait species. Make a note on the form confirming when overlimits are recorded.	
RELS alive	Enter the whole number of fish by species reported as released alive by the angler(s) contributing to this bag. This includes fish released with and without a descending device. Fish must have been landed and intentionally released. Probe for catch that may not be remembered. "Alive" means the fish was not dead upon release, did not have significant wounds, and swam away after release.	Example: 3 = three specimens of this species were reported by the angler as released with no mortal injuries 0 = no fish released alive Refused or Barrier: Code an R or B in the Sample # field, respectively, and terminate the interview
RELS dead	Enter the whole number of fish by species reported as released dead by the angler(s) contributing to this bag. Fish must have been landed and intentionally released. Probe for catch that may not be remembered. "Dead" means the fish was not moving upon release, had significant wounds, and could not swam away after release.	Example: 2 = two specimens of this species were reported by the angler as released with mortal injuries, were floating, and/or unable to swim away 0 = no fish released dead Refused or Barrier: Code an R or B in the Sample # field, respectively, and terminate the interview

Field Name	Instructions	Coding Examples and Formats
	BIO DATA	
Fork length(mm)	In the top box, enter the catch species' fork length or carapace size or mantle length in millimeters.	Example: 321 = 321 mm fork length. If no length can be obtained, leave blank
Sex (M/F/T)	Record M, F, or T after the length for sexed species. Do not collect lengths from salmonids with an intact adipose fin. Make a note on the form confirming when sublegal fish are recorded.	F = female M = male T = transitional If no sex can be obtained, omit sex Example: 443F = the fish was 443 mm and a female
Weight (decimal kg) or Tag#	In the box below the length, enter the catch species' weight in kg. Do not weight filleted, beheaded, or gutted fish. You may weigh bled fish. For salmon, enter the 5 digit numeric headtag number below the length in place of the weight and circle the tag number.	Example: 5.3 = 5.3 kg weight. If no weight can be obtained, leave blan. Example: 12345 = tagged salmon where head was collected Do not weight salmon

Shore Form Example (MM Mode)

No Catch		V12_11262018 SITE NAME	V12_11262018	V12_11262018
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	5	Srban	STBAN	zp dz
	۵	N H N	Ξ	I Z
	9	35548	geoge	93021 ^{zp}

9-22

GEAR: Hook & line, Spear, Bait Net Invert gear only: Pot #, Flat # or Rigid # hoop net, snarE, sCuba, free Dive

Additional Time: Incomplete trips-record angler's estimate of additional time at the site in hours & minutes. Complete trips-record zero in the ADD-hrs & ADD-min boxes.

Arrival Time: Record the time the angler(s) arrived at the site.
Additional Time: Incomplete trips-record angler's estimate of adWATER AREA: Nearshore (<3mi), enclosed Bay/estuary/harbor

Primary Private and Rental Boat (PR1) Mode Sampling

Introduction

Although all fishing modes are sampled, CRFS puts more emphasis on fishing from boats, where the majority of managed fish species are caught compared to other modes. The <u>private and rental boat</u> (PR) mode fishery is the largest in the state in terms of total catch. The PR fishery is also seasonally and geographically irregular. The publicly accessible sites where private and rental boats launch are stratified into primary sites (PR1) and secondary sites (PR2). To divide sites into the two strata (PR1 and PR2), data for "important management species" were analyzed separately for sites north and south of Point Conception. Important management species were defined as those with active fishery management plans and include salmon, groundfish (e.g., rockfishes, Lingcod, Cabezon, California Scorpionfish, flatfishes, and some sharks and rays), highly migratory species (tunas, billfishes, Dolphinfish, and certain oceanic sharks), and species in the California Nearshore Fishery Management Plan.

PR1 sites are defined as publicly accessible launch facilities (e.g., launch ramps, hoists, beach tractors, rental shops) where at least 90 percent of fishing effort and catch of "important management species" by private or rental boats occurs. The PR1 survey estimates total effort and catch for each individual primary site and month. The data from this survey, the secondary survey (PR2) and the telephone survey of licensed anglers (ALDTS for night and private access fishing) is used to make total private and rental boat (PR) effort and catch estimates for the CRFS program. The sampling procedures for PR1 and PR2 are similar, and the same forms will be used for both PR1 and PR2. Differences in the sampling procedures for the two strata are listed in the table below.

Differences between PR1 and PR2 sampling procedures.

Sampling Procedure	PR1	PR2
Time on site when trailers present	From the return of the first boat until the last boat returns or sunset (whichever is first)	Approximately 6 to 7 hours during daylight hours
Scheduled start time	Lead will assign a start time	Early or late start as defined by Lead each month
Count boats launching	No	Yes
Count offsite missed boats	Counted at some PR sites	No

PR1 GOALS

The primary goals for PR assignments are to:

- ✓ Obtain accurate counts of the boats and anglers using the site
- ✓ Obtain high quality interviews and catch data
- ✓ Observe all kept salmon
- ✓ Collect heads of all adipose fin-clipped salmon

Effort Data

The goal is to estimate total fishing effort for the day. This is done by counting trailers and returning boats and determining the number of anglers on each fishing boat. For each boat we must determine the primary activity. If the boat is fishing, we determine the target fish species and the number of anglers per boat. The monthly random sample selects 20% or more of the days each month for each PR1 sample site. Effort is expanded to account for weekend (and holidays) and weekday days not sampled (the remaining 80%). The effort estimate is calculated in angler trips by target fishery group.

Catch Data

An additional goal is to estimate catch per angler-trip. Catch per angler-trip is determined by counting the number of each species that is kept and recording the number of each species that are reported as released or otherwise unavailable to examine. Estimation of effort and estimation of catch per angler-trip are each calculated for a PR1 site, month, kind of day (weekend/weekday), water area, and trip type (target). Effort is calculated as the total number of anglers sampled during the time period, adjusted for un-sampled anglers, and expanded for the total available weekend or weekdays per month. Catch rate per angler-trip is calculated from the sum of catch recorded from sampled anglers divided by the total sampled anglers. Total catch is the product of estimated effort and estimated catch rate.

Observe Kept Salmon

Another goal of PR1 sampling is to observe all kept salmon for adipose finclipped fish and head removal of adipose fin-clipped fish for Coded Wire Tag (CWT) recovery. CWT recoveries are important because they enable managers to 1) track fishery harvest rates, 2) manage fisheries by time and area to target abundant stocks while minimizing the impact on stocks of special concern and, 3) calculate hatchery/natural contributions to the fishery. Ideally, CRFS Samplers will not miss any boats at PR1 sites during salmon season.

Location of Catch Data

Another goal is to collect data on the location and depth of catch. These data are determined by showing the boat operators maps of the area and asking them to point out specific locations and average depths of their catch. The data is used to apply depth-based mortality estimates to some released species and summarize the catch estimates in depth ranges and by

geographic areas. The data can also be viewed in a GIS for trends in catch. This information is required to manage the fisheries by depth and geographic area.

Biological Data Collection

Another goal is to sample lengths and weights of landed catch. Lengths will be used in a regression to calculate a predicted weight for fish without a sampled weight, and to examine the size distribution of the landings. Sampled weights are used to calculate average weights by species. These average weights are multiplied by estimated total catch by species in numbers of fish to estimate total catch in metric tons. Metric ton estimates are used to evaluate catch quotas and the rebuilding status of some distressed species. Numbers of fish for quotas and evaluating status of ESA listed stocks is used in salmon management. Note: do not measure or weigh non-adipose fin-clipped salmon; only adipose fin-clipped fish need to be measured for fork length, prior to head removal. Salmon are managed in numbers of fish, and not by weight.

Sample Selection

Sampling days are selected each month with a set number of weekdays and weekends/holidays based on sample rate objectives. Weekends and holidays are scheduled separately from weekdays. PR1 samples are selected to ensure statistically valid estimates of fishery catch and angler effort, and representative sampling of salmon CWTs. Assignments are scheduled one to two weeks before the first of the month. During ocean salmon season, CDFW's Ocean Salmon Project (OSP) creates the PR1 schedule for those Districts with salmon effort. OSP's sampling schedule is additionally stratified by half-month periods to ensure representative CWT recovery.

Scheduling

The Lead will schedule the random selection of days for sampling for each month in advance. PR1 sites are sampled on several days per month by kind of day. The two kinds of days are weekends/holidays and weekdays. Effort and catch rate are expected to be different for these kinds of days. Rescheduling PR1 days will reduce the statistical validity of the random selection of samples and should be avoided. If it is necessary, the Leads will reschedule and observed the originally scheduled kind of day. Zero effort days are included in computation of the effort, but do not require that a Sampler stay at the site all day to be complete, contact the lead for direction if zero trailers are present during the start count. Samplers should expect an erratic schedule as PR1 sites can have varying effort dependent on the fishing seasons, ocean conditions, etc.

PR1 SURVEY PROCEDURES

Effort Data Collection

During salmon season, the primary goal is to determine the activity, i.e. effort, of every boat returning to the site that day. A specific set of data must be

collected for every boat that returns to the PR1 site for a robust sample. For every boat intercepted the intercept time, number of anglers (licensed and unlicensed), and the target(s) (species or activity) should be recorded. For non-fishing (NF) boats (recreational or commercial activity type), record the specific non-fishing activity as the primary target. See Non-Fishing (NF) Boat Types.

Boats targeting invertebrates are sampled as well, just like finfish boats, regardless of whether they had finfish bycatch. See the Species Sampling Chapter for more information on invertebrate trips.

Arrival and Trailer Counts

Primary sites will be sampled for effort and catch during daylight hours. The Sampler will arrive early enough to sample the first boat returning to the site and depart after the last boat returns, the sun sets, or the departure time scheduled by the Lead. When more than one Sampler is assigned a PR1 assignment, Samplers will stagger their arrivals so that a Sampler is present when the first boat returns to the PR1 site, and a Sampler is present when the last boat returns or sunset. The Lead may schedule arrival times, or they may leave it up to the Samplers to arrange among themselves.

Trailer counts are used to estimate effort for the day. A starting trailer count will be conducted upon arrival of the first Sampler. All boats returning to the site during sampling hours will be intercepted. A final trailer count will be conducted upon departure of the last Sampler. Counts of "trailers" include traditional boat trailers. Personal watercraft (PWC) trailers, car top boat carriers, boats loaded into the beds of trucks, non-trailered inflatable boats, and kayak and canoe carriers are excluded. See the Summary of PR Trailer Counts table on page 10-36 for more information. Trailer counts are not conducted at some PR1 sites (CRD and TRD, for example). Some PR1 sites have buddy sites that are not sampled, but trailer counts are still conducted. These counts are used to determine effort and catch rates are assumed to be the same as the sampled PR1 site. These counts are considered offsite trailer counts.

Low Effort Protocol

If after arrival the first Sampler determines that fishing effort for the day is low, the Sampler will follow the low effort protocols (with regard to additional Samplers assigned to the site) provided by the Lead, which may vary by District. If there is known/suspected finfish effort, the Sampler will remain until the last boat returns or sunset.

No Boats in PR1 Mode

Occasionally, a PR1 site may have no effort, due to weather, etc. If the first Sampler to arrive determines that the start count is zero, the Sampler should notify any other Samplers assigned to the site that day that they will not be needed, and stay on site for a minimum of two hours to see if effort develops. If no effort develops after two hours, the assignment will be considered complete.

Sub-Sampling

The goal of CRFS is to produce high quality data for both catch and effort. Typically, Leads schedule enough Samplers to sample every boat, especially during salmon season. Sub-sampling during salmon season should not occur. During salmon season it might be necessary to skip the collection of biological data on non-salmon boats in order to sample all salmon boats. When salmon fishing is closed, sub-sampling boats can become a necessity to retain the collection of high-quality catch data. It is OK to miss a boat to observe catch and collect biological data for CRFS priority species. Boats should not be canvassed for target to determine if they are to be missed.

Onsite Missed Boats

Avoid missing boats at the PR1. If a boat is completely missed while sampling other boats, it is considered an onsite missed boat. Onsite missed boats do not have a time, target species, or number of anglers recorded; they are simply tallied with the current boat the Sampler is interviewing in the left-most missed boat column on the PR Form. Record "K" next to the number of missed fishing kayaks. Page totals for onsite missed boats are tallied at the bottom of each PR page, and assignment totals for all onsite missed boats are tallied at the bottom of the ASF.

Offsite Missed Boats

During salmon season in northern California, it is important to count sport fishing boats going past certain PR1 sites into a marina or harbor/mooring as "offsite missed boats." Do not include boats returning to adjacent or alternate sites (aka buddy sites) as offsite missed boats unless instructed to do so. Specifics are given for each site below. Offsite missed boats are tallied in the right most missed boats column with the current boat the Sampler is interviewing at the time. Record "K" next to the number of offsite missed fishing kayaks. Page totals for offsite missed boats are tallied at the bottom of each PR page, and assignment totals for all offsite missed boats are tallied at the bottom of the ASF. If the vessel returns to the PR1 site, remove one boat from the offsite missed boat count.

PR Sites: Offsite Missed Boats and/or Offsite Trailer AreasPR1 Site	Offsite Missed Boats	Offsite Trailer Count /Offsite trailer area
Fields Landing LR (FLD)	King Salmon marina	<none></none>
Noyo River LR (FTB)	Dolphin Isle marina	South Harbor District LR
Westside LR (BOD)	Bodega Bay marinas	Doran LR
Berkeley Marina LR (BER)	Berkeley marina	Emeryville LR
Pillar Point LR (PRI)	Pillar Point (Princeton) marina	<none></none>
Santa Cruz Marina LR (SCR)	Upper marina	<none></none>
Moss Landing LR (MOS)	North and South marinas	Woodward Boat Ramp
Monterey Marina LR (MOH)	Monterey marina	<none></none>
Dana Basin LR and	<none></none>	Seaforth Boat Rentals (put counts on Dana Landing Rentals PR form)

Specific Offsite Count Instructions

Fields Landing Launch Ramp (FLD): Recreational fishing boats that are seen going into the King Salmon marina are to be counted as offsite missed boats.

Noyo River Launch Ramp (FTB): A trailer count is made at the South Harbor District Launch Ramp before and after sampling at the Noyo River Launch Ramp (PR1 site). Recreational fishing boats that pass the Noyo River Launch Ramp on their way to Dolphin Isle Marina are counted as offsite missed boats.

Westside Launch Ramp (BOD): A trailer count is made at the Doran Launch Ramp before and after sampling at the Westside Launch Ramp (PR1 site). Recreational fishing boats that go by the Westside Launch Ramp into Bodega Bay marinas are counted as offsite missed boats.

Berkeley Marina Launch Ramp (BER): A trailer count is made at the Emeryville Launch Ramp before and after sampling at the Berkeley Marina Launch Ramp (PR1 site). Recreational fishing boats that go by the Berkeley Marina Launch Ramp into the marina are counted as offsite missed boats.

Pillar Point Launch Ramp (PRI): Recreational fishing boats that are seen going into the marina are to be counted as offsite missed boats. Note that anglers using the Pillar Point LR park their trailers in the upper lot or along the highway; these are considered onsite trailers.

Santa Cruz Marina Launch Ramp (SCR): Recreational fishing boats are counted as offsite missed if they pass the launch ramp and head to the upper harbor. Boats interviewed at the launch ramp are asked if they went toward the upper harbor prior to landing. Boats answering yes are adjusted with a (-1) in the offsite missed boat count.

Moss Landing Launch Ramp (MOS): A trailer count is made at the Woodward Boat Ramp before and after sampling at Moss Landing Launch Ramp (PR1 site). Recreational fishing boats that head towards Moss Landing Marina and the North Harbor Marina are both counted as offsite missed boats.

Monterey Marina Launch Ramp (MOH): Recreational fishing boats that are seen going into the marinas are to be counted as offsite missed boats.

Catch Data Collection

All private boats that have completed a fishing trip should be sampled for catch. Catch includes landed catch (fish brought ashore) and reported catch such as discards or other catch not available. The Sampler may have to interview all anglers on the boat to determine total catch since anglers may not be aware of each other's catch. This determination may need to be done before the driver leaves to get the trailer. The goal is for the Sampler to observe all finfish catch to identify to species, recover heads from all adipose fin-clipped salmon, measure and weigh as many fish as possible, and document all unobserved catch for each boat.

Q. What if too many salmon boats are coming in for me to key out all rockfish species and also take heads? Can I code all the rockfish to genus?

A. No, you are to avoid coding observed kept rockfish to the genus level. Work with the other CRFS and OSP Samplers present to avoid missing boats while still identifying retained catch to the species level. It may be necessary to drop the collection of weights and lengths from landed catch to avoid missing boats.

Biological Data Collection

After determining the catch by species for the boat, the Sampler will measure and weigh as much of the catch as possible. It is important to the CRFS program to measure fish that are under active management, especially species of concern. A prioritized list of species to preferentially sample is provided (see Priority Species). Lengths can be used to predict weights and to examine length classes; however, recording length-weight pairs is the goal

for bio data collection. Do not weigh any salmon species, and only record lengths of adipose fin-clipped salmon.

Sub-sampling Lengths and Weights

There may be times when the level of activity at a site is too high to sample the lengths and weights of every fish on every incoming boat. The Sampler should attempt a random sample of fish in this case, following the priority list. Lengths are required for all adipose fin-clipped salmon. Refer to the section, General Onsite Procedures: Catch Measurement.

Catch Location and Average Depth Data Collection

The Sampler will attempt to determine the location and average depth of catch by species, or the location and depth of the majority of the boat's fishing effort if there is no catch. Maps with depth contour lines are provided to assist the angler in determining the catch location(s) and depths. If all species were caught within the same location and depth, then only one location and depth may need to be reported. Often, locations and corresponding depths may need to be reported separately for individual species or species groups. For suspect data, rare species, and especially for prohibited species, double check the catch location and average depth with the angler. For trips with large areas of trolling (for non-bottomfish species), a general area can be used. Catch location is used to manage fisheries by geographic boundaries.

Sub-sampling Locations and Depths

There may be times when the level of activity at a site is too high to sample the locations and average depths of all catch on every boat. In these cases, the Sampler should attempt a random sample of more specific locations and depths for bottom-fishing boats. This allows some boats to give a single more general location to save time. Boats targeting surface fishes (tuna, salmon, seabass, etc.) may be coded with the general locations and depths as well, when time is short. It is important to document location and average depth for non-retention species and species on the Priority Species List.

Q. What if a salmon boat comes in with a few rockfish but no salmon catch, do I code the salmon effort or the bottomfish catch location and depth when I'm in a hurry?

A. Do not code the location of fishing effort if there is catch; code the location specific to the catch species. In this instance, it is more important to code the location for the rockfish catch.

Minimum PR Sample

A CRFS sample is defined as a boat which has been sampled for both effort and catch. Catch locations, average depths, and bio data are not required for a valid CRFS sample. Note: the data elements below are the minimum required for a valid sample; Samplers should make every effort to collect the data needed to complete EVERY field.

The following data elements are the minimum requirements for a useable CRFS interview:

- Total number of anglers who fished
- Number of days fished
- Target
- Area fished (water area, e.g., Nearshore=N)
- All catch, unobserved number of fish by species

Minimum requirements for a valid salmon sample usable by OSP:

- Total number of anglers who fished
- Gear
- Number of kept salmon by species
- All kept salmon must be observed for adipose fin -clips
- All adipose fin-clipped salmon receive a unique headtag, even if the head cannot be collected (i.e., NRS)
- Boats that provide only the minimum requirements for a valid salmon sample usable by OSP shall be flagged with "RS" in the sample # field and summed to the page tally as a salmon boat.

The minimum items for this interview are listed above. Fish measurements may be omitted but fish counts may not. Never code rockfish to the genus level to save time. If the minimum requirements cannot be met, the boat will be considered a Barrier or Refusal or tallied as a missed onsite boat depending on the nature of the interaction. If only the salmon requirements are met it will receive a "RS" code.

Screening Divers

In addition to hook-and-line anglers, divers may qualify for the CRFS interview. If a diver carries a spear gun with them, they can be interviewed as 'anglers.' If they spear a fish or intended to spear a fish they are considered eligible anglers and can be interviewed with gear code "S." Divers taking or intending to take invertebrates are also eligible to be sampled (see General Onsite Procedures under invertebrate sampling section). Divers entering the water from the shore using fins and a flotation device (such as a dive tube) to fish are considered either BB or MM anglers. Divers who enter the water from a boat or other craft are considered PR anglers. This includes kayaks, stand up paddleboards (SUPs) and pontoon boats with 'oars.' In effect, having a paddle is what designates the mode as PR.

Fishing Tournaments

For the purposes of CRFS sampling, a tournament is defined as a site and date specific fishing contest. Contestants usually must return to tournament head quarters by a certain time for the "weigh in" which limits how far they can travel, and only specific species of fish may be taken on the date of the tournament event. Generally, tournaments are not sampled. Once the Sampler determines that a tournament is taking place, the Sampler should contact the Lead immediately to discuss the situation. It is important to notify the Lead in advance when a tournament date and location is discovered so that the Lead can make appropriate arrangements, if necessary. If the Lead

determines to sample as scheduled, a sample flag of "T" should be used for all boats sampled that are participating in the tournament.

Informal 'pools', such as those arranged on CPFVs (jackpot contests), are not considered tournaments—anglers participating in these types of contests should be sampled as usual. Some fishing contests are regional (encompassing a large area) and span a long period of time (a week, a month, or an entire fishing season). Participants usually pre-register at a location, such as a tackle or bait store, and may bring qualifying fish in during the entire time the contest is running. For the purposes of CRFS sampling, these types of contests are called derbies. Anglers participating in derbies should be sampled as usual.

THE PR FORM (PR1 MODE)

The PR Form collects total boat effort for the day by counting trailers and intercepting returning boats. Each boat is screened as fishing or non-fishing. For fishing boats, determine target fish species and number of anglers per boat. In northern California during salmon season, the form will also count all retained and released salmon as well as record lengths of adipose fin-clipped salmon head tag numbers. For boats with catch, all the fish are counted by species along with location(s) and average depth(s). When time allows, detailed catch locations and average depths are recorded and observed finfish are measured and weighed.

Questionnaire Usage

Samplers are given a laminated copy of the **questionnaire** used with the PR Form. The questions for the interview are written out, in full for standardization. The Sampler should word each question specifically <u>as it is written in the questionnaire</u>. In order to have meaningful comparative data, each angler should respond to a standardized stimulus. <u>Methodological studies have shown that even slight changes in questionnaire wording, for example "should" versus "could," drastically influence responses.</u>

Introduction to the PR1 Interview

Tasks while sampling boats are generally done in this order:

- 1. Determine if anyone on the boat has fished
- Determine the total number of anglers and of those, the number unlicensed
- 3. Determine the launch time of the boat
- 4. Determine zip code of one random angler
- 5. Determine total days fished on the trip
- 6. Determine if night fishing occurred
- 7. Determine the 12-month avidity for one random angler
- 8. Determine the target species and gear (or non-fishing activity)
- 9. Determine the primary area fished for the fishing target(s)
- Determine any catch (including discards) or marine mammal losses (salmon only)
- Determine how many of each rockfish species were released using a descending-device

- 12. Count catch by species (mandatory for all salmon species)
- 13. Determine the location and average depth of the catch, or location of majority of effort if no catch
- 14. Record finfish length measurements and weights of the catch (prefer length-weight pairs)
- 15. Depending upon region: collect salmon and/or White Seabass heads and Yelloweye Rockfish

Before the Assignment

The Sampler should check their equipment and forms before leaving for the site. This will ensure that the Sampler has enough forms and other supplies to complete the assignment. Be aware of the weather forecast and prepare for conditions. In northern California during salmon season, be sure additional salmon equipment and tags are on hand. In southern California, make sure to have a White Seabass wand if one has been issued. Double check the date, site, port and assignment ID. Record site information, Sampler name, and ID number on the PR Form and on the Assignment Summary Form (ASF). Plan to arrive onsite at a time given by the Lead.

Arrival on Site

Upon arrival at the PR1 site, count the number of trailers (if applicable) in the parking lot and any adjacent streets or parking lots (consult the CRFS Wiki site or the site description book to determine the count area for each site). Record the arrival time on the ASF and the arrival trailer count in the start count box on the first PR Form. During salmon season in northern California, call the Lead if you think help will be needed from additional Samplers in order to not miss any boats.

Sampler Location Onsite

There are differences among PR sites. Onsite positioning procedures for obtaining interviews will vary slightly by site. For example, boats might be interviewed while they are waiting for a boat hoist, while they are cleaning their boat at the wash down station, at the dock, on the beach, or at the ramp. The Sampler will use discretion in determining the best approach at a particular site. For most PR1 sites, the best spot to sample is where the boats are waiting for their turn to exit the ramp. If boat traffic is heavy, do not conduct interviews on the dock or ramp, as this may delay the trailering process which may result in unhappy anglers.

Multiple Samplers on One Assignment

In some cases, the Lead will schedule two or more Samplers to work at a PR1 site due to expected high effort. Samplers may work shifts that overlap. A common sampling strategy is one Sampler will arrive first and work until the second Sampler arrives, generally just prior to peak activity. Both Samplers then work the peak period together until activity drops off and the first Sampler departs. The second Sampler then works until all the activity is complete for the day or sunset. The Lead will advise as to which methodology to use based on the season, fishing effort, District, etc. In northern California,

Samplers should coordinate onsite arrival times with each other when working on the same assignment.

Avoiding Duplication and Sharing Counts

It is important that Samplers working together not duplicate or omit any data in the field and when submitting forms and summaries. Each Sampler edits and submits a separate set of forms. The Assignment ID is the same for both Samplers. Be sure to record the last names and Sampler ID number of all Samplers working the assignment at the top of the first PR page and on the ASF, and circle "Y" or "N" if they have data or not. Each Sampler numbers their boats separately, so there may be two or more boats labeled #1 for the assignment. The <u>start count</u> will be performed by the Sampler who arrives on site first, while the <u>stop count</u> will be performed by the Sampler who leaves the site last. These two counts will be on different form sets and specific to the Sampler for the assignment. The start and stop count will be reported in the Weekly Report by the Sampler who conducted the respective count. Each Sampler will have their own separate PR form and ASF form subtotals. These totals will be summed after data entry to compute totals for the entire assignment with multiple Samplers. The data will be merged in the database.

Onsite Trailer Counts

Trailer counts are made when the first Sampler arrives and when the last Sampler leaves. Counts of "trailers" include traditional boat trailers. Personal watercraft (PWC) trailers, car top boat carriers, boats loaded into the beds of trucks, non-trailered inflatable boats, and kayak and canoe carriers are excluded. Do not count trailers not attached to vehicles, or known non-fishing, commercial or CPFV trailers. See the Summary of PR Trailer Counts table on page 10-43 for more information. Immediately before leaving the site at the end of the day, the Sampler will count the number of trailers remaining in same area. Known commercial, non-fishing or CPFV trailers should not be included; all others are included in the stop count Trailer counts are not conducted at some PR1 sites (CRD and TRD, for example).

Offsite Trailer Counts

Trailer counts for buddy sites are recorded during certain PR1 samples. This data is recorded on the first page of the PR Form in the offsite start and stop trailer count boxes. These counts are recorded on the Assignment Summary Form too. If the "count area" (ramp parking lot) is full and trailers are forced to be (that are active at the PR1 site) parked on the street or outside the normal "count area", include those trailers in the onsite trailer count, <u>not</u> in the offsite trailer count.

Offsite start and stop trailer count coding example.

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Monitoring Boats

When a boat arrives at the PR1 site, a new sample is created with the time of arrival. During very busy times, a boat may arrive and will not be sampled because the Sampler(s) are busy with other boats. This boat will be tallied on an existing boat row as an onsite missed boat in the onsite missed boat column. An onsite missed boat may be either a non-fishing boat (NF) or a fishing boat. The proportion of fishing to non-fishing sampled boats is applied to the count of onsite missed boats to estimate several additional fishing boats. It is expected that missed boats will have the same proportion of NF to fishing boats as the boats sampled. This assumption is a potential source of bias. For example, if all the missed boats are fishing boats, but half the boats actually sampled were NF boats, then the estimate of fishing boats missed will be underestimated by 50% because missed boats were not representative of the boats sampled. Therefore, onsite missed boats should be a representative selection of all boats, not just fishing boats or boats that look like a lot of work to sample. During salmon season every effort should be made to avoid missing boats. Ideally, there should not be any missed boats. With two or more Samplers working each PR1 assignment, it should be possible to sample every boat. Once a boat has been canvassed and the target is either finfish or invertebrate the minimum CRFS interview is required. Contact your Lead immediately if additional help is needed to avoid missing boats.

Multiple PR Trips on the Same Day

Occasionally PR boats will make more than one trip per day; sometimes the skipper drops off passengers from a morning trip and takes a new crew out on a second trip in the afternoon, or the crew may remain the same after returning from the first trip of the day. The Sampler may recognize the boat as having been sampled earlier in the day, or the crew may point out that they have already been sampled at the completion of their first trip. Regardless of how this second (or subsequent) trip is discovered, the Sampler is to treat these trips separately, and attempt to sample both as distinct trips each with unique data — separate sample numbers, different launch times, segregated catch, etc. Do not combine both trips into one sample. If the catch from both trips is still onboard at the completion of the second trip and the crew is unable to separate catch by trip, the Sampler is

to record catch from the second trip as angler reported (kept unobserved). If anglers are reluctant to participate in the survey again, point out that each of their trips is unique, and it's important for CRFS to capture data from each and every unique trip — perhaps the boat had different targets, fished in a different location, or caught a different composition of species.

Determination of Boat Type

A category based on activity must be assigned for each boat intercepted. Ask a passenger on the boat as to its activity for the day. There are, essentially, two types of boats in the PR survey: Fishing and Non-fishing (NF). A fishing boat is defined as a boat, either privately owned or rented, upon which recreational fishing effort (for finfish OR invertebrates) occurred. Boats that targeted invertebrates only are considered fishing boats. Catch is not necessary to be considered a fishing boat. Boats that intended to fish but did not put gear in the water are NF boats. A CPFV carrying passengers paying to fish is not considered a fishing boat for the purposes of PR mode sampling.

Non-Fishing (NF) Boat Types

There are three NF codes currently being used:

- **1. NFCOM** a commercial fishing boat targeting finfish or invertebrates (note: occasionally a commercial fishing boat may be fishing recreationally that day the boat would be sampled just like any other PR boat).
- **2. NFPC6** Commercial Passenger Fishing Vessels, also called party/charter (PC) boats, vessels that are permitted to take paying passengers fishing. This includes smaller, trailered "6-pack" boats. The Sampler may have to inquire with the operator to determine if the boat was a regular PR boat or was fishing as a CPFV that trip.
- **3. NFOTH** all other non-fishing boats fall into this category. This includes boats that intended to fish but for whatever reason had no wet-gear time, cruises, sailboats that did not fish, bird watching, whale watching, burials at sea, enforcement, research, etc.

CPFV and Commercial Boats

Commercial Passenger Fishing Vessels (CPFV) are coded as "NFPC6" on the PR Form. If the Sampler encounters a CPFV at the PR1 site, the boat is coded as "NFPC6" in the Target field of the PR Form. The boat should then be sampled opportunistically using the appropriate PC dockside sampling. Commercial fishing boats are coded as NFCOM in the Target field of the PR Form; commercial fisheries are sampled using other non-CRFS surveys.

Opportunistic PC Sampling

Commercial Passenger Fishing Vessels (CPFV) that utilize a PR site are coded as "NFPC6" on the PR Form. Monitoring PR effort during a PR assignment is a priority; if time allows and without missing any PR effort, the Sampler should sample the CPFV using the appropriate PC dockside sampling form – the CRFS-OSP SALMON CPFV DOCKSIDE form for trips that targeted salmon only, and the CRFS PC (CPFV) DOCKSIDE form for

trips that targeted something other than salmon. If the boat targeted both salmon and non-salmon on the same trip, sample the boat using both forms, recording data on the appropriate form. Report all CPFV activity to the PEC Port Lead (Districts 3-6) or record the vessel's effort on a PEC form (Districts 1-2). See CPFV Dockside Sampling sections in this manual for more information on sampling CPFVs dockside.

Q. What if I see a PC (party or charter) boat returning to the PR1 site?

- **A.** Determine if the boat was fishing recreationally (PR trip), or if it was carrying passengers paying to fish (CPFV trip). If the former, sample the boat as a PR boat on the PR form. If the latter, code the boat as an NFPC6 boat on the PR form and do one or both of the following:
- 1) If the boat was targeting salmon, sample the boat using the CRFS-OSP SALMON CPFV DOCKSIDE form.
- If the boat was targeting anything besides salmon, sample the boat using the CRFS PC (CPFV) DOCKSIDE form.
- 3) If the boat was targeting both salmon and non-salmon species, sample the boat using both dockside forms.

Refused Boats

Participation in this survey is voluntary. An angler may refuse to participate. However, this data is crucial to sustainable fisheries management, so the Sampler should try to get as many questions answered as possible. Some anglers on the boat may be more receptive than others.

Although refusal to answer key CRFS questions will be coded as a refusal, salmon minimum data element requirements will allow for saving a sample when CRFS minimum interview requirements are not met. Anglers are required to make kept salmon available for sampling (Title 14, CCR, Section 1.73(b)); minimum requirements for a valid salmon sample include number of anglers, kept salmon by species, and salmon with adipose fin clips—code these boats as RS in the Sample #. Zip code, avidity, location, and depth are not necessary for a "valid" sample; however, these items are important. If you cannot get all the required questions answered, you will have to record the boat as a refusal; code an "R" in the Sample # field. Refusals do not get a sample number, just an "R." Refused boats tallies are not inclusive of the "Total Boats" subtotal on the bottom of the PR form but should be included in the salmon boats subtotal. If you can collect the minimum salmon requirements the boat would be coded "RS" and included in the total boats on the PR1 page totals.

Language Barrier Boats

Anglers that cannot speak English may not be able to effectively answer survey questions. If there is too much of a language barrier, the Sampler should stop the CRFS interview. If all the required questions are not answered, the boat is recorded as a barrier; code a "B" in the Sample # field. Barriers do not get a sample number, just a "B." Language-barrier boats are

<u>not</u> tallied into the total boats field on the PR1 page totals but are tallied in the Refu + Barrier total.

Anglers, Zip Code and Days Fished

Once the Sampler determines the boat is an eligible fishing boat and willing and able to participate, they determine the angler effort on the boat. Some of the passengers may not be anglers. The Sampler will determine the number of anglers who actually fished. Next, the Sampler determines the number who fished without a valid CA fishing license. The number of unlicensed anglers will always be equal to or less than the total number of anglers on the boat. It is best to determine this indirectly by asking what type of fishing license the anglers used. Often, the anglers will want to show their licenses—Samplers do not need to see their licenses to code them as licensed anglers. The number of unlicensed anglers is used to adjust effort from the licensed angler telephone survey; children are not eligible to participate in the telephone survey, and some anglers are not required to have a license and so would not be a part of the telephone survey.

The final item required to estimate effort on the boat is the number of days fished. Usually this will be one day; however, some boats, especially in southern California, may have taken multi-day trips. The "N" box will be checked if the boat fished at night (after dark the night before until dawn of the current day). If only night fishing occurred, the "N" box will be checked and "0" days fished will be recorded. If it is a multi-day trip, record the number of days fished, leave the launch time blank, and leave a note on the data sheet.

One of the anglers on the boat will need to provide a zip code. This is the zip code of the permanent residence of the angler, not temporary lodging. If the angler is from a foreign country, use the applicable foreign country code. The zip code is used primarily to help quantify the contribution of sportfishing to the economy. The angler asked should be at random, not biased by boat ownership, fishing skill, age, gender, etc.

Determination of Catch

The Sampler will determine if any fish were caught by the boat. Each fishing boat will need a complete census of catch. The term "catch" includes observed and unobserved kept fish and released fish. Catch includes landed fish, fish given away, taken by marine mammals (salmon only), used for bait, filleted or eaten, <u>AND</u> fish purposely released, thrown back alive (shakers) or dead. Anglers may report that they have no fish on the boat. However, a boat may still have catch if they caught and released fish or lost a fish to a marine mammal. Be sure to inquire about anything that was caught and then used for bait or any other fish that were caught but not available for the Sampler to observe.

Examining Catch

The Sampler will examine all landed catch for each fishing boat. Examined or observed finfish are the most robust because the Sampler actually saw,

counted, and identified the catch to species. If the angler(s) refuses to have the landed catch examined, all catch are coded as "kept unobserved". It is more important to count and identify rockfish to the species level than to get lengths and weights from those fish.

Q. What if the ramp is busy and I don't have time to count each rockfish to species. Can I just code rockfish genus "RFGEN"?

A. No, you must record catch to species. The only time you should be using the RFGEN code is for unobserved catch that the angler simply cannot identify, even with identification guides. There will often be at least one other Sampler there to help you avoid missing boats; if you are unable to keep up with the boats as they come in, stop collecting bio data.

Salmon Head Recovery

All kept salmon shall be examined for the presence of an adipose fin. Title 14, California Code of Regulations Section 1.73(b) require anglers to show

their salmon catch to Department representatives, and to relinquish the heads of all adipose fin-clipped salmon to the State at no charge. When the Sampler encounters an adipose fin-clipped salmon, notify the angler that the salmon head will be removed for recovery of the CWT. Apply a headtag to the lower jaw of the



salmon using the attached wires, measure the fork length, and record these numbers on the PR Form. Remove the salmon's head using the knife and cutting board provided, taking as little flesh and gills as possible. Place the tagged head in the clear bag with the headtag numbers facing outward and freeze as soon as possible. A headtag is issued to every adipose fin-clipped salmon, even if the head cannot be recovered. See the Species Sampling section for complete details regarding salmon sampling.

Observed Catch (Sampler-Examined)

The Sampler will attempt to observe and examine all retained finfish catch, recording the number of fish kept and observed by species in the appropriate box on the PR Form. It is important to note that only fish that the Sampler sees <u>and</u> counts can be recorded as "kept observed". Fish not able to be physically viewed and counted by the Sampler must be recorded in the "kept unobserved" box. It is important to the CRFS program to differentiate between Sampler-examined and angler-reported fish counts.

Estimates of total harvest are summarized separately for the Sampler-examined and angler-reported catches.

Sampler may identify fillets with skin patches, being careful not to double count fish (i.e. two fillets equals one fish). Fish identified by skins are considered "kept observed." Anglers may not want the Sampler examining fish that have been filleted. These fillets are someone's dinner, and they may not want to get their food dirty or they may be hesitant to open a tied bag. Ask the angler before attempting to examine fillets.

Unavailable Catch (Angler-Reported)

In addition to any fish the Sampler sees, each fishing boat will be polled for any fish caught that are not available for examination. Unavailable catch are usually fish that have been thrown back, given away, packed away, used for bait, filleted (not identified by skins), eaten or taken by marine mammals (salmon only). Unavailable fish are reported by the entire group of anglers on the boat. The anglers are asked to separately report any unobserved fish in four categories; kept, released alive, released dead, and seal take (salmon only). If no fish were caught (kept or released), a NO CATCH code is recorded in the Species code box and the catch boxes are zeroed out.

Kept Unobserved Catch

Fish that are not thrown back, but otherwise are not available for examination will be separately recorded on the PR Form. Kept unobserved fish include fish given away, packed away, used for bait, filleted (not identified by skins), or eaten. Kept fish that the angler refuses to show to the Sampler are included as "kept unobserved." These fish are counted separately from fish which the Sampler personally examines and counts (kept observed). Be persistent with anglers that have unavailable rockfish catch. Use your best effort to gain access to the catch for species identification.

Released Alive

The released alive catch category is the total number of fish by species that were released alive in swimming condition. Released alive includes fish intentionally landed and subsequently released, those that are purposely shaken off the hook boat-side, and any rockfish that are released using a descending device. The Sampler and anglers are not to judge the likelihood of survival of a swimming fish. Fish that 'got away' are not considered purposely released and are not included as released-alive.

Released Alive with Descending Device (DD)

This is a subset of released-alive and includes the total number of rockfish by species that were released alive using a descending device. Rockfish brought up from depth suffer from barotrauma from gas expansion as a result of decreasing pressure. Stomachs protruding from mouths, eyes popped out of their orbits, and "crystallized" corneas are all symptoms of barotrauma. Use of a descending device to send rockfish back down to depth can greatly reduce discard mortality. A descending device can be a professionally fabricated store-bought lip-gripping contraption; it can be a line tied to the

bend of a hook with a heavy lead sinker tied to the eye of the hook; or it can be an inverted, weighted milk crate with a rope tied to the bottom (now the top) — anything used to send a fish back to depth can be considered a descending device. Use of a needle to vent the swim bladder of a fish is not considered a descending device. Released-alive with descending device is coded only for rockfish species. Released-alive with descending device is a subset of the released-alive total; the number of released-alive with descending device will always be less than or equal to the released-alive total.

Released Dead

The released dead category includes fish landed or purposely shaken off the lines which are returned to the water in dead condition. Fish that are technically alive but are obviously not going to survive (due to severe wounds or inability to swim down) may be coded as dead. The Sampler and angler are to judge that the non-swimming fish is dead or will be shortly. The survival of all fish returned is determined by application of mortality rates. These rates are determined by scientific studies of hooking and depth-based mortality.

Seal Take

The seal take category includes any <u>salmon</u> that were known to have been taken by any marine mammal (seals, sea lions or other marine mammals). <u>Seal take should only be determined for salmon catch</u>. Anglers must be certain and have seen the marine mammal take the salmon from their line. The Sampler should inquire further with those anglers who say 'I think' or 'maybe' a fish was lost to a pinniped. The Sampler should not include fish that naturally escaped or were naturally caught and eaten by a pinniped.

Catch Location and Average Depth

All CRFS boats are sampled for the catch location and average bottom depth. For boats with catch, a catch location will be recorded. Location and an average bottom depth may be recorded for all catch together or by species when determined and time allows. For boats with no catch, location and average bottom depth for the majority of fishing effort is recorded. The majority of effort is defined as where most of the boat's time was spent with gear in the water. Average bottom depth is used to put the catch estimates into depth zones and compare with locations. It is also used to help estimate depth dependent mortality rates for some groundfish species.

Q. If the PR anglers do not have any catch, should the catch location be left blank?

A. No, still code a location. In this case, code to the major area fished (where effort mostly occurred).

Coding Location of Catch for Multiple Species on the Same Trip

Frequently, PR boats will fish in several locations for different species/species groups on the same trip. It is important for the Sampler to recognize when this occurs and code distinct locations of catch for each species/species group. If anglers do not give some indication that their catch for the day came from more

than one location and the Sampler is either not paying attention or not familiar with local fisheries, then incorrect location information will be collected that may bias CRFS data. Taken to the extreme, when a boat's catch comes from more than one location and the Sampler does not collect location of catch data for each species/species group, it may appear that the boat was fishing in an illegal area, at an illegal depth or with illegal gear.

TARGET	1	3200		KEPT	RE	LS	SPECIES LOC	DEPTH			
1st	AREA	GEAR	SPECIES	obs	alive	(widd)	or effort loc if no catch	Average			
2nd	I N H	CODE	unobs	dead	seal take	Block-box; Lat / Lon	Bottom (ft)				
RFGEN	N	н	RFBLK	6	2	(o)	222-23	40			
HALPA	0	н	RFBLK	unebs O	dead O	seal					
			HALPA	2	olive O	()					
			HALPA	0	dead O	seal					
				3	alive O	(o)					
			RFBLU	Unebs O	dead O	seal					
			LNOOD	4	2	()					
			LNGCD	0	O O	seal					

This is an example of incorrect coding of location of catch. As is, the Pacific Halibut catch location will be attributed to the one recorded location of catch; this is incorrect, as HALPA are rarely taken in 40 feet of water, and the water area for the HALPA target was coded as offshore – this location is clearly within three miles of shore.

TARGET				KEPT	RE	LS	SPECIES LOC	DEPTI
1st	AREA	GEAR	SPECIES	obs	alive	(wiDD)	or effort loc if no catch	Average
2nd	A	B	CODE	unobs	dead	seal take	Block-box; Lat / Lon	Bottom (ft)
RFGEN	N	н	LIALDA	2	olive O	()	223-25	350
HALPA	0	н	HALPA	0	tent O	seal		
		П		6 6	alive 2	(0)		
		П	RFBLK	unebs O	dead O	seal		
		П		3	alve O	(o)		
		П	RFBLU	unebs O	dead O	seal		
		П	100 4 144 204 2000	obs 4	elive 2	()		
П			LNGCD	unobs O	test O	seal		

This is another example of incorrect coding of location of catch. As is, the rockfish and Lingcod catch will be attributed to the one recorded location of catch. This example is typical of catch seen in District 6; current groundfish regulations include depth restrictions of 120 to 180 feet. If left as is, it will appear that the bottomfish were taken at

an illegal depth.

TARGET				KEPT	RE	LS	SPECIES LOC	DEPTH
1st	AREA	GEAR	SPECIES	obs	alive	(wiDD)	or effort loc if no catch	Average
2nd	4	5	CODE	unobs	dead	seal take	Block-box; Lat / Lon	Bottom (ft)
RFGEN	N	Н	RFBLK	6	2	(0)	222-23	40
HALPA	0	н	KFBLK	Unobs O	dead O	seal		
		П		obs 2	alive O	()	223-25	350
	Т	П	HALPA	unobs O	deed O	seel		
			RFBLU	3	alive O	(0)	222-23	40
			KFBLU	0	dead O	seal		
			LUGGE	4	alive 2	()		
			LNGCD	0	dead O	seal		

This is an example of correct coding of location of catch.

Note that for the Blue Rockfish and Lingcod catch to be associated with the location of the Black Rockfish catch, the location and depth needs to be repeated after the Pacific Halibut row.

Measuring Catch

For each CRFS boat with observed catch, the Sampler should sample the catch for species composition and bio data: lengths, weights, and sex for some fish. The priority is to document and measure the priority species and, in northern California, adipose fin-clipped salmon. Do not measure salmon with intact adipose fins. A secondary priority is to weigh important management species. Please see the General Onsite Procedures section for a complete list of priority species.

Time allowing, all fish except salmon may be measured and weighed. The goal is for paired lengths and weights, if possible. Paired lengths and weights allow for a regression equation to check for sampling error. If time is short, 5 paired length and weights should be collected per species. Lengths are used to predict weights using a regression and to examine length classes. Weights are used to calculate more precise metric ton estimates and are used with the length to estimate fish condition.

Some fish may be sexed using external characteristics. Please see the Species Sampling section for complete details on which species may be sexed.

Interview Priorities

Samplers should be aware that some of the PR data is required for a valid CRFS interview, while sub-sampled data may be of high or lower priority.

Required Fishing Effort Data

Count offsite trailers upon arrival (where applicable)

Intercept all onsite boats

Determine if the boat is fishing or not

Determine the number of anglers

Determine the target species (or non-fishing activity)

Count all missed onsite and offsite (where applicable) boats

Count all fishing boat trailers at departure

Count all off-site fishing boat trailers after departure (where applicable)

Required Catch Data

Determine if any catch (including unobserved/unavailable catch)

Count catch by species (not higher-level taxa)

Examine salmon for adipose fin-clips and collect heads

Determine the location and average bottom depth of the majority of the catch (or effort if no catch)

Sub-sampled Data (Priority Order)

- 1. Record length measurements of priority species
- 2. Record weights of priority species
- 3. Record length and weight pairs of other species
- 4. Determine the location and average bottom depth of each species

2020 CRFS PR Form Questionnaire

It is important to use the wording of questions as stated in the PR questionnaire because slight changes in wording can result in different responses.

INTRODUCTION: Hello, my name is ____ and I represent CDFW. I am interviewing marine recreational anglers for the California Recreational Fisheries Survey. May I ask you a few questions?

PRIVACY ACT STATEMENT: This study is being conducted in accordance with the Privacy Act of 1974. You are not required to answer any question that you consider to be an invasion of your privacy.

BOAT ROW, EFFORT COLUMNS:

Sample # [or R or B or RS]: In sequence, the boat number for all boats returning to the site during the sample, including non-fishing boats, but excluding missed boats, initial refusals (R) and language barriers (B). Fishing boats that do not provide the minimum data elements (# anglers, # days fished, water area, targets, gear, catch #s by species) are also coded with R and do not get a sample #. Code boats that refuse all data elements other than those required for a minimum salmon sample (# angs, # kept/obs salmon and all must be observed. #ad clips) with "RS" in the sample number field.

Time: Enter the time in the 24 hr format when the vessel interview was started. Times are unique for each Sampler's data.

SCREENING: Did anyone on the boat do any sport fishing?

YES:	Go to next
NO:	Record appropriate NF (non-fishing) code in
	target box, and conclude the interview
Refused:	Code Sample # as R, terminate interview
NOTE: If the boat is going	back out for more fishing skip till next return.
	f you had gear in the water? (on vessel)
	nglers on the vessel that fished (gear in the
water)	
Refused:	Code Sample # as R, terminate interview
7 1	sport fishing license does each of you have?
	IGS (above) who fished on the boat without a
current California sport fisl	ning license.
Refused:	R

PRIMARY TARGET: What were y the boat's primary target. Anything: Not fishing: Refused:interview	UNIFH Approp	riate NF code ample # as R, terminate	of
SECONDARY TARGET: What we taxon of the boat's secondary target	-	econdarily after? Code the	;
Anything: Refused:	Leave b Leave b		
EFFORT AREA: Was your <prima (="" (<="" 3="" ask:="" bay?="" if="" in="" miles):="" nearshore="" ocean,="" offshore="" that="" the="" was=""> 3 miles): Bay/Estuary/Harbor: Mexico: Refused: Offshore islands have separate contacts.</prima>	N O B Be av M Code Sa intervie	within 3 miles of land? vare of freshwater cutoffs ample # as R, terminate w	
EFFORT AREA: Was your <secon (<="" 3="" ask:="" bay?="" if="" in="" miles):<="" nearshore="" ocean,="" td="" that="" the="" was=""><td>N O B Be av M Leave b</td><td>within 3 miles of land? vare of freshwater cutoffs blank</td><td>or</td></secon>	N O B Be av M Leave b	within 3 miles of land? vare of freshwater cutoffs blank	or
GEAR: What gear did you use fo	r <primar< td=""><td></td><td></td></primar<>		
Finfish Hook & Line:	Н	Shellfish Pot #:	Pn
Spear:	S	Flat Hoop Net #:	F _n
Troll:	Ť	Rigid Hoop Net #:	Rn
Bait Net:	N	Snare:	Ε
Mooch: (salmon only)	M	SCUBA:	С
Both M & T (salmon only): Refused:	B R	Free Diving:	D
GEAR: What gear did you use fo	r <secon< td=""><td>dary target>? Shellfish</td><td></td></secon<>	dary target>? Shellfish	
Hook & Line:	Н	Pot #:	P_n
Spear:	S	Flat Hoop Net #:	Fn
Troll:	Т	Rigid Hoop Net#:	R_n
Bait Net:	N	Snare:	E
Mooch: (salmon only)	M	SCUBA:	С
Both M & T (salmon only):	В	Free Diving:	D

Refused:	R
DAYS FISHED trip: Record number of daylight Check the N box if any fist Refused:	
today, within the past 12	sk a randomangler on the vessel. Not counting 2 months, how many days have you gone ng in this state or from a boat launched in this
Refused:	R
Don't know	DK
Sampler didn't ask	DA
Launch Time: What time hr format) the boat left the Refused:	
Don't know	DK
Sampler didn't ask	
your residence? If ZIP u Refused:	DK DA
	total and RELS dead. If salmon were targeted, proceed to SEAL TAKE
Refused:	Code Sample # as R, terminate interview
KEPT OBSERVED: May Yes:species	I see the catch? Sampler will identify and count all fish by
No: Unobserved	Enter zero and code numbers of Kept
Fillets: Unobserved	Enter zero and code numbers of Kept
Refused:	If there is salmon catch, code Sample # as R, terminate interview. If no salmon catch, go to

next

KEPT UNOBSERVED: Did the boat retain any other catch? Probe for any catch given away, filleted, used for bait or trashed. Yes:----Record species and number of fish No:----Enter zeros in Kept Unobserved boxes for species recorded Kept Observed Refused:-----If both Kept Observed and Kept Unobserved are refused, code Sample # as R, terminate interview Don't Know:-----Code Sample # as R, terminate interview RELEASED ALIVE TOTAL: Were any fish released alive? Probe for any fish that were purposely released alive. Yes:----Record species and number of fish No:----Enter zeros in Released Alive Total boxes for species recorded Kept Observed or Unobserved Code Sample # as R, terminate interview Refused:-----Code Sample # as R, terminate interview Don't Know:-----RELEASED WITH DESCENDING DEVICE: Ask only if any species of rockfish were reported as Released Alive. Of those <# released alive> <rockfish species> released alive, were any released using a descending device? Yes:-----Record number released using a descending device in (w/DD) Don't know-----DK No:----Record zero in (w/DD) Sampler didn't ask----DA Refused:----R No Rockfish Catch:---Leave blank RELEASED DEAD: Were any fish released dead? Probe for any fish that were thrown back dead. Yes:----Record species and number of fish No:----Enter zeros in Released Dead boxes for species recorded Kept Observed or Unobserved Code Sample # as R, terminate interview Refused:----Code Sample # as R, terminate interview Don't Know:-----SEAL TAKE: Ask only if boat targeted salmon. Did you see any seals or sea lions take your fish from your line? Yes:----Record number of fish lost to pinnipeds in the seal take box in the same row with the salmon catch

Enter zero in seal take box in the same row with No:----

the salmon catch

Refused:----R Don't know-----DK

No Salmon Catch:----Leave blank

Sampler didn't ask----DA CATCH LOCATION: Where were most of the <species> caught?

NO CATCH: Where did the boat spend most of its time fishing today?

The priority order of the location is for 1) landed fish, 2) reported fish, or 3) majority of fishing time. If the anglers report locations by species and time allows, record the location for each species observed or reported.

 Refused:----- R

 Don'tknow----- DK

 Sampler didn'task--- DA

Block-Box:---- BBB-bb-bb-bb (up to three boxes for one block)

Lat & Lon:---- Enter the latitude above the longitude.

1) Degrees, minutes and grid

(DD.MM/DD.MM+GG)

2) Degrees, minutes and seconds

(DD.MM.SS/DD.MM.SS) where D=degrees, M=minutes, S=seconds, G=area in minutes

NOTE: If the location is above a freshwater cutoff, the boat is not eligible and should be coded as NFOTH.

BOTTOM DEPTH: What was the bottom depth at that location? Record mean depth

 Don't know---- DK

 Depth in Feet:---- FFF

 Sampler didn't ask--- DA

 Refused:------ R

PR Form Layout

Boat samples are recorded in rows with data fields arranged by columns. Each boat row has two sub-rows to record two observations for each item in some fields. Boat sample data may span multiple rows and sub-rows as needed to document additional catch species, fish counts, catch location(s) and depths, and fish bio data. Fish records for a boat may also be continued on the next page; the PR form is double-sided to reduce waste and the front and back of the form are the same.

The form is subdivided into four sections; the header row (sample day), individual boat data (effort), individual fish data (catch and bio data) and subtotal/totals (page summary).

Header Row Items

The header row records data for the sample day. The header includes a unique assignment ID number, date, site information (county, site, port), Sampler ID number and name, additional Samplers present at the site and their ID numbers and if they have data or not, start and end times, and trailer counts. All these items are required.

CRFS PR	RFS PR FORM (V8 11/08/2014)		FORM (V8 11/08/2014)				Tees		Page of		Trailer Counts		
		S		K1 L			Other Samplers:Name & #	(w/da	ta) Time	onsite	offsite		
ASSN ID	Date (MM/DD/YY)	CNTY	SITE	OSP Port	Sampler #	Sampler Last Name			Start 1				
									•)	_	-		
								· v	Stop		1		

Interview Effort Items

Individual boat data include boat sample number, time, total anglers (licensed and unlicensed), days fished, night fishing check box, 12-month avidity, zip code, target species (primary and secondary), water area and gear (for each target). Onsite and offsite missed boats (for select PR1) are tallied on the right side of the form. Launching vessels are tallied only in PR2 sampling mode; leave blank for PR1 mode.

		EFFORT					- ID4	100
Sample # Time	ANGS Total (unlic)	DAYS fished Zip Code	TARGET 1 st 2 nd	AREA	GEAR	onsite	edBt	per tauched
		N 12mos				ons	offsite	0,0
	()	Zip						

Individual Fish Data: Catch and Biological Data

Individual fish data recorded include the species, number landed examined (kept obs), number landed unobserved (kept unobs), number released alive, number of barotrauma-sensitive species released with a descending device, number of fish released dead, number lost to pinnipeds (salmon only), species catch location, average bottom depth, lengths, weights, sex, and head tag numbers.

	65	CATC	li	9	200			BIO DATA				
	KEPT	RE	LS	SPECIES LOC	DEPTH	Fork length / carapace size (mm), sex (M/F/T)						
SPECIES	total		or effort if no catch	воттом	Weight (decimal kg) or (tag ⊭)							
code	unobs	dead	seal take	Block-box Lat / Lon	(ft)	1	2	3	4	5		
	abr	alive	()									
	unabr	dead	soal /						!	-		
	unabr	dead	real						<u> </u>			

Footer Totals

At the bottom of each page, sum the number of refusals and language barriers, total boats (includes fishing and non-fishing), boats targeting salmon or with kept salmon, anglers targeting salmon or with kept salmon, the number of king salmon kept and released (Chinook Salmon, SALCK), the number of silver salmon kept and released (Coho Salmon, SALCO), the number of Pacific Halibut kept and released (HALPA), the number of Yelloweye Rockfish (RFYEY) kept and released, the number of Cowcod (RFCOW) kept and released, the number of Canary Rockfish (RFCAN) kept and released, the number of Black Rockfish (RFBLK) kept and released, and the number of onsite and offsite missed boats. The summary of effort and catch from each page is used to facilitate completion of the assignment summary form and weekly summary report; the data is also used to verify

data entry. The salmon, Pacific Halibut, and overfished rockfish totals allow for in-season estimates of catch and effort to monitor catch quotas.

_																				
	_								$\overline{}$							$\overline{}$			$\overline{}$	
Refu +	Total	Roate	Anac	Kent	Relc	Kent	Rele	Head	Kent	Rele	Kent	Relc	Kent	Rele	Kent	Relo	Kent	Rele	On	Off
ricia -	rotai	Douts	ruiga	rept	IVOIS	rept	Ittels	Houd	recpt	ricia	rept	IVOIS	recpt	Ittis	rept	11010	recpt	Ittoia	Oii	Oil
Barrier	Dt-	Sali		1/2	ngs	0.	ho	T	LIA	LPA	DE	YEY	DEC	OW	DEC	CAN	RF	DLV	1.6	sed
barrier	Dodls	Odli	non	KII	iys	U	MU	Tags	ПА	LITA	INF.	101	REC	/UW	KE	UNIN	KF	DEN	IVIIS	seu

PR Form Item-by-Item Instructions

PR Form Item-by-Item Instructions Field Name Instructions Coding		
Field Name	Instructions	Coding
	HEADER	Examples and Formats
	HEADER	I III DD4
□PR1 □PR2	Check the box for the assigned survey mode.	⊠PR1
Page of	Enter, in sequence, the page number of the	Example: Page 2 of 7
	form and the total number of pages on all pages.	
ASSN ID	Enter the six-digit assignment ID number on all pages.	Assignment ID in the MMDXNN format, where MM is the month ranging from 01-12, D is the CRFS District from 1 to 6, X is the assignment mode and NN is the sequence from 01 to 99.
		Example: 074510 This is the 10 th PR1 assignment drawn in July in CRFS District 4.
Date	Enter the date of the assignment on all pages.	Use the MM/DD/YY format. Example: 07/14/20 = July 14, 2020
CNTY	Enter the 3-digit numeric county code on all pages.	Example: 045 = Mendocino County
SITE	Enter the 3-digit numeric site code on all pages.	Example: 100 = Noyo River Launch Ramp
OSP port (PR1)	For all PR1 assignments, enter the 3-letter alpha code on all pages.	Example: FTB = Fort Bragg Noyo River Launch Ramp
Sampler #	Enter your 3-digit Sampler identification number on all pages.	3-digit numeric code = 305

Field Name	Instructions	Coding Examples and Formats
Sampler Last Name	Write out your last name completely on all pages.	
Other Samplers: Name & #	Write out last name and Sampler # for other Samplers working on this assignment. Circle Y (yes) or N (no)	Example: Smith 132 (<u>Y</u>)
	to indicate if the Sampler has a separate set of data to submit.	
	First PR page only. TRAILER COUNT	
Notes on Trailer Counts	Upon arrival, the first Sampler will enter the total number of trailers in the established trailer count area for that site for onsite and offsite (if applicable) under "Start." At the end of the assignment, the last Sampler will enter the total number of trailers on site upon departure under "Stop."	NOTE: When conducting trailer counts, it is important to include all effort for the site. If the "count area" (ramp parking lot) is full and there are trailers (that are active at the site) parked on the street or other parking area, it is important to include that effort in the counts.
Time [Trailer Counts, Start and Stop]	First PR page only. Record the time you began counting the onsite trailers (Start is upon arrival, and Stop is at the end of the assignment). The times of Sampler arrival and departure from locations where offsite counts are conducted will also be recorded on the ASF.	Use 24-hour military time format. Example: 9:00 AM = 0900

Field Name	Instructions	Coding
Onsite	Onsite refers to trailer	Examples and Formats See the table titled
[Trailer Counts,	count occurring at the	"Summary of PR Counts"
Start and Stop]	assigned site.	for specifics on what to
Clart and Clop	assigned site.	include in the counts.
Offsite	At certain PR1 sites	See the table titled
[Trailer Counts,	count the fishing trailers	"Summary of PR Counts"
Start and Stop]	at a nearby PR site. The	for specifics on what to
	"offsite trailer count	include in the counts.
	area" is listed on the	
	monthly site list or the	
	Lead will provide a list.	
	The Start Count should	
	take place before going	
	to the assigned PR1	
	site. The Stop Count	
	should take place after	
	sampling is complete at	
	the assigned PR1 site. EFFORT	
Sample # [or R	Record a sample	Sample # = 1, 2, 3
or RS or Bl	number in consecutive	Sample # = 1, 2, 3
OT NO OT D	order (starting with 1)	REFUSALS and
	for every boat	LANGUAGE BARRIERS
	intercepted (except for	are NOT issued a sample
	refusals or language	number. Record an "R" or
	barriers). See the table	"B" in the Sample # box.
	titled "Summary of PR	Do not list a target. Do not
	Counts" for specifics.	record as a missed boat.
	For refusals (R) or language barriers (B)	SAMPLE FLAGS
	record an R or B without	KAYAK - record a "K" after
	a sample number. For	the sample number.
	refusals where	PWC, canoes, other small
	minimum salmon data	non-trailered boats, and
	elements are available,	"non-traditional boats" with
	record RS and record	trailers - record a "P" after
	the salmon data	the sample number
	elements.	TOURNAMENT - record a "T" after the sample
	Flag special types of	number
	boats using letter codes	SAILBOAT - record a "S"
	(see right column) after	after the sample number
	the sample number.	·
		NON-FISHING TRIPS:
		Record a sample number,
		and the non-fishing type

Field Name	Instructions	Coding Examples and Formats
		under TARGET. See the
		table titled "Summary of
		PR Counts" for specifics.
Time	Enter a time stamp for	Use 24-hour military time
	every boat that is given a sample number or is a	format.
	refusal or language	Example: 5:00 PM = 1700
	barrier.	
ANGS total	Enter the total number	0 = NF boat only
	of anglers on the boat	3 = three anglers fished
	regardless of license status (licensed	total R or B: code the Sample #
	anglers+unlicensed	box with "R" or "B" and #
	anglers). Code zero for	of anglers if known,
	NF boats.	terminate the interview
ANGS (unlic)	Enter the number of	0= all anglers were
	anglers out of the total anglers fishing who do	licensed
	NOT have a current CA	R or B: code the Sample #
	fishing license of any	box with "R" or "B" and #
	type.	of anglers if known. Leave
	AL CULTURE	unlic. blank. Continue
	Note: unlicensed is a subset of total anglers,	interview if possible, otherwise terminate the
	therefore unlicensed ≤	interview
	total anglers	
DAYS fished	Enter the total number	Example: fishing during
(left column) =	of days the boat fished	daylight hours the evening
trip effort	on this trip. This is recorded as the number	of one day and the morning of the next day =
N = Night	of daylight fishing days	2 days of fishing effort.
fishing	for the boat without	
	returning to port. Some	R or B: code the Sample #
	boats launched from ramps will have the	box with "R" or "B" and terminate the interview
	capability to fish	terminate the interview
	multiple days.	
	Boats that engaged in	
	any night fishing (non- daylight) will be	
	identified by checking	
	the "N" box. If only night	
	fishing occurred, record	
	0 days fished.	

Field Name	Instructions	Coding Examples and Formats
Launch Time	Enter the time that the boat launched from the PR1 site for this fishing trip.	Use 24-hour military time format. Example: 6:15 AM = 0615 If a trip is greater than 1 day in duration, leave the
		launch time blank and make a note on the data sheet.
DAYS fished (right column) = 12-month avidity	Select a random angler on the boat and ask, "Not counting today, how many days have you gone saltwater sport finfishing in California in the last 12	52 days = fishing 1 day/week over the last 12 months Refused = R Don't know = DK Sampler didn't ask = DA
	months?" Use a random method of selection to avoid bias (do not always pick the boat operator).	Note: the largest number entered would be "364."
Zip Code	Select a random angler on the boat and request the zip code of their residence.	Example: 90210 = Beverly Hills Refused = R
	Use a random method of selection to avoid bias (do not always pick the boat operator). May be the same angler that answered the 12-month avidity question.	Don't know = DK Sampler didn't ask = DA Foreign country = 3-letter country code e.g., Ireland = FIE
Target Primary = the	Each intercepted boat is screened to determine the primary and	Example: HALCA = targeting California Halibut
main target/activity for the trip Secondary = the secondary target/activity	secondary activity/target, including fishing and nonfishing activity. Activities/targets are coded using 5 letter alpha codes.	Non-Fishing Codes: NFCOM = commercial fishing trip (non-CPFV). Record F&G numbers for commercial salmon vessels.
for the trip		NFPC6 = CPFV trip

Field Name	Instructions	Coding Examples and Formats
AREA	Targets may be determined by asking the angler(s) "what was the number one and number two fish you were fishing for?" Anglers who don't have specific targets after probing are recorded as UNIFH. If the anglers refuse to provide a target, then code the Sample # box with "R" and terminate the interview. If the anglers cannot speak English, then code the Sample # box with "B" and terminate the interview. Record the water area code where the majority of fishing effort (fishing time) occurred for each primary and secondary target. Note that the AREA of effort and SPECIES location can differ for the same target. Area is left blank for NF trips or blank secondary targets.	*Do NOT record CPFV trips as a PR; record the NF code then sample using the appropriate PC dockside form. NFOTH = Any other boating activity, including maintenance, enforcement, research, sailing, etc. Do not record NF kayaks or personal watercraft. NF sailboats are recorded as NF boats with an "S" flag. R or B: code the Sample # box with "R" or "B" and terminate the interview. N = (nearshore ocean > 3 mi) O = (offshore ocean > 3 mi) B = enclosed bay or estuary M = Mexico Island Codes: F = Farallones 1 = Coronados 2 = San Clemente 3 = Santa Catalina 4 = Santa Barbara 5 = San Nicolas 6 = Anacapa 7 = Santa Cruz 8 = Santa Rosa 9 = San Miguel
		R or B: code the Sample # box with "R" or "B" and terminate the interview.
GEAR	Enter single letter code for the fishing gear used	H = Hook-and-line S = Spear

Field Name	Instructions	Coding Examples and Formats
	by the boat for each target. The gear should be determined and recorded for each primary and secondary target identified. Gear is left blank for NF trips or blank secondary targets. There are two special gears for salmon fishing. The gear should be determined and recorded for each primary and secondary target identified.	T = Troll M = Mooch (salmon only) B = Both M and T(salmon only) N = Bait Net Invert Only Pn = Pot and # Fn = Flat hoop net and # E = Snare C = SCUBA diving D = Free diving Unspecified invert. gear (shovel, rake, gun, etc.) – leave blank and make note on form.
	CATCH	
SPECIES	Enter the 5-letter alpha code for each species or taxon of all fish examined or reported by the boat. Additional rows are used for boats with multiple species catch.	No catch: write "NO CATCH" in the SPECIES box and zeros in catch boxes: - KEPT obs - KEPT unobs - RELS alive total - RELS dead If the anglers refuse to let you see the catch or provide information on the fish caught or released, code the Sample # box with "R" and terminate the interview.
KEPT obs (observed)	Enter the number of fish by species examined for this boat. If no fish of a species are examined, record a zero. Sampler will identify and count each species retained by the boat.	Only fish that the Sampler can see, and count are recorded here. May include fillets that can be counted and identified. Make a note on the form if the daily bag limit is exceeded for a species or group of species.

Field Name	Instructions	Coding
KEPT unobs (unobserved)	Enter the number of kept fish by species reported by the boat that the Sampler was not able to see and identify or count. If no fish of a species are reported as landed but unavailable to examine, record a zero. Probe for catch that may not be remembered, such as bait species.	If the boat refuses both KEPT obs and KEPT unobs, then code Sample # box with "R" and terminate the interview This includes fish used for bait, thrown away as trash, given away, and fillets that are not identifiable or countable, or any other fish that are not available for sampling. This also includes fish that the Sampler can see, but for whatever reason, is not able to count. Make a note on the form if the daily bag limit is exceeded for a species or group of species. If the boat refuses both KEPT obs and KEPT unobs, then code Sample # box with "R" and
RELS alive total	Enter the number of fish by species reported as released alive by the boat. This includes both fish released with a descending device and without a descending device Fish must have been landed first or have been intentionally released. Probe for catch that may not be remembered. If no fish of a species are reported as	Fish appeared alive with no mortal injuries upon release No fish released = zero R or B: code the Sample # box with "R" or "B" and terminate the interview.

Field Name	Instructions	Coding Examples and Formats
	released alive record a zero.	
RELS alive (w/DD)	Enter the number of rockfish by species that were released alive using a descending device. Venting the fish is not a descending device. This field does not apply to non-rockfish species. Note: Rockfish released using a descending device are considered alive.	This field is only applicable for rockfish that are released alive. No rockfish catch = leave blank. Code this box for all rockfish species. If RELS alive total = 0 then (w/DD) = 0 Refused = R Don't know = DK Sampler didn't ask = DA Note: RELS alive (w/DD) is a subset of RELS alive total, therefore RELS alive (w/DD) ≤ RELS alive total
RELS dead	Enter the number of fish by species reported as released dead by the boat. If no fish of a species are reported as released dead, record a zero. Probe for catch that may not be remembered.	Refused: code the Sample # box with "R" and terminate the interview
Seal take	Enter the number of salmon reported taken by pinnipeds for the trip. The angler must have seen the pinniped take the salmon from the line.	This question is only asked if salmon was targeted. No salmon target = leave blank Refused = R Don't know = DK Sampler didn't ask = DA No salmon lost = 0

Field Name	Instructions	Coding Examples and Formats
SPECIES LOC	Enter the location where the majority of the catch species were caught.	Block- Box: BBB-bb-bb-bb or BBB-bbb-bbb-bbb
	If no catch, record the location where the majority of fishing effort occurred.	718-106-107-108 = block and 3 boxes (inland)
	A separate location may be recorded for each species observed or	235-12-14-15 = block and 3 boxes (ocean) 252 = block only
	reported. For trips with large areas of trolling for non-bottomfish species, record a general area.	Block-Box-Grid Size: BBB-bb+g: 212-01+3 = block and one box plus grid size (in nautical miles)
		Lat/Long: Latitude in upper box and longitude in the lower box. Only use whole degrees and minutes (no seconds or decimals). Grid size can also be used.
		37,30+3/118,57 = lat 37 degrees, 30 minutes and long 118 degrees and 57 minutes with a grid size of 3
		37,30/118,57 = lat 37 degrees, 30 minutes and long 118 degrees and 57 minutes
		Refused = R Don't know = DK Sampler didn't ask = DA
DEPTH	Enter the average bottom depth in feet for the catch location. This is not a mid-water depth of capture.	100 = 100 feet 100 min/120 max = enter as mean depth 110 feet = 110

Field Name	Instructions	Coding Examples and Formats
	Enter a single depth or if a range is given enter the mean depth.	Refused = R Don't know = DK Sampler didn't ask = DA
	The depth should be recorded by species when possible.	
	BIO DATA	
Fork Length (mm)	In the top row enter the fork length for the fish in mm. Add an M, F, or T after	321 = fork length in mm F = Female M = Male T = Transitional (CA
Sex	the length for sexed species. Do not measure a	Sheephead) 333F = female fish 333 mm fork length
	salmon with an intact adipose fin.	Ü
Weight (decimal kg) or Head Tag #	Below the length, enter the weight of the fish in kg. Do not weigh salmon.	5.3 = weightin kg
	For salmon, Yelloweye Rockfish, and White Seabass enter the headtag number below the length and circle the headtag number.	12345 NRS = adipose fin- clipped salmon head not recovered
	For salmon heads not recovered or lost, record the head tag number and code NRS (non-recoverable species).	
	Salmon and groundfish headtag numbers are 5 digits.	
	MISSED BOATS	
Missed Bt onsite	Enter the number of boats that returned to the sample site that	This includes un-sampled or missed boats

Field Name	Instructions	Coding Examples and Formats
	were not sampled since the last sampled boat.	See the table titled "Summary of PR Counts" for specifics
	Tally marks can be recorded in the box, then the total is recorded when the next	Refusals are NOT missed boats
	sampled boat comes in.	Language barriers are NOT missed boats
		SAMPLE FLAGS K = KAYAKS P = PWC, paddle boards, canoes, other small nontrailered boats, and "nontraditional boats" with trailers S = SAILBOAT
		Example: 2K = 2 fishing kayaks It is OK to put multiple numbers and flags in a row (line). For example, you could list: 2 = 2 recreational fishing boats missed AND 1K = one fishing kayak missed. Page Tot = 3
Missed Bt offsite	Enter the number of fishing boats that returned to an offsite boat area since the last sampled boat.	See the table listing sites with associated offsite missed fishing boat counts
	See the table titled "Summary of PR Counts" for specifics.	
PR2 Launched (PR2)	This data is <u>only</u> needed for PR2 samples.	Leave blank for PR1 sampling mode
FOOTER The footer contains the sum of the page totals for each category below.		
Refu + Barrier	Enter the sum of refusals and language barriers for the page	Count the number of R and B entries in the

Field Name	Instructions	Coding Examples and Formats
		sample# column. Do not
		include RS samples
Total Boats	Enter the sum of intercepted boats on the page.	Total Boats = sampled finfish boats + eligible invertebrate only boats+ non-fishing boats
		Does NOT include missed boats or refusals/language barriers, but does include RS samples
Salmon Boats/Angs	Enter the sum of number of boats that targeted and/or kept salmon on the page/sum of anglers for these boats.	A boat/angler(s) that kept salmon while targeting other species would be tallied as a salmon boat with salmon angler(s)
Kings kept/rels	Enter the sum of observed and reported kept and released alive and dead king (Chinook) salmon on the page.	
Coho kept/rels	Enter the sum of observed and reported kept and released alive and dead silver (Coho) salmon on the page.	
Pacific Halibut kept/rels	Enter the sum of observed and reported kept and released alive and dead Pacific Halibut on the page.	
Yelloweye kept/rels	Enter the sum of observed and reported kept and released alive and dead Yelloweye Rockfish on the page.	
Cowcod kept/rels	Enter the sum of observed and reported kept and released alive and dead Cowcod on the page.	
Canary kept/rels	Enter the sum of observed and reported kept and released alive	

Field Name	Instructions	Coding Examples and Formats
	and dead Canary Rockfish on the page.	
Black kept/rels	Enter the sum of observed and reported kept and released alive and dead Black Rockfish on the page.	
Missed boats on/off	Enter the sum of missed onsite and offsite boats on the page.	Do NOT sum by sample flag type (i.e., K, P, S). Sum all missed boats together

Specific editing checks:

- Check that offsite start and stop counts and/or offsite missed boats are appropriately present or not present depending on the PR1 site sampled.
- Check that missed boats are coded on each <u>boat</u> row. NOT on rows with just catch and bio data.
- 3. Check that all pages are present and numbered sequentially.
- 4. Check that there are no missing gears and that catch location coordinates are coded in the correct format.
- Check that all fish of a species are listed consecutively (if more than 5 measurements) and, if not, that there is clear indication of where the rest of the measurements are so that the data can be entered consecutively.
- Make sure fish sex is in correct position (after length). Do not circle fish sex code.

Summary of PR Counts

Type of Boat, Angler or Trailer	Interviews and Sample Number	Onsite Missed	Offsite Missed (PR1 sites only)	Trailer Counts: Onsite, Offsite and Pressure Checks
General Rule		NEVER A	DJUST THES	E COUNTS
non-fishing boats (NFPC6, NFCOM, NFOTH) See below for info on kayaks, PWC and sailboats	Interview & record sample #	Do NOT include boats that can be identified as NFPC6 or NFCOM. Include all other traditional trailerable boats	Do NOT include in count	Do NOT include trailers that can be identified as NFPC6 or NFCOM or NFOTH Include all other traditional trailers
kayaks, PWCs, canoes, other small non-trailered boats, and "non- traditional boats" with trailers (e.g. jet skis, dinghies) Use the flag K for kayaks. Use the flag P for PWCs, canoes, other small non- trailered boats, and "non- traditional boats".	Interview fishing boats & record sample # with K or P flag Do NOT interview non-fishing boats & do NOT give them a sample #.	boats & flag		Do NOT include in count. This means, do NOT include: jet ski trailers trailers that can be identified as for sailing dinghies vehicle with racks for boats vehicles without trailers Do NOT adjust counts for kayaks etc. interviewed

Type of Boat, Angler or Trailer	Interviews and Sample Number	Onsite Missed	Offsite Missed (PR1 sites only)	Trailer Counts: Onsite, Offsite and Pressure Checks
sailboats	Do NOT interview non-fishing dinghies. Treat like kayaks or PWC. For larger sailboats, interview & record sample # with S flag for both fishing and non-fishing (NFOTH)	Include in count & flag with S if fishing		Do NOT include trailers that can be identified as sailboat trailers in count If you can't determine or don't know that a trailer belongs to a sailboat, then include it in the count
trailers with no vehicle attached or "abandoned" trailers	Not Applicable	Not Applicable	Not Applicable	Do NOT include in count
refusals (R) & language barriers (B)	Interview & record R or B (no sample #)	Not Applicable	Not Applicable	Not Applicable
Vehicles with no trailers				Do NOT include in count. Do NOT adjust counts for boats that have been interviewed that are not trailered

PR1 Form Example - Salmon

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Page 1 of 5 Other Samplers:Name & # (w/data)			BIO	Fork length / carapace size (mm), sex (M/F/T)	Weight (decimal kg) or (tag #	m									y sho	fish w	(*2 NRS* Angler took fish.		(No Catch)	give	0	Kept Rel	guel,
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PR1 Form Example - Non-Salmon

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Secondary Private and Rental Boat (PR2) Mode Sampling

Introduction

Although all fishing modes are sampled, CRFS puts more emphasis on fishing from boats, where the majority of managed fish species are caught compared to other modes. The <u>private and rental boat</u> (PR) mode fishery is the largest in the state in terms of total catch. The PR fishery is also seasonally and geographically irregular. The publicly accessible sites where private and rental boats launch are stratified into primary sites (PR1) and secondary sites (PR2). To divide sites into the two strata (PR1 and PR2), data for "important management species" were analyzed separately for sites north and south of Point Conception. Important management species were defined as those with active fishery management plans and include salmon, groundfish (e.g., rockfishes, Lingcod, Cabezon, California Scorpionfish, flatfishes, and some sharks and rays), highly migratory species (tunas, billfishes, Dolphinfish, and certain oceanic sharks), and species in the California Nearshore Fishery Management Plan.

PR2 sites are defined as publicly accessible launch facilities where less than 10 percent of the private and rental boat catch of "important management species" has been landed historically. The sampling procedures for PR1 and PR2 are similar, and the same forms (PR Form and ASF) are used for both PR1 and PR2. Differences in the sampling procedures for the two strata are listed in the table below.

Differences between PR1 and PR2 sampling procedures.

Sampling Procedure	PR1	PR2
Time on site when trailers present	From the return of the first boat until the last boat returns or sunset (whichever is first)	Approximately 6 to 7 hours during daylight hours
Scheduled start time	Lead will assign a start time	Early or late start as defined by Lead each month
Count boats launching	No	Yes
Count offsite missed boats	Counted at some PR sites	No

PR2 GOALS

The primary goals for PR assignments are to:

- ✓ Obtain accurate counts of the boats and anglers using the site.
- ✓ Obtain high quality, interviews and catch data
- ✓ Observe all kept salmon
- ✓ Collect heads of all adipose fin-clipped salmon

Effort Data

The goal is to estimate total fishing effort for the day. This is done by counting trailers and returning boats and determining the number of anglers on each fishing boat. For each boat we must determine the primary activity. If the boat is fishing, we determine the target fish species and the number of anglers per boat. Effort is expanded to account for weekend (and holidays) and weekday days not sampled. The effort estimate is in angler trips by target fishery group. Additionally, in PR2 mode we collect the number of boats launching. These data along with the trip length data is used to generate better effort profiles (i.e., effort by time of day).

Catch Data

An additional goal is to estimate catch per angler-trip. Catch per angler-trip is determined by counting the number of each species that is kept and recording the number of each species that are reported as released or otherwise unavailable to examine. Estimation of effort and estimation of catch per angler-trip are each calculated for a PR1 site, month, kind of day (weekend/weekday), water area, and trip type (target). Effort is calculated as the total number of anglers sampled during the time period, adjusted for un-sampled anglers, and expanded for the total available weekend or weekdays per month. Catch rate per angler-trip is calculated from the sum of catch recorded from sampled anglers divided by the total sampled anglers. Total catch is the product of estimated effort and estimated catch rate.

Location of Catch Data

Another goal is to collect data on the location and depth of catch. These data are determined by showing the boat operators maps of the area and asking them to point out specific locations and average depths of their catch. The data is used to apply depth-based mortality estimates to some released species and summarize the catch estimates in depth ranges and by geographic areas. The data can also be viewed in a GIS for trends in catch. This information is required to manage the fisheries by depth and geographic area.

Biological Data Collection

Another goal is to sample lengths and weights of landed catch. Lengths will be used in a regression to calculate a predicted weight for fish without a sampled weight, and to examine the size distribution of the landings. Sampled weights are used to calculate average weights by species. These average weights are multiplied by estimated total catch by species in numbers of fish to estimate total catch in metric tons. Metric ton estimates are used to evaluate catch quotas and the rebuilding status of some distressed species. Numbers of fish for quotas and evaluating status of ESA listed stocks is used in salmon management. Note: do not measure or weigh non-adipose fin-clipped salmon; only adipose fin-clipped fish need to be measured for fork length, prior to head removal. Salmon are managed in numbers of fish, and not by weight.

Sample Selection

The PR2 sampling schedule takes into consideration the effort of all PR2 sites within the District. Sites with high effort have a higher probability of being drawn for sampling than those with low effort. Based on random chance, all PR2 sites may not be sampled every month. Some PR sites may change from PR2 to PR1 and back to PR2 on a monthly basis, depending on fishery season openings and closings and effort changes throughout the year. Be sure to check the District site list every month.

Scheduling

The Lead will schedule the random selection of days for sampling for each month in advance. The number of PR2 assignments sampled each month depends on the number of active PR2 sites in the District and the number of kind of days available. The two kinds of days are weekends/holidays and weekdays. Effort is expected to be different for these kinds of days. Rescheduling of PR2 assignments will disrupt the random selection of samples reducing the statistical validity and representation, and if allowed, will respect separation of the kinds of days and may be done only with the Lead's approval. Zero effort days are included in computation of the effort, but do not require that a Sampler stay at the site all day to be complete (see Assignment Duration). Samplers should expect an erratic schedule as PR2 sites can have varying effort dependent on the fishing seasons, ocean conditions, etc.

PR2 SURVEY PROCEDURES

Effort Data Collection

One goal is to determine the activity, i.e. effort, of every boat returning to the site during the PR2 shift. A specific set of data must be collected for every boat that returns to the PR2 site. For every boat intercepted the time, number of anglers (licensed and unlicensed), and the target(s) (species or activity) should be recorded. For non-fishing (NF) boats (recreational or commercial activity type), record the specific non-fishing activity for the primary target. See Non-Fishing (NF) Boat Types.

Boats targeting invertebrates are sampled as well, just like finfish boats, regardless of whether they had finfish bycatch. See the Species Sampling Chapter for more information on invertebrate trips.

PR2 Assignment Duration

Up to eight hours is allotted for each PR2 assignment. This includes travel time to and from the site to your headquarters and should allow the Sampler to be on site for six to seven hours.

On average, the highest number of boats returning per hour is between 1300 and 1600 hours. This varies by time of the year, location, target, and weather conditions, but the sampling duration at the site should include the time period when most of the boats are returning. Sampling early and late returning boats is also important as the species composition for those boats

may differ from the boats returning during the peak period. The Monthly Schedule will list **early** or **late** start times for each PR2 assignment for the purpose of varying the time on site. The Lead will set the time for the early and late start times each month based on knowledge of the fisheries and the daylight hours available. An early start time might be 0900 hours, and a late start time might be 1100 hours.

Low Effort Protocol

The Sampler will stay onsite if there is known/suspected finfish effort, until eight hours has been spent on the assignment (including travel time), or sunset.

No Boats in PR2 Mode

If there is no effort at a PR2 site, the Sampler should stay for a minimum of 2 hours to see if effort develops. If no effort develops on a zero-effort day, the Sampler terminates the assignment and it is considered complete.

Arrival and Trailer Counts

PR2 sites will be sampled for effort and catch during daylight hours, at a single site. The Sampler will arrive based on the start time defined in the Monthly Schedule and depart after 6-7 hours of sampling.

Trailer counts are used to estimate effort for the day. A starting trailer count will be conducted upon arrival of the first Sampler. All boats returning to the site during sampling hours will be intercepted. Counts of "trailers" include traditional boat trailers. Personal watercraft (PWC) trailers, car top boat carriers, boats loaded into the beds of trucks, non-trailered inflatable boats, and kayak and canoe carriers are excluded. See the Summary of PR Trailer Counts table on page 11-38 for more information.

Onsite Trailer Count

The Sampler should count the number of trailers in the parking lot and any adjacent streets or parking lots (consult the Wiki or a site description book to determine the count area for each site). Do not count trailers not attached to vehicles, or known non-fishing, commercial or CPFV trailers. Some PR2 sites consist of a boat rental shop where there are no traditional trailers; in these instances, ask the rental shop about how many boats were rented. Immediately before leaving the site at the end of the day, the Sampler will count the number of trailers remaining in same area. Known commercial or CPFV trailers should not be included; all others are included in the stop count

Onsite Missed Boats

Avoid missing boats at the PR2. If a boat is completely missed while sampling other boats, it is considered an onsite missed boat. Once a boat is canvassed for activity it is not considered missed and a conservative attempt should be made to conduct a CRFS interview. Refusals and language barriers do not count as missed boats. Onsite missed boats do not have a time, target species, or number of anglers recorded; they are simply tallied with the current boat the Sampler is interviewing in the left-most missed boat column on the PR Form. Record "K" next to the number of missed fishing kayaks. Page totals for onsite missed boats are tallied at the bottom of each PR page, and assignment totals for all onsite missed boats are tallied at the bottom of the ASF.

Limited Activity at the Site

Since many of the PR2 sites have relatively low effort, especially in the winter, Samplers should be prepared to conduct other work while waiting for boats to return. Work that might be completed includes editing forms, reviewing the manual, studying fish identification, and reviewing outreach materials.

Offsite Trailer Counts

Several PR2 sites are near one another, and catch, effort, and species composition are similar. These are nearby launch ramps or boat rental facilities where the sampler will obtain start and stop counts of trailers or fishing boats out at a rental facility. Before and after the PR2 assignment you will stop by the offsite count site and get a start count before your PR2 assignment and a stop count after your PR2 assignment. You will not be obtaining any interviews at the offsite count site. If it is a rental facility the sampler may be able to call and see how many fishing boats are out and use these for a start and stop count. These counts are recorded in the offsite trailer count box in the upper right hand of the PR form.

District	Sampled PR2 Site	Offsite Start and Stop Counts Obtained
1	073-310 Chula Vista Launch Ramp	073-310 National City Launch Ramp
1	073-112 Glorietta Launch Ramp	073-118 Coronado Boat Rentals
1	073-018 Seaforth Boat Rentals	073-119 Dana Landing Rentals
3	053-104 Moss Landing Launch Ramp	053-105 Woodward Boat Ramp
4	001-100 Berkeley Launch Ramp	001-107 Emeryville Launch Ramp

4	097-100 Westside Launch Ramp	097-105 Doran Park Launch Ramp
4	097-108 Ocean Cove Launch Area	097-109 Salt Point Launch Ramp
4	097-107 Timber Cove Launch Area	097-110 Stillwater Cove
5	045-100 Noyo River Launch Ramp	045-104 South Harbor Launch Ramp

Launched Boats

In PR2 mode, boats that launch are also tallied. **Record all boats that launch**, regardless of activity (except non-fishing kayaks or PWC). Fishing kayaks, fishing PWCs, and sailboats are flagged with a "K", "P", and an "S" respectively. PWCs include canoes, other small non-trailered boats, and non-traditional boats with trailers. Launching boats are tallied in the "PR2 Launched" column. Do not include known commercial and CPFV boats in this count.

Catch Data Collection

All private boats that have completed a fishing trip should be sampled for catch. Catch includes landed catch (fish brought ashore) and reported catch such as discards or other catch not available. The Sampler may have to interview all anglers on the boat to determine total catch since anglers may not be aware of each other's catch. This determination may need to be done before the driver leaves to get the trailer. The goal is for the Sampler to observe all finfish catch to identify to species, measure and weigh as many fish as possible, and document all unobserved catch for each boat.

Biological Data Collection

After determining the catch by species for the boat, the Sampler will measure and weigh as much of the catch as possible. It is important to the CRFS program to measure fish that are under active management, especially species of concern. A prioritized list of species to preferentially sample is provided (see Priority Species). Lengths can be used to predict weights and to examine length classes; however, recording length-weight pairs is the goal for bio data collection. Do not weigh any salmon species, and only record lengths of adipose fin-clipped salmon.

Sub-sampling Lengths and Weights

There may be times when the level of activity at a site is too high to sample the lengths and weights of every fish on every incoming boat. The Sampler should attempt a random or systematic sample of fish in this case, following the priority list. Refer to the section, General Onsite Procedures: Catch Measurement.

Catch Location and Average Depth Data Collection

The Sampler will attempt to determine the location and average depth of catch by species, or the location and depth of the majority of the boat's fishing effort if there is no catch. Maps with depth contour lines are provided to assist the angler in determining the catch location(s) and depths. If all species were caught within the same location and depth, then only one location and depth may need to be reported. Often, locations and corresponding depths may need to be reported separately for individual species or species groups. For suspect data, rare species, and especially for overfished species, double check the catch location and average depth with the angler. For trips with large areas of trolling (for non-bottomfish species), a general area can be used. Catch location is used to manage fisheries by geographic boundaries.

Sub-sampling Locations and Depths

There may be times when the level of activity at a site is too high to sample the locations and average depths of all catch on every boat. In these cases, the Sampler should attempt a random or systematic sample of more specific locations and depths for bottom-fishing boats. This allows some boats to give a single more general location to save time. Boats targeting surface fishes (tuna, salmon, seabass, etc.) may be coded with the general locations and depths as well, when time is short. It is important to document location and average depth for trips with catch of non-retention species and species on the Priority Species List.

Minimum PR Sample

A CRFS sample is defined as a boat which has been sampled for both effort and catch. Catch locations, average depths, and bio data are not required for a valid CRFS sample. Note: the data elements below are the minimum required for a valid sample; Samplers should make every effort to collect the data needed to complete EVERY field.

The following data elements are the minimum requirements for a useable CRFS interview:

- Total number of anglers who fished
- Number of days fished
- Target
- Area fished (water area, e.g., Nearshore=N)
- All catch, unobserved number of fish by species

The minimum items for this interview are listed above. Fish measurements may be omitted but fish counts may not. Never code rockfish to the genus level to save time. If the minimum requirements cannot be met, the boat will be considered a Barrier or Refusal or tallied as a missed onsite boat depending on the nature of the interaction.

Screening Divers

In addition to hook-and-line anglers, divers may qualify for the CRFS interview. If a diver carries a spear gun with them, they can be interviewed as an 'angler/s.' If they spear a fish or intended to spear a fish they are

considered an eligible angler and can be interviewed with gear code "S." Divers taking or intending to take invertebrates are also eligible to be sampled (see the General Onsite Procedures section). Divers entering the water from the shore using fins and a flotation device (such as a dive tube) to fish are considered either BB or MM anglers. Divers who enter the water from a boat or other craft are considered PR anglers. This includes kayaks, stand up paddleboards (SUPs) and pontoon boats with 'oars.' In effect, having a paddle is what designates the mode as PR.

Fishing Tournaments

For the purposes of CRFS sampling, a tournament is defined as a site and date specific fishing contest. Contestants usually must return to tournament headquarters by a certain time for the "weigh in" which limits how far they can travel, and only specific species of fish may be taken on the date of the tournament event. Generally, tournaments are not sampled. Once the Sampler determines that a tournament is taking place, the Sampler should contact the Lead immediately to discuss the situation. It is important to notify the Lead in advance when a tournament date and location is discovered so that the Lead can make appropriate arrangements, if necessary. If the Lead determines to sample as scheduled, a sample flag of "T" should be used for all boats sampled that are participating in the tournament.

Informal 'pools', such as those arranged on CPFVs (jackpot contests), are not considered tournaments—anglers participating in these types of contests should be sampled as usual. Some fishing contests are regional (encompassing a large area) and span a long period of time (a week, a month, or an entire fishing season). Participants usually pre-register at a location, such as a tackle or bait store, and may bring qualifying fish in during the entire time the contest is running. For the purposes of CRFS sampling, these types of contests are called derbies. Anglers participating in derbies should be sampled as usual.

THE PR FORM (PR2 MODE)

The PR Form collects total boat effort for the day by counting trailers and sampling returning boats. Each boat is screened as fishing or non-fishing. For fishing boats, determine target fish species and number of anglers per boat. In northern California during salmon season, the form will also count all retained and released salmon as well as record lengths of adipose fin-clipped salmon head tag numbers. For boats with catch, all the fish are counted by species along with location(s) and average depth(s), and observed fish are measured and weighed.

Questionnaire Usage

Samplers are given a laminated copy of the **questionnaire** used with the PR Form. The questions for the interview are written out, in full for standardization. The Sampler should word each question specifically <u>as it is written in the questionnaire</u>. In order to have meaningful comparative data, each angler should respond to a standardized stimulus. <u>Methodological</u>

studies have shown that even slight changes in questionnaire wording, for example "should" versus "could," drastically influence responses.

Introduction to the PR2 Interview

Tasks while sampling boats are generally done in this order:

- 1. Determine if anyone on the boat has fished
- Determine the total number of anglers and of those, the number of unlicensed
- 3. Determine the launch time of the boat
- 4. Determine zip code of one random angler
- 5. Determine total days fished on trip
- 6. Determine if night fishing occurred
- 7. Determine the 12-month avidity for one random angler
- 8. Determine the target species and gear (or non-fishing activity)
- 9. Determine the primary area fished for the fishing target(s)
- Determine if any catch (including discards) or marine mammal losses (salmon only)
- Determine how many of each rockfish species a descendingdevice was used for release
- 12. Count catch by species (mandatory for salmon species)
- Determine the location and depth of the catch, or if no catch, where the majority of fishing effort occurred
- Record finfish length measurements and weights of the catch (prefer length-weight pairs)
- 15. Depending upon region: collect salmon and/or White Seabass heads and Yelloweye Rockfish

Before the Assignment

The Sampler should check their equipment and forms before leaving for the site. This will ensure that the Sampler has enough forms and other supplies to complete the assignment. Be aware of the weather forecast. In northem California during salmon season, be sure additional salmon equipment and tags are on hand. In southern California, make sure to have a white seabass wand if one has been issued. Double check the date, site, port and assignment ID. Record site information, Sampler name, and ID number on the PR Form and on the Assignment Summary Form (ASF). Plan to arrive on site at the time designated by your Lead.

Arrival on Site

Upon arrival at the PR2 site, count the number of trailers in the parking lot and any adjacent streets or parking lots (consult the CRFS Wiki site or the site description book to determine the count area for each site). Record the arrival time on the ASF and the arrival trailer count in the start count box on the first PR Form.

Sampler Location Onsite

There are differences among PR2 sites regarding onsite positioning for obtaining interviews. For example, boats might be interviewed while they are waiting for a boat hoist, while they are cleaning their boat at the wash down

station, at the dock, on the beach, or at the ramp. The Sampler will use discretion in determining the best approach at any particular site. For most PR2 sites, the best spot to sample is where the boats are waiting for their turn to exit the ramp. If boat traffic is heavy, do not conduct interviews on the dock or ramp, as this may delay the trailering process which may result in unhappy anglers.

Monitoring Boats

When a boat arrives at the PR2 site, a new sample is created with the time of arrival. During very busy times, a boat may arrive and will not be sampled because the Sampler(s) are busy with other boats. This boat will be tallied on an existing boat row as an onsite missed boat in the onsite missed boat column. An onsite missed boat may be either a non-fishing boat (NF) or a fishing boat. The proportion of fishing to non-fishing sampled boats is applied to the count of onsite missed boats to estimate several additional fishing boats. It is expected that missed boats will have the same proportion of NF to fishing boats as the boats sampled. This assumption is a potential source of bias. For example, if all the missed boats are fishing boats, but half the boats actually sampled were NF boats, then the estimate of fishing boats missed will be underestimated by 50% because missed boats were not representative of the boats sampled. Therefore, onsite missed boats should be a representative selection of all boats, not just fishing boats or boats that look like a lot of work to sample. During salmon season every effort should be made to avoid missing boats. Ideally, there should not be any missed boats. Once a boat has been canvassed and the target is either finfish or invertebrate the minimum CRFS interview is required. Contact your Lead immediately if additional help is needed to avoid missing boats.

Multiple PR Trips on the Same Day

Occasionally PR boats will make more than one trip per day; sometimes the skipper drops off passengers from a morning trip and takes a new crew out on a second trip in the afternoon, or the crew may remain the same after returning from the first trip of the day. The Sampler may recognize the boat as having been sampled earlier in the day, or the crew may point out that they have already been sampled at the completion of their first trip. Regardless of how this second (or subsequent) trip is discovered, the Sampler is to treat these trips separately, and attempt to sample both as distinct trips each with unique data - separate sample numbers, different launch times, segregated catch, etc. Do not combine both trips into one sample. If the catch from both trips is still onboard at the completion of the second trip and the crew is unable to separate catch by trip, the Sampler is to record catch from the second trip as angler reported (kept unobserved). If anglers are reluctant to participate in the survey again, point out that each of their trips is unique, and it's important for CRFS to capture data from each and every unique trip - perhaps the boat had different targets, fished in a different location, or caught a different composition of species.

Determination of Boat Type

A category based on activity must be assigned for each boat intercepted. Ask a passenger on the boat as to its activity for the day. There are, essentially, two types of boats in the PR survey: Fishing and Non-fishing (NF). A fishing boat is defined as a boat, either privately owned orrented, upon which fishing effort (for finfish OR invertebrates) occurred. Boats that targeted invertebrates only are considered fishing boats. Catch is not necessary to be considered a fishing boat. Boats that intended to fish but did not put gear in the water are NF boats. A CPFV carrying passengers paying to fish is not considered a fishing boat for the purposes of PR mode sampling.

Non-Fishing (NF) Boat Types

There are three NF codes currently being used:

- NFCOM a commercial fishing boat targeting finfish or invertebrates (note: occasionally a commercial fishing boat may be fishing recreationally that day – the boat would be sampled just like any other PR boat).
- 2. NFPC6 Commercial Passenger Fishing Vessels, also called party/charter (PC) boats, vessels that are permitted to take paying passengers fishing. This includes smaller, trailered "6-pack" boats. The Sampler may have to inquire with the operator to determine if the boat was a regular PR boat or was fishing as a CPFV that trip.
- 3. **NFOTH** all other non-fishing boats fall into this category. This includes boats that intended to fish but for whatever reason had no wet-gear time, cruises, sailboats that did not fish, bird watching, whale watching, burials at sea, enforcement, research, etc.

CPFV and Commercial Boats

Commercial Passenger Fishing Vessels (CPFV) are coded as "NFPC6" on the PR Form. If the Sampler encounters a CPFV at the PR2 site, the boat is coded as "NFPC6" in the Target field of the PR Form. The boat should then be sampled opportunistically using the appropriate PC dockside sampling. Commercial fishing boats are coded as NFCOM in the Target field of the PR Form; commercial fisheries are sampled using other non-CRFS surveys.

Opportunistic PC Sampling

Commercial Passenger Fishing Vessels (CPFV) that utilize a PR site are coded as "NFPC6" on the PR Form. Monitoring PR effort during a PR assignment is a priority; if time allows and without missing any PR effort, the Sampler should sample the CPFV using the appropriate PC dockside sampling form – the CRFS-OSP SALMON CPFV DOCKSIDE form for trips that targeted salmon only, and the CRFS PC (CPFV) DOCKSIDE form for trips that targeted something other than salmon. If the boat targeted both salmon and non-salmon on the same trip, sample the boat using both forms, recording data on the appropriate form. Report all CPFV activity to the PEC Port Lead (Districts 3-6) or record the vessel's effort on a PEC form (Districts

1-2). See CPFV Dockside Sampling sections in this manual for more information on sampling CPFVs dockside.

Refused Boats

Participation in this survey is voluntary. An angler may refuse to participate. However, this data is crucial to sustainable fisheries management, so the Sampler should try to get as many questions answered as possible. Some anglers on the boat may be more receptive than others.

Although refusal to answer key CRFS questions will be coded as a refusal, salmon minimum data element requirements will allow for saving a sample when CRFS minimum interview requirements are not met. Anglers are required to make kept salmon available for sampling (Title 14, CCR, Section 1.73(b)); minimum requirements for a valid salmon sample include number of anglers, kept salmon by species, and salmon with adipose fin clips—code these boats as RS in the Sample #. Zip code, avidity, location, and depth are not necessary for a "valid" salmon sample; however, these items are important. If you cannot get all the required questions answered, you will have to record the boat as a refusal; code an "R" in the Sample # field. Refusals do not get a sample number, just an "R." Refused boats are not tallied into the total boats on the PR2 page totals. If you can collect the minimum salmon requirements the boat would be coded "RS" and included in the total boats on the PR2 page totals.

Language Barrier Boats

Anglers who cannot speak English are usually not able to effectively answer survey questions. If there is too much of a language barrier, the Sampler should stop the CRFS interview. If all the required questions are not answered, the boat is recorded as a barrier; code a "B" in the Sample # field. Barriers do not get a sample number, just a "B." Language-barrier boats are not tallied into the total boats on the PR page totals.

Anglers, Zip Code and Days Fished

Once the Sampler determines the boat is an eligible fishing boat and willing and able to participate, determine the angler effort on the boat. Some of the passengers may not be anglers. The Sampler will determine the number of anglers who actually fished. Next, the Sampler determines the number who fished without a valid CA fishing license. The number of unlicensed anglers will always be equal to or less than the total number of anglers on the boat. It is best to determine this indirectly by asking what type of fishing license the anglers used. Often, the anglers will want to show their licenses—Samplers do not need to see their licenses to code them as licensed anglers. The number of unlicensed anglers is used to adjust effort from the licensed angler telephone survey; children are not eligible to participate in the telephone survey, and some anglers are not required to have a license and so would not be a part of the telephone survey.

The final item required to estimate effort on the boat is the number of days fished. Usually this will be one day; however, some boats, especially in

southern California, may have taken multi-day trips. The "N" box will be checked if the boat fished at night (after dark the night before until dawn of the current day). If only night fishing occurred, the "N" box will be checked and "0" days fished will be recorded. If it is a multi-day trip, record the number of days fished, leave the launch time blank, and leave a note on the data sheet.

One of the anglers on the boat will need to provide a zip code. This is the zip code of the permanent residence of the angler, not temporary lodging. If the angler is from a foreign country, use the applicable foreign country code. The zip code is used primarily to help quantify the economic role of sportfishing. The angler asked should be at random, not biased by boat ownership, fishing skill, age, gender, etc.

Determination of Catch

The Sampler will determine if any fish were caught by the boat. Each fishing boat will need a complete census of catch. The term "catch" includes observed and unobserved kept fish and released fish. Catch includes landed fish, fish given away, taken by marine mammals (salmon only), used for bait, filleted or eaten, <u>AND</u> fish purposely released, thrown back alive (shakers) or dead. Anglers may report that they have no fish on the boat. However, a boat may still have catch if they caught and released fish. Be sure to inquire about anything that was caught and then used for bait or any other fish that were caught but not available for the Sampler to observe.

Examining Catch

The Sampler will examine all landed finfish catch for each fishing boat. Examined or observed fish are the most robust because the Sampler actually saw, counted, and identified the catch to species. If the boat refuses to have the landed catch examined, all catch are coded as "kept unobserved". It is more important to count and identify rockfish to the species level than to get lengths and weights from those fish.

Q. What if the ramp is busy and I don't have time to count each rockfish species? Can I just code rockfish genus "RFGEN"?

A. No, you must record catch to species. The only time you should be using the RFGEN code is for unobserved catch that the angler simply cannot identify, even with identification guides. There will often be at least one other Sampler there to help you avoid missing boats; if you are unable to keep up with the boats as they come in, stop collecting bio data.

Observed Catch (Sampler-Examined)

The Sampler will attempt to observe and examine all retained finfish catch, recording the number of fish kept and observed by species in the appropriate box on the PR Form. It is important to note that only fish that the Sampler sees, <u>and</u> counts can be recorded as "kept observed". Fish not able to be physically viewed and counted by the Sampler must be recorded in the "kept unobserved" box. It is important to the CRFS program to differentiate

between Sampler-examined and angler-reported fish counts. Estimates of total harvest are summarized separately for the Sampler-examined and angler-reported catches.

The Sampler may identify fillets with skin patches, being careful not to double count fish (i.e. two fillets equals one fish). Fish identified by skins are considered "kept observed." Anglers may not want the Sampler examining fish that have been filleted. These fillets are someone's dinner, and they may not want to get their food dirty or open a tied bag. Ask the angler before attempting to examine fillets.

Unavailable Catch (Angler-Reported)

In addition to any fish the Sampler sees, each fishing boat will be polled for any fish caught that are not available for examination. Unavailable catch are usually fish that have been thrown back, given away, packed away, used for bait, filleted (not identified by skins), eaten, or taken by marine mammals (salmon only). Unavailable fish are reported by the entire group of anglers on the boat. The anglers are asked to separately report any unavailable fish in four categories; kept, released alive, released dead, and seal take (salmon only). If no fish were caught (kept or released), a NO CATCH code is recorded in the Species Code box and zeros recorded in the catch boxes.

Kept Unobserved Catch

Fish that are not thrown back, but otherwise are not available for examination will be separately recorded on the PR Form. Kept unobserved fish include fish given away, packed away, used for bait, filleted (not identified by skins), or eaten. Kept fish that the angler refuses to show to the Sampler are included as "kept unobserved". These fish are counted separately from fish which the Sampler personally examines and counts (kept observed). Be persistent with anglers that have unavailable rockfish catch. Use your best effort to gain access to the catch for species identification.

Released Alive

The released alive catch category is the total number of fish by species that were released alive in swimming condition. Released alive total includes fish intentionally landed and subsequently released, those that are purposely shaken off the hook boat-side, and any rockfish that are released using a descending device. The Sampler and anglers are not to judge the likelihood of survival of a swimming fish. Fish that 'got away' are not considered purposely released and are not included as released alive.

Released Alive with Descending Device (DD)

This is a subset of released alive and includes the total number of rockfish by species that were released alive using a descending device. Rockfish brought up from depth suffer from barotrauma from gas expansion as a result of decreasing pressure. Stomachs protruding from mouths, eyes popped out of their orbits, and "crystallized" comeas are all symptoms of barotraumas. Use of a descending device to send rockfish back down to depth can greatly reduce discard mortality. A descending device can be a professionally

fabricated store-bought lip-gripping contraption; it can be a line tied to the bend of a hook with a heavy lead sinker tied to the eye of the hook; or it can be an inverted, weighted milk crate with a rope tied to the bottom (now the top) — anything used to send a fish back to depth can be considered a descending device. Use of a needle to vent the swim bladder of a fish is not considered a descending device. Released alive with descending device is coded only for rockfish species. Released alive with descending device is a subset of the released-alive total; the number of released alive with descending device will always be less than or equal to the released alive total.

Released Dead

The released dead category includes fish landed or purposely shaken off the lines which are returned to the water in dead condition. Fish that are technically alive but are obviously not going to survive (due to severe wounds or inability to swim down) may be coded as dead. The Sampler and angler are to judge that the non-swimming fish is dead or will be shortly. The survival of all fish returned is determined by application of mortality rates. These rates are determined by scientific studies of hooking and depth-based mortality. However, CRFS may decide to use different capture mortality rates or compare computed mortality with observed mortality.

Seal Take

The seal take category includes any <u>salmon</u> that were known to have been taken by any (seals, sea lions or other marine mammals). <u>Seal take should only be determined for salmon catch</u>. Anglers must be certain and have seen the marine mammal take the salmon from their line. The Sampler should inquire further with those anglers who say 'I think' or 'maybe' a fish was lost to a pinniped. The Sampler should not include fish that naturally escaped or were naturally caught and eaten by a pinniped.

Catch Location and Average Depth

All CRFS boats are sampled for the catch location and average depth. For boats with catch, a fishing location will be recorded. Location and the average depth may be recorded for all catch together or by species when determined and time allows. For boats with no catch, the location and average depth where the majority of fishing effort occurred is recorded. The majority of effort is defined as where most of the boat's time was spent with gear in the water. Average depth is used to put the catch estimates into depth zones and compare with locations. It is also used to help estimate mortality rates for some groundfish species.

Q. If the PR anglers do not have any catch, should the catch location be left blank?

A. No still code a location. In this case, code to the major area fished (where effort mostly occurred).

Coding Location of Catch for Multiple Species on the Same Trip

Frequently, PR boats will fish in several locations for different species/species groups on the same trip. It is important for the Sampler to recognize when this occurs and code distinct locations of catch for each species/species group. If anglers do not give some indication that their catch for the day came from more than one location and the Sampler is either not paying attention or not familiar with local fisheries, then incorrect location information will be collected that may bias CRFS data. Taken to the extreme, when a boat's catch comes from more than one location and the Sampler does not collect location of catch data for each species/species group, it may appear that the boat was fishing in an illegal area, at an illegal depth or with illegal gear.

TARGET		1		KEPT	RE	LS	SPECIES LOC	DEPTH
1st	AREA	GEAR	SPECIES	obs	alive	(wiDD)	or effort loc if no catch	Average
2nd	Ā	95	CODE	unobs	dead	seal take	Block-box; Lat / Lon	Bottom (ft)
RFGEN	N	н		6	2	(o)	222-23	40
HALPA	0	н	RFBLK	unebs O	dead O	seal		
				2	ofive O	()		
			HALPA	0	dead O	seat		
			RFBLU	3 unobs	O desc O	(0)		
				0 4	2	()		
			LNGCD	0	O O	seal		

This is an example of incorrect coding of location of catch. As is, the Pacific Halibut catch location will be attributed to the one recorded location of catch; this is incorrect, as HALPA are rarely taken in 40 feet of water, and the water area for the HALPA target was coded as offshore – this location is clearly within three miles of shore.

TARGET				KEPT	RE	LS	SPECIES LOC	DEPTH
1st	AREA	GEAR	SPECIES	obs	alive	(wiDD)	or effort loc if no catch	Average
2nd	¥	95	CODE	unobs	dead	seal take	Block-box; Lat / Lon	Bottom (ft)
RFGEN	N	н	HALPA	2	0	()	223-25	350
HALPA	0	н	HALPA	0	teet 0	seel		
				6	alve 2	(0)		
		П	RFBLK	unebs O	test O	seal		
		П	0	obs 3	alive O	(o)		
			RFBLU	0	tast 0	seal		
				obs 4	alive 2	()		
Ī			LNGCD	unobs 0	test O	seal		

incorrect coding of location of catch. As is, the rockfish and Lingcod catch will be attributed to the one recorded location of catch. This example is typical of catch seen in District 6; current groundfish regulations include depth restrictions of 120 to 180 feet. If left as is, it will appear that the bottomfish were taken at

This is another example of

an illegal depth.

TARGET				KEPT	RE	LS	SPECIES LOC	DEPTH
1st	AREA	GEAR	SPECIES	obs	alive	(wiDD)	or effort loc if no catch	Average
2nd	¥	95	CODE	unobs	dead	seal take	Block-box; Lat / Lon	Bottom (ft)
RFGEN	N	Н	RFBLK	6	2	(0)	222-23	40
HALPA	0	н	KFBLK	unobs O	dead O	seal		
		П	101010111111111111111111111111111111111	obs 2	alive O	()	223-25	350
	Г	П	HALPA	unobs O	deed O	sed		
		П		3	olive O	(o)	222-23	40
			RFBLU	unobs 0	dead O	seal		
			LNOOD	4	alive 2	()		
			LNGCD	0	dead O	seal		

This is an example of <u>correct</u> <u>coding</u> of location of catch.

Note that in order for the Blue Rockfish and Lingcod catch to be associated with the location of the Black Rockfish catch, the location and depth needs to be repeated after the Pacific Halibut row.

Measuring Catch

For each CRFS boat with observed catch, the Sampler should sample the catch for species composition and bio data: lengths, weights, and some fish may be sexed. The priority is to document and measure the priority species and, in northern California, adipose fin-clipped salmon. Do not measure non-adipose fin-clipped salmon. A secondary priority is to weigh important management species. Please see the General Onsite Procedures section for a complete list of priority species.

Time allowing, all fish may be measured and weighed. The goal is for paired lengths and weights, if possible. Paired lengths and weights allow for a regression equation to check for sampling error. Lengths are used to predict weights using a regression and to examine length classes. Weights are used to calculate more precise metric ton estimates and are used with the length to estimate fish condition.

Some fish may be sexed using external characteristics. Please see the Species Sampling section for complete details on which species may be sexed.

Interview Priorities

Samplers should be aware that some of the data is required while subsampled data may be high priority or low priority.

Required Counts

Count boat trailers upon arrival Count all boat trailers at departure Count all boats missed

Required Boat Records

Monitor all intercepted boat return times
Determine if the boat is fishing or not
Determine the target species and gear (or non-fishing activity)

Required Catch Data

Determine if any catch (including unobserved/unavailable catch)

Count catch by species (not higher-level taxa) Examine salmon for adipose fin-clips and collect heads Determine the location and average bottom depth of each species

Sub-sampled CRFS Data (Priority Order)

- Record length measurements of priority species
- 2. Record weights of priority species
- 3. Record length and weight pairs of priority species
- Determine the location and depth of each species

2020 CRFS PR Form Questionnaire

It is important to use the wording of questions as stated in the PR questionnaire because slight changes in wording can result in different responses.

INTRODUCTION: Hello, my name is ____ and I represent CDFW. I am interviewing marine recreational anglers for the California Recreational Fisheries Survey. May I ask you a few guestions?

PRIVACY ACT STATEMENT: This study is being conducted in accordance with the Privacy Act of 1974. You are not required to answer any question that you consider to be an invasion of your privacy.

BOAT ROW, EFFORT COLUMNS:

Sample # [or R or B or RS]: In sequence, the boat number for all boats returning to the site during the sample, including non-fishing boats, but excluding missed boats, initial refusals (R) and language barriers (B). Fishing boats that do not provide the minimum data elements (# anglers, # days fished, water area, targets, gear, catch #s by species) are also coded with R and do not get a sample #. Code boats that refuse all data elements other than those required for a minimum salmon sample (# angs, # kept/obs salmon and all must be observed, #ad clips) with "RS" in the sample number field

Time: Enter the time in the 24 hr format when the vessel interview was started. Times are unique for each Sampler's data.

SCREENING: Did anyone on the boat do any sport fishing? YFS:----Go to next

NO:----Record appropriate NF (non-fishing) code in

target box, and conclude the interview

Code Sample # as R, terminate interview Refused:----NOTE: If the boat is going back out for more fishing skip till next return.

ANGS Total: **How many of you had gear in the water?** (on vessel)

Enter the total number of anglers on the vessel that fished (gear in the water)									
Refused: Co	de Sample # as R, terminate interview								
Enter the number of the ANG current California sport fishin	ort fishing license does each of you have? (above) who fished on the boat without a plicense. de Sample # as R, terminate interview								
PRIMARY TARGET: What we the boat's primary target. Anything: Not fishing: Refused: interview	UNIFH Appropriate NF code Code Sample # as R, terminate								
SECONDARY TARGET: What taxon of the boat's secondary Anything:	t were you secondarily after? Code the target. Leave blank Code Sample # as R, terminate interview								
bay? If in the ocean, ask: Wa Nearshore (< 3 miles): Offshore (> 3 miles): Bay/Estuary/Harbor: Mexico: Refused:	orimary target> fishing in the ocean or sthat mostly within 3 miles of land? N O B Be aware of freshwater cutoffs M Code Sample # as R, terminate interview the codes – see bottom of PR form								
	O B Be aware of freshwater cutoffs								
Refused: Offshore islands have separa	Code Sample # as R, terminate interview te codes – see bottom of PR form								
GEAR: What gear did you u Finfish Hook & Line:	se for <primary target="">? Shellfish</primary>								
Spear: Troll: Bait Net:	S Flat Hoop Net #: F _n - T Rigid Hoop Net #: R _n N Snare: E								
	5552								

Both M & T (salmon only): Refused:		B R	Free Diving:	D
GEAR: What gear did you Finfish Hook & Line: Spear:		<second b<="" h="" m="" n="" s="" t="" td=""><td>lary target>? Shellfish Pot #: Flat Hoop Net #: Rigid Hoop Net #: Snare: SCUBA: Free Diving:</td><td>P_n F_n R_n E C</td></second>	lary target>? Shellfish Pot #: Flat Hoop Net #: Rigid Hoop Net #: Snare: SCUBA: Free Diving:	P _n F _n R _n E C
(in 24 hr format) the boat le	eft the ran	np. Reco	ve the ramp? Record the rd number of daylight DAYS ck the N box if any fishing s R, terminate interview	Sthe
today, within the past	12 mont g in this	hs, how	er on the vessel. Not coun many days have you g from a boat launched in	one
	known, a		sel. What is the ZIP code o city or town do you live ir	
BOAT ROW, CATCH COL	UMNS:			
SPECIES CODE: Did the Yes:	Record of Record I zeros in total and record z	code in S No Catch KEPT ob I RELS dero in sea	pecies Code and go to nex in Species Code box and is, KEPT unobs, RELS alive ead. If salmon were targete	Э
KEPT OBSERVED: May I Yes:	see the o	catch?	tify and count all fish by	
species No: Unobserved	Enter ze	ro and co	ode numbers of Kept	

Fillets:-----Enter zero and code numbers of Kept Unobserved Refused:-----If there is salmon catch, code Sample # as R, terminate interview. If no salmon catch, go to next KEPT UNOBSERVED: Did the boat retain any other catch? Probe for any catch given away, filleted, used for bait or trashed. Yes:----Record species and number of fish Enter zeros in Kept Unobserved boxes for No:---species recorded Kept Observed Refused:----If both Kept Observed and Kept Unobserved are refused, code Sample # as R, terminate interview Don't Know:----Code Sample # as R, terminate interview RELEASED ALIVE TOTAL: Were any fish released alive? Probe for any fish that were purposely released alive. Record species and number of fish Yes:----No:----Enter zeros in Released Alive Total boxes for species recorded Kept Observed or Unobserved Refused:-----Code Sample # as R, terminate interview Don't Know:----Code Sample # as R, terminate interview RELEASED WITH DESCENDING DEVICE: Ask only if any species of rockfish were reported as Released Alive. Of those <# released alive> <rockfish species> released alive, were any released using a descending device? Yes:----Record number released using a descending device in (w/DD) Don't know-----DK No:----Record zero in (w/DD) Sampler didn't ask----DA Refused:----No Rockfish Catch:---Leave blank RELEASED DEAD: Were any fish released dead? Probe for any fish that were thrown back dead. Yes:----Record species and number of fish

No:----Enter zeros in Released Dead boxes for species

> recorded Kept Observed or Unobserved Code Sample # as R, terminate interview

Refused:-----Don't Know:-----Code Sample # as R, terminate interview

SEAL TAKE: Ask only if boat had salmon catch. Did you see any seals or sea lions take your fish from your line?

Yes:----Record number of fish lost to pinnipeds in the

seal take box in the same row with the salmon

catch

No:----- Enter zero in seal take box in the same row with

the salmon catch

Refused:----- R Don't know----- DK

No Salmon Catch:---- Leave blank

Sampler didn't ask---- DA

CATCH LOCATION: Where were most of the <species> caught?

NO CATCH: Where did the boat spend most of its time fishing today?
The priority order of the location is for 1) landed fish, 2) reported fish, or 3) majority of fishing time. If the anglers report locations by species and time allows, record the location for each species observed or reported.

Refused:----- R
Don'tknow----- DK
Sampler didn'task--- DA

Block-Box:---- BBB-bb-bb-bb (up to three boxes for one block)

Lat & Lon:----- Enter the latitude above the longitude.

1) Degrees, minutes and grid

(DD.MM/DD.MM+GG)

2) Degrees, minutes and seconds

(DD.MM.SS/DD.MM.SS) where D=degrees, M=minutes, S=seconds, G=area in minutes

NOTE: If the location is above a freshwater cutoff, the boat is not eligible and should be coded as NFOTH.

BOTTOM DEPTH: What was the bottom depth at that location? Record average bottom depth

PR Form Layout

Boat samples are recorded in rows with data fields arranged by columns. Each boat row has two sub-rows to record two observations for each item in some fields. A boat sample data may span multiple rows and sub-rows as needed to document additional catch species, fish counts, catch location(s) and depths, and fish bio data. Fish records for a boat may also be continued on the next page; the PR form is double-sided to reduce waste and the front and back of the form are the same.

The form is subdivided into four sections; the header row (sample day), individual boat data (effort), individual fish data (catch and bio data) and subtotal/totals (page summary).

Header Row Items

The header row records data for the sample day. The header includes a unique assignment ID number, date, site information (county, site, port), Sampler ID number and name, additional Samplers present at the site and

their ID numbers and if they have data or not, start and end times, and trailer counts. All these items are required.

CRFS PR	FORM (V8 11/08/20	14)	ПР	R1 [PR2		Page of Other Samplers: Name & # (w /data) Time	Trailer	Counts
ASSN ID	Date (MM/DD/YY)	CNTY	SITE	OSP Port	Sampler #	Sampler Last Name	Other Samplers Name & # (W/data	Start	onsite	Offsite
							(Y N	Stop		

Individual Effort Items

Individual boat data include boat sample number, time, total anglers (licensed and unlicensed), days fished, night fishing check box, 12-month avidity, zip code, target species (primary and secondary), water area and gear (for each target). Onsite and offsite missed boats (for select PR1) are tallied on the right side of the form. Launching vessels are tallied only in PR2 sampling mode; leave blank for PR1 mode.

		EFFORT				Miss	edBt	7-
Sample # Time	ANGS Total (unlic)	DAYS fished Zip Code	TARGET 1 st 2 nd	AREA	GEAR	onsite	offsite	PR2 Launched
A	()	N 12mos				uo	off	Ы

Recording Launched Boats

In PR2 mode you will also be tallying any boats that are launching. In the left-most "PR2 Launched" enter the number of boats that launched since you completed sampling the last boat. In addition, these fishing kayaks, fishing PWCs, and sailboats are to be flagged "K", "P", and "S" respectively.

Individual Fish Data: Catch and Biological Data

Individual fish data recorded include the species, number landed examined (kept obs), number landed unavailable (kept unobs), number released alive, number of barotrauma-sensitive species released with a descending device, number of fish released dead, number lost to seals (salmon only), species catch location, average bottom depth, lengths, weights, sex, and head tag numbers.

		li .		- 20			BIO DATA			
KEPT	REI	LS	SPECIES LOC	DEPTH	Fork	length / ca	arapace size (mm), sex (M	/F/T)	
obs	alive total	(widd)	or effort if no catch	воттом		Weight ((decimal kg)	or (tag#)	200	
unobs	ode unobs	dead	seal take	Block-box Lat / Lon	(ft)	1	2	3	4	5
abr	alive	()								
nobr	doad	real						<u> </u>		
	obs unobs	obs alive total unobs dead	obs alive (w/DD) unobs dead seal take	obs alive (w/DD) or effect if no catch unobs dead seal block-box Lat / Lon te elice ()	obs alive (w/DD) confliction catch bottom unobs dead seal block-box Lat / Lon (ft) **Total Late Control of the catch Control of the	obs alive total (w/DD) or effort if no catch BOTTOM unobs dead seal take Block-box Lat / Lon (ft) 1	obs alive total (wrD) or effort/if no cetch BOTTOM Weight (unobs dead seal Block-boxLat / Lon (ft) 1 2	obs alive total (w/DD) or effort if no catch BOTTOM Weight (decimal kg) unobs dead seal take Blook-box Lat / Lon (ff) 1 2 3	obs alive total (w/DD) or effect if no catch BOTTOM Weight (decimal kg) or tag # unobs dead seal take Block-box Lat / Lon (ff) 1 2 3 4 to alive ()	

Footer Totals

At the bottom of each page, sum the number of refusals and language barriers, total boats (includes fishing and non-fishing), boats targeting salmon or with kept salmon, anglers targeting salmon or with kept salmon, the number of king salmon kept and released (Chinook Salmon, SALCK), the

number of silver salmon kept and released (Coho Salmon, SALCO), the number of Pacific Halibut kept and released (HALPA), the number of Yelloweye Rockfish (RFYEY) kept and released, the number of Cowcod (RFCOW) kept and released, the number of Canary Rockfish (RFCAN) kept and released, the number of Black Rockfish (RFBLK) kept and released, and the number of onsite and offsite missed boats. The summary of effort and catch from each page is used to facilitate completion of the assignment summary form and weekly summary report; the data is also used to verify data entry. The salmon, Pacific Halibut, and overfished rockfish totals allow for in-season estimates of catch and effort to monitor catch quotas.

_																				
Refu +	Total	Boats	Angs	Kent	Rele	Kent	Rels	Head	Kent	Rels	Kent	Relo	Kent	Rels	Kent	Rels	Kent	Rels	On	Off
				reope	11010															
Rarrie	Boats	Salı	mon	Kir	ngs	Co	ho	Tags	I HA	LPA	RF'	YEY	REC	COW	RF(CAN	RF	RIK	Mis	ced
Dunio	Docto	ū	11011	100	igo	Š	110	ugo	110					,011	-	27 11 4		JEIN	14110	oou

PR Form Item by Item Instructions

Field Name (noted if exclusively PR1 or PR2)	Instructions	Coding Examples and Formats
	HEADER	
□PR1 □PR2	Check the box for the assigned survey mode.	⊠PR2
Page of	Enter, in sequence, the page number of the form and the total number of pages on all pages.	Example: Page 2 of 7
ASSN ID	Enter the six-digit assignment ID number on all pages.	Assignment ID in the MMDXNN format, where MM is the month ranging from 01-12, D is the CRFS District from 1 to 6, X is the assignment mode (PR2 assignments are numbered 301-399), and NN is the sequence from 01 to 99 Example: 072311 This is the 11 th PR2 assignment in July in CRFS District 2
Date	Enter the date of the assignment on all pages.	Use the MM/DD/YY format. Example: 07/14/20 = July 14, 2020

CNTY	Enter the 3-digit numeric county code on the first page only.	Example: 045 = Mendocino County
SITE	Enter the 3-digit numeric site code on the first page only.	Example: 100 = Noyo River Launch Ramp
OSP port (PR1)	For all PR1 assignments, enter the 3-letter alpha code on all pages. The codes are in Sampler Manual, behind the tab Other Codes.	Example: FTB = Fort Bragg
Sampler #	Enter your 3-digit Sampler identification number on all pages.	3-digit numeric code = 305
Sampler Last Name	Write out your last name completely on all pages.	
Other Samplers: Name & #	Write out last name and Sampler # for other Samplers working on this assignment.	Example: Smith 132 (Y)
	Circle Y (yes) or N (no) to indicate if the Sampler has a separate set of data to submit.	
	First PR page only.	9
Notes on Trailer Counts	TRAILER COUNTS Upon arrival enter the total number of boat trailers in the established trailer count area for that site on the first page only for onsite and offsite (if applicable) under "Start". At the end of the sample day, enter the total number of boat trailers on site upon departure under "Stop". First PR page only.	NOTE: When conducting trailer counts, it is important to include all effort for the site If the "count area" (ramp parking lot) is full and there are trailers (that are active at the site) are parked on the street or offsite, it is important to include that effort in the counts
Time [Trailer Counts, Start and Stop]	Record the time you began counting the onsite trailers (Start is	Use 24-hour military time format

	upon arrival, and Stop is at the end of the assignment). The times of Sampler arrival and departure from locations where offsite counts are conducted will also be recorded on the ASF.	Example: 9:00 AM = 0900 hours
Onsite [Trailer Counts, Start and Stop]	Onsite refers to trailer count occurring at the assigned site.	See the attached table titled "Summary of PR Counts" for specifics on what to include in the counts
Offsite [Trailer Counts, Start and Stop]	At certain PR sites count the fishing trailers at a nearby PR site. The "offsite trailer count area" is listed on the monthly site list or the Lead will provide a list.	See the attached table titled "Summary of PR Counts" for specifics on what to include in the counts
	The Start Count should take place <u>before</u> going to the assigned PR site. The Stop Count should take place after sampling is complete at the assigned PR site.	
	EFFORT	
Sample # [or R or B]	Record a sample number in consecutive order (starting with 1) for every boat intercepted (except for refusals or language barriers). See the attached table titled "Summary of PR Counts" for specifics. For refusals, record an R without a sample number. For language barrier, record a B without a number. Flag special types of boats using letter codes	Sample # = 1,2,3 REFUSALS and LANGUAGE BARRIERS are NOT issued a sample number. Record an "R" or "B" in the Sample # box. Do not list a target. Do not record as a missed boat SAMPLE FLAGS KAYAK - record a "K" after the sample number PWC, canoes, other small non-trailered boats, and "non-traditional boats" with trailers -

	(see right column) and a sample number.	record a "P" after the sample number TOURNAMENT - record a "T" after the sample number SAILBOAT - write a "S" after the sample number NON-FISHING TRIPS: Record a sample number in the box, and the non-fishing type under TARGET. See the attached table titled "Summary of PR Counts" for specifics.
Time	Enter a time stamp for every boat that is given a sample number or is a refusals or language barriers.	Use 24-hour military time format Example: 5:00 PM = 1700 hours
ANGS total	Enter the total number of anglers on the boat regardless of license status (licensed anglers+ unlicensed anglers). Code zero for NF boats.	0 = NF boat only 3 = three anglers fished total R or B: code the Sample # box with "R" or "B" and # of anglers if known, terminate the interview
ANGS (unlic)	Enter the number of anglers out of the total anglers fishing who do NOT have a current CA fishing license of any type. Note: unlicensed is a subset of total anglers, therefore unlicensed ≤ total anglers.	0 = all anglers were licensed R or B: code the Sample # box with "R" or "B" and # of anglers if known. Leave unlic. blank. Continue interview if possible, otherwise terminate the interview
DAYS fished (left column) = trip effort N = Night fishing	Enter the total number of days the boat fished on this trip. This is recorded as the number of daylight fishing days for the boat without returning to port. Some boats launched from	Example: fishing during daylight hours the evening of one day and the morning of the next day = 2 days of fishing effort

	ramps will have the capability to fish multiple days. Boats that engaged in any night fishing (nondaylight) will be identified by checking the "N" box. If only night fishing occurred, record 0 days	R or B: code the Sample # box with "R" or "B" and terminate the interview
Launch Time	fished. Enter the time that the boat launched from the PR2 site for this fishing trip.	Use 24-hour military time format Example: 6:15 AM = 0615 hours If a trip is greater than 1 day in duration, leave the launch time blank and make a note on the data sheet
DAYS fished (right column) = 12-month avidity	Select a random angler on the boat and ask, "Not counting today, how many days have you gone saltwater sport finfishing in California in the last 12 months?" Use a random method of selection to avoid bias (do not always pick the boat operator).	52 days = fishing 1 day/wk over the last 12 months Refused = R Don't know = DK Sampler didn't ask = DA Note: the largest number entered would be "364"
Zip Code	Select a random angler on the boat and request the residence zip code. Use a random method of selection to avoid bias (do not always pick the boat operator). May be the same angler that answered the 12-month avidity question.	Example: 90210 = Beverly Hills Refused = R Don't know = DK Sampler didn't ask = DA Foreign country = 3-letter country code e.g., Ireland = FIE
Target Primary = the main	Each boat not missed is screened to determine the primary and secondary activity/target,	Examples: HALCA = targeting California Halibut

target/activity for the trip Secondary = the secondary target/activity for the trip	including fishing and nonfishing activity. Activities/targets are coded using 5 letter alpha codes. Targets may be determined by asking the angler(s) "what was the number one and number two fish you were fishing for?" Anglers who don't have specific targets after probing are recorded as UNIFH. If the anglers refuse to provide a target, then code the Sample # box with "R" and terminate the interview. If the anglers cannot speak English, then code the Sample # box with "B" and terminate the interview.	Non-Fishing Codes: NFCOM=commercial fishing trip (non CPFV) NFPC6 = CPFV trip *Do NOT record CPFV trips as a PR; record the NF code then sample using a PC dockside form NFOTH = Any other boating activity, including maintenance, enforcement, research, sailing, etc. Do not record NF kayaks or personal watercraft. NF sailboats are recorded as NF boats with an "S" flag R or B: code the Sample # box with "R" or "B" and terminate the interview
AREA	Record the water area code where the majority of fishing effort (fishing time) occurred for each primary and secondary target. Note that the AREA of effort and SPECIES location can differ for the same target. Area is left blank for NF trips or blank secondary targets.	N = (ocean < 3 mi) O = (ocean > 3 mi) B = enclosed bay or estuary M = Mexico Sland Codes: F = Farallones

		R or B: code the Sample # box with "R" or "B" and terminate the interview
GEAR	Enter single letter code for the fishing gear used by the boat for each target. The gear should be determined and recorded for each primary and secondary target identified. Gear is left blank for NF trips or blank secondary targets. There are two special gears for salmon fishing.	H = Hook-and-Line S = Spear T = Troll M = Mooch (salmon only) B = Both M and T (salmon only) N = Bait Net Invert Only Pn = Pot and # Fn = Flat hoop net and # Rn = Rigid hoop net and # E = Snare C = SCUBA diving D = Free diving
	The gear should be determined and recorded for each primary and secondary target identified.	Unspecified invert. gear (shovel, rake, gun, etc.) – leave blank and make note on form.
	CATCH	
SPECIES	Enter the 5-letter alpha code for each species or taxon of all fish examined or reported by the boat. Additional rows are used for boats with multiple species catch.	No catch: write "NO CATCH" in the SPECIES box and zeros in catch boxes: - KEPT obs - KEPT unobs - RELS alive total - RELS dead If the anglers refuse to let you see the catch or provide information on the fish caught or released, code the Sample # box with "R" and terminate the interview
KEPT obs (observed)	Enter the number of fish by species examined for this boat. If no fish of a species are examined, record a zero.	Only fish that the Sampler can see, and count are recorded here; may include fillets that can be counted and identified: make a note

	Sampler will identify and count each species retained by the boat.	on the form if the daily bag limit is exceeded for a species or group of species If the boat refuses both KEPT obs and KEPT unobs, then code Sample # box with "R" and terminate the interview
KEPT unobs (unobserved)	Enter the number of fish by species reported by the boat that the Sampler was not able to see and identify or count. If no fish of a species are reported as landed but unavailable to examine, record a zero. Probe for catch that may not be remembered, such as bait species.	This includes fish used for bait, thrown away as trash, given away, and fillets that are not identifiable or countable; this also includes fish that the Sampler is able to see, but for whatever reason, is not able to count; make a note on the form if the daily bag limit is exceeded for a species or group of species If the boat refuses both KEPT obs and KEPT
		unobs, then code Sample # box with "R" and terminate the interview
RELS alive total	Enter the number of fish by species reported as released alive by the boat. This includes both fish released with a descending device and without a descending device. Fish must have been landed first or have been intentionally released. Probe for catch that may not be remembered. If no fish of a species are reported as released	Fish appeared alive with no mortal injuries upon release No fish released = zero R or B: code the Sample # box with "R" or "B" and terminate the interview
	alive record a zero.	

RELS alive (w/DD)	Enter the number of rockfish by species that were released alive using a descending device. Venting the fish is not a descending device. This field does not apply to non-rockfish species. Note: Rockfish released using a descending device are considered alive.	This field is only applicable for rockfish that are released alive No rockfish catch = leave blank Code this box for all rockfish species If RELS alive total = 0 then (w/DD) = 0 Refused = R Don't know = DK Sampler didn't ask = DA Note: RELS alive (w/DD) is a subset of RELS alive total, therefore RELS alive (w/DD) ≤ RELS alive (w/DD) ≤ RELS alive total
RELS dead	Enter the number of fish by species reported as released dead by the boat. If no fish of a species are reported as released dead, record a zero. Probe for catch that may not be remembered.	R or B: code the Sample # box with "R" or "B" and terminate the interview
Seal take	Enter the number of salmon reported taken by pinnipeds for the trip. The angler must have seen the pinniped take the fish from the line.	This question is only asked if salmon catch was targeted No salmon target = leave blank Refused = R Don't know = DK Sampler didn't ask = DA No salmon lost = 0
SPECIES LOC	Enter the location where the majority of the catch species were caught.	Block- Box: BBB-bb-bb-bb or BBB-bbb-bbb-bbb

	If no catch, record the location where the majority of fishing effort occurred. A separate location may be recorded for each species observed or reported. Refer to the manual for codes. For trips with large areas of trolling for non-bottomfish species, record a general area.	718-106-107-108 = block and 3 boxes (inland) 235-12-14-15 = block and 3 boxes (ocean) 252 = block only Block-Box-Grid Size: BBB-bb+g: 212-01+3 = block and one box plus grid size Lat/Long: Latitude in upper box and longitude in the lower box; Only use whole degrees and minutes (no seconds or decimals). Grid size can also be used 37,30+3/118,57 = lat 37 degrees, 30 minutes and long 118 degrees and 57 minutes with a grid size of 3 37,30/118,57 = lat 37 degrees, 30 minutes and long 118 degrees and 57 minutes with a grid size of 3 Refused = R
		Don't know = DK Sampler didn't ask = DA
DEPTH	Enter the bottom depth in feet for the catch location. This is not a mid-water depth of capture. Enter a single depth or if a range is given enter the mean depth. The depth should be recorded by species when possible.	100 = 100 feet 100 min/120 max = then enter as mean depth 110 feet = 110 Refused = R Don't know = DK Sampler didn't ask = DA

	BIO DATA	
Fork Length	In the top row enter the	321 = FL in mm
(mm)	fish's fork length.	
Sex	_	F = Female
	Add an M, F, or T after	M = Male
	the length for sexed	T = Transitional (CA
	species.	Sheephead)
	·	• ,
	Do not measure a	321F = female fish 321
	salmon with an intact	mm FL
	adiposefin.	
Weight	Below the length, enter	5.35 = weight in kg
(decimal kg) or	the weight for the fish or	
Head Tag #	eligible invertebrate in	
	kg.	12345 NRS = adipose
		fin-clipped salmon head
	Do <u>not</u> weigh headed or	notrecovered
	gutted fish.	
	3	
	For salmon and	
	Yelloweye Rockfish,	
	enter the headtag	
	number below the length	
	and circle the headtag	
	number.	
	For salmon heads not	
	recovered or lost, enter	
	the head tag number and	
	code NRS (non-	
	recoverable specimen).	
	Salmon and Groundfish	
	headtag numbers are 5	
	digits.	
	l anguar	
	Do not weigh salmon.	
	MISSED AND LAUNCHED	BOATS
Missed Bt	Enter the number of	This includes un-sampled
onsite	boats that returned to the	or missed boats
	sample site that were not	See the table titled
	sampled since the last	"Summary of PR Counts"
	sampled boat.	for specifics
	· ·	Refusals are NOT
	Tally marks can be	missed boats
	recorded in the box, then	Language barriers are
	the total is recorded	NOT missed boats
	when the next sampled	
	boat comes in.	SAMPLE FLAGS
	שטמו לטוווכט ווו.	SAMELE FLAGS

Minned Dt		K = KAYAKs P = PWC, paddle boards, canoes, other small non- trailered boats, and "non- traditional boats" with trailers S = SAILBOAT Example: 2K = 2 fishing kayaks It is OK to put multiple numbers and flag in a row (line). For example, you could list: 2 = 2 recreational fishing boats missed AND 1K = one fishing kayak missed. Page Tot = 3
Missed Bt offsite	Enter the number of boats that returned to an offsite boat area since the last sampled boat. See the attached table titled "Summary of PR Counts" for specifics on what to include in the counts.	PR1 only, leave blank for PR2
PR2 Launched (PR2)	This data is only needed for PR2 samples. Enter the number of boats that launched since you completed sampling the last boat. Include all boats regardless of activity (except do not record NF kayaks or NF PWC).	See the attached table titled "Summary of PR Counts" for specifics on what to include in the counts SAMPLE FLAGS K = KAYAKS P = PWC, canoes, other small non-traditional boats" with trailers S = SAILBOAT It is OK to put multiple numbers and flag in a row (line). For example, you could list: 2 = 2 recreational fishing boats launched

	ı	=						
		AND 1K = one fishing kayak						
		launched						
	FOOTER	launcheu						
The feeter center	FOOTER The footer contains the sum of the page totals for each category below.							
Refu + Barrier	Enter the sum of refusals	Count the number of R						
	and language barriers for	and B entries in the						
	the page.	sample# column Do notinclude RS						
		samples						
Total Boats	Enter the sum of	Total Boats = sampled						
Total Boats	intercepted boats on the	finfish boats +						
	page.	invertebrate only boats+						
	p ago.	non-fishing boats						
		Does NOT include						
		missed boats or						
		refusals/language						
		barriers, but does include						
		RS samples						
Salmon	Enter the sum of number	A boat/angler(s) that kept						
Boats/Angs	of boats that targeted	salmon caught						
	and/or caught salmon on	incidentally while						
	the page/sum of anglers for these boats.	targeting other species would be tallied as a						
	for these boats.	salmon boat/angler(s)						
Kings kept/rels	Enter the sum of	saimon boarangier(s)						
Tungo Kop troid	observed and reported							
	kept and released alive							
	and dead king (Chinook)							
	salmon on the page.							
Coho kept/rels	Enter the sum of							
	observed and reported							
	kept and released alive							
	and dead silver (Coho)							
Pacific halibut	salmon on the page. Enter the sum of							
kept/rels	observed and reported released alive and dead							
	Pacific Halibut on the							
	page.							
Yelloweye	Enter the sum of							
kept/rels	observed and reported							
	kept and released alive							
	and dead Yelloweye							
	Rockfish on the page.							
Cowcod	Enter the sum of							
kept/rels	observed and reported							

	kept and released alive and dead Cowcod on the page.	
Canary kept/rels	Enter the sum of observed and reported kept and released alive and dead Canary Rockfish on the page.	
Black kept/rels	Enter the sum of observed and reported kept and released alive and dead Black Rockfish on the page.	
Missed boats on/off	Enter the sum of missed onsite boats for the page. The missed boats offsite is for the PR1 mode only.	Do NOT sum by sample flag type (i.e., K, P, S). Sum all missed boats together

Specific editing checks

- Check that offsite start and stop counts are appropriately present or not present depending on the PR2 site sampled.
- Check that onsite missed boats are coded on each <u>boat</u> row. NOT on rows with just catch and bio data.
- 3. Check that all pages are present and numbered sequentially.
- Check that there are no missing gears and that catch location coordinates are coded in the correct format.
- Check that all fish of a species are listed consecutively (if more than 5 measurements) and, if not, that there is clear indication of where the rest of the measurements are so that the data can be entered consecutively.
- Make sure fish sex is in correct position (after length). Do not circle fish sex code.

Summary of PR Counts

Summary of PF		0::::	04-14-	T.,, '1
Type of Boat, Angler or Trailer	Interviews and Sample Number	Onsite Missed	Offsite Missed	Trailer Counts: Onsite, Offsite and Pressure Checks
General Rule non-fishing boats (NFPC6, NFCOM, NFOTH) See below for info on kayaks, PWC and sailboats	Interview & record sample #.	NEVER ADJUST THESE COUNTS Do NOT include boats that can be identified as NFPC6 or NFCOM. Include all other traditional trailerable boats	Not Applicable Not Applicable	NEVER ADJUST THESE COUNTS Do NOT include trailers that can be identified as NFPC6 or NFCOM or NFOTH. Include all other traditional trailers
kayaks, PWCs, canoes, other small non-trailered boats, and "non- traditional boats" with trailers (e.g. jet skis, dinghies) Use the flag K for kayaks. Use the flag P for PWCs, canoes, other small non- trailered boats, and "non-	Interview fishing boats & record sample # with K or P flag Do NOT interview non-fishing boats & do NOT give them a sample #.	Only count fishing boats & flag with K or P.	Not Applicable	Do NOT include in count. This means, do NOT include: jet ski trailers trailers trailers that can be identified as for sailing dinghies vehicle with racks for boats vehicles without trailers Do NOT adjust counts for kayaks etc. interviewed.

Type of Boat, Angler or Trailer	Interviews and Sample Number	Onsite Missed	Offsite Missed	Trailer Counts: Onsite, Offsite and Pressure Checks
traditional boats".				
sailboats	Do NOT interview non-fishing dinghies. Treat like kayaks or PWC. For larger sailboats, interview & record sample # with S flag for both fishing and non-fishing (NFOTH).	with S.	Not Applicable	Do NOT include trailers that can be identified as sailboat trailers in count. If you can't determine or don't know that a trailer belongs to a sailboat, then include it in the count.
trailers with no vehicle attached or "abandoned" trailers	Not Applicable	Not Applicable	Not Applicable	Do NOT include in count
refusals (R) & language barriers (B)	Interview & record R or B (no sample #).	Not Applicable	Not Applicable	Not Applicable
Vehicles with no trailers				Do NOT include in count. Do NOT adjust counts for boats that have been interviewed that are not trailered.

PR2 Form Example

Comparison Com		<u></u>
NIT SITE OSP PORT Sampler # Sampler Last Name Other Sampler # Time One		On Off Missed
Moory Carry Street Sampler Last Name Carry Carry Sampler Last Name Carry Car	0	0
Carry Carr		0 0 spt Rels
ANDONY ONLY SITE OSP PORT Sampler # Sampler Last Name		2
ANDONY ONLY SITE OSP PORT Sampler # Sampler Last Name		0 0 ept Rels RFCAN
ANDONY ONLY SITE OSP PORT Sampler # Sampler Last Name	(se)	Nept Rept
AND CAN SITE OSP PORT Sampler # Sampler Last Name	Bay Cruise)	0 0 cept Rels RFCOW
AND CAN SITE OSP PORT Sampler # Sampler Last Name	Вау	Kept Kept
AND CAN SITE OSP PORT Sampler # Sampler Last Name		ppt Rels RFYEY
AND CAN AND CAN AND CAN AND CAN AND CAN AND CAN		Kept Kept
AND CAN AND CAN AND CAN AND CAN AND CAN AND CAN		Kept Rels HALPA
ANDONY CRIT SITE OSP Port Sampler # Sampler Least Sampler Least		C Kept
ANDONY CRIT SITE OSP Port Sampler # Sampler Least Sampler Least		Head Tags
ANDONY ONLY SITE OSP Post Sampler #		ot Rels
ANDONY ONLY SITE OSP Pout Sampler #		Kept Coh
ANCHORN CAITY SITE CSP POAT Sampler # CAITCH		ot Reis
ANCORA Samples Sampl		Kept Ning
TARGET STE OSP PORT SAME STE OSP PORT SA	~ <u>'</u>	0 0 Boats Angs Salmon
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2/17 023 107 TARGET & & & CODE 1st & & & & & CODE 2nd & & & & & CODE TARGET & & & & & CODE 1st & & & & & & CODE TARGET & & CODE TARGET & & CODE TARGET & & CODE TARGET &		3 + Total
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2/17 023 такет та		40
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	Ŧ	ge Ba numan hore (3 irbor,
	NFOTH	angua at, To Offst ary/ha
(A) C C C C C C C C C	N one of	d or Li
O5/22		befuse betc, to hore (*
HE LE	₽	1002
	0 _	(a) (b) (c) (c) (c) (c) (c) (c) (c) (c) (c) (c
0563 0563 SMIPLE # Time 1119 ZK 1208	20	3 mmple: ags: R ater Ar

PR2 Form Example

Party/Charter Boat Onboard (PCO) Mode Sampling

CPFV Survey Procedures

The PC boat survey samples Commercial Passenger Fishing Vessels (CPFVs) by sampling onboard or sampling dockside to get various data elements related to catch. CPFV log data and Pressure Effort Checks (PECs) are used for effort estimates.

The primary goal for PC sampling is to obtain the catch per unit effort (CPUE). CPUE is determined from the total kept and released fish for each angler per trip type. Other data relevant to angler catch and effort, such as location, depth, and fish measurements will be recorded. Catch estimates are produced for each CRFS District each month. Estimated mean catch per angler will be calculated and multiplied by total effort derived from the CPFV logs that operators are required to submit for each fishing trip and adjusted for compliance using PEC data to estimate total catch. Since CPUE is not calculated for fishing in Mexican waters, no PC trips fishing in Mexico are sampled.

A secondary goal for onboard PC sampling is to collect discard fish lengths and record detailed locations and depths. Fish that will be released to the water are measured. Discard data is important for the weight calculation of catch thrown back alive and dead.

PC Assignments

In general, PC assignments are single site samples. They may be assigned as onboard or dockside samples. Your Lead will assign the location of the sample. Rescheduling these assignments is not desirable to the survey, and you will contact your Lead if the assignment needs to be rescheduled.

PC Definitions

Party and charter boats that take passengers fishing in saltwater must be permitted by the state as a CPFV, operated under a Coast Guard license and be inspected.

- Party boat A CPFV on which fishing space and privilege are provided for a fee. The vessel is operated by a licensed skipper (guide) and crew. Anglers on a party boat are usually not associated with one another but may be in small groups.
- Chartered boat A CPFV that is hired by a single group of anglers for exclusive use. The purpose of chartering a vessel is to gain privacy, increased deck space, and/or control in the operation of the vessel's fishing activity and fishing locations. Party boats operate under charter for a specified price, time, etc. Charters are usually closed parties, as opposed to the open status of party boats all-day and half-day party boats. The terms "charter boat" and "party boat" can be used interchangeably in different parts of the Pacific coast.

Six-pack – Informal term for small party or chartered boats that carry up
to six paying passengers, either due to Coast Guard license
requirements or boat capacity. Due to limited passenger capacity, these
trips will frequently need to be sampled dockside. Some six-pack vessels
launch from public launch ramps and may be encountered during PR
mode assignments.

When to Ride or Sample Dockside

Priority should be given to bottom fishing trips, especially rockfish and Lingcod. The entire trip should be sampled, regardless if the boat changes targets after leaving the dock. Overtime issues may prevent Samplers from riding long-range trips, so most long-range trips are sampled dockside. In Districts 3-6 dockside sampling is most often exercised on lower-priority species to ensure CPUE is obtained from all trip types. Six-pack vessels are to be sampled dockside, unless special circumstances allow for Samplers to sample onboard. In some instances, your Lead will schedule dockside sampling assignments. Salmon trips will be sampled dockside exclusively, however salmon-rockfish combos may be sampled onboard. Samplers may encounter Ocean Salmon Project (OSP) Samplers on the dock. If this occurs, be sure to work with the OSP sampler in order to intercept as many boats as possible. Note that all finfish catch, including salmon, for all anglers sampled, goes on the PCO forms.

Your Lead may assign the type of boat trip to sample, such as $\frac{1}{2}$ day, $\frac{3}{4}$ day, full day, twilight, or overnight. Your Lead may also assign the target species, such as bottomfish, Lingcod, salmon, or bass. Long range boats arrive at odd hours so you will need to check with the landing for the boat's arrival time to sample dockside.

Opportunistic PC Sampling

You may encounter CPFVs while sampling in another mode (e.g. PR1). These trips can be sampled opportunistically using the appropriate dockside form (PCS for salmon or PCD for non-salmon). However, the assigned mode takes priority unless directed otherwise by the Lead — do not miss anglers or boats in your assigned mode to opportunistically sample CPFVs. See Opportunistic PC Sampling in the PR1 and PR2 sections of this manual.

Introduction to Onboard Sampling

This onboard data collection program has been conducted since 1999.

Since many CPFVs fillet their catch at sea, Samplers must ride onboard in order to collect important data on retained catch such as species composition, discard measurements, and species targets. In addition, most CPFVs maintain an array of electronics, which allows Samplers to collect detailed information regarding location and depth. Moreover, Samplers carry a handheld GPS receiver (with the captain's permission). This location and depth data are used to assess depth-based mortality rates of encountered species.

Additional Data Collected Onboard

- Species targeted, area fished, and duration of each fishing stop
- Species kept and released for 'observed' anglers at each stop
- Measurement of returned fish by fishing stop
- Whether or not a descending device (DD) was used for each species

Unbiased Angler Sampling

Many potential biases are avoided by going onboard while some new potential biases are created. The behavior of the anglers and crew may be altered by the presence of the Sampler. For example, the Sampler may be perceived as an enforcement officer when dressed in a uniform. One study has shown that the returned catch rate of rockfish can decrease for observed trips. Due to these potential biases, the Sampler should avoid actions that alter fishing behavior at sea. Some of these actions include drawing attention to over limits, illegal fish, and illegal fishing practices.

Some difficulties arise as the number of anglers on the boat increases beyond a reasonable number which can be observed. Therefore, sampling a subset of anglers is allowed. Generally, a subset greater than 10 is not advised. When observing fewer than the total anglers on the boat, the Sampler should vary the group of anglers by position on the boat and by composition of individual anglers. This is required so that the sample you take is random with respect to the position on the boat (e.g., stern, bow or side) and the skill of the anglers. This is important on trips utilizing live bait where the live bait is also chummed in the stern of the boat. High catch rate anglers tend to congregate near the bait box. Avoid continuous sampling of the stem area by sampling in proportion the 'numbers of anglers' not the amount of catch. Contact your Lead if there is any question or concerns about how to sample or observe fewer than the total number of anglers on the boat.

Onboard Observer Protections

Under California law, CPFVs are required to carry Samplers. However, it is important to work with the vessel and landing operators as this will develop a positive relationship. Positive relationships reduce sampling bias and prevent unnecessary hostility. Samplers are there to observe normal fishing activities, and not to enforce rules or alter angler behavior. Below is a copy of Title 14 which each Sampler should have when sampling CPFVs.

Title 14, California Code of Regulations Excerpts COMMERCIAL PASSENGER FISHING VESSEL LICENSES

§105.5. Cooperation with State and Federal Fishery Observers. (a) Owners or operators of commercial fishing vessels permitted under regulations of the Commission, and commercial passenger fishing vessels licensed pursuant to Fish and Game Code Section 7920, will, as a condition of permit or license issuance, cooperate with Department or Federal fishery observers, or observers collecting data for the Department, when asked to carry and accommodate an observer on fishing trips at no charge to the sponsoring agency.

- (b) If observer coverage of a trip is denied by the owner or operator of a vessel, the Department may require an explanation in writing from the owner or operator. This explanation shall be received by the Department within 15 days of written request by the Department for an explanation.
- (c) The Department may request revocation of fishing permits or licenses to the Commission for denials that it deems to be uncooperative in nature, after first allowing the owner or operator to meet with the Manager of Marine Region, or his representative, to provide an explanation for the denial.
- (d) The Department or Federal agency requesting cooperation under subsection (a) shall not require the vessel operator or owner to provide an observer with meals or a subsistence allowance on observed fishing trips, but shall accommodate the observer with regard to reasonable eating and working conditions and access to pertinent fishing information and fishery data while aboard the vessel.
- (e) Failure to provide reasonable eating and working conditions or access to pertinent fishing information or fishery data to observers, or actions taken by a vessel owner or operator against an observer that is prohibited pursuant to subsection (f), on observed fishing trips may lead to revocation of the vessel's fishing permits or licenses issued under regulations of the Commission following the procedure outlined in subsections (b) and (c) above.
- (f) To ensure that observer objectives may be reasonably and safely achieved, consistent with federal groundfish observer rules, it is unlawful for any person to do any of the following:
- (1) forcibly assault, resist, oppose, impede, intimidate, sexually harass, bribe, or interfere with an observer,
- (2) interfere with or bias the sampling procedure employed by an observer, including physical, mechanical, or other sorting or discarding of any catch before sampling.
- (3) tamper with, destroy or discard an observer's collected samples, equipment, or personal gear, without the express consent of the observer,
- (4) prohibit or bar by command, impediment, threat, coercion, or refusal of reasonable assistance, an observer collecting samples, making observations, or otherwise performing the observer's duties,
- (5) harass an observer by conduct that has sexual connotations, has the purpose or effect of interfering with the observer's work performance, or otherwise creates an intimidating, hostile or offensive environment,
- (6) require, pressure, coerce, or threaten an observer to perform duties normally performed by crew members

Sampling Chartered Trips

You should be able to sample chartered boat trips along with open-party trips. Chartered trips can make up a large proportion of the total CPFV fishing trips, especially during the summer. It is very important to sample chartered trips, as well as open-party trips in order to accurately estimate CPFV catch and effort.

Our policy is to sample chartered trips with consent from the charter master (the charter master is the private party individual who has paid for a private group to charter the vessel for fishing), however we do have the authority to sample chartered trips that are not filled to Coast Guard rated maximum capacity.

When you call the landing to make arrangements, you should introduce yourself as CRFS Sampler and ask about <u>all the scheduled trips</u> going out for the assigned trip type including chartered trips. Be sure to confirm any chartered trips and get the name of the contact. If there is no party boat trip going out for the scheduled trip type, but there is a charter for that trip type, you should request to sample that trip with consent from the charter master.

You should ask the landing for the charter master's contact information, or if the charter master can contact you. If you are unable to confirm with the charter master, you should show up an hour before the trip is scheduled to leave so that you can have the opportunity to explain onboard sampling to the charter master, and request permission to sample onboard. Furthermore, you should occasionally attempt to sample chartered trips (even though there is a party boat trip available) when there is the opportunity to get on a boat that is rarely sampled.

Always keep an eye out for information on CPFVs when in the field. It is important for you to introduce yourself to the crew and captain especially on vessels that are not normally sampled. For vessels that are not normally sampled, you should ask about the trips that they are running and the best way to get in contact with them. Some CPFVs may not book trips through the landing office or may be overlooked by office personnel because they are not running the typical "party boat" trips that are sampled.

CPFV Refusals

Under section 105.5 (Title 14 CCR) Samplers have authority to access all CPFV boats. However, you may need to explain the survey and provide evidence that you are a CRFS Sampler. Always be prepared with copies of Title 14, section 105.5, your CDFW ID, a CRFS handout, and your Lead's business card so that you are prepared to demonstrate the legitimacy of the sampling program, and explain the survey. You should be familiar with the relevant sections in the CDFW regulation booklet on CRFS cooperation and have a copy to show to charter masters and landing personnel.

It is very important to document all attempts (successful or unsuccessful), to sample chartered trips on the <u>Assignment Summary Form</u>. Make sure that you indicate that the trip was either a charter or a party boat in the comment section of the form. If the attempt to sample was not successful, explain in the comment section why.

If you are outright refused by landing personnel or encounter any hostility or difficulties, leave a copy of Title 14, section 105.5 with the landing manager and contact call your Lead. Please provide your Lead with detailed documentation (date, name of individuals and vessels concerned, details of refusal or problem and how you dealt with it). Provide this information the same day of the event. Your Lead will initiate procedures to follow-up with the vessel.

Chartered Trip Refusals

If the charter master of a chartered boat declines, it will be considered an acceptable 'unable to sample' event. If this occurs, try to sample an alternate trip or contact your Lead to reschedule. However, if the landing or captain of the chartered vessel refuses you or does not allow access to the charter master who should be asked directly by the Sampler for their decision, the act will be deemed an "illegal refusal". For illegal refusals, you are to contact your Lead as soon as possible. Document everything that occurred (individuals involved, time, etc.), and be as descriptive and precise as possible.

Alternate PC Trips

Occasionally, your scheduled PC trip won't go out due to low effort, boat maintenance, Coast Guard capacity, etc. This is one reason why it is important to call ahead and show up at least a half an hour before the trip's scheduled departure. It is important to follow this hierarchy of steps when choosing an alternate trip and/or landing for your assignment:

- 1. Sample your assigned trip type at your assigned landing.
- 2. Sample a different trip type at your assigned landing.
- 3. Sample your assigned trip type at an adjacent landing. Landings are considered "adjacent" when they are close enough to be targeting the same fishing grounds.
- 4. Sample a different trip type at an adjacent landing.
- 5. Reschedule your assignment to another day within that week (weekday assignment) or weekend (weekend assignment). This move requires approval from the Lead.

Onboard Fishing Locations

Each "stop" the boat makes where anglers are allowed to drop their lines into the water is a separate fishing location. At each stop, the Sampler will select a subset of anglers onboard to monitor for kept and released fish. If the number of observed anglers changes within a stop, a new stop should be created, with the same location coordinates, and the new number of

observed anglers reasons for this include an angler taking a break for lunch or stopped fishing due to sea sickness.

When the boat is not anchored and the anglers drop their lines, the location is termed a "drift" if the engine(s) (running or not) are not engaged into gear to provide power. As the boat drifts along anglers continue to fish the "drift" and cover an area over the bottom dependent on currents and wind. Once the anglers are told by the captain or crew to pull up their lines the "drift" ends when all anglers have their gear out of the water.

Sometimes, the boat will reposition or "station" over a productive fishing location. In this case, the anglers may or may not pull up their gear and the boat may be under power (gears engaged) in order to maintain or slowly move into a favorable location. In this situation, the sampler would record this as a single stop, even if the anglers needed to pull in their lines temporarily while the boat moved (usually relatively slowly) back into position. Often this "re-location" event is announced to the passengers in advance.

Since a fishing location may be a drift or troll with starting and ending points, two locations need to be recorded, one for when the anglers put their "lines down" and a second for when they pull their "lines up". Each starting and ending location will have a set of geographic coordinates (deg,xx.01 min) and a time (in 24-hour format) in order to map the extent of travel over the bottom and calculate direction and average speed. If the drift was only a very short distance and time (less than 3 minutes or 300 feet) then the ending location geographic coordinates does not need to be recorded. However, the ending time should always be recorded.

Often the captain will be "prospecting" for fish when he asks the anglers to drop their lines into the water because there is some evidence of fish on the electronics. This may result in very short unproductive stops. Record these locations. There is biological interest in locations where fish are unavailable or not catchable.

Onboard Catch by Location

For each fishing stop or location, you will keep a count of the fish caught by species and the disposition (kept, released alive, or released dead) of each fish for the observed subset of anglers.

When the catch rates are very high, you may find it necessary to monitor fewer anglers for the catch count. It is acceptable to monitor different numbers of anglers at each location; however, the preference is to monitor the same number of anglers throughout the trip.

PC Sampling Scheduling

PC assignments are selected to sample at least 2% of non-salmon fishing trips. Sampling will occur on weekends and weekdays throughout the month based on historical finfish fishing trips. If effort is low or absent at the assigned site, then follow the Alternate PC Trips protocol previously described in this section.

Scheduling PC Trips

Your Lead will provide you with a list of charter boats and landing sites with contact information. You will call ahead of time to determine the availability of PC boats for sampling onboard or dockside. You may use alternate sites if sampling cannot be conducted at the assigned site. In this case, you must still list the assigned site as the first site visited on the Assignment Summary Form. For PC dockside assignments, you must use as many PC sites in addition to the primary assigned site to attempt to obtain interviews in the assigned mode for PC.

Call the assigned site landing(s) prior to your assignment and ask if any charters or open party boats are going out on your assigned date. When you contact the office introduce yourself as a CDFW CRFS Sampler. Going to the landing is preferred to calling the landing as you'll have a better chance of getting on a boat. You may need to contact the landing closer to the trip departure to determine the number of angler reservations and how many are required to send the trip out. Keep in mind that many landings receive 'walk-up' anglers right before departure that don't make reservations. Since you may be contacting a number of different people at different times, you may want to keep a contact log with numbers, names, dates, times, and messages you may have left so that you don't duplicate or omit contact efforts include this information on the ASF if the trip is not successful.

It is important to remember that different boats from the same landing may fish different methods or different locations. If you have the option, try to sample boats that are infrequently sampled, and always inquire about charted trips, as these trips are just as important as open party. Your Lead may assign certain trip types, either by trip duration or target species. If the assigned trip is unavailable to sample for some reason (i.e. not enough passengers, vessel dry docked), then refer to the Alternate PC Trips protocol previously described in this section.

Contact your Lead for instructions when assignments and boat scheduling is unsuccessful, and assignments are not getting completed in timely manner. If a trip is going to go out and you suspect you will not obtain at least one interview per hour (i.e. 4 anglers on a 6-hour trip), contact your Lead before sampling. Your Lead may reassign an assignment to a specific site, boat or trip type in an attempt to represent the fisheries in your area with a limited number of assignments.

Arrival at a PC Site

Show up at least a 1/2 hour before the boat is scheduled to leave. Sometimes party boats are full to Coast Guard capacity and you will be denied boarding. If this is the case, you will try another boat. If the landing says that the boat is "chartered", ask if you can get permission from the charter trip leader (charter master) to ride the chartered trip. Be sure and get the captain's permission to board the boat and never board the boat without his permission. Some boats will require you to sign in on a sheet, as such it is a

good practice to confirm with the deck hand or captain if it is needed. Good rapport with the captain will often result in increasing the cooperation of the party boat patrons.

The operator must allow you free boarding privileges, if not, inform your Lead immediately and attempt to board another boat. Refusals are illegal. Document these actions. Since you are an unpaid passenger and most boats have a legal capacity you may be unable to board at the time of the trip if the boat is full of paying passengers. It may not be legal for them to take another passenger due to Coast Guard regulations. If you are told that the boat is at Coast Guard capacity, politely ask for the number of passengers and crew on the boat, so this claim can be verified by the Lead. Many vessels have a fishing capacity which is lower than the Coast Guard capacity, confirm with the Captain about the capacity limit. Just because a vessel is at fishing capacity does not mean they are at Coast Guard capacity and they may be able to take another non-fishing passenger.

Onboard the Boat Trip

The onboard Sampler has different procedures to follow before, during, and after fishing. These procedures are designed to optimize your time and reduce potential bias. Samplers will use available time to collect avidity and demographic data from anglers in advance, since that is difficult once the fishing starts.

On the Way Out

Once the boat gets underway, the captain will give a speech about life jackets, fishing procedures etc. After this speech is a good time to introduce yourself what you are doing and start collecting angler data. It is helpful to start on one side and work around the boat, this will allow you to be able to figure who is who at the end of the day. It is better to ask most questions in a pre-fishing interview as the anglers are in a good mood as opposed to asking on the way back when they may be sick or tired. Make sure to record good angler reference notes, as you will be revisiting these anglers after fishing has concluded. You can make a note of the angler's appearance, such as blue Nike shoes or tattoos. Recording easily removable articles of clothing such as jackets and hats are not recommended since the angler may remove them as the temperature changes. Try and choose features that cannot be changed such as facial hair, piercings etc.

Boats that assign numbers to anglers and keep their fish in numbered gunny sacks provide an ideal way to sample because the catch and angler are tied together by this number, and you can keep track of their catch. Make sure to record these on the CPFV Angler form, and remember that there could be duplicate numbers with different colors and multiple anglers for a bag.

Under optimum circumstances, all anglers on the boat will be interviewed. However, some form of angler sub-sampling may be necessary if the boat holds many anglers, there is a large number of fish or if the time required for

travel back to the dock is minimal. Generally, attempt to sub-sample at least 30 anglers aboard the boat.

During Fishing

The CPFV onboard location form is used to monitor the start and stop, time, and depths for each fishing location. You will also be monitoring a subset of the anglers (observed anglers) for kept and released numbers of fish by species for each fishing location. You will also be taking measurements of returned fish on the CPFV Onboard Catch and Discard form when time allows. Details of this procedure and items to collect are in the detail section for those forms below.

CPFV crew members who fish with the intention of keeping their catch, or who are putting their catch in a separate "crew bag" are considered eligible anglers and can be interviewed. Conversely, crew members who are fishing to add catch to the bags of paid passengers are not considered eligible (note: this practice is illegal under Title 14, CCR Section 195(e)(2)). The fish that the crew catches and gives to paid passengers belong in the receiving angler's data as KEPT catch (as if the angler caught the fish). It can be too difficult to track fish that are distributed among anglers by the crew, so always follow this procedure.

If you witness illegal fishing practices, do nothing. Let the captain and the crew police the boat if they choose to. Your job is to sample, not to police illegal activity. Do not alter angler fishing behavior in any way. Do not act as a deck hand by helping passengers land fish or provide advice to increase the catch rate. Our workers' compensation insurance does not cover you if you are injured while doing any deckhand duties. Stay out of the way as much as possible. Use your spare time to edit data from the trip, key out any unusual fish, etc. Don't make comments about other party boats and their success at catching fish: keep a low profile.

Remember, we want to foster a good working relationship between CRFS and the CPFV industry; having the cooperation of the crews and landings is important. Don't do anything to jeopardize the relationship. Some of these boats have secret fishing spots or secret methods of catching fish. Don't reveal any boat secrets to others. It is best not to discuss your party boat trips with anyone. If anyone asks you questions about where you fished, what kinds of fish were caught, or how the fishing was, politely explain that the data is confidential and refer the person to the captain. Any cooperation problems with deck hands should be referred to the captain.

On the Way Back

Allow plenty of time on PC trips to identify fish before the filleting process begins. This means you may have to stop observing a bit earlier. Try to judge when the anglers will stop fishing (you can ask the captain). You'll also need to determine how long the boat ride back to the dock will be and estimate how long it will take you to work up each bag. This will give you an idea of

when you need to stop your observations of catch and start collecting biological data from the catch. Ask the filleter where he would prefer you to measure and which bags he will do first; also ask if any bags or anglers are not having fish filleted as these can be left for last. Filleters may have preferential treatment of some anglers or bags. Count and measure fish in the bag that is associated with each angler # or bag # of your interviewed anglers. While the filleter is cutting, count and measure the next bag. Attempt to keep ahead of the filleter, and do not interfere with the filleting process. You may have to skip the measurements for some fish. For safety reasons, stand clear of the filleter and fillet knives. Coordinating and communicating with the crew will allow you to collect the data we require and minimize your impact on the boat's operations.

Ask each interviewed angler about any unobserved catch. This includes any fish kept for bait, released fish and disposition (released alive or released dead). You may have to remind anglers about fish you saw thrown back or used for bait. For rockfish, try to probe to identify the released catch to the species level and avoid grouping at a higher level (e.g., RFGEN"). You can use your field guides (time permitting) or reference catch that they kept and are in front of them. If you encounter a bag of fillets the angler won't open or can't enumerate to species level, it is best to skip this interview and move to the next angler bag. The point is to get high-quality bag census to species level rather than many bags of higher-level taxa.

Due to boat limits and fish-shuffling, do not attempt to record catch given-away to another angler, as these are now in another angler's bag (and that angler may not even know it). We don't want to double count the catch.

Boat Limits

With the CDFW boat limit regulations, open party and chartered boats can continue to fish until limits have been caught for all authorized anglers and crew onboard. Even seasick anglers who do not wet a line all day may leave the boat with fish, provided they have a fishing license. The crew may be interviewed if they kept fish towards the boat limit. The crew might distribute their personal catch to other anglers (note: this practice is illegal under Title 14, CCR Section 195(e)(2)).

Occasionally, the skipper or crew will want to include you when calculating the boat limit. This is not appropriate, as you will not be fishing, and you are not allowed to accept gifted fish. If you find out that the crew is including you in their boat limit calculation, tactfully inform them that you are not allowed to leave the boat with any fish in your possession and may not be included in the boat limit calculation.

When the boat has exceeded boat limits, there will be unclaimed fish. If these fish are to be thrown overboard, the Sampler should obtain a total count (and any measurements, if possible), by species of all fish discarded after the kept 'boat fish' have been distributed amongst anglers. All extra fish that are left

over, whether kept onboard or thrown over the side (another illegal practice – Section 1.87 Title 14 CCR), should be coded on the Catch/Discard form as "Boat Fish". Of note, do not inform the crew or anglers when you see illegal practices.

Please see example at the end of the PC Onboard Catch and Discard form for how to code "Boat Fish".

NO CATCH Bags

Occasionally there will be anglers who do not catch fish and do not accept fish from other anglers as part of the boat limit. In this instance they are a 'NO CATCH' bag and must be recorded as such. Try to pay attention to this situation because these anglers (often seasick) will not line up at the fillet station. If we ignore these no catch bags, and leave them off the PCO forms, fish will be expanded to those anglers based on the interviews that are obtained from successful anglers. All eligible anglers, with or without catch, should be interviewed. Do not interview only the anglers with catch.

Overnight Trips

You may occasionally be asked to sample a trip that departs at night and fishes the next day. When filling in the ASF the date of the assignment will need to match the date of the interviews. The date of the trip is the day the trip ended (fishing concluded). Only record one row with one date for the boat trip on the ASF. If departing before midnight, record the actual departure time in the comments section and put down 0000 for the "departure time". Record the total number of hours you were on the boat-sleeping hours are included and are compensable. Sleep should be limited to nighttime and when no anglers are actively fishing.

Two PC Assignments in One Day

Occasionally a Sampler may be given two PC assignments on the same day. The Lead should specify which assignment to work first. The Sampler must attempt that assignment first before the second assignment is completed.

Special Assignment Summary Form and PC Effort Check Instructions

- The Assignment Summary Form (ASF) will be coded for each SITE scheduled and visited.
- 2. The PC Effort Check (PEC) form will be completed for each BOAT sampling attempt that provides information about CPFV activities (including no activity).
- A PEC can only be recorded if information is obtained about the site effort/vessel activity. Information is obtained from phone calls, onsite visits, and published information. Nothing will be recorded on the PEC if no contact is made or no information is collected (e.g., unreturned messages and unanswered phone calls).
- 4. If you later determine that the site or CPFV did not have PC effort and you had been provided contrary information, modify the ASF and PEC forms to show the change in effort at the site for that date.

- 5. If instructed to sample a specific boat or trip-type, only one ASF is needed to record the assignment when complete or canceled (ASF assignment disposition = 1 or 6).
- 6. Record attempted/unsuccessful sampling when the original assignment could not be completed as scheduled (i.e., the boat is full, trip is canceled, etc.; ASF assignment disposition = 2).

No Anglers in PC Mode

If you go to your assigned PC site as scheduled and no anglers are observed, refer to the Alternate PC Trips protocol described earlier in this section, unless your Lead has given you specific landings to sample as an alternative. If no effort in the assigned mode is found at the primary site and alternate sites, contact your Lead to determine the assignment's final disposition.

Sampling Dungeness Crab Onboard

Crab biological data will be ignored on PC trips, only record CRBDG as a target, area fished, and gear type (including the number of pots pulled).

Onboard Sampling Tips

- Many booking offices have an automated phone tree or website that includes the fishing schedule for the week, this can be a helpful tool when planning a PC assignment. However, the phone tree or website is not a replacement for actually speaking to the booking office or captain.
- 2. All eligible anglers, with or without catch, should be interviewed. Do not just interview the anglers with catch.
- 3. Seasick anglers may be eligible since wet gear hours include any 'rod time' provided by others in the boat limit and catch may be shared. Include the sick angler as an eligible angler.
- Anglers who are too seasick to fish but, due to boat limits, have catch
 can still be included on the CPFV Onboard Angler and Catch/Discard
 forms.
- 5. Include fish caught by the crew and given to the customers.
- 6. Make sure you don't measure the same fish twice. Occasionally an angler may have more than their limit in the bag. If you measure all the fish and the angler decides to keep their legal limit and give away their extras, make sure the fish you have already measured don't go into someone else's bag that you have not measured, as they could potentially get measured a second time.
- 7. Fish filleted at sea count as "Kept unobserved" unless identified by the Sampler (identified from skin patches left on the fillet).
- 8. Do not interfere with the filleting process. Try not to hold up the filleters.
- 9. Do not record fish to be released as KEPT records. Discarded fish measurements are recorded on the CPFV Onboard Catch and Discard form. These discards are also recorded as RELS fish by that particular angler (the angler should report this at the end of the trip).
- If the sea conditions prevent accurate measurements, do not attempt to weigh the catch. Weigh especially unusual or important management species when sea conditions allow.
- 11. Do not take your friends along with you on the trip.

- 12. Do not sleep on day trips. Sleep is permitted on overnight trips, but only at night and while no anglers are actively fishing.
- 13. Document, in detail, if you are refused access to a boat. Similarly, document any action by the crew that impedes your duties.
- 14. Thank the captain and crew.
- 15. Be courteous, you should be the last off the boat allowing, within reason, all paying passengers off the boat first.
- Additional storage space and seats within cabins should go to paying passengers first.
- 17. Do not fish while onboard.
- 18. Do not accept free fishing trips.
- 19. Do not accept any gifts while onboard. This includes fish, food, drinks.
- 20. The wheelhouse is typically off limits to everyone but the crew. If you need to see a GPS or depth-finder, ASK the captain first before entering the wheelhouse or have him report the numbers to you.
- 21. Do not bring a banana on board they are considered bad luck. It is best to be considerate of anglers' superstitions.

PC Scheduling Questions and Answers

Q. I keep calling the booking office and there is no answer. How do I code the forms?

A. Code nothing; you have no information. If possible, go the booking office in person or to the slips before or after an assignment in the area. Additionally, you can show up on the morning of the PC assignment and try to get on the boat or reschedule the assignment. Contact your Lead in this instance.

Q. I call around and no boats are going out at the assigned or alternate sites on that date. What do I do?

A. You code the assigned sites and alternates on your Assignment Summary Form (ASF) for the assigned date with a reschedule. Contact your Lead immediately to reschedule.

Q. I leave messages, but they don't call back. Do I code a refusal?

A. Code nothing; you have no contact and no information. You can either: show up on the morning of the assigned PC assignment and try to get on the boat or reschedule the assignment. Contact your Lead in this instance.

Q. I'm told that no boats are going out, but later find out that was a lie.
A. Code a refusal for that date and boat(s) on the ASF and REPORT this to your Lead or Supervisor. Contact your Lead immediately to reschedule.

Q. I'm told earlier that no boats are going out, but later find that a boat went out because the weather was nice.

A. This is something that could possibly have been anticipated. Code the boats activity on your PEC for the trip date. Contact your Lead to reschedule.

Q. I'm told that no boats are going out. Do I code an attempt?

A. Code this on your ASF and your PEC. Contact your Lead to reschedule.

Q. I'm told by the office that no boats went out, but later find that one went out and the captain would have let me ride. Do I code a refusal?

A. Yes, code the refusal. Remember to always note "who" did the refusing on the ASF. Also include your comments in the follow-up email to your Lead. Contact your Lead to reschedule.

Q. The office refused to talk to me. What do I do?

A. Code a refusal on your ASF and contact an alternate boat or site. Contact your Lead with the refusal details and reschedule the assignment.

Q. The office scheduled me on a boat, but the captain refused me. I ride one of the other boats at the site. Do I code a refusal?

A. Yes, code the boats for that site and date on your PEC. Indicate who refused on which boat and detail the event to your Lead and reschedule the assignment.

Q. I call and schedule to ride a boat three days before the trip. The trip is completed on the assigned date. Do I code the date of the phone call?

A. No just code the assignment as complete on the assignment date.

Q. No boats are going out on my assigned PC day; I schedule the boat for a later date. Do I code a reassignment?

A. Yes, if the alternate PC trip protocol was followed and the reschedule has been approved by your Lead.

THE CPFV ONBOARD ANGLER FORM

The CPFV Onboard Angler form is used to collect the CPFV trip details, as well as connecting an angler's catch with the angler, and the angler's avidity and zip code. This is the form that will be utilized at the beginning of the trip, before any fishing occurs. The CPFV Onboard Angler form has a front and back side which can capture data for 38 anglers, so only one form is needed per assignment.

CPFV Onboard Angler Form Layout

The form has two major areas for data from the PC assignment: Boat trip data, and angler information.

Boat Trip Data

There are 25 boat trip data items, which are used to link the data to the other PC forms and to provide some unique information about the CPFV trip. All these items are required.

ASSN ID	Date (MM/DD/YY)	CNTY	SITE	OSP F	ort Sampler #	# Sampler Last Name						
			Duration	PC Mode	Departure &	Return	DAYS	TOTAL	TARGET	EA	AR	DD3
DFG Boat #	Boat Name		Type	(P or C)	Date (MM/DD/YY)	Time	fished	ANGS	TARGET	AREA	GE	DD?
					Depart				1st			Y/N
Captain:					Return				2nd			

Catch Recorded obs reported	ANGLER #	BAG#	Angler REFERENCE (angler name, description, etc.)	DAYS fished (12 mo) Zip Code
unobs & RELS	_			12 mos Zip

Angler Data

There are seven items for each angler. All the data except for the Catch Recorded column can be collected on the way out to the

fishing grounds. The Catch Recorded fields will be used after fishing has stopped and catch is being recorded. This will help ensure that each type of catch is recorded for each angler.

CPFV Onboard Angler form Item by Item Instructions

Field Name	Instructions	Coding Examples and Formats
	HEADER	
Page _ of _	Enter, in sequence, the page number of the form and the total number of pages with data.	Example: Page 2 of 2
ASSN ID	Enter the 6-digit assignment ID number on all pages.	Assignment ID in the format MMDNNN where MM is the month ranging from 01-12, D is the geographic District from 1 to 6 and NNN is the sequence number from 001 to 999 (PCO assignments are numbered 600-699)
Date	Enter the date. Use MM/DD/YY format.	07/14/20 = July 14, 2020
CNTY	Enter the 3-digit numeric county code.	037 = Los Angeles County
SITE	Enter the 3-digit numeric site code.	103 = Ventura Sportfishing
OSP port	Enter the 3-letter alpha code used by the Ocean Salmon Project for this port.	FTB = Fort Bragg
Sampler #	Enter your 3-digit Sampler identification number.	3-digit numeric code = 132

Field Name	Instructions	Coding Examples and Formats
Sampler Last Name	Write out your last name completely.	"Smith"
CDFW Boat #	This is the CDFW vessel ID number of the boat (permit number).	12345 = 'Fish Hoover'
Boat Name	Enter the name of the boat.	'Fish Hoover'
Duration Type	Record the trip duration type.	½ = half day trip
	duration type.	$\frac{3}{4} = \frac{3}{4}$ or full day trip
		T = twilight
		O = overnight trip
PC Mode	Enter the appropriate PC mode.	P = open party trip
		C = boat was chartered to a private party
Departure & Return	Record the date and time of the departure	Depart 10/26/20 0700
	and return of the	Return 10/26/20 1700
Date = (MM/DD/YY)	CPFV.	
Time = Military hours		
DAYS Fished	Enter the days fished for this trip.	1 = all fishing within one day
BOAT ANGS	Enter the total # of anglers on the sampled trip (include any crew that take fish).	25 = 25 eligible anglers

Field Name	Instructions	Coding Examples and Formats
Target 1st = primary target 2nd = secondary target	Record both the primary and secondary targets of that trip. If the angler states "any" as a target, then record the targets of the boat. Targets will be recorded using the 5-digit alpha codes.	HALCA = targeting California Halibut
AREA	Record the water area code where the	N = (< 3 mi)
	majority of fishing effort (fishing time)	O = (> 3 mi) B = enclosed bay
	occurred for each primary and	Island Codes:
	secondary target.	F = Farallones
		1 = Coronados
		2 = San Clemente
		3 = Santa Catalina
		4 = Santa Barbara
		5 = San Nicolas
		6 = Anacapa
		7 = Santa Cruz
		8 = Santa Rosa
		9 = San Miguel
GEAR	Enter single letter code for the fishing	H = Hook and Line
	Ů	T = Troll

Field Name	Instructions	Coding Examples and Formats								
	gear used by the boat for the target.	S = Spear N = Bait Net								
	The gear should be determined and recorded for each primary and secondary target identified.	TY = Bait Not								
DD?	If the boat used a descending device of	Y = descending device was used on this trip								
	any kind to release discarded fish, record a Y; otherwise, record N. Descending devices can increase the survival rate of fish that are suffering from barotrauma, by returning them to the proper depth. Examples of descending devices include inverted/weighted milk crates and specially designed quick-release hooks.	N = No device was used								
	ANGLER									
Catch Recorded	Indicate here when you have confirmed that the obs and unobs/RELS catch	Y = catch was recorded for angler								
obs= observed	has been recorded on the Catch and	12.20								

Field Name	Instructions	Coding Examples and Formats
reported=unobs & RELS	Discard Form for this angler.	N = no catch was recorded for angler
		BLANK or DK = catch was missed for some reason; details must be provided on the ASF
Angler#	Record a number in consecutive order (starting with 1) for every angler	REFUSALS/LANGUAGE BARRIERS: do NOTissue sample number
	interviewed (except refusals/barriers).	Record an "R" (refusal) or "B" (language barrier) in the Angler # box
Bag #	Enter the bag # used by this angler, if one is issued.	32 Blue = bag number of the angler
Anger REFERENCE	Use this space to record notes that	'Bob' 'Father with son'
	may help you identify the angler. This field will not be used by data entry, so the format is open.	'Kid with cowboy hat'
DAYS fished (12 mo)	Ask angler how many saltwater finfishing trips within the last 12 months that	52 trips = fishing 1 day/wk over the last 12 months

Field Name	Instructions	Coding Examples and Formats
	occurred in or departed from CA, excluding today.	
Zip Code	Record the angler's permanent residence zip code.	90210 = Beverly Hills Refused = R Don't know = DK Sampler didn't ask = DA Foreign codes are in the back of this manual

PC Angler Form: Specific Editing Checks Header:

- Make sure to use the correct F/G boat number and vessel name in the header.
- 2. If the trip is a full day, still code the trip as ¾ because the duration is defined as ¾ to full day trip.
- 3. Twilight trips are often called 'sundowners'.
- 4. Include crew members in the total anglers count when crew take fish home.
- 5. The gear codes M (mooch) and B (both troll and mooch) are SALMON gear types ONLY.
- 6. If there is no secondary target, leave blank or line-out that field.

Main Form:

- Make sure each angler who is interviewed is assigned a unique angler # and has their own separate row on the PC Angler Form.
- 8. While anglers have their own unique number, there may be more than one angler contributing to a bag (i.e. bag #s may be repeated if they include more than one anglers' catch, but angler #s are not repeated).
- 9. Anglers may only attribute their catch to ONE bag.
- 10. Check the PC Catch and Discard Form to make sure the anglers and bags recorded on the PC Angler Form matches up.
- 11. The Catch Recorded column must be filled out for each angler interviewed. Fill out both the observed and unobserved/released boxes for each angler. If these boxes are left blank, it is considered an incomplete interview, a (DK) bag, and it will be discarded. Remember "DK" means that the interview is unusable.

12. Refusals and language barriers do not get an angler #. An "R" or "B" should be put in the angler #box. Verify that no refusals or language barriers have received a sample #.

Example of Onboard Angler Form

ASSN	IID	Date (MM/DD/	YY)	CNTY	SITE	OSP	Port	Sar	mpler#			Sampl	er Last Name			
112601		11/08/18		83	400	SBA	١	- 10	02		SN	IITH				
DFG B	oat#	Boat	Name		Duration Type	PC Mode (P or C)			arture &	Return Time	DAYS fished	TOTAL	TARGET	AREA	GEAR	DE
39022	-	Stardust					Depa	rt .	8/18	0600			1st RFGEN	7	н	Υ,
aptain	. Jaso	n			3/4	Р	Retur	11/0	8/18	1605	1 1	10	^{2nd} LNGCD	7	н	١
atch		_	_	Angler		DAYS			Catch		_	$\overline{}$	Angler		DAY	S
corded	ANGLER	# BAG#		REFEREN		ished (12 m	0)		Recorded	ANGLER	BAG #	.	REFERENCE	fis	ned (1	2 mc
obs ported	ANOLLIN	" DAG"		(angler nar lescription,		Zip Co	ie		obs reported	#	DAG II	(ang	ler name, description, etc.)	z	їр Со	ode
porteu		_	۳	rescription,	etc.)				obs	-		+	610.)	\vdash		_
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`Y 👍						10			ous							
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ıration	П		_				Zφ			P.				_		

AREA (Water Area & Island): Water Area: Nearshore (<3mi), Offshore (>3mi), enclosed Bay/estuary/harbor, Mexico

Island: 1-Coronado, 2-San Clemente, 3-Catalina, 4-Santa Barbara, 5-San Nicolas, 6-Anacapa, 7-Santa Cruz, 8-Santa Rosa, 9-San Miguel, F-Farallones

GEAR: Hook & line, Spear, Bait Net, Troll Salmon gear only: Mooch, Both (mooch & troll)

Invert gear only. Pol.#. Flat # or Rejdid # hoop net, snar#, sGuba, free Diving

DD? = Was a descending device used on this trip? Yes or No

Catch Recorded: Y = Yes, type of catch (obs or unobs/RELS) occurred and was recorded. N = No, Type of catch did not occur

DK = Don't know (didn't examine catch or didn't interview angler)

Angler #: Number or Refusal or Language Barrier Angler # Flag: Crew

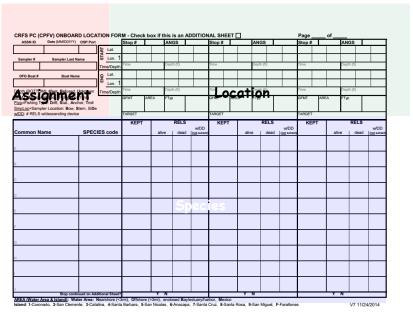
PC (CPFV) ONBOARD LOCATION FORM

The CPFV Onboard Location form collects the fishing locations, depths, times, and species counts for observed anglers.

The CPFV Onboard Location form has a front and back side to cover several fishing locations. For trips that use additional sheets, you will code the location number or species numbers for those observations on an additional location. Information from the top of the additional sheet will be used to link the data with the primary sheet and other PC forms that contain data collected on that trip.

CPFV Onboard Location Form Layout

The form has three major areas for data on the boat trip, the locations fished in columns and the catch species in rows. The location columns have two sub-areas for coordinate and physical data.



- Boat assignment data (top left of form)
- Fishing location data (top right columns)
- Species count data (bottom rows)

ASSN ID	Date (MM/DD/YY)	OSP Port
Sampler #	Sampler Last N	ame
DFG Boat #	Boat Nam	e

Boat Assignment Data

There are seven boat assignment data items, which are used to link the data to the other PC forms and to provide some unique information about the CPFV trip. All these items are required.

Fishing Location Data

There are 16 data items for

each location. There are three columns on each side of the form allowing you to record a total of six stops per sheet. Each fishing stop will have its own column. Not all the items are required at each stop.

Coordinate Data – latitude, longitude, geographic format (use default mode (1 = deg,xx.01 min)), area fished, and start and end times.

	Stop #		ANGS	
Lat.				
Lon. 1				
e/Depth	Time		Depth (ft)	
Lat.				
Lon. 1				
ne/Depth	Time		Depth (ft)	
	GFMT	AREA	FTyp	SmpLoc
	TARGET			
	Lon. 1 e/Depth Lat. Lon. 1	Lat. Lon. 1 e/Depth Lat. Lon. 1 image: Time Con. 1 image: Time GFMT	Lat. Lon. 1 e/Depth Lat. Lon. 1 e/Depth Time GFMT AREA	Lat. Lon. 1 e/Depth Time Depth (ft) Lat. Lon. 1 ie/Depth Time Depth (ft) AREA FTyp

Physical Data – depths, observed anglers, fishing type, Sampler location and primary target. Depth is used to help allocate catch and effort into depth zones. Depth is also used to estimate mortality of released catch.

Depths may be obtained from the skipper. Based on section 105.5, the Sampler is allowed to view the vessel depth

finder. Should you think the depth information from skipper is inaccurate, leave the location information blank and inform your Lead.

Q. What if the batteries on my GPS fail?

A. Put in the spare batteries. If the GPS fails, ask the captain for locations from the vessel GPS, if refused, leave the location blank. Record all other items, including times. If there is no GPS on the vessel, leave the location blank and write a comment about what happened.

Q. What if the captain does not want this location recorded?

A. Ask if we can record the location without the seconds (e.g., within one mile, 3232___ 1910__), otherwise leave the location blank. Record all other items, including times.

Species Count Data

There are 10 rows for species counts for each location column. There are two items to identify the species on each row: the common name and the 5-character alpha species code.

		KEPT		RELS		KEPT		RELS		KEPT		RELS	
I					w/DD				w/DD				w/DD
Common Name	SPECIES code		alive	dead	(not subset)		alive	dead	(not subset)		alive	dead	(not subset)
l	1												
A	1			l									

For each location column there are four items to record for each species row: the number of fish kept, released alive, released dead, and released with a descending device (w/DD). The counts of fish must be for the number of observed anglers in that location column.

Q. If a fish is returned alive with a DD, do I tally it in both the RELS alive and RELS w/DD fields?

A. No. The RELS w/DD field is <u>not a subset</u> of the other RELS fields. Code the fish as descended.

Recording Numbers Kept and Returned

	dot-line system								
1	•	6	ľ.						
2	:	7	П						
3	: .	8	П						
4	::	9	Ŋ						
5	1:	10	×						

The method used for recording the count for fish kept or returned is called the "dot-line system". The system allows for a count to ten in less space than the more common "count-mark" (i.e.) system does going to five. When editing your forms for the day, decode the dot-line system by writing the actual number to the right, and circling it. The key to this system is printed on the back of the CPFV Form.

Refused Items

The items that may be "refused" are depth and location. The captain may decide that a location is 'secret' and not want you to record it or the depth. Document all such refusals and contact your Lead.

All other items are dependent on the Sampler monitoring activity on the boat and may not be coded as 'refused' (i.e. fish counts). In cases where the Sampler is unable to determine Sampler-dependent information, the item(s) may be coded as "don't know" with an explanation on the Assignment Summary form. It is expected that Sampler-dependent data will be collected.

CPFV Onboard Location Form Item by Item Instructions

PFV Onboard Location Form Item by Item Instructions							
Field Name	Instructions	Coding Examples and Formats					
BOAT ASSIGNMENT							
Check box if ADDITIONAL SHEET	This is used to indicate whether this is the primary sheet, or an additional sheet.	Box checked if this is not the first sheet used for this assignment					
Page of	This is used to indicate total number of pages. Each side of the form is considered a page. The assignment data must be the same on all forms.	"1 of 2" on the first page "2 of 2" on the second page					
ASSN ID	This is the same as on the Assignment Summary Form and is used for data tracking. Enter the 6-digit assignment ID number on all pages. (Refer to your schedule)	Enter assignment ID in the format MMDNNN where MM is the month ranging from 01-12, D is the geographic District from 1 to 6 and NNN is the sequence from 001 to 999 (PCOs are numbered 601-699.)					
Date	This is the same as on the Assignment Summary and is used to classify and track the data.	MM/DD/YY 01/01/20 = New Year's Day, 2020					
OSP Port	Enter the 3-letter alpha code used by the Ocean Salmon Project for this port.	FTB = Fort Bragg					
Sampler #	Use your 3-digit Sampler ID code.	100 = Joe Sampler					
Sampler Last Name	Print your last name to the right of your code.	"Smith"					
CDFW Boat #	This is CDFW vessel id number of the boat (permit number).	12345 = 'Fish Hoover'					
Boat Name	Enter the name of the boat.	'Fish Hoover'					
	FISHING LOCATI						
Stop#	Record the Stop # that is associated with the	1 = First fishing stop for this trip					

Field Name	Instructions	Coding Examples and Formats
	location data in this column.	
ANGS	Record the number of anglers observed for the catch counts at this location. When feasible, 10 anglers should be the target number of observed anglers, and a different set of anglers should be observed at each stop.	10 = ten anglers observed for catch at this location
Start Latitude	North latitude in one of the valid formats at the start fishing time.	334996 = 33 degrees 49.96 minutes north latitude (GFMT=1) R = Captain refusal Blank = Don't know
Start Longitude	West longitude in one of the valid formats at the start fishing time. The hundreds place is pre- coded to 100 with a "1".	182474 = 118 degrees 24.75 minutes east longitude (GFMT=1) R = Captain refusal Blank = Don't know
Start Time	This is "lines down" time. Record the time in 24-hour format when fishing started at a new location.	0000 = midnight 0001 = one minute after midnight Blank = Don't know
Start Depth	Record the start bottom depth in feet, 1 fathom = 6 feet.	60 = sixty feet Blank = same as start R = Captain refusal Blank = Don't know
End Latitude	North latitude is one of the valid formats at the end fishing time. An ending location is not necessary if the boat did not travel more than 300 feet or fish for more than three minutes.	335002 = 33 degrees 50.02 minutes north latitude (GFMT = 1) R = Captain refusal Blank = same as start (i.e., anchored stop), or don't know
End Longitude	West longitude is one of the valid formats at the end fishing time. The hundreds place is precoded to 100 with a "1". An ending location is not necessary if the boat did not travel more than 300 feet or fish for more than 3 minutes.	182461 = 118 degrees 24.61 minutes east longitude (GFMT = 1) R = Captain refusal Blank = same as start (i.e., anchored stop) or don't know

Field Name	Instructions	Coding Examples and Formats
End Time	This is "lines up" time. Record the time in 24-hour format when fishing ended for this location.	0500 = 5am 1800 = 6pm
End Bottom Depth	Record the end bottom depth in feet.	50 = fifty feet BLANK = same as start (i.e., anchored stop) or don't know
GFMT	Geographic Format – The measurement units used to record the latitude and longitude coordinates at the start and end fishing times. All four position records must be in the same units. For longitude all fishing locations the hundreds place has been pre-coded with a "1".	The four geographic formats (GFMT) expected to be read from boat GPS and loran equipment (with proper punctuation): 1 = Degrees, minutes – DDMM.MM 3 = Degrees, minutes, seconds – DDMMSS
AREA	Distance from shore where the majority of fishing occurred.	N = Nearshore (< 3 mi) O = Offshore (> 3 mi) B = enclosed bay/estuary/harbor Island Codes: F = Farallones 1 = Coronados 2 = San Clemente 3 = Santa Catalina 4 = Santa Barbara 5 = San Nicolas 6 = Anacapa 7 = Santa Cruz 8 = Santa Rosa 9 = San Miguel
FTyp	Fishing Type- This is one of the four predefined types of boat movement used for the fishing activity.	D = Free drift (engine not in gear) S = Stationed (engine in/out of gear to maintain position) A = Anchored (boat attached to the bottom) T = Troll (engine in gear and powered to trolling speed)

Field Name	Instructions	Coding Examples and Formats
SmpLoc	It is important to observe anglers from different parts of the vessel, as catch rates can differ. Record where on the boat the observed anglers are located.	B = Bow S = Stern D = Side
TARGET	Record the 5-digit alpha code for the target species for this stop.	HALCA = California Halibut
	SPECIES CATC	Н
Common Name	This is the approved AFS common name.	'BROWN ROCKFISH'
SPECIES Code	Use the 5 letter CRFS alpha code.	RFBRN = Brown Rockfish
KEPT	Record the number of fish of species <i>kept</i> at this location by the observed anglers.	2 = two kept Blank = None kept
RELS alive	Record the number of fish of species <i>released alive</i> at this location by the observed anglers.	1 = one released alive Blank = None released alive
RELS dead	Record the number of fish of species released dead at this location by the observed anglers. Fish that are alive but are obviously not going to survive (due to severe wounds or inability to swim down) may be coded as dead.	10 = ten released dead Blank = None released dead
RELS w/DD (<u>not</u> subset)	Record the number of fish of each species that were released with the aid of a descending device.	1 = one released w/DD Blank = None released w/DD
Additional Sheet?	Indicate here if you require another sheet to capture all species for this stop.	Y(circled) = YES, Add'l sheet N(circled) = NO Add'l sheet

PC Onboard Location Form Coding Tips

Trolling between Locations

Trolling is common for tuna and salmon. The boat will troll until a 'hook-up' occurs and then stop to have anglers reel in their fish. On the boat location

form, you should be recording a new "stop" for every drift and troll. When the boat stops, this is the end position of the troll. Make sure that you bring extra data forms to record locations. For trolled locations, the number of observed anglers is the number of trolling rods you can observe.

Non-Stop trolling

For an entire trip of trolling continuously, the Sampler may record starts and stops hourly or when the boat makes a major change in heading, such as when reversing direction along a stretch of coast

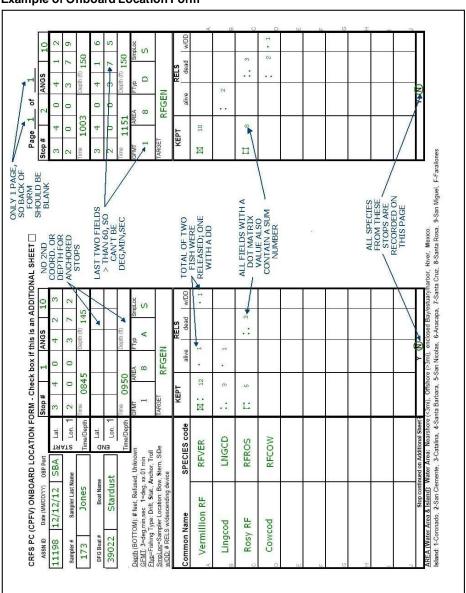
PC Location Form Specific Editing Checks Header:

- Make sure the header information is completely filled out and matches all other PC forms of an assignment.
- 2. OSP Port codes are never left blank.
- Boat name and CDFW Boat # cannot be left blank.

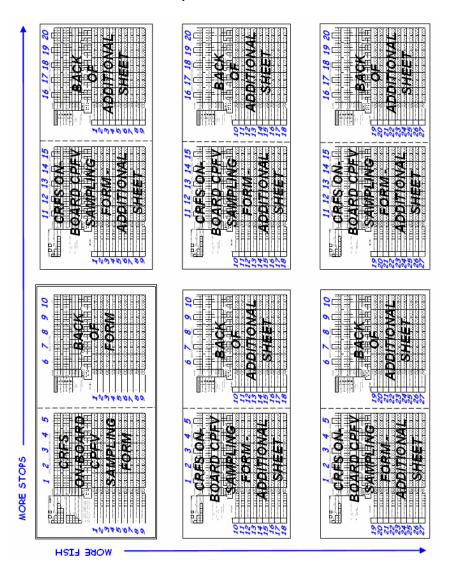
Main Form:

- 4. Species codes and names must be listed on first page.
- Make sure the GFMT matches the location in 'seconds' (or '100th minute'). Coordinates in deg.min.sec format can't end in a number greater than '59'.
- Make sure the GPS unit is not set to decimal degrees; this is not a valid format.
- 7. GPS devices should be set to deg,xx.01 min. as the preferred format.
- 8. Start time and depth should not be left blank.
- Start and end coordinates and times must be provided for all troll and drift trips.
- Do not leave location fields blank, except the end location when anchored or if the GPS unit fails and cannot get coordinates from crew.
- 11. If no fish were caught, leave field blank. Do not code as "0" easier for data-entry.
- 12. Fill in stop numbers on any additional sheets.
- 13. Check that the GFMT is correct (or does not conflict) with the stop location coordinates. FType must agree with the coordinates given (i.e. two different sets of coordinates can't be on an anchored stop).
- 14. Make sure that the fish counts are clear and legible for entry staff and that each field with a tally mark also has a sum total recorded and circled.
- 15. When recording fish released with a DD (descending device) make sure to remember it is not a subset of released fish.

Example of Onboard Location Form



Onboard Location Form - Multiple Sheets



CPFV ONBOARD CATCH AND DISCARD FORM

The CPFV Onboard Catch and Discard form collects all of the biological data from the PC trip. Effort, length and weight of obtained catch and reported catch are all captured on this form. Discarded fish that are opportunistically measured will also be recorded here.

The CPFV Onboard Catch and Discard form has front and back sides to capture a number of species records. For trips that use additional sides and/or sheets the Sampler will utilize the Page __ of __ fields at the top of each form. Information from the top of each additional sheet will also be used to link the data with the primary sheet and other PC forms that contain data collected on that trip.

CPFV Onboard Catch and Discard Form Layout

The form has four major areas for data on the boat trip: Boat assignment data, effort, discard fish data and catch/biological data.

Boat Assignment Data

There are seven boat assignment data items, which are used both to link the data to the other PC forms and to provide some unique information about the CPFV trip. All these items are required to be completed for the form to be acceptable.

CRFS PC	(CPFV) ONBOAR	D CAT	CH AND	DISCARD FORM V9 11/19/12	Page _	of
ASSN ID	Date (MM/DD/YY)	OSP Port	Sampler #	Sampler Last Name	DFG Boat #	Boat Name

EFFORT						
ANGLER #(s) from Angler Form	ANGS Total	BAG # (Sample #)				
A						

Effort Data

There are three effort data items, which are used in calculation of catch per unit effort (CPUE). The unit of effort used in this calculation is the angler bag, but the angler # field is also used as a point of reference.

Discard Fish Data

The primary purpose of this <u>measurement</u> data collection is to estimate the total metric tons of fish *discarded*. In the past, the mean weights of kept fish were used to calculate all weight estimates. However, the size of discarded fish may differ from retained catch, leading to a potential bias if only kept fish sizes are used to estimate discarded catch.

Location of discard onboard CPFV trips ('stop #' on the form) is collected because management methods include latitude, distance from shore and depth criteria. The CPFV stop number links the fish size to these criteria for management analysis. For example, bottom depth

may be used to apply additional mortality to the rockfish released alive that are susceptible to barotrauma.

To capture these data, the goal for onboard CPFV discard measurement is to get a number of measurements that is at least equal to 20% of the counts of 'observed' returned catch, per stop, on the CPFV Form. Discard lengths from unobserved anglers count towards this goal as well, so consider all anglers equally when measuring discards. You will find it easier to get your discarded fish measurements by mentioning your intention to anglers during the pre-fishing interview.

Handling Live Fish

Do not allow live fish to remain aboard waiting to be measured before discard, as this may give the impression that we are allowing fish to sustain trauma or die needlessly. We don't want to increase the chance of mortality of released fish by obtaining our discard measurements. Here are some tips that will minimize the stress on the fish:

- 1. Handle fish with a wet rag or a glove
- 2. Avoid sticking fingers in the gill chamber
- 3. Avoid touching the eyes
- 4. Make sure hands and measuring board are cool and wet
- 5. Return the fish to the water as soon as possible
- 6. Do not ask the crew to bring aboard large giant sea bass, oversize sturgeon, etc., just so they can be measured before release. These fish should not be removed from the water and released boat-side.
- 7. Do not measure released salmon brought aboard during the onboard fishing location survey.

Discard Data Coding Tips

- 1. 100% of discarded non-retention species that are brought on deck should be measured.
- Lengths are required for discard fish records, but weights should only be collected on fish that are already dead.
- 3. Unusually small or large size fish should not affect your decision to measure the discard. Keep it random.
- 4. Discarded fish can also be recorded as RELS by an angler, but never as KEPT. If measured discards were released alive and dead for the same species, record the fish released alive in one row and the fish released dead in another row. Double counting should not happen.
- 5. Fish that are cut up for bait, filleted, taken home or given to others are NOT discarded fish.
- Discarded fish are <u>not</u> connected to individual anglers. For each row, if the Discard field has a value, then the Effort fields should be blank, and vice versa.

Catch and Bio Data

This section will include the catch type, number and biological data for each species encountered. Multiple rows may be used to capture the numbers of different types of catch for the same species in a bag.

CAT	СН		BIO DATA				
	KEPT	RELS	Fork length / Carapace size (mm), sex (M/F/T)			r)	
SPECIES	obs	alive	Weight (decimal kg) or tag # (circle tag #)				
	unobs	dead	1 2 3 4 5				5
	obs	alive					
	unobs	dead					

Rockfish-Combo Trips

All finfish catch must be included in the angler's bag for a valid sample. Do not record invertebrate data on the Catch and Discard form. The Sampler may need to interview anglers about their catch twice during the trip if the boat targets separate species in different locations. Focus on collecting bio data on groundfish, before collecting bio data on other finfish species. If salmon are landed, include the catch in the angler's bag, but collecting salmon heads is not a priority in this mode. It is not necessary to complete a PCS sample on a Salmon/Rockfish trip. However, if you visually inspect/count all salmon and collect heads from ad-clipped salmon dockside, this data needs to be recorded on a Party/Charter Boat Sample (PCS) form. See chapter "PCS Sampling" for further details.

Total Items

At the bottom of each page, tally the number of Cowcod, Yelloweye Rockfish and Pacific Halibut encounters. Don't forget to notify your Lead on the same day of these encounters.

Kept	Rels								
HAI	_PA	RF'	ΥΕΥ	RFC	OW	RF(CAN	RF	3LK

PC Onboard Catch and Discard Form Item by Item Instructions

PC Onboard Catch and D		
Field Name	Instructions	Coding Examples
		and Formats
	<u>I</u> HEADER	
	HEADER	
Page _ of _	Enter, in sequence, the page number of the form and the total number of pages with data.	Example: Page 2 of 7
ASSN ID	Enter the 6-digit assignment ID number on all pages.	Assignment ID in the format MMDNNN where MM is the month ranging from 01-12, D is the geographic District from 1 to 6 and NNN is the sequence number from 001 to 999 (PCO assignments are 600-699)
Date	Enter the date in the MM/DD/YY.	01/01/20 = January 1, 2020
OSP port	Enter the 3-letter alpha code used by the Ocean Salmon Project for this port.	FTB = Fort Bragg
Sampler #	Enter your 3-digit Sampler identification number.	3-digit numeric code = 132

Field Name	Instructions	Coding Examples and Formats
Sampler Last Name	Write out your last name completely.	"Smith"
CDFW Boat #	This is the CDFW vessel ID number of the boat (permit number).	12345 = 'Fish Hoover'
Boat Name	Enter the name of the boat.	'Fish Hoover'
	EFFORT	
Angler #(s)	Enter the Angler # from the Onboard Angler Form of the angler(s) who are responsible for the catch.	1 = Angler # 1 from the Angler Form 1,3 = Angler #s 1 and 3 from the Angler Form 2-5 = Angler #s 2, 3, 4 and 5 from the Angler Form BLANK = discard measurement or boat fish; EFFORT column left blank
ANGS total	Enter the total number of anglers associated with this catch (licensed anglers+unlicensed anglers). This number should correspond with the	3 = three total anglers associated with this catch BLANK = discard measurement or boat fish; EFFORT column left blank

Field Name	Instructions	Coding Examples and Formats
	number of ANGLER #(s).	
BAG #	Enter the bag #(s) used by the Angler who are associated with this catch.	32 Blue = bag number of the angler
		BLANK = discard measurement; EFFORT column left blank
	DISCARDS	
Stop#	Enter the stop # from the Onboard Location Form where the fish was discarded.	5 = stop number '5'
	CATCH	
SPECIES	Enter the alpha code for each species or taxon of all fish examined or reported by the angler(s). Additional rows are used for anglers with multiple catch species. NOTE: If the angler is unavailable at this time to report unobserved catch, this data can be collected later.	"NO CATCH" No catch: enter zeros for numbers of fish Refused: This is a refusal, terminate interview

Field Name	Instructions	Coding Examples and Formats
KEPT Obs =	Kept Observed: Enter the number of fish examined for this angler(s). Sampler will identify and count each species retained by	Includes fished used for bait, thrown away, and fillets that are not identifiable
observed/verifiable by the Sampler	the angler(s). May include fillets with identifiable skin. Bags of	0 = None
Unobs = retained but not verifiable/available for the Sampler	unidentifiable fillets, fish not seen, or fish not counted by the Sampler get recorded as "kept unobserved" here.	Refused/don't know: interview is incomplete and should be terminated
RELS Alive = fish appeared	Enter the total number of fish reported as released alive and/or dead	Record species and number of fish ALIVE and/or DEAD
alive with no mortal injuries upon release	by the angler(s). Fish must have been landed and intentionally	0 = None
Dead = fish was thrown back dead/dying	released. Probe for catch that may not be remembered, such as bait species.	Refused / don't know = the interview is incomplete and should be terminated
	If measured discards were released alive and dead for the same species, record the fish released alive in one row and the fish	

Field Name	Instructions	Coding Examples and Formats
	released dead in another row.	
	BIO DATA	
Fork Length Size (mm), Sex	In the top row enter the fish's fork length	321 = FL in mm
	Add an M, F, or T after the length for sexed species.	F = Female M =Male T = Transitional (CA Sheephead)
		321F = female fish 321 mm FL
Weight/Head Tag #	Below the length, enter the weight in kg of the fish	5.35 = weightin kg
	For salmon and other relevant species, enter the head tag number below the length. Circle the tag number. For salmon heads not recovered or lost, enter the head tag number and code NRS (non-	12345 NRS = tagged head not recovered

Field Name	Instructions	Coding Examples and Formats
	recoverable specimen).	
	Salmon head tag numbers are 5-digit.	
	FOOTER	
Pacific Halibut Kept/Rels	Enter the sum of kept and released Pacific Halibut on the page.	
Yelloweye Kept/Rels	Enter the sum of kept and released Yelloweye Rockfish on the page.	
Cowcod Kept/Rels	Enter the sum of kept and released Cowcod on the page.	
Canary Kept/Rels	Enter the sum of kept and released Canary Rockfish on the page.	
Black Kept/Rels	Enter the sum of kept and released Black Rockfish on the page.	

PC Catch and Discard Form Specific Editing Checks

- Make sure all boxes are filled out in the catch section (including zeros).
 Stop # is used for discard measurements only. When obtaining discard measurements leave the effort section blank.

- 3. Make sure the cowcod, yelloweye rockfish and Pacific halibut boxes (bottom of the page) are filled out on each sheet.
- 4. Make sure all tag #s are circled.
- 5. Make sure all headers are filled out and nothing is left BLANK.
- 6. Make sure to fill out Angler Total on all bags.

Onboard Catch & Discard Form Example

ASSN ID		Date (MM/D	D/YY)	OSP Port	Sampler #	S	ampler Last N	lame	DFG B	loat#	Boat	Name									
071605	07/13/		07/13/		19	WAR	102		SMITH		1053	32	Seeke	er							
EF	EFFORT SOARS			EFFORT 5			EFFORT SO				САТСН		BIO DATA								
ANGLER #(s) from Angler Form	ANGS Total	BAG # (Sample #)	# Stop SPECIE		2770	/ Carapace siz ght (decimal kg															
FORM		Tracelle Strategy	, seein	20000000	unobs obs O	dead	268	311	: 159		4 :	5									
			1	SBKL	unata 0	dead ()	200	. 311	. 100	-3-											
Š.					ota O	C avis	309	. 315	8/	20	99										
			3	RFVE	R unote 0	dead ()			9	-											
2	1	1 4		RFBR	obs	O	309	315		100											
2	1	1 4		KEDK	Unote	dead 0	.49	.51	Ů.												
			0 0	CODO	obs 3	alve 2	302	320	330		ii ii										
				SCRO	A unotes	dead 0	.50	: .60	.65	20											
				SBKL	D 058	1	359	375	- N	10	- 12										
			e - 8	SDILL	0	dead 0	.66	.75	Ž.	- N	i.										
		2	\$3	ocwi	-IT 000 2	0	309	: 304	- 2	1											
				OCW	O Uniobia	dead 0	.41	.39	- 20	20	- 100										
7	1	1		SBKI	P 3	5	360	359	380		- 1										
			4	00,0	0	dead 0	.66	.66	.78	- 8	ii.										
				SBBA	AR unota_	alive 2	402	399													
			Ш	100000000000000000000000000000000000000	0	0	.88	.88		1											
				SCRO	CA unotes	o dead	275	255	300	30											
			Ш	CTM TO SEC	0		.40	.32	53	202 63	55 .										
5	1	3		SBKL	P unots	dead _	420	: 400	: 405	41.	40 :										
		255		ODINE	0	0	1.1	: .95	.99		1.3										
				SBBA	,	alve 2	400	70 2-22-23		-1-											
S.	-		-		ohe	alve 3	.89	. 86		18	16										
				SCR	CA unate 0	deed 0	301	315	-	-1-	- 2										
4					obs	o order	.53	317	-	-	- 1										
				REK	1 P 2	U	329	. 317			- 3										

ANGLER #(s): List the Angler # or #s from the Angler Form for all anglers contributing to the bag.

ANGS Total: Number of anglers associated with this bag.

Bag (=Sample): Record the bag number.

Boat Fish: Leave ANGLER # blank; write Boat Fish for BAG #.

For finfish, ANGS Total: TOTAL ANGS from the PCO Angler Form (i.e., number of eligible anglers incl. crew if they take fish home). Record obs. DISCARDS: Record the \$top # for measured discards; leave EFFORT columns blank; complete CATCH & BIO DATA columns.

0 .57

> 0 0 0 0 0

Kept Rels

HALPA

.51

Kept Rels RFYEY

0

Kept Rels

0

0

Kept Rels

0 0

Kept Rels RFBLK

Coding Boat Fish on the PC Onboard Catch & Discard Form

- ANGS Total is equal to the number of eligible anglers (i.e., the BOAT ANGS). This will include the crew and captain if they are keeping fish. Additional fish or fish that do not belong to a specific person are termed boat fish. The boat fish will be coded to the crew until each crew member has limits. Only then will left over fish be coded as boat fish.
- The bag # will be "Boat Fish."
- List the species and the number as kept-observed.
- If the Sampler has time, the fish should be measured and recorded in the BIO DATA section of the form. If the Sampler doesn't have time to measure the fish, then the species and number should be recorded.

	FORT		DISCARDS	CAT	СН		BIO DATA							
ANGLER #(s)	ANGS	BAG#	Stop	000000	KEPT	RELS			rapace size (m)			
from Angler Form	Total	(Sample #)	#	SPECIES	obs	alive dead	Weight (decimal kg) or tag # (circle tag #) 1 2 3 4 5							
					obs	alive	-	- 2	3	4	5			
/	40	Boat		25606	4	0	215	231	240	215				
/	42	Fish		RFSQS	unobs	dead								
					0	0								
					obs	alive	250	207						
				RFGRN	2 unobs	O dead	259	207						
					0	0								
—	-				obs	alive			-	-				
					3	0								
				RFSTA	unobs	dead								
					0	0								

Party/Charter Boat Non-Salmon Dockside (PCD) Mode Sampling

The CRFS PC (CPFV) Dockside form collects catch and effort data from non-salmon trips (PCD) that will be used to supplement data from onboard trips (PCO).

When to Sample Dockside

The goal of this sampling mode is to supplement onboard data with data from boat trips the Sampler usually cannot ride. Some examples are trips that target California or Pacific Halibut, Albacore Tuna, White Seabass, Dungeness Crab-sanddab combo trips, and trips conducted by six-pack vessels where there is no room for an onboard observer. In Districts 3-6, dockside sampling is most often conducted on lower-priority species to ensure CPUE is obtained from all trip types. Six-pack vessels are to be sampled dockside, unless special circumstances allow for Samplers to ride onboard. Overtime issues may prevent Samplers from riding long-range trips, so most are sampled dockside.

Scheduled vs. Opportunistic

PCDs can be assigned on the monthly CRFS schedule, or they may be done opportunistically during other assignments, if you happen to see PC boats come into port after fishing. Opportunistic PCDs are encouraged; however, do not miss interviews from your assigned mode in order to sample a PCD opportunistically. This commonly happens at PR ramps where a PC boat (NFPC6) comes in. For opportunistic PCDs, leave the ASSN ID blank (on both the ASF and PCD forms); the Lead will fill in an ASSN ID when the data are received at the office.

Sampling Unit

The sampling unit for PCD sampling is all catch and effort from one or more angler(s) bags) from a CPFV non-salmon trip. Collection of these data from at least one angler-bag constitutes a complete PCD sample. The Sampler should attempt to collect catch and effort data from as many angler-bags from as many boats as possible. For each new boat sampled during the assignment, use a new form.

PC Assignments

In general, PC assignments are single site samples. They may be assigned as onboard or dockside samples. Your Lead will assign the location of the sample. Rescheduling these assignments is not desirable to the survey, and you will contact your Lead if the assignment needs to be rescheduled or moved to an alternate PC site.

Data Collection

Information collected during a PC non-salmon dockside assignment includes: boat name and number, trip type and duration, departure and return times, number of anglers, targets with area and gear, descending device usage, avidity and zip code from as many angler-bags as possible and the corresponding catch and fish bio data. It is important to note that there is no "maximum" sample, meaning the Sampler should try to interview

as many boats and angler-bags from those boats as possible at the landing; however, one "sample" consisting of one angler-bag from one PC boat will fulfill the assignment.

The most important items to collect are the catch and effort data. It is important that you also report to your Lead any harbor closures, launch ramp closures, road closures or other incidents that prevent you from sampling or restrict or prevent fishing effort.

Combo-Trips: PCD versus PCS Sampling

CPFVs that have multiple targets including salmon may be sampled in both PCD <u>and</u> PCS modes (both forms are required). Note that all catch, including salmon, for all angler-bags sampled, goes on the PCD form; be sure to check the "OSP Form also completed" box on the top of the PCD form if the requirements for a PCS are met. Only salmon catch goes on the PCS form. Salmon-only boats may not be sampled with the PCD form, even if there was non-salmon bycatch while targeting salmon. In order to be eligible for PCD sampling, the boat must have had at least one non-salmon finfish target. Samplers should focus on collecting the PCD information prior to any PCS information. If time allows, then collect the information relevant to complete the PCS form. Remember all anglers should be asked about kept and released catch. Even if the Sampler does not think they will have time to collect PCS information, they still need to ask interviewed anglers about all kept and released fish (including salmon) for an interview to be complete.

Sampling Chartered Trips

You should be able to sample chartered boat trips along with open-party trips as they come back to the dock. Chartered trips can make up a large proportion of the total CPFV fishing trips, especially during the summer. Sixpacks are predominantly chartered trips. It is very important to sample chartered trips, as well as open-party trips in order to accurately estimate CPFV catch and effort.

Sampling Guidelines and Procedures

Plan to arrive at the port with adequate time to meet the first boat. You can estimate the time boats are going to return by looking at the previous day's report or calling the landing or booking agent. It will not always be 100% accurate, but it is the best way to anticipate the landing time. Most landings have set times that boats intend to return, depending on the duration type of the trip. Return times are also influenced by catch and weather. As you work in the port you will get a feel for the typical routine of the boats.

As the boat pulls up to the dock, identify yourself as a CDFW employee conducting CRFS. Count the number of anglers on the boat and verify this number with the crew before you leave. Ask the crew and captain if they contributed to the boat limit (took fish as part of the boat limit). If so, crewmembers are counted in the Boat Anglers field on the header of the data sheet. Try and intercept as many anglers (angler-bags) as possible as they leave the boat and ask if you can get some information on what they

caught and released. Some of the information on your data sheet will have to be provided by the captain or crew, such as Area Fished and catch location, if any. To save time and maximize the number of interviews, get the boat and trip information from the captain or landing agent after you have interviewed as many angler-bags as possible.

Try to sample as many boats as you can (note, each new boat sampled will require a new form but will have the same ASSNID). Check your data for any errors while at the dock before departure.

CPFV Refusals

Under section 105.5 (Title 14 CCR) Samplers have authority to access all CPFV boats. However, you may need to explain the survey and provide evidence that you are a CRFS Sampler. Always be prepared with copies of Title 14, section 105.5, your CDFW ID, a CRFS handout, and your Lead's business card so that you are prepared to demonstrate the legitimacy of the sampling program, and explain the survey. You should be familiar with the relevant sections in the CDFW regulation booklet on CRFS cooperation and have a copy to show to charter masters and landing personnel.

It is very important to document all attempts (successful or unsuccessful), to sample chartered trips on the <u>Assignment Summary Form</u>. Make sure that you indicate that the trip was either a charter or a party boat in the comment section of the form. If the attempt to sample was not successful, explain in the comment section why.

If you are outright refused by landing personnel or encounter any hostility or difficulties, leave a copy of Title 14, section 105.5 with the landing manager and contact call your Lead. Please provide your Lead with detailed documentation (date, name of individuals and vessels concerned, details of refusal or problem and how you dealt with it). Provide this information the same day of the event. Your Lead will initiate procedures to follow-up with the vessel.

Interviewing Anglers

Ask each angler about kept and unobserved catch. Unobserved catch includes any fish kept for bait and fish released alive or dead. You may have to remind anglers about any fish released or used for bait. For rockfish, try to probe to identify the released catch to the species level and avoid grouping at the higher level (e.g., "RFGEN"). You can use your field guides (time permitting) or reference catch that they kept and are in front of them. If an angler has a bag of rockfish fillets that he won't open or can't enumerate to species-level, it is best to skip this interview and move to the next angler-bag. The point is to get high-quality bag census to species-level rather than many bags of higher-level taxa.

Due to boat limits and fish-shuffling, do not attempt to record catch given-away to another angler, as these are now in another angler's bag (and that angler may not even know it). We don't want to double count the catch.

Boat Limits

With the CDFW boat limit regulations, open party and chartered boats can continue to fish until limits have been caught for all authorized anglers and crew onboard. Even seasick anglers who do not wet a line all day may leave the boat with fish, provided they have a fishing license. The crew may be interviewed if they kept fish towards the boat limit. The crew might distribute their personal catch to other anglers (note: this practice is illegal under Title 14, CCR Section 195(e)(2)).

NO CATCH Bags

Occasionally there will be anglers who do not catch fish and also do not accept fish from other anglers as part of the boat limit. In this instance they are a 'NO CATCH' bag and must be recorded as such. Try to pay attention to this situation because these anglers (often seasick) will try to slip by you at the dock or think that because they did not catch fish, they are unimportant. If we ignore these no catch bags, and leave them off the PCD forms, fish will be expanded to those anglers based on the interviews that are obtained from successful anglers. All eligible anglers, with or without catch, should be intercepted. Do not just interview the anglers with catch.

No Anglers in PC Mode

If you go to your assigned PC site as scheduled and no anglers are observed, refer to the Alternate PC Trips protocol described earlier in this section, unless your Lead has given you specific landings to sample as an alternate. If no effort in the assigned mode is found at the primary site and alternate sites, contact your Lead to determine the assignment's final disposition.

Sampling Dungeness Crab

Crab biological data will be ignored on PC trips, only record CRBGN as a target, area fished and gear type (including the number of pots pulled).

When sampling a PC non-salmon dockside (PCD):

- 1. Record all data on the CRFS PC (CPFV) Dockside Form.
- 2. Complete an ASF for both scheduled and opportunistic PCDs.
- Interview all angler bags, if possible, even if they did not catch any fish.
 However, one complete angler interview constitutes a valid (complete)
 sample from the boat.
- 4. For opportunistic PCDs during a PR assignment, list the CPFV as a NFPC6 boat on the PR form and conduct interviews with the CRFS PC (CPFV) Dockside Form. Write a comment on the ASF listing any dockside sampled PC boats. You should always have a PCD form with you at PR assignments.
- 5. If the PC boat is not listed on your PEC form (if prefilled) or its information has changed, get the boat name, boat number (if present), a vessel contact name (landing office, captain or owner) and telephone number for the phone survey. Inform your Lead.
- 6. If all the fish on the vessel are filleted, try to count fillets and, if possible, ID the species of fish based on attached skins.

- For fish that are reported to you, or fillets that you did not count or ID, 7. the fish should be recorded under "kept unobserved".
- 8. Ask crew and any anglers interviewed about descending device usage.
- Do not measure 'trophy fish' landed whole when the angler had all the 9. small fish of the same species filleted. Doing so can bias the average size of the landed catch. Code the trophy fish as "kept observed" omitting the bio data. Code the fillets separately as "kept unobserved" with the number reported by the angler (unless they can be identified and counted, then they would be "kept observed").
- 10. Do not record 'boat fish' during a PCD sample. All fish on the PCD form must be associated with an angler bag interview.
- 11. Gifts of fish are not to be accepted. Enforcement may find that you are either contributing to or helping the boat avoid an over-limit.
- 12. Do not sample salmon-only boats (only targeted salmon) that have by catch of another species as a PCD sample - only boats that targeted finfish other than salmon are sampled as a PCD.
- 13. Please pay attention to high priority species such as Yelloweye Rockfish, Cowcod and Pacific Halibut.
- 14. Crab catch and biological data is ignored; only include CRBGN as a target, location and gear type (including the number of pots pulled).

	Dockside (PCD) form Item by It	
Field Name	Instructions	Coding Examples and
		Formats
	HEADER	
OSP Form also	Check this box if there is a	This may happen if the
completed	PC Salmon Dockside form	boat did a combo
	that also has data from this	Rockfish/Salmon trip and
	boat.	the salmon data was put on
		the OSP Salmon dockside
		form
Page_of _	Enter, in sequence, the page	Example: Page 2 of 7
	number of the form and total	
	number of pages with boats.	
ASSN ID	Enter the six-digit	Enter assignment ID in the
	assignment ID number on all	format MMDNNN where
	pages.	MM is the month ranging
		from 01-12, D is the
	(Refer to your schedule)	geographic District from 1
		to 6 and NNN is the
		sequence from 001 to 999
		(PCDs are numbered 701-
		799 for scheduled PCD
		sampling assignments, and
		901-999 for opportunistic
		PCD sampling)
Boatof	Each boat sampled for the	Boat: (Boat #) of (Total # of
	dockside assignment	boats sampled per
	requires a new PCD Form	assignment)

Field Name	Instructions	Coding Examples and Formats
	and a unique boat number, starting with 1.	Boat : 2 of 4
	Enter in chronological order the boat number and the total number of boats sampled for the assignment. Enter boat number info on all pages.	
Date (MM/DD/YY)	Enter the numeric date as 2- digit month/2-digit day/2-digit year.	MM/DD/YY 01/01/15 = January 1, 2015
CNTY	Enter the 3-digit numeric county code.	037 = Los Angeles
SITE	Enter the 3-digit numeric site code.	014 = 22 nd Street Landing Sportfishing
OSP Port	Enter the 3-letter alpha code used by the Ocean Salmon Project for this port.	FTB = Fort Bragg
Sampler #	Enter your 3-digit Sampler identification number.	3-digit numeric code = 132
Sampler Last Name	Write out your last name.	"Smith"
Ohtain trir	TRIP INFORMATION information by interviewing the	-
CDFW Boat #	Enter the Fish and Wildlife CPFV Permit number for the vessel. Refer to the list provided by your Lead and verify by observing the number that is posted on the vessel or from crew/landing.	CDFW Boat # = 22776
Boat Name	Write out the name of the vessel. Observe the name that is painted on the vessel and refer to the list provided by your Lead.	Vessel Name = Monte Carlo
Duration Type	Use the coding at the bottom of the sheet to categorize the length of trip or provide a description of the trip type.	 ½ = half day ¾ = ¾ to full day T = twilight O = overnight Other = write in a description
PC Mode	Determine by asking crew or landing if the trip was open party or chartered to a	P = open party trip C = boat was chartered to a private party

Field Name	Instructions	Coding Examples and Formats
	private group. Enter the appropriate PC mode.	
Depart and Return Time and Date	Record the time and date that the vessel departed and returned to the dock for this trip.	1400 = 2 pm Date = MM/DD/YY Most trips will have the same depart and return date. Overnight trips are the exception
DAYS fished	Record the number of calendar days in which fishing effort occurred on the trip.	1 day = fishing occurred from 3 am to 7 pm within 1 calendar day
BOAT ANGS	Record the number of anglers, including crew, who would qualify for a CRFS interview and verify with crew/captain.	30 = thirty anglers Include the crew in the total if they fished (include anglers who did not fish but took fish home)
TARGET	Record the primary and secondary target for the trip.	RFGEN = Rockfish
AREA	Record the water area code where the majority of fishing effort (fishing time) occurred.	N = Nearshore (< 3 mi) O = Offshore (> 3 mi) B = enclosed bay/estuary/harbor Island Codes: F = Farallones 1 = Coronados 2 = San Clemente 3 = Santa Catalina 4 = Santa Barbara 5 = San Nicolas 6 = Anacapa 7 = Santa Cruz 8 = Santa Rosa 9 = San Miguel
GEAR	Enter the single letter code for the fishing gear used by the boat. Codes can be referenced at the bottom of the page.	H = Hook and Line T = Troll S = Spear N = Bait Net
DD?	Determine if any descending device was <u>used</u> to release fish on this trip. Record the appropriate designation.	Y = Yes, a descending device was used on this trip N = No, none used

Field Name	Instructions	Coding Examples and Formats
LOCATION	Determine the general location of where the majority of the fish were caught. If no catch, code the primary location of the boat effort. Samplers should use their maps and have the captain show where fishing took place. Latitude and longitude from the captain are also acceptable.	Block-Box method is preferred: 212-01 (block & one box) 235-12-14-15 (block & up to 3 boxes or two 3- digit boxes for inland marine waters bbb-bbb) 252 (block only)
DEPTH	Enter the mean bottom depth in feet for the catch location obtained from the captain.	100 = 100 feet
	EFFORT Interview anglers to obtain the	nie data
Sample #	Record a sampler number for this individual angler or group interview record (angler-bag). If the angler refuses to be interviewed or refused key data (catch and effort information) then an "R" should be recorded in the box with no sample number. A language barrier that prevents an interview should be recorded as a "B" with no sample number. Record the number of	1 = first interview R = refusal B = language barrier Do not record "boat fish" on the PCD form; if it is not assigned to an angler, do not record it
ANGS	individuals who fished for this angler-bag.	2 = 2 anglers fished for this sample #
DAYS	Randomly select one angler for this angler-bag. For this angler, record the number of days this individual has been saltwater sportfishing in	12 = angler fished 12 days within the last 12 months Refused = R Don't know = DK Sampler didn't ask = DA

Field Name	Instructions	Coding Examples and Formats									
	California (or on trips departing from California) within the last 12 months not including today's trip.										
Zip Code	Determine the residential zip code for a randomly selected angler contributing to the bag.	90720 = angler resides in Los Alamitos, CA Refused = R Don't know = DK Sampler didn't ask = DA Foreign country = 3-letter country code, e.g. Ireland = FIE									
CATCH Interview anglers to obtain this data											
SPECIES	Use the 5-letter alpha code to record the catch species.	HALCA = California Halibut									
KEPT obs/unobs	Enter the number of fish landed and retained for this interview record. Examined catch is tallied under "obs" (observed), while unavailable catch such as fish used for bait is tallied under "unobs (unobserved)"	Includes whole fish examined by the Sampler, fish used for bait, thrown away, given away, and fillets. No catch = zero									
	under unobs (unobserved)	Refused/don't know: interview is incomplete and should be terminated									
RELS alive/dead	Enter the total number of fish reported as released alive or dead by the angler(s) for this interview. Fish must have been landed or have been intentionally released.	Record by species the number of fish released alive and/or dead No catch = zero Refused/don't know = the interview is incomplete and									
	Probe for catch that may not be remembered, such as	should be terminated									
	bait species.										
Fork	BIO DATA	321 = FL in mm									
Fork length/carapace size (mm), Sex (M/F/T)	In the top row enter the fish's fork length or the carapace length for crab/lobster in mm. Add an M, F, or T after the length for sexed species.	F = Female M = Male T = Transitional (Ca sheephead)									
	iong an ion do kod dp do los.	321F = female fish, 321 mm FL									

Field Name	Instructions	Coding Examples and Formats
Weight (decimal kg) or tag#	Below the length, enter the weight in kg of the fish	5.35 = weight in kg 12345 NRS = tagged
	Do not weigh headed or gutted fish.	salmon head not recovered
	For salmon and other relevant species, enter the head tag number below the length. For salmon heads not recovered or lost, enter the head tag number and code NRS (Non-Recovered Specimen). Salmon head tag numbers are 5 digits.	
	FOOTER	
HALPA	Enter the sum the number of kept and released Pacific Halibut from the page.	
RFYEY	Enter the sum the number of kept and released Yelloweye Rockfish from the page.	
RFCOW	Enter the sum the number of kept and released Cowcod from the page.	
RFCAN	Enter the sum the number of kept and released Canary Rockfish from the page.	
RFBLK	Enter the sum the number of kept and released Black Rockfish from the page.	

Specific Editing Checks

- The PCD interview is not complete until you have asked all anglers contributing to the bag about what was discarded. Data are unusable unless BOTH retained catch and discards are recorded.
- If there are more than five fish of one species measured, go to the second row and repeat the species code in the species box, but do not repeat catch totals. All catch and discard information for a species should go on the first line only.

- 3. For an opportunistic sample (6-pack PC trip at PR1 site, for example), leave the assignment ID blank on the PC Dockside form an assignment ID will be given to it by your lead.
- 4. Fillets that you see but can't identify the number of fish or the species, are considered unobserved, even if you looked at them.
- 5. If there is salmon aboard a PC boat, please also fill out an OSP Salmon Dockside form with that information on it and check the "OSP Form Also Completed" box at the top of the CRFS PC Dockside form. It will count as a sample for OSP; even if it was a combo trip (RF/Salmon trip, for example).

Example of PC Non-Salmon Dockside Form

ASSNI		oat of		M/DD/YY)				Port		ampl		npleted	=		ge _ pler La				_
11275		2 4		30/18	111	43	С	IS		20					SMI				_
DFG Boa	4.#	Boat N		Duration	PC Mo	de	Depa	ture & I	Retur	n	DAYS	TOTA	L			GEAR		D?	
				Туре	(P or C	Depa		DD/YY)	Ti	me	fished	ANGS		TARG	EI	AREA	Ŗ		7N
37693		ALOHA S	SPIRIT	3/4	Р	Retu	11/	30/18	05	03	1	23	П	RFG	EN	7	Н		γ
aptain:	10	HN DOE		Í		Retu	11/3	30/18	16				ľ	LNG		7	Н		
		ATION 8				\vdash	LOC	ATION		k-box	or Lat/L	.on)	+	DEP	TH Av	_		ttom ((ft)
		pth where t if no catc		he fish ca	ught, c	" <u> </u>							+			31	ь		
	EFFC	ORT			CATC	н			/08	-16-1	15		BIC	DATA					_
ample#	ANGS	DAYS FIS				KEPT	RELS			F				e size (m			/T)		
r R, B, Boat Fish	Total	12 mon		SPECIE	S	obs	alive	-1	1		2	Weight ((dec	imal kg) o 3	r (tag #) 4			5
		18				obs	alive	1				Fillet	ed	in the	າລອ	-	_		
1	2	9303	12 mo	RFVE	R	unobs	dead			+		Tillet	.cu	III tile i	Jag		-		_
		9303	zip			obs	2 alive	-		_							_		_
			12 mo	RFBC	C	2 unobs	0 dead			_									
			zip			0	1												
_		5	12 mo	DECT		obs 3	alive 1	3	12		276		- 2	291					
2	1	9301	3 zip	RFSTA		unobs	dead 0	0.	.72		0.51		().62					
						obs 2	alive 3	66	1M		711	F							_
			12 mo	LNGC	D	unobs	dead	,	2.8	-	3.5						-		
			zip			O obs	0 alive	-	0	-	3.3		_				-		_
R	3		12 mo			unobs	dead	-		_		Ret	us	ed Surv	ey		_		
			zip																
3	1	26	12 mo	RFCAI	NI.	obs 1	alive 0	3	29										
3	1	9031	.0 _{zip}	KFCAI	V	unobs 0	dead 0	0	.62										
			12 mo			obs 9	alive	4	21	1 407			3	391		401		37	71
				RFCOI	P	unobs	dead 0	1	.2		1.1			1.0	1	1.1		1	.0
		0	zip			obs	alive	╁		+			_	2.0	<u> </u>		-		
4	1		12 mo	RFCOV	N	unobs	1 dead	-		_							_		
		9032	6 zip			0 obs	0 alive												
			12 mo	RFVE	R	4	0	Filleted in bag but able to ID and o			nd c	oun	t						
			zip	IN VE		unobs 0	dead 3												
			12 mo			obs 2	alive 0			Fille	ted i	n bag b	out	able to	ID ar	nd c	oun	t	_
			12 1110	RFCOI	P	unobs	dead	1		+							-		_
los CREC C	OD CALL	MON CPFV D	zip	AMDIE FO	DM for -	O Iman tric	0	0	0	÷	0	0	0	1	1	_	0	0	Γ

Boat: (Boat #) of (Total # sampled per assignment)

Duration Type: 1/2 day, 3/4 to full day, Twilight, Overnight, Other-describe

HALPA RFYEY PC Mode: Open Party, Charter DD?: Was a descending device used on this trip? Yes or No TOTAL ANGS: all eligible anglers (including crew if they take home fish) AREA (Water Area & Island): Water Area: Nearshore (<3mi), Offshore (>3mi), enclosed Bay/estuary/harbor, Mexico.

Kept Rels

Kept Rels

Invert gear only: Pot #, Flat # or Rigid # hoop net, snarE, sCuba, free Diving

Sample #: # of interview, OR Refusal, Language Barrier, or Boat Fish. Sample # Flag: Crew

Rels

RFCAN

Kept

Kept Rels

RFCOW

Kept R Rels

Island: 1-Coronado, 2-San Clemente, 3-Catalina, 4-Santa Barbara, 5-San Nicolas, 6-Anacapa, 7-Santa Cruz, 8-Santa Rosa, 9-San Miguel, F-Farallones GEAR: Hook & line, Spear, Troll, Bait Net. Salmon gear only: Mooch, Both (mooch & troll).

CRFS PC	` ,									K	Page _	1	of _	2
ASSN ID	Boatof	ofDate (MM/DD/YY) CNTY S			SITE	SITE OSP Port Sampler #			Sampler Last Name					
116719	1 2	2 11/30/18 023		121	121 EUR 203		3	SMITH						
DEG Boat #	DFG Boat # Boat Name		Duration	PC Mode) De	eparture & I	Return	DAYS	TOTAL	Γ.	TARGET	AREA	GEAR	DD?
DI O Boat #			Туре	(P or C)	Date	(MM/DD/YY)	Time	fished	ANGS		IAKOLI	AR	픙	55.
71226	REEL STE	EEL	1/2	1/2 C		11/30/18	0700	1			SALCK	N	Т	Y/N
Captain:	TIM KLASS	EN	1/2		Return 1	11/30/18	1130	1	6	2nd	вотом	N	н	N
SPECIFIC LOCATION & DEPTH INFO:						LOCATION (block-box or Lat/Lon)					DEPTH Average Bottom (ft)			
Location and depth where most of the fish caught, or						217-95-85					90			
most of the effort if no catch.														

EFFORT CA				:н	H BIO DATA										
Sample #	ANGS	DAYS FISHED		KEPT	RELS	Weight (decimal kg) or (ag #)									
or R, B, Boat Fish	Total	12 months	SPECIES	obs	alive										
71311		Zip Code		unobs obs	dead	┝┷			2		3		4		5
1	2	1 12 mo	RFBLK	10	2	39	92	41	l1	38	39	4	42	39	99
		95503 _{zip}		unobs	dead 0	1.	.0	1.	.2	0.	92	1	.4	1	.1
		12 mo		obs	alive	38	38	40)7	43	34	4	23	52	21
В		zip	RFBLK	unobs	dead	0.9	98	1.	.1	1	.4	1	.3	2.	.4
c		12 mo		obs 1	alive	50	2F								
		zip	SCCAB	unobs	dead 0	2.	.2								
		12 mo		obs 1	alive	612M									
D		zip	LNGCD	unobs	dead 0	2.	2								
2	1	20 _{12 mo}	RFBLK	obs 5	alive 3	407		40)5	505		481		390	
		95501 _{zip}		unobs 0	dead 0	1.1		1.	1	2.2		1.9		0.98	
		12 mo		obs 2	alive 1	661M		71	1F						
		zip	LNGCD	unobs 0	dead 0	2.8		3.	5						
		0 12 mo	DEDLIK	obs 3	alive 5	400		412		399					
3	1	85754 _{zip}	RFBLK	unobs 0	dead 0	1.	1	1.2		1.1					
		12 mo		obs 1	alive 0										
Н		zip	RFCHN	unobs 0	dead	0.75									
		12 mo	INCCD	obs 1	alive 2	696F									
		zip	LNGCD	unobs	dead 3	3.3									
		12 mo	SALCK	obs 1	alive 0	701									
J		zip	SALCK	unobs 0	dead 0										
			DE SAMPLE FORM for s	almon trip	S.	0	0	0	0	0	0	0	0	18	10
Boat: (Boat #) of (Total # sampled per assignment)							_	,		_	_	_	_		
<u>Duration Type</u> : 1/2 day, 3/4 to full day, T wilight, O vernight, Other-describe PC Mode: Open P arty, C harter							Rels PA	Kept RF\	Rels YEY	Kept RF0	Rels COW	Kept RF	Rels CAN	Kept RFI	Rels BLK
			n this trip? Yes or No)									nome fish)		J_1\

AREA (Water Area & Island): Water Area: Nearshore (<3mi), Offshore (>3mi), enclosed Bay/estuary/harbor, Mexico.

Island: 1-Coronado, 2-San Clemente, 3-Catalina, 4-Santa Barbara, 5-San Nicolas, 6-Anacapa, 7-Santa Cruz, 8-Santa Rosa, 9-San Miguel, F-Farallones

<u>GEAR:</u> Hook & line, Spear, Troll, Bait Net. Salmon gear <u>only:</u> Mooch, Both (mooch & troll). Invert gear only: Pot #, Flat # or Rigid # hoop net, snarE, sCuba, free Diving

Sample #: # of interview, OR Refusal, Language Barrier, or Boat Fish. Sample # Flag: Crew

Party/Charter Boat Salmon Dockside (PCS) Mode Sampling

The CPFV salmon dockside form (PCS) collects catch and effort data that will be used to create in-season and post-season estimates of the recreational salmon harvest. This is done by examining at least 20% of the CPFV salmon trips in each port area during each bimonthly sampling period and collecting the heads from all adipose fin-clipped fish. There are two sampling periods each month: 1st to the 15th and the 16th to the end of the month.

Sampling Unit

The sampling unit for PCS sampling is all catch and effort from a CPFV salmon trip. All data must be collected in the sample unit to be considered a valid (complete) sample.

Data Collection

In the CPFV fishery, information collected beyond the header information includes: number of anglers, number of salmon landed by species, fishing method, number of salmon released by species, number of salmon taken by pinnipeds, fork length of adipose fin-clipped fishin mm, and assigned OSP headtag number for adipose fin-clipped fish. Also, CPFV names, CPFV numbers and sample time are collected in the course of sampling.

The most important items to collect are the catch and effort numbers, and the heads of all adipose fin-clipped salmon. You must count and visually inspect every salmon landed by the CPFV for an adipose fin-clip to obtain a valid sample from the boat.

Sampling Guidelines and Procedures

In the CPFV fishery, each salmon CPFV trip constitutes a sample. Each port will have a Sampler in charge of making sure the sampling goals are met. The Port Lead Sampler must ensure that a minimum of 20 percent of all salmon CPFV trips made in their assigned port during each bimonthly sampling period are sampled. Sampling days are not usually assigned and Samplers will have to determine when they need to be at their assigned port to sample boats. Try to distribute your samples throughout the sampling period. Do not leave your samples until the end of the period; weather can be unpredictable and can prevent boats from going out causing you to miss the minimum 20% sample rate. Aiming for a 25-30% sampling rate protects against activity towards the end of the period. There is also the possibility that there may be unknown activity from the launch ramp or by a transient CPFV in a berth. These both count toward the number of CPFV trips. Other CRFS/OSP Samplers may be sampling CPFVs at the port throughout the season. Coordinate with these other Samplers to obtain information about port activity, sampled boats, etc.

Plan to arrive at the port with adequate time to meet the first charter boats. You can estimate the time boats are going to return by looking at the previous day's report. It will not always be 100% accurate, but it is the best

way to anticipate the landing time. Most boats tend to return around the same time every day. These times are influenced by catch and weather. As you work in the port you will get a feel for the typical routine of the boats.

As the charter boat pulls up to the dock, identify yourself as a Sampler for the Department of Fish and Wildlife (CDFW), Count the anglers and fish as they pass by you on the dock. Look at each person's catch for adipose finclipped salmon. You must visually inspect every salmon landed by the CPFV for an adipose fin-clip to obtain a valid sample from the boat. If found, ask those anglers to wait aside and explain that you need to measure the fish and remove the head. Work quickly to attach the headtag, record the length of each fish and remove the head. Using your headtags in order will speed up data recording, but do not make the assumption that you are using your headtags in order. Verify that the correct salmon length is recorded with the correct headtag number on your data sheet. Once you have processed all fish and talked to each passenger, ask the deckhand and captain if they have any fish; if they do, process those fish, add them to the total kept and add the crew to the count of total anglers. Then ask the captain or deckhand the questions required to fill out your daily data form. Sample all the fish on one boat even if you have to miss another boat to do so. Try to sample as many boats as you can. When you are finished, make sure all heads are in your possession and noted on your data sheets. Check your data for any errors while at the dock.

When sampling a PC Salmon dockside:

- Get to the landing site in time to sample the CPFVs. The specific time will vary depending on the weather and effort. Use your best judgment; the pattern of the CPFVs from the last few days should give you an idea of when to arrive.
- Wear the proper, clean attire provided by CDFW. Clean your gear after each boat if time allows.
- 3. When you approach the deckhand or captain identify that you work for CDFW. Let them know you intend to sample the boat. After a short period, your presence will be routine to them. However your presence to each angler will not be routine and you will need to identify yourself as a CDFW employee. You must observe and count every salmon, checking for species, as well as counting the anglers.
- Try not to miss any boats. If two salmon CPFVs come into port at the same time, be random about which boat you choose to sample.
- 5. Record all header information; Date, OSP Port, Sampler ID Sampler Last Name, Other Samplers, and the page #.
- 6. Determine how many salmon were caught and retrieve salmon heads. As each angler disembarks observe their fish. Count the fish, checking to see whether the fish are Chinook or Coho and whether the salmon are missing their adipose fins. Record the number of Chinook and Coho on the data sheet.

If any fish are missing the adipose fin, explain to the angler that their fish contains a coded wire tag (CWT) and that you need to remove the head. Securely fasten a headtag through the lower jaw of the fish. Measure the fish, record the fork length in millimeters and headtag number on your salmon dockside form and then remove the head. Cut the head so the cut ends approximately two inches behind the eyes. Do not take the gills, collar or any flesh. Put the head in a plastic bag with the number on the tag facing out.

If the angler does not allow you to take the head, explain the importance of CWTs to salmon management. If they persist in refusing to relinguish the head, remind them that the law requires tagged salmon heads to be relinquished upon request by an authorized agent or employee of the Department. Show them Title 14. CCR Section 1.73(b). If you still cannot retrieve the head. attempt to get a length and attach a headtag to the fish. Explain that the angler can call the toll-free number on the headtag and the OSP will coordinate retrieval of the head. Record this Non-Recovered Species (NRS) on the data sheet. If attaching a headtag to a salmon is not possible, the head is still assigned a headtag. Place the headtag (with no head) in a plastic bag. Record this information on your data sheet and put "NRS" on the back of the corresponding headtag, and on the headtag report form. Be sure to correctly complete the NRS column on your data sheet. Lastly, inform your Supervisor about the refusal and they will take the appropriate action. If a Wildlife Officer confiscates any adipose fin-clipped salmon make sure that you put a headtag on the head and note any information that will help us retrieve the head at a later date, such as the Officer's name and contact information.

- If time permits, ask the anglers if they would like to receive information about their fish. Write the tag number (or series of tag numbers) on an orange information request card. Refer the angler to the card for instructions on submitting their contact information to OSP.
- 8. Ask the captain or deckhand how many Chinook Salmon were kept and released ("shakers") and if they released any Coho Salmon. Record the appropriate information in the correct box. Unknown shakers are recorded in the "Released Kings" column.
- 9. Count the number of people on the boat. The deckhand or captain can clarify if all were fishing (including the deckhand and the captain). Record the number of anglers in the appropriate column.
- 10. Ask the captain or deckhand if they mooched or trolled. Circle the appropriate letter, "M" = mooch, "T" = troll. If both methods were used, circle both "M" and "T".
- 11. Ask the captain or deckhand how many fish were lost to sea lions. Record the number of salmon actually taken by sea lions.
- 12. Be sure to record the vessel name and Fish and Wildlife number ("boat number") of the CPFV as well as the time of return. The

- boat number should be displayed on the wheel house, if you cannot locate it, ask the captain or deckhand.
- 13. Add up the totals at the bottom of the page.
- 14. Go over your data sheets as time permits and at the end of your sampling day. The captain should have most of the information in a log book if a data field was missed.
- 15. Fill out your headtag report sheet with the date, port and sampling mode of the headtags collected. Make sure that the headtags in your bag match the headtags written on the data sheets and the headtag report sheet.
- 16. Inventory the heads before you put them in the freezer.

CRFS-OSP Salmon CPFV Dockside Form (PCS) Item by Item Instructions

Field Name	Instructions	Coding Examples and Formats							
HEADER									
Page _ of _	Enter, in sequence, the	Page 1 of 2							
	page number of the form and the total number of								
Date	pages with data. Enter the numeric date	05/10/13 = May 10,							
Date	as 2 digit month, 2 digit	2017							
	_	2017							
OSP Port	day, 2 digit year. Enter the 3 letter alpha	SCR = Santa Cruz							
USP POIL	code used by the Ocean	SCR = Santa Cruz							
	Salmon Project for this								
	port.								
Sampler ID	Enter your 3 digit	3 digit numeric code =							
	Sampler identification	207							
	number.	207							
Sampler Last Name	Write out your last name	"DaSilva"							
	completely.								
Other Samplers, ID	Write the last name of the	Example: Phillips 302							
(w/data)	Sampler(s) you worked	(Y) = James Phillips,							
,	with and their Sampler	Sampler 302 also has							
	IDs, if known.	data sheets for this							
	·	PCS assignment.							
	Circle "Y" or "N" if other								
	Sampler(s) have data	If you sampled alone,							
	sheets.	leave blank.							
EFFORT									
Boat Name	Enter the name of the	Becky Ann							
	boat.								
Boat #	This is the CDFW vessel	32965 = 'Becky Ann'							
	ID number of the boat.								
		Never leave blank.							
Time Sampled	Enter a time stamp for	Use 24 hour format:							
	every CPFV boat that is	1700 hours = 5:00 PM							
	intercepted at the dock.	Never leave blank.							

Field Name	Instructions	Coding Examples and Formats
		Each returning boat should have a time stamp.
Gear	Circle "T" if the boat trolled for salmon. Circle "M" if the boat mooched for salmon. Circle both "T" and "M" if the boat used both types of gear.	(T) = Troll (moving/under power) (M) = Mooch (static/drifting)
Total Angs	Enter the total number of anglers (licensed and unlicensed), including crew if they fished.	30 = there were thirty people fishing on this boat
	CATCH	
Kings Kept	Enter the sum of King Salmon kept for each boat trip.	0 = No kings kept # = Number of Kings kept
Kings Rels	Enter the sum of King Salmon released for each boat trip.	0 = No kings released # = Number of Kings released
Coho Kept	Enter the sum of Coho Salmon kept for each boat trip. Clearly note any kept Coho and notify your Lead as soon as possible.	0 = No Coho kept # = Number of Coho kept
Coho Rels	Enter the sum of Coho Salmon released for each boat trip.	0 = No Coho released # = Number of Coho released
Sea Lion Take	Enter the number of salmon reported taken by pinnipeds for the trip. The angler, deckhand, or skipper must have seen the pinniped take the fish.	0 = No salmon lost # = Number of salmon lost
	BIO DATA	
Headtag # 1, 2, 3	Enter the headtag number assigned to ad- clipped fish#1,#2, #3 (for each boat).	Example: 50001 = the headtag number assigned
	Use additional rows for multiple ad-clipped fish from each boat.	

Field Name	Instructions	Coding Examples and Formats
FL (mm)	Enter the fork length (in mm) of ad-clipped fish #1, #2, #3corresponding to the headtag number.	Example: 695 = the fork length of the ad- clipped salmon corresponding to headtag 50001
NRS*	Check this box when you are unable to recover the head of an ad-clipped salmon. NRS heads should have a headtag number assigned to them. Try to attach the headtag to the NRS salmon but if you are unable to, put the headtag by itself in a baggie and process as usual.	= This head was recovered. This head was not recovered. Note: write "NRS" on the back of the assigned headtag if unable to attach to the NRS salmon. Write "NRS" next to this headtag number of your Headtag Report.
	FOOTER	
Comments	Use this section to write any important comments.	Example: Doble and Becky Ann came in at the same time; so I randomly chose Becky Ann. Doble was missed.
Page Totals - # Boats	Report the number of salmon boats sampled.	Example: 3 = three salmon boats were sampled
Page Totals - # Anglers	Report the number of salmon anglers sampled.	Example: 35 = thirty- five salmon anglers were sampled
Page Totals - # Kings Kept	Report the number of King Salmon kept.	Example: 20 = twenty King Salmon were kept
Page Totals - # Kings Rels	Report the number of King Salmon released.	Example: 9 = nine King Salmon were released
Page Totals - # Coho Kept	Report the number of Coho Salmon kept.	Example: 0 = no Coho salmon were kept
Page Totals - # Coho Rels	Report the number of Coho Salmon released.	Example: 4 = four Coho Salmon were released
Page Totals - SL Take	Report the number of salmon taken by pinnipeds.	Example: 5 = five salmon were taken by pinnipeds
Page Totals # ad- clips	Report the number of ad- clipped salmon sampled.	Example: 7 = seven salmon were adipose fin-clipped

Field Name	Instructions	Coding Examples and Formats
Page Totals-# sal heads	Report the number of salmon heads recovered.	Example: 6 = six salmon heads were recovered
Page Totals - # NRS	Report the number of salmon heads that were non-recovered species.	Example: 1 = one head was not recovered and received an "NRS" status

Specific Editing Checks

- Make sure each boat that is sampled has the boat name and number recorded on the form.
- Try to use headtag numbers in order. If not possible, use the margin of the form to note tags were used out of sequence.
- 3. Double check that the headtag number given to each fish matches the length measurement for that fish.
- 4. Do not assume you are pulling your headtags out in order; look at the number on the headtag as you attach it to a salmon head.
- 5. Be sure to check the "NRS" box if a salmon head is not recovered.
- 6. Always circle a gear: Mooch, Troll, or Mooch AND Troll.
- 7. Please report who worked with you and if they have data or not.
- 8. Make sure your writing is legible.
- 9. Clearly delineate which headtags came from which boat.
- At the end of the sample day, inventory your heads. Make sure the heads you collected match the headtags on your data sheet and on your headtag report form.
- 11. Check your data sheets for duplicate headtag numbers, missing headtag numbers and non-sequential headtag numbers.

CRFS-OSP SALMON CPFV Dockside Form Example

Date (MWDDYY)	os	P Port	Samp	oler ID	Sa	mpler L	ast Nan	Other Samp	iers, ID (w/d	
07/21/15	E	UR	312	2	Ti	roxel				(Y N) (Y N)
EFFORT	_				CATCH			BIO	DATA	
Boat Name Boat #	Gear		Ki	ng		ho	Sea Lion Take		lmon	
time sampled	(circle)	Total Angs	# Kept	# Rels	# Kept	# Rels	# Salm	Headtag#	FL(mm)	NRS*
Reel Steel	Ð	6	4.0			2		99101	711	
71226 110	M M	0	12	3	0	2	0	99102	691	
at Name.	Т							99103	595	
nat# In	M				out of	seque	ence	99105	702	
at Name	Т							99106	688	
oat# In	M as									
Fishy Business	Ŧ	,			_			99104	687	
71284 112	M M	6	12	4	0	4	1	99107	697	
at Name	т							99108	750	X
oat# <u>Tin</u>	M M							accidentally dro	pped into	bay
Sea Weasel II	G			2				99109	722	
70586 113	9 M	6	12	3	0	5	0	99110	699	
Silver Star	G	_						99111	787	
70786 11 ^{Td}	В м	5	9	3	0	2	0	99112	715	
oat Name	Т							99113	692	
eat# <u>Tir</u>	M M							99114	748	
oat Name	т									
oat# In	M									
at Name	т									
ostë In	M as									
omments: Reel Steel head he's going to be out of t trip.	led back the wate	out for t	or PM tr	ip. For l couple	Reel II days. S	did per hellbac	sonal t k still	rip. Gary on Sea out, doing SALCk	Weasel s (/HALPA	ays combo
OTAL SAMPLED EFFORT & CATFO	H FOR THE	DAY:	Ki	ng	Co	ho				
4	2	3	45	13	0	13	1	14	13	1
#Boats NRS (non-recovered species): check		nglers	# Kept	#Rels	# Kept		SL Take		# Sal Heads	# NRS

PC EFFORT CHECKS

Need and Purpose

The PC effort checks (PECs) are needed to produce monthly estimates of PC catch and effort. The purpose of the PEC is to determine whether a PC boat fished on a particular day. These data will be used to validate log information submitted by each CPFV. PEC data is used along with the logs to estimate fishing effort for the PC mode in California. All PC owner/operators in California are required by law to submit to the Department an activity record, or log, for each fishing trip. However, compliance is less than 100 percent for the fleet overall and not all logs are submitted on time. CRFS uses the PEC to verify fishing trips and estimate the fraction of CPFV logs submitted by the time we make the monthly estimates. In addition, during salmon season the PEC data are used in conjunction with CPFV logbooks to determine total salmon fishing trips, and to ensure achievement of the minimum 20% sample rate.

The PEC data are as important as dockside and onboard PC sampling; all CPFV sample mode data is used together to estimate total effort and catch.

Methods

There are two methods for conducting PC effort checks. The method employed is dependent on the District and whether or not the ocean salmon fishery is open. During salmon season, the sampling responsibility for PECs will vary between CRFS and OSP depending on port logistics and staffing availability. Where CRFS is conducting PECs, the Lead will task a Sampler with PEC sampling.

1) During salmon season in Districts 2-6 activity for every PC boat should be recorded for every day. During this time, a Sampler may be designated in each port to collect effort information for all PC vessels in their designated port (i.e. the Port Lead Sampler). The best way to get an accurate number of salmon trips is to contact the vessel owners directly, but the bait shops/landings that book the trips may also have this information. Confirm information gathered from the bait shop/landing with the CPFV captain whenever possible. The Port Lead Sampler must keep track of all PC effort to maintain the required minimum salmon PC sampling rate of 20 percent per half-month period. The Port Lead Sampler will conduct most of the salmon PC dockside assignments. The CRFS-OSP PC (CPFV) Effort Check Form should be filled out by the Port Lead Sampler every week. Get info on these vessels whenever you are at the docks. This is a good time to collect effort information from previous days when effort was not checked. It's advised that effort is checked at least three times a week to adequately capture all PC effort. Sometimes boat operators are difficult to contact if they are not present at the dock during sampling or they do not immediately return your call. Do not wait until the end of the week to collect effort information. If you wait longer than a few days, you may not be able to collect all of the effort information needed. The Port Lead Sampler should also look for transient boats that may use a slip for a short period or use the

launch ramps in the port (e.g., trailered 6-packs). Samplers conducting PR assignments should collect effort information as they encounter these boats at the PR site. If you are not the Port Lead Sampler, please report all sampled or otherwise known PC effort in the body of the weekly email to the Lead.

2) For District 1 and when salmon season is closed in District 2, PECs are scheduled based the minimum sample number needed per District. In Districts 3-6 when salmon season is closed, Leads will assign Samplers as Port Leads and determine the sampling rate. Use the PEC form for your District's sub region (either Cen/Nor Cal PEC Form or So Cal PEC Form). Within each District, a sufficient number of confirmed PC trips are needed to compare with the logs for each landing with an active PC. If it has been confirmed that all PCs at a landing or in a District have stopped fishing, then no PECs are needed at that landing or in that District. However, the landing/District should still be monitored to confirm no trips are taking place.

Type of Assignments

In Districts 1 & 2 a PEC assignment <u>must be scheduled</u> as a part of each PC onboard or dockside assignment. The Sampler uses the *CRFS PC (CPFV) Effort Check Form* (Southern California version) to gather information about the daily activities of all (or as many as possible) of the PC boats that use that landing.

Additional PC Assignments

In Districts 1 & 2 additional assignments that are needed to meet the minimum number of PECs per landing may be scheduled by the Lead in the following ways:

- Specific PC Effort Check Assignments: Samplers are given an assignment to check on the activity at a number of PC landings. Checks can be conducted either by going to the landing or calling the landing. Specific PEC assignments are scheduled separately from other sampling assignments. The Sampler uses the CRFS-OSP PC (CPFV) Effort Check Form (Southern California version).
- PC Effort Check Assignments as Part of a Non-PC
 Assignment: Samplers are assigned to go to a specific PC landing in conjunction with an MM, BB, PC, PR2 or PR1 assignment. The Sampler uses the CRFS PC (CPFV) Effort Check Form (Southern California version).

The Lead will show all PC effort check assignments (specific PEC assignments as well as those done in combination with a PC onboard/dockside, BB, MM or PR assignment) on the Monthly Schedule. PECs do not have an assignment ID. However, time spent doing PECs can be included in the "edit" hours on an ASF if the PEC is done in conjunction with a CRFS assignment. PECs should also be recorded on the Weekly Report.

CRFS-OSP PC (CPFV) Effort Check Form Item by Item Instructions

Field Name	Instructions	Coding Examples and
		Formats
Page _ of _	Fill in the consecutive number of pages used to document all the PC boats that use that landing.	Page 1 of 1
OSP Port	Fill in (or make sure you are using the correct pre-filled form) for the landing you are documenting.	AVI = Avila
Sampler ID	Enter your 3-digit Sampler Identification number.	Example: 207 = DaSilva
Sampler Last Name	Write out your last name.	"DaSilva"
CNTY	Fill in (or make sure you are using the correct pre-filled form) for the 3-digit numeric county code.	079= San Luis Obispo County
SITE	Enter the 3-digit numeric site code, unless the form has been prefilled for you.	101 = Patriot's Landing
Site Name/CDFW Port	Enter (or make sure you are using the correct pre-filled form) for the port name and CDFW port number.	Avila (602)
Week Starting Mon.	Enter the date (MM/DD/YY) of the Monday starting the sample week.	10/21/17 = October 21, 2017
ASSN ID	Enter the 6-digit assignment ID number if the PEC is in conjunction with a BB/MM/PR/PC assignment. Leave blank otherwise.	Assignment ID in the format MMDNNN where MM is the month ranging from 01-12, D is the CRFS District from 1 to 6, N is the mode and NN is the sequence number from 01 to 99.
Date	Enter the MM/DD of the date that corresponds to Monday, Tuesday, etc. for the sample week	10/21 = MON 10/22 = TUES, etc.
CPFV Boat Name	Enter the boat name	'Patriot'

Field Name	Instructions	Coding Examples and Formats
CDFW Boat #	This is the 5-digit Fish and Wildlife vessel ID number (permit number).	02214 = 'Patriot'
Target or Status	Determine the target (fishing) or status (nonfishing) from the list of codes at the bottom of the form and record it for each boat. A maximum of two fishing targets can be recorded. If a boat targets salmon and rockfish for example, record "SR". If the boat was targeting salmon, circle T for troll or M for mooch. If both types of gear were used circle both. Circle D if the trip was a spearfishing dive trip.	Fishing Target: A = Pacific Halibut S = Salmon R = Rockfish L = Lingcod Z = Striped Bass T = Tuna N = Sturgeon H = CA Halibut K = Shark O = Other D = Crab Non-Fishing Status: 1 = boat docked (trailered) 2 = non-fishing trip 3 = non-CPFV fishing trip Gear: T = Troll, salmon
		M = Mooch, salmon D = Dive
Source	Record your source for the information you recorded.	Initials = record the Sampler's initials who sampled the boat P = Personal Observation C = Captain/deckhand O = Office contact W = Website
Total salmon CPFVs sampled per day	Record the number of salmon trips sampled out of the total salmon trips by day. This will make it easier to tally the weekly totals at the bottom of the form.	Example: "1/2"
Notes:	Each landing or port has a list of PC boats that the Samplers should lookout for. Notes about those boats should go	Example: "While you are checking CPFV activity, check for these boats: BBQ (7404), Liberty (11635), RG

Field Name	Instructions	Coding Examples and Formats
	here. This info is already listed on the pre-filled form. Use additional space as needed to record relocations of CPFVs.	Spot 2 (5392)" "Look out for any 'new' trailered 6-packs."
Comments	Provide any necessary comments.	Example: "Patriot is expected to be in dry dock for 1 month"
Total Salmon CPFVs	Report the total number of salmon trips for the sample week.	10 = ten trips where salmon were targeted or kept
Salmon CPFVs Sampled	Report the number of salmon trips sampled by CRFS and/or OSP for the sample week.	5 = five salmon- targeting trips were sampled
% Salmon CPFVs Sampled	Report the percentage of salmon trips sampled for the sample week.	50.0% = Fifty percent of the trips were sampled

Note: Districts 2-6, use additional rows on the PEC form as needed to document when boats do additional trips in the same day or when transient CPEV effort is documented at the site

Specific Editing Checks

- Make sure the ASSN ID is filled out if the PEC was done in conjunction with another CRFS assignment. Leave blank otherwise.
- 2. If a new boat started using a particular landing, write in the new boat's name, number and the daily trip information. Just because the boat is not listed on the form does not mean that we should n't be tracking it. D1 PECs are not pre-filled with boat names and numbers. Boat numbers can be found in prominent lettering on the wheelhouse, or on the master vessel list for your District provided by your Lead.
- Make sure to fill in the boat numbers. We track the vessels' activities by their boat numbers; so make sure that information is always filled in.
- 4. Multiple days' worth of information can go on one PEC form provided those days fall within the same sample week.
- 5. Up to two targets may be reported for each trip.

- 6. All salmon trips need to have a gear type circled T (troll), M (mooch) or circle both if both gear types are used on the same trip.
- 7. Make sure that when striped bass is recorded as a target that your writing is clear and legible as a "Z" and not a "2".
- 8. Do not remove boats that leave the port to fish elsewhere notify your Lead. Include a note in the comments section about any boats that have moved or are fishing in another port.

PEC Form Example - Non Salmon

Sampler #	Sa	mple	r La	st Nai	ne			CNT	Υ		SITE	Ε	Site	nam	e (C	DFW	port)	Wee	k sta	rting l	Mon
344		В	ond					15			301	1	Inne	r Bo	at Ba	asin	(201)	(06/0)5/1	7
	ASSN ID	0	660	05													(066	07			
Da	te (MM/DD)	0	6/0)5	(06/0	6		06/0	7		06/0	9	(06/1	0	(06/1	1	(06/1	2
		١	NOI	V	0.000,900	TUE			WED)		THU	J		FRI			SAT			SUN	l
CPFV Boat Name	CDFW Boat #	Tar Sta		So.		rget atus	So.		rget atus	So.		rget atus	So.		rget itus	So.		rget atus	So.		rget itus	S
	Dout #	Gea	ır (ci	rcle)	Ge	ar (cir	cle)	Ge	ar (cir	cle)	Ge	ar (ci	cle)	Ge	ar (cir	cle)	Ge	ar (cir	cle)	Ge	ar (cir	cle)
Tally Ho II	21352	R	L	DT	F	RL	C		1	0		1	0	F	RL	С	RI	D	С	R	LD	(
		Т	М	D	Т	М	D	Т	М	D	Т	М	D	Т	М	D	Т	М	D	Т	М	
Dancing Hooker	70199	RI	D	C		3	C	R	LD	С	100075	1	С		1	С	F	RL	ТВ	F	RL	C
		Т	М	D	Т	М	D	Т	М	D	Т	М	D	Т	М	D	Т	М	D	Т	М	D
Jolly Dolly	71430	3	3	С		1	С		1	С	F	RL	W	F	RL	W	R	RL.	ТВ	7	2	٧
		Т	М	D	Т	М	D	Т	М	D	Т	М	D	Т	М	D	Т	М	D	Т	М	C
Shannon L' Ree	71614	R	L	DT	RI	LD	C	R	LD	С		3	С		1	С	R	RL	С	R	L	C
		Т	М	D	Т	М	D	Т	М	D	Т	M	D	Т	М	D	Т	М	D	Т	М	C
		т			T			т			т					L						L
			М	D		М	D		М	D	<u> </u>	М	D	Т	М	D	Т	М	D	Т	М	
		T	M	D	Т	М	D	т	М	D	Т	M	D	т	М	D	Т	М	D	T	М	
		T	М	D	Т	М	D	Т	М	D	Т	М	D	Т	М	D	Т	М	D	Т	М	0
		т	М	D	T	М	D	т	М	D	т	M	D	т	М	D	Т	М	D	т	M	
		T	М	D	Т	М	D	Т	М	D	Т	M	D	T	М	D	T	М	D	T	М	D
		T	M	D	т	M	D	T	М	D	Т	M	D	T	M	D	т	M	D	т	M	
Total salmon CPFVs sampled per day:			0/0)		0 / 0			0,0			0 / 0)		0,0	y i		0,0			0,0	
Notes:												-										
Comments:																						
															0 al Saln			O non CF			0 Salmo	
Fishing Target		T - T							hing						1		s	non CF ample		CPF	Salm	

Fishing Target

S = Salmon (circle Troll or Mooch)

R = Rockfish L = Lingcod

Z = Striped bass N = Sturgeon

K = Shark

T = Tuna

O = Other H = CA Halibut

A = PA Halibut

Note: Record the fishing target and circle gear D for dive trips; Record non-take dive trips (e.g., wildlife viewing) as 2.

1 = boat docked (trailered)

3 = non-CPFV fishing trip

2 = non-fishing trip

Initials = sampled by CDFW P = Personal observation C = Captain / deckhand

O = Office contact W = Website

20151112

303	1	A C II				23 307 T						Week starting Mor			
		Nalken	hau	er		23		307	Trin	idad (23	1)		0	7/02/1	7
	ASSN ID	0766	02					0766	604						
		07/	12	07/	03	07/0	14	07/	05	07/0	16	07/0	07	07/0	na.
Dat	e (MM/DD)													 	
		MO	N .	TU	=	WEI	,	TH	U	FR	1	SAT	_	SUI	4
CPFV Boat Name	CDFW Boat #	Target Status	So.	Target Status	So.	Target Status	So.	Target Status	So.	Target Status	So.	Target Status	So.	Target Status	Sc
	Dout #	Gear (ci	rcle)	Gear (c	ircle)	Gear (cir	rcle)	Gear (c	ircle)	Gear (ci	rcle)	Gear (ci	rcle)	Gear (ci	rcle)
Jumpin Jack	35910	RL	SW	RL	Р	RL	С	1	P	RL	С	RL	С	RL	C
		T M	D	T M	D	т м	D	T M	D	T M	D	т м	D	т м	D
Toni Rae II	42146	1	P	S	MS	RL	С	S	ER	RL	С	Α	С	3	C
		т м	D	M	D	т м	D	O M	D	T M	D	T M	D	т м	D
Wind Rose	46519	SA	P	S	MS	3	С	1	P	S	MS	1	С	R	C
		(D) M	D	O M	D	т м	D	Т М	D	(D) M	D	т м	D	т м	D
White Cadillac	71623	SA	P	RL	P	L	C	1	P	1	С	S	С	S	С
		O M	D	T M	D	т м	D	т м	D	T M	D	O M	D	O M	D
Reel Therapy II	71654	S	P	1	Р	S	С	S	ER	S	MS	RL	С	RL	C
Walter Company		O M	D	T M	D	T M	D	T 🐠	D	T 🐠	D	T M	D	T M	D
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		I M	D.	1 M	i b	T M	D	T M	D	T M	D	T M	D	T M	D
Total salmon CPFVs		T M	D	T M	D	T M	D	т м	D	T M	D	T M	D	ТМ	D
sampled per day:		0/	3	2/	2	0 / 1		2/	2	2/	2	0 / 1		0 /1	l.
Notes:					75 75										_
Comments: Walken	hallara		Live	onin la	als as	ah a and	(7/00 T	1						not be only
Toni Ra										12		6		5	0%
Tonicka	e ii ariu	vviilu r	1056	LOOK II	OII-C	rrv ui	ps t	ills wee	K.						
										Total Sali CPFV		Salmon Cl sample		% Salm CPFVs sar	
Fishing Target					Nor	-Fishing	Sta	tus				ort Source			приос
S = Salmon		T = Tun	а			boat doc			1)					by CDF\	N
(circle Troll or Mood		O = Oth				non-fishi						Persona			
R = Rockfish		H = CA		ut		non-CPF						Captain			
L = Lingcod		A = PA I										Office c			
Z = Striped bass		D = Cral		700							//	= Website		2070)	
N = Sturgeon			-	ne fishina	tarnet	and circle	nea	r D for div	a trina	Pecord		3.0010			

Site Effort Checks

Need and Purpose

A Site Effort Check (SEC) is an instantaneous count of finfish anglers/trailers at a specific MM, BB, or PR site. SECs allow for the detection of changes in effort at sites made inactive based on historic data and may detect new fishing sites that might have otherwise been overlooked. The SEC counts are used to update baseline site effort data for weighted probability sampling, to determine if the effort level warrants the site being surveyed and to determine the proportionate level of effort for under-coverage adjustments.

Scheduling

SECs may be scheduled as routes or conducted, when possible, in conjunction with any CRFS assignment. Samplers are provided with a list of sites in a set SEC route, as well as a list of sites where adjacent SEC counts should be obtained. SEC routes are scheduled monthly. All sites in the route should be sampled for SEC counts.

Adjacent SEC counts are opportunistic and include counting anglers or trailers in a different mode at the same site being sampled or counting anglers or trailers at a nearby site. Adjacent SEC counts should only be conducted at sites or in modes that are easily accessible and close to the site being sampled and only performed as time allows. Do not miss CRFS interviews in the assigned mode to get adjacent SEC counts. Additionally, adjacent counts should not be prompted by changes in effort. Examples of adjacent SECs include a BB section next to an MM site, or an MM structure next to a PR launch ramp.

Methods

There are two methods for conducting SECs. The method employed is dependent whether an SEC route has been assigned as a standalone assignment or if adjacent counts are being obtained in conjunction with a CRFS assignment.

1) SEC Routes: MM, BB, and PR sites in a District are grouped into routes based on location and a Sampler's ability to visit and count anglers or trailers in a typical workday. Leads will provide a list of sites for each route and the Sampler uses the CRFS Wiki or District site list to visit all sites to get accurate counts of anglers or

trailers. For BB sites, this includes visiting all access points of a BB site. Start times will be determined by the Lead using local knowledge and available daylight hours

To promote sampling efficiency, the direction routes are sampled will be determined by randomly selecting between two predetermined starting points and generally proceed in a northerly or southerly direction. Start times will be classified as either late or early and will be randomly selected with equal probability. A simple random selection from all days in a month will be surveyed for each route or may be stratified by KOD. The number of routes and samples will vary by District.

2) Adjacent SEC: These counts should be obtained, when possible, in conjunction with other CRFS assignments (MM, BB, PCO, PCD and PR). Leads will provide a list of sites where adjacent counts are feasible that will not drastically increase sampling time of the assignment. Obtaining these counts should not cause the Sampler to incur much additional sampling time or mileage, nor should they be prompted by changes in effort.

Sampling SEC

For each site visit all finfish anglers or trailers to be counted. SEC counts are considered instantaneous and should be conducted as quickly as possible. For BB and MM modes, an angler is defined as a person actively fishing, taking a break from fishing, or having the intent to fish during the survey day. Only finfish anglers are to be counted, the Sampler will use their expert local knowledge and discretion to determine target. Canvassing anglers for target should not be performed unless it can be done with minimal effort or time.

The BB section is further divided to include kayak and PWC vessels that are fishing. This section includes PR vessels that have accessed the ocean or bay from the BB section of the site being sampled.

For "beach and bank launched PR" at BB sites, binoculars should be used to count fishing <u>kayaks and/or PWCs vessels</u> and make a judgment call as to which site those boats launched from for inclusion in the BB observed PR count. Remember that fishing kayaks can be launched from almost any shore site and their SEC count goes in the "BB OBS" section of the ASF. Please note, the PR section is to be used for traditional boats at established PR

sites, since CRFS protocols do not count kayak trailers or car tops as a 'boat'.

At PR sites, trailer counts will be used for fishing effort. Count any trailer that could potentially be recreationally fishing. As with all CRFS trailer counts, do NOT count PWC trailers, car top boat carriers, boats loaded into the beds of trucks, non-traditional inflatable boats, trailers not attached to a vehicle, or known non-fishing boats. If you observe a non-traditional fishing vessel (e.g., PWC or kayak) launching or believed to be launched from the PR site, record them in the "BB OBS PR". For the San Francisco District, consult with the D4 lead.

If you cannot accurately obtain a count for an entire site due to fog or access points that were not visited, leave the SEC box blank. Provide a note in the site comment box if fog or another factor prevented you from conducting the SEC.

Specific Editing Checks

- Make sure counts are obtained for all applicable adjacent modes. If a mode does not have possible adjacent counts, the Site Effort Check box should be left blank.
- 2. Contact the Lead if a new site(s) is discovered or a site has effort in a mode not currently being sampled.
- 3. When conducting SEC routes, all sites should be visited during daylight hours. If an entire route cannot be completed, contact the Lead and they will determine how to obtain counts at the remaining unchecked sites.
- 4. Make sure the site disposition for any SEC site is (0) zero.
- Pay special attention to the hours onsite for each SEC count. This is important for data-entry.

SEC Route Example- ASF Form

	R ON/BY:	UPLD ON/BY: _					ILE ON/BY:							
Acen #			ASSIGNA						V15 1/	3/2017				
	SAMPLER NAME:	CAMPI ED #	DATE (MA)	DD000	40	CHID	HODE	DO	DT/ 0/			IDO		
1	Ryan Denton	SAMPLER #	06/03/1		AS	SSN ID	MODE		OS		HOL 8.		тоти	ΔΙ
\dashv	OTHER SAMPLER(S): NAME & # (w/data)	OTHER SAMI			//data)		1	_	N DIS	-	-	75	SAM	
	,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,		3.2/					SALT COM	Car					
	(Y/N) circle MARINE CONDITIONS: Sunny, clear. Calm		n air tamn		(Y/N) circle on	e PV	Pers	Vehic	le	1.	.25	TOT	
	Low tide @ 1305.		n air temp	s.			10500	ODO START		रा	.7	75	TRV HQ to	o site
	EFFORT: Moderate effort for a summer	weekend.					10580	10580 ODO				.5	TRV last s	TIME site to
ENTS	CATCH: HALCA, SPFAM. Many anglers	reported low	atch toda	y.			80	MILE	AGE		(0	EDIT	
OMM	PC ACTIVITY: PPT had 1/2 and 3/4 day b	oats out. LBM	had 1/2 an	nd 3/4 b	oats o	out.				1	P	ound	all he	oure
8	OTHER PERTINENT INFORMATION:												st 0.2	
											0.	.00hr	= 53-0)7 mi
													= 08-2	
													= 23-3 = 38-5	
	HEAD TAGS USED (PR/PC):			HE	AD TAC	GS USED	(MM/BB):				100	10.50010	2000	
Assignr	ment dispositions: 1=Complete, 2=Reassigned, 6=0	Canceled								- 3		Traile	er Cou	ınts
									te Effe Check		Start	Start	Stop	Stop
Assn # SAMPI Rya OTHER Rya PC AC OTHER PC									pplica		Onsite	Offsite (Onsite (Offsite (
Row#	SITE NAME / SITE COMMEN	Т	Francisco (10000000			TIME		ANGS	OBSPR	ō	6	ō	o
ŀ	Palos Verdes		CNTY	37		ARRV	0735	BB	18	0				
1			SITE	211		START		ММ						
Assn# 1 SAMI RY OTHE MARI EFFC CATC OTHE *Assignment of Row # Part Ab Cal Cor Cor Cor Cor Cor Cor Cor Co			DISPO**	0	_	STOP		PR		_				ĺ
	N-2 10 2210		HRS	1.0	2 2	DEPR	0836	PK	ANGS	ORSPR	_	_		\vdash
	Abalone Cove		CNTY	37	_	ARRV	0845	BB	0	0				ĺ
2	No anglers present		SITE	205	-	START		ММ						ĺ
_			DISPO**	0	1	STOP				\rightarrow				ĺ
OTHE MARINEFFO CATC CATC OTHE HEAD OTHE Assignment d Row # Pa 1 Ab. Roy Roy Roy Roy Roy Pricable Cable Pricable Cable Pricable Pric			HRS	.75		DEPR	0922	PR		OBSPR				
	Royal Palms		CNTY	37	_	ARRV	0926	BB	9	0				ĺ
3			SITE	206	5	START		ММ	\					ĺ
•			DISPO**	0		STOP		200000	_	\				
	3		HRS	.75		DEPR	1014	PR		\				L
	Cabrillo Beach		CNTY	37		ARRV	1018	ВВ	0	OBS PR				
	Counts obtained at MM and BB	1 12 1 17 2 2 1 10	SITE	110		START		ММ	1000	g and				
100	PR is currently PR1 site-no counts Cabrillo Pier very busy today	s obtained	DISPO**	0	0 STOP			50.0000	54	SANS I				
	, , , , , , , , , , , , , , , , , , , ,		HRS	.25	1	DEPR	1030	PR	N	/A				L
	Pier J/South Shores LR		CNTY	37	8	ARRV	1052	ВВ	36	OBS PR				
_			SITE	201	5	START				-				
O			DISPO**	0		STOP		MM	L.	\				
	3		HRS	.5		DEPR	1117	PR	3	9				
\neg			-		. 3		30	T			T		- 1	
- 1				ı I		ı I	- 1	- 1		1	1		1 1	í
Refusal	Total Boats Angs Kept Rels Kept	Dale	ept Rels	Kept	D.:	Kept	Date: 11	pt F	2-1	10	t B		0	OFF

	Shoreline Village	CNTY	37	ARRV	1126	ВВ	ANGS OBSPR	Т	Т	
6		SITE	202	START			4 0			
0		DISPO**	0	STOP		MM				
		HRS	.5	DEPR	1150	PR				
	Shoreline Village Piers 4,5	CNTY	37	ARRV	1128	BB	ANGS OBSPR			
7		SITE	221	START		ММ	5			
L		DISPO**	0	STOP			,			
		HRS	0	DEPR	1134	PR				
	Shoreline Village Piers 1,2,3	CNTY	37	ARRV	1144	ВВ	ANGS OBSPR			
8		SITE	220	START		ММ	14			
U		DISPO**	0	STOP						
		HRS	0	DEPR	1145	PR				
	Shoreline Village E Jetty	CNTY	37	ARRV	1159	ВВ	ANGS OBSPR			
9		SITE	216	START		ММ	10			
J		DISPO**	0	STOP		-	10			
		HRS	.25	DEPR	1220	PR				$oxed{oxed}$
	Cherry Beach	CNTY	37	ARRV	1222	BB	2 0			
10	Counts obtained for Cherry Beach	SITE	402	START		ММ	83			
10	and Belmont Pier	DISPO**	0	STOP			03			
		HRS	.75	DEPR	1302	PR				
	Alamitos Bay	CNTY	37	ARRV	1304	BB	3 0			
11	Counts obtained for Alamitos Bay	SITE	214	START		ММ	17			
1 1	and Alamitos Bay W Jetty	DISPO**	0	STOP			- 17			
		HRS	.5	DEPR	1340	PR				
	Seaport Village Jetty	CNTY	37	ARRV	1341	ВВ	ANGS OBSPR			
12		SITE	204	START		ММ	15			
14		DISPO**	0	STOP		100000	13			
		HRS	.25	DEPR	1352	PR				
	Marine Stadium	CNTY	37	ARRV	1400	ВВ	0 0			
13	Lots of boat traffic! Most likely cause for low effort	SITE	215	START		ММ				
10		DISPO**	0	STOP		-	_ \			
		HRS	.25	DEPR	1414	PR				
		CNTY		ARRV		ВВ	ANGS OBSPR			
14		SITE		START		ММ	17.00			
		DISPO**		STOP						
		HRS		DEPR		PR				
		CNTY		ARRV		BB	ANSS I OBS PR			
15		SITE		START		ММ				
.0		DISPO**		STOP		3				
		HRS		DEPR		PR				
		CNTY		ARRV		BB	ANGS OBSPR			
16		SITE		START		мм				
10		DISPO**		STOP		3				
		HRS		DEPR		PR				

SEC Adjacent Example

	R ON/BY:	UPLD ON/BY: _				FILE ON/BY:	9					
ssn#		CRFS	ASSIGNN	IENT SU	MMARY	FORM		V14 11/18/	2016			
2001	SAMPLER NAME:	SAMPLER#	DATE (MM/I	DD/YY)	ASSN ID	MODE	POF	RT/ CLUS	но	URS		
1	Ryan Denton	116	01/01/16	- 1	012345	ВВ		HS 5		3.0	TOTA	L
	OTHER SAMPLER(S): NAME & # (w/data)	OTHER SAM	PLER(S): NAM	ИЕ & # (w/da	ta)	1	ASSI	N DISP*	(6.5	SAME	PLI
	(Y/N) circle	000		(V	/ N) circle on	PV		Car or Vehicle	3.4	1.5	TOT	TR
	MARINE CONDITIONS: Sunny, clear. Cali		tide @ 1		/ IN) CITCLE OF	10000	100000	START	-	.5	TRV 1	
	EFFORT: Moderate effort.					10100	SCHEO!	STOP	-	1.0	HQ to	TIN
TS	CATCH: SPFAM, HALCA					100	MILE		\vdash	0	last si EDIT	te
COMMENTS	PC ACTIVITY: Ventura SF had no boat	ts out				100	IVIILL	AGE	-	-	lane ne	
8	OTHER PERTINENT INFORMATION: Adjacent SECs conducted at sites		95 and 402	2.					0	0.00hr 0.25hr 0.50hr	= 53-0 = 08-2 = 23-3 = 38-5	7 2
	HEAD TAGS USED (PR/PC):			HEAD	TAGS USED	(MM/BB):				7.7 OHII	- 00-0	-
ssign	ment dispositions: 1=Complete, 2=Reassigned, 6=C	Canceled					0	o Effort		Total Control	er Cou	_
ow#	OUT NAME (OUT COMMENT	.				TIME	(te Effort Check pplicable)	Onsite Start	Offsite Start	Onsite Stop	Official Chan
OW #	SITE NAME / SITE COMMENT	1	CNTY	111	ARRV	0715	BB	ANGS OBS	R			_
				7.7.7.7						1		
			SITE	22	START				-			
1			SITE DISPO**	7	START		ММ	- 1				
1			(500000)	20000	(Excellence)	0830	MM PR					
1	Solimar Beach		DISPO**	7	STOP	0830 0840		ANGS OBS	· R			
	Solimar Beach		DISPO**	7 1.25	STOP DEPR	0.0000000000000000000000000000000000000	PR BB	ANGS OBS	R			
2	Solimar Beach		DISPO** HRS CNTY	7 1.25 111	STOP DEPR ARRV	0.0000000000000000000000000000000000000	PR BB	ANGS OBS	88			
	Solimar Beach		DISPO** HRS CNTY SITE	7 1.25 111 209	STOP DEPR ARRV START	0.0000000000000000000000000000000000000	PR BB	AM3S ORS	R			
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16-6

SPECIES CODES

SPECIES CODES Sorted by Species Code		
SP CODE	COMMON NAME	SCIENTIFIC NAME
ABALO	abalone	Haliotis
ANCDB	anchovy, deepbody	Anchoa compressa
ANCFM	anchovy family	Engraulidae
ANCGN	anchovy genus	Anchoa spp.
ANCNO	anchovy, northern	Engraulis mordax
ARGNT	argentine, Pacific	Argentina sialis
BARPA	barracuda, Pacific	Sphyraena argentea
BFFFM	butterflyfish family	Chaetodontidae
BIVAL	bivalves	Bivalvia
BLKSJ	skipjack, black	Euthynnus lineatus
BLKSM	blacksmith	Chromis punctipinnis
BLNBY	blenny, bay	Hypsoblennius gentilis
BLNRP	blenny, rockpool	Hypsoblennius gilberti
BOGBY	goby, bay	Lepidogobius lepidus
BOGYL	goby, yellowfin	Acanthogobius flavimanus
BONEF	bonefish	Albula vulpes
BONPA	bonito, Pacific	Sarda chiliensis
BOTOM	bottomfish (groundfish)	Car da Crimoriolo
BOXSP	boxfish, spiny	Ostracion diaphanum
BULBR	bullhead, brown	Ictalurus nebulosus
BUTFM	butterfish family	Stromateidae
CARPC	carp, common	Cyprinus carpio
CASTG	smoothtongue, California	Leuroglossus stilbius
CATCN	catfish, channel	Ictalurus punctatus
CBFLS	combfish, longspine	Zaniolepis latipinnis
CBFSS	combfish, shortspine	Zaniolepis frenata
CLAMS	clams, unspecified	Bivalvia
CLMBK	clam, basket cockle	Clinocardium nuttallii
CLMGD	clam, geoduck	Panopea generosa
CLMGP	clam, gaper	Tresus nuttallii
CLMLM	clam, common littleneck	Protothaca staminea
CLMNR	clam, northern razor	Siliqua patula
CLMPO	clam, Pismo	Tivela stultorum
CLMWA	clam, common	Saxidomus nuttalli
	Washington	
CLNGN	clingfish, nothern	Gobiesox maeandricus
CODFM	cod family	Gadidae
CODPA	cod, Pacific	Gadus macrocephalus
CODTC	tomcod, Pacific	Microgadus proximus
COROM	corvina, orangemouth	Cynoscion xanthulus
CORSF	corvina, shortfin	Cynoscion parvipinnis
CRABS	crab tribe, true	Brachyuratribe
CRBCA	corbina, California	Menticirrhus undulatus
CRBBR	crab, brown rock	Cancer antennarius

Sorted by Species Code		
SP CODE	COMMON NAME	SCIENTIFIC NAME
CRBDG	crab, Dungeness	Metacarcinus magister
CRBGN	crab genus, cancer	Cancer
CRBGR	crab, graceful rock	Cancer gracilis
CRBPR	crab, pelagic red	Pleuroncodes palnipes
CRBRR	crab, red rock	Cancer productus
CRBSH	crab, sheep	Loxorhynchus grandis
CRBYR	crab, yellow rock	Cancer anthonyi
CRKBK	croaker, black	Cheilotrema saturnum
CRKSF	croaker, spotfin	Roncador stearnsi
CRKYF	croaker, yellowfin	Umbrina roncador
CROWT	croaker, white	Genyonemus lineatus
CRUST	crustaceans	Crustacea
CSHFM	shark family, cow	Hexanchidae
CSKFM	eel family, cusk	Ophidiidae
CTFPE	catalufa, popeye	Pristigenys serrula
CTSFM	shark family, cat	Scyliorhinidae
CUCUM	sea cucumbers	Holothuroidea
CUTLP	cutlassfish, Pacific	Trichiurus nitens
DABGN	sanddab genus	Citharichthys
DABLF	sanddab, longfin	Citharlchthys xanthostigma
DABPA	sanddab, Pacific	Citharichthys sordidus
DABSP	sanddab, speckled	Citharichthys stigmaeus
DAMFM	damselfish family	Pomacentridae
DRADO	dolphinfish	Coryphaena hippurus
DRGFM	dragonfish family	Stomiidae
DRMFM	drum family	Sciaenidae
DSSFM	smelt family, deepsea	Bathylagidae
EELOR	eel order	Anguilliformes
ELPFM	eelpout family	Zoarcidae
ERYPA	ray, Pacific electric	Torpedo californica
FLLFN	flounder family, lefteye	Bothidae
FLNFM	blenny family, combtooth	Blenniidae
FLRAR	flounder, arrowtooth	Atheresthes stomias
FLRFM	flounder family, righteye	Pleuronectidae
FLRKM	flounder, Kamchatka	Atheresthes evermanni
FLRST	flounder, starry	Platichthys stellatus
FLTOR	flatfish order	Pleuronectiformes
FLYCA	flyingfish, California	Cypselurus californicus
FLYFM	flyingfish family	Exocoetidae
FRSFM	shark family, frill	Chlamydoselachidae
FTRIG	triggerfish, finescale	Balistes polylepis
GAPOD	sea slug, sea snail	Gastropoda
GARIB	garibaldi	Hypsypops rubicundus
GNTFM	gruntfamily	Haemulidae
GNTSB	seabass, giant	Stereolepis gigas

Sorted by Species Code		
SP CODE	COMMON NAME	SCIENTIFIC NAME
GOBAR	goby, arrow	Clevelandia ios
GOBBE	goby, blackeye	Coryphopterus nicholsi
GOBFM	goby family	Gobiidae
GRNFM	greenlingfamily	Hexagrammidae
GRNGN	greenlinggenus	Hexagrammos
GRNKP	greenling, kelp	Hexagrammos
		decagrammus
GRNMA	greenling, masked	Hexagrammos
		octogrammus
GRNPT	greenling, painted	Oxylebius pictus
GRNRK	greenling, rock	Hexagrammos
		lagocephalus
GRNWT	greenling, whitespotted	Hexagrammos stelleri
GRPBT	grouper, broomtail	Mycteroperca xenarcha
GRPGC	gulfconey	Hyporthodus acanthistius
GRPGF	grouper, gulf	Mycteroperca jordani
GRPGN	grouper,genus	Epinephelus
GRPSC	cabrilla, spotted	Epinephelus analogus
GRPSS	Grouper, star-studded	Hyporthodus niphobles
GRUCA	grunion, California	Leuresthes tenuis
GUIBD	guitarfish, banded	Zapteryx exasperata
GUIFM	guitarfish family	Rhinobatidae
GUISN	guitarfish, shovelnose	Rhinobatos productus
GUNCR	gunnel, crescent	Pholis laeta
GUNFM	gunnel family	Pholidae
GUNPP	gunnel, penpoint	Apodichthys flavidus
GUNSB	gunnel, saddleback	Pholis ornata
HAGBK	hagfish, black	Eptatretus deani
HAGFM	hagfish order	Myxinidae
HAGPA	hagfish, Pacific	Eptatretus stouti
HALCA	halibut, California	Paralichthys californicus
HALFM	halfmoon	Medialuna californiensis
HALGL	halibut, Greenland	Reinhardtius
		hippoglossoides
HALPA	halibut, Pacific	Hippoglossus stenolepis
HERFM	herring family	Clupeidae
HERPA	herring, Pacific	Clupea pallasi
HERRD	herring, round	Etrumeus teres
JACFM	jack family	Carangidae
JACMK	mackerel, jack	Trachurus symmetricus
KAWAK	kawakawa	Euthynnus affinis
KLFCA	killifish, California	Fundalus parvipinnis
KLPCR	kelpfish, crevice	Gibbonsia montereyensis
KLPFM	clinid family	Clinidae
KLPGT	kelpfish, giant	Heterostichus rostratus

Sorted by Species Code		
SP CODE	COMMON NAME	SCIENTIFIC NAME
KLPOF	fringehead, onespot	Neoclinus urinotatus
KLPRB	blenny, reef	Paraclinus integripinnis
KLPSF	fringehead, sarcastic	Neoclinus blanchardi
KLPSP	kelpfish, spotted	Gibbonsia elegans
KLPST	kelpfish, striped	Gibbonsia metzi
KOSAL	king-of-the-salmon	Trachipterus altivelis
LANLN	lancetfish, longnose	Alepisaurus ferox
LJMUD	mudsucker, longjaw	Gillichthus mirabilis
LMPAR	lamprey, Arctic	Lampetra japonica
LMPFM	lamprey family	Petromyzontidae
LMPPA	lamprey, Pacific	Entosphenus tridentatus
LNGCD	lingcod	Ophiodon elongatus
LOBSP	lobster, spiny	Panulirus interruptus
LUVAR	louvar	Luvarus imperialis
LZDCA	lizardfish, California	Synodus lunioceps
LZDFM	lizardfish family	Synodontidae
MACBL	mackerel, bullet	Auxis rochei
MACFM	mackerel family	Scombridae
MACFR	mackerel, frigate	Auxis thazard
MACPA	mackerel, chub (Pacific)	Scomber japonicus
MANTA	manta	Manta birostris
MARBK	marlin, black	Makaira indica
MARBL	marlin, blue	Makaira nigricans
MARFM	billfish family	Istiophoridae
MARST	marlin, striped	Tetrapturus audax
MIDGN	midshipman genus	Porichthys
MIDPF	midshipman, plainfin	Porichthys notatus
MIDSP	midshipman, specklefin	Porichthys myriaster
MOJFM	mojarra family	Gerreidae
MOLLU	mollusks	Mollusca
MORAY	moray, California	Gymnothorax mordax
MSCAD	scad, Mexican	Decapterus scombrinus
NEDCA	needlefish, California	Strongylura exilis
ОСТОР	octopods	Octopoda
OCWHT	whitefish, ocean	Caulolatilus princeps
OPAHS	opah	Lampris guttatus
OPALE	opaleye	Girella nigricans
PERFM	perch family	Percidae
PERZB	perch, zebra	Hermosilla azurea
PHAKE	hake, Pacific	Merluccius productus
PILTF	pilotfish	Naucrates ductor
PIPEB	pipefish, bay	Syngnathus leptorhynchus
POLWE	pollock, walleye	Theragra chalcogramma
POMDO	dolphin, pompano	Coryphaena equisetis
POMFM	pomfret family	Bramidae

Sorted by Species Code		
SP CODE	COMMON NAME	SCIENTIFIC NAME
POMPA	pompano,Pacific (butterfish)	Peprilus simillimus
PRKBK	prickleback, black	Xiphister atropurpureus
PRKFM	prickleback family	Stichaeidae
PRKMK	prickleback, monkeyface	Cebidichthys violaceus
PRKRK	prickleback, rock	Xiphister mucosus
PRKSN	prickleback, snake	Lumpenus sagitta
PUFFM	puffer family	Tetraodontidae
QUEEN	queenfish	Seriphus politus
RAGFS	ragfish	Icosteus aenagmaticus
RAJOR	order, skate and ray	Rajiformes
RATFS	ratfish, spotted	Hydrolagus colliei
REMFM	remora family	Echeneidae
REMWS	whalesucker	Remora australis
RFAUR	rockfish, aurora	Sebastes aurora
RFBAY	rockfish, black and yellow	Sebastes chrysomelas
RFBKG	rockfish, blackgill	Sebastes melanostomus
RFBLK	rockfish, black	Sebastes melanops
RFBLU	rockfish, blue	Sebastes mystinus
RFBNK	rockfish, bank	Sebastes rufus
RFBOC	rockfish, (bocaccio)	Sebastes paucispinis
RFBRN	rockfish, brown	Sebastes auriculatus
RFBSP	rockfish, bronzespotted	Sebastes gilli
RFCAN	rockfish, canary	Sebastes pinniger
RFCHN	rockfish, China	Sebastes nebulosus
RFCLO	rockfish, calico	Sebastes dalli
RFCMA	rockfish, chameleon	Sebastes phillipsi
RFCOP	rockfish, copper	Sebastes caurinus
RFCOW	rockfish, (cowcod)	Sebastes levis
RFDBL	rockfish, darkblotched	Sebastes crameri
RFDUS	rockfish, dusky	Sebastes ciliatus
RFFLG	rockfish, flag	Sebastes rubrivinctus
RFFRK	rockfish, freckled	Sebastes lentiginosus
RFGBL	rockfish, greenblotched	Sebastes rosenblatti
RFGEN	rockfish genus	Sebastes
RFGOP	rockfish, gopher	Sebastes carnatus
RFGRN	rockfish, greenspotted	Sebastes chlorostictus
RFGRS	rockfish, grass	Sebastes rastrelliger
RFGST RFHBD	rockfish, greenstriped rockfish, halfbanded	Sebastes elongatus Sebastes semicinctus
RFHNC	rockfish, honeycomb	Sebastes semicinctus Sebastes umbrosus
RFKLP	rockfish, kelp	Sebastes umbrosus Sebastes atrovirens
RFLST	thornyhead, longspine	
RFMEX	rockfish, Mexican	Sebastolobus altivelis Sebastes macdonaldi
RFOLV	rockfish, olive	Sebastes macdonaidi Sebastes serranoides
KFULV	TOURIISH, UNVE	SENASTES SELFATIOIDES

Sorted by Species Code		
SP CODE	COMMON NAME	SCIENTIFIC NAME
RFPEP	rockfish, (chilipepper)	Sebastes goodei
RFPNK	rockfish, pink	Sebastes eos
RFPOP	perch, Pacific ocean	Sebastes alutus
RFPRS	rockfish, pinkrose	Sebastes simulator
RFPSD	rockfish, Puget Sound	Sebastes emphaeus
RFPYG	rockfish, pygmy	Sebastes wilsoni
RFQIL	rockfish, quillback	Sebastes maliger
RFRBD	rockfish, redbanded	Sebastes babcocki
RFRGH	rockfish, rougheye	Sebastes aleutianus
RFROS	rockfish, rosy	Sebastes rosaceus
RFRST	rockfish, redstripe	Sebastes proriger
RFRTN	rockfish, rosethom	Sebastes helvomaculatus
RFSCN	rockfish, sharpchin	Sebastes zacentrus
RFSDS	rockfish, swordspine	Sebastes ensifer
RFSHB	rockfish, shortbelly	Sebastes jordani
RFSLG	rockfish, silvergray	Sebastes brevispinis
RFSNS	rockfish, splitnose	Sebastes diploproa
RFSPK	rockfish, speckled	Sebastes ovalis
RFSQS	rockfish, squarespotted	Sebastes hopkinsi
RFSRK	rockfish, shortraker	Sebastes borealis
RFSST	thornyhead, shortspine	Sebastolobus alascanus
RFSTA	rockfish, starry	Sebastes constellatus
RFSTR	rockfish, stripetail	Sebastes saxicola
RFTIG	rockfish, tiger	Sebastes nigrocinctus
RFTRE	rockfish, (treefish)	Sebastes serriceps
RFVER	rockfish, vermilion	Sebastes miniatus
RFWID	rockfish, widow	Sebastes entomelas
RFWTB	rockfish, whitebelly	Sebastes vexillaris
RFYEY	rockfish, yelloweye	Sebastes ruberrimus
RFYMN	rockfish, yellowmouth	Sebastes reedi
RFYTL	rockfish, yellowtail	Sebastes flavidus
RNQBB	ronquil, bluebanded	Rathbunella hypoplecta
RNQFM	ronquil family	Bathymasteridae
RNQNO	ronquil, northern	Ronqilus jordani
ROCKH	rockhead	Bothragonus swani
RYBAT	ray, bat	Myliobatis californica
RYFLY	butterflyray, California	Gymnura marmorata
SABFM	sablefish family	Anoplopomatidae
SABLE	sablefish	Anoplopoma fimbria
SAILF	sailfish	Istiophorus platypterus
SALAC	trout, Arctic char	Salvelinus alpinus
SALAT	salmon, Atlantic	Salmo salar
SALCK	salmon, chinook	Oncorhynchus tshawytscha
SALCM	salmon, chum	Oncorhynchus keta
SALCO	salmon, coho	Oncorhynchus kisutch

Sorted by Species Code		
SP CODE	COMMON NAME	SCIENTIFIC NAME
SALCT	trout, cutthroat	Oncorhynchus clarki
SALDV	Varden, Dolly	Salvelinus malma
SALEM	salema	Xenistius californiensis
SALFM	salmon family	Salmonidae
SALGN	salmon genus	Oncorhynchus spp.
SALPK	salmon, pink	Oncorhynchus gorbuscha
SALRB	trout, rainbow	Oncorhynchus mykiss
SALSE	salmon, sockeye	Oncorhynchus nerka
SALTR	trouts, sea run	
SARGO	sargo	Anisotremus davidsoni
SARPA	sardine, Pacific	Sardinops sagax
SAUPA	saury, Pacific	Cololabis saira
SBBAR	sandbass, barred	Paralabrax nebulifer
SBFAM	bass family, sea	Serranidae
SBGEN	sandbass genus	Paralabrax
SBKLP	bass, kelp	Paralabrax clathratus
SBSPT	sandbass, spotted	Paralabrax maculatofascia
SBTHF	bass, threadfin	Pronotogrammus
		multifasciatus
SBWHT	seabass, white	Atractoscion nobilis
SCANT	sculpin, antlered	Enophrys diceraus
SCASH	sculpin, Arctic staghorn	Gymnocanthus tricuspis
SCBBS	chub, bluestriped	Sectator ocyurus
SCBFM	chub family, sea	Kyphosidae
SCBIL	lord, brown Irish	Hemilepidotus spinosus
SCBKF	sculpin, blackfin	Malacocottus kincaidi
SCBLD	sculpin, bald	Clinocottus recalvus
SCBNH	sculpin, bonehead	Artedius notospilotus
SCBRZ	scabbardfish, razorback	Assurger anzac
SCBUF	sculpin, buffalo	Enophrys bison
SCBUL	sculpin, bull	Enophrys taurina
SCCAB	cabezon	Scorpaenichthys
		marmoratus
SCCRG	sculpin, coastrange	Cottus aleuticus
SCDSK	sculpin, dusky	Icelinus burchani
SCFAM	sculpin family	Cottidae
SCGRT	sculpin, great	Myoxocephalus
		polyacanthocep
SCGRU	sculpin, grunt	Rhamphocottus richardsoni
SCILG	lord genus, Irish	Hemilepidotus
SCLST	sculpin, leister	Enophrys lucasi
SCNTH	sculpin, northern	Icelinus borealis
SCPAD	sculpin, padded	Artedius fenestralis
SCPRK	sculpin, prickly	Cottus asper
SCPRO	scallop, giant rock	Crassadoma gigantea

Sorted by Species Code		
SP CODE	COMMON NAME	SCIENTIFIC NAME
SCPSH	sculpin, Pacific staghorn	Leptocottus armatus
SCPUS	scallop, unspecified	Pectinidae
SCRCA	scorpionfish, California	Scorpaena guttata
SCRFM	scorpionfish family	Scorpaenidae
SCRIL	lord, red Irish	Hemilepidotus
		hemilepidotus
SCRRB	scorpionfish, rainbow	Scorpaenodes xyris
SCRSL	sculpin, rosylip	Ascelichthys rhodorus
SCSCL	sculpin, scaled	Archaulus biseriatus
SCSCT	sculpin, scissortail	Triglops forficata
SCSFN	sculpin, sailfin	Nautichthys oculofasciatus
SCSHN	sculpin, sharpnose	Clinocottus acuticeps
SCSLH	sculpin, scalyhead	Artedius harringtoni
SCSPT	sculpin, spotfin	Icelinus tenuis
SCTDP	sculpin, tidepool	Oligocottus maculosus
SCTRF	sculpin, threadfin	Icelinus filamentosus
SCWOL	sculpin, wolly	Clinocottus analis
SELFM	eel family, snake	Ophichthidae
SELYL	eel, yellow snake	Ophichthus zophochir
SENOR	senorita	Oxyjulis californica
SERLT	searobin, limptail	Prionotus stephanophrys
SGDIA	stingray, diamond	Dasyatis dipterura
SGFAM	stingray family	Dasyatidae
SGGEN	stingray genus	Dasyatis spp.
SGPEL	stingray, pelagic	Dasyatis violacea
SGRND	stingray, round	Urolophus halleri
SHADA	shad, American	Alosa sapidissima
SHANG	shark, Pacific angel	Squatina californica
SHBCS	shark, brown cat	Apristurus brunneus
SHBLU	shark, blue	Prionace glauca
SHBNH	shark, bonnethead	Sphyrna tiburo
SHBSM	smoothhound, brown	Mustelus henlei
SHBUL	shark, bull	Carcharhinus leucas
SHDFM	shark family, dogfish	Squalidae
SHDKY	shark, dusky	Carcharhinus obscurus
SHEEP	sheephead, California	Semicossyphus pulcher
SHFIN	shark, soupfin	Galeorhinus zyopterus
SHGSM	smoothhound, gray	Mustelus californicus
SHHRN	shark, horn	Heterodontus francisci
SHLEP	shark, leopard	Triakis semifasciata
SHMFM	shark family, mackerel	Lamnidae
SHNTH	shark, narrowtooth	Carcharhinus brachyurus
SHRFM	shark family, requiem	Carcharhinidae
SHRMP	shrimp	Caridea
SHSAL	shark, salmon	Lamna ditropis

Sorted by Species Code		
SP CODE	COMMON NAME	SCIENTIFIC NAME
SHSDG	shark, spiny dogfish	Squalus acanthias
SHSEV	shark, seven gill	Notorynchus maculatus
SHSGN	smooth hound genus	Mustelus
SHSIX	shark, six gill	Hexanchus griseus
SHSLP	shark, Pacific sleeper	Somniosus pacificus
SHSMK	shark, shortfin mako	Isurus oxyrinchus
SHSSM	smoothhound, sicklefin	Mustelus lunulatus
SHSWL	shark, swell	Cephaloscyllium ventriosum
SHTHR	shark, thresher	Alopias vulpinus
SHTIG	shark, tiger	Galeocerdo cuvieri
SHINS	Unidentified inshore sharks	
SHOFF	Unidentified offshore	
311011	sharks	
SHWHT	shark, white	Carcharodon carcharias
SKALT	skate, Aleutian	Bathyraja aleutica
SKBFM	stickleback family	Gasterosteidae
SKBGN	skipback genus	Euthynnus
SKBIG	skate, big	Raja binoculata
SKBTS	stickleback, threespine	Gasterosteus aculeatus
SKFAM	skate family	Rajidae
SKLGN	skate, longnose	Raja rhina
SKSTY	skate, starry	Raja stellulata
SKTCA	skate, California	Raja inornata
SMCAP	capelin	Mallotus villosus
SMEUL	eulachon	Thaleichthys pacificus
SMFAM	smelt family	Osmeridae
SMJAK	smelt, (jacksmelt)	Atherinopsis californiensis
SMLGF	smelt, longfin	Spirinchus thlaeichthys
SMNGT	smelt, night	Spirinchus starksi
SMSUR	smelt, surf	Hypomesus pretiosus
SMTOP	smelt, (topsmelt)	Atherinops affinis
SMWTB	smelt, whitebait	Allosmerus elongatus
SNDFM	sandfish family	Trichodontidae
SNDPA	sandfish, Pacific	Trichodon trichodon
SNFFM	sunfish family	Centrarchidae
SOLAF	flounder, Arctic	Pleuronectes glacialis
SOLBF	flounder, Bering	Hippoglossoides robustus
SOLBG	sole, bigmouth	Hippoglossina stomata
SOLBT	sole, butter	Isopsetta isolepis
SOLCF	sole, curlfin	Pleuronichthys decurrens
SOLCO	sole, C-O	Pleuronichthys coenosus
SOLDS	sole, deepsea	Embassichthys bathybius
SOLDT	turbot, diamond	Pleuronicthys guttulatus
SOLDV	sole, Dover	Microstomus pacificus

Sorted by Species Code		
SP CODE	COMMON NAME	SCIENTIFIC NAME
SOLEG	sole, English	Parophrys vetulus
SOLFH	sole, flathead	Hippoglossoides elassodon
SOLFT	sole, fantail	Xystreurys liolepis
SOLHT	turbot, hornyhead	Pleuronichthys verticalis
SOLPA	lance, Pacific sand	Ammodytes hexapterus
SOLPL	plaice, Alaska	Pleuronectes
		quadrituberculatus
SOLPT	sole, petrale	Eopsetta jordani
SOLRK	sole, rock	Lepidopsetta bilineatus
SOLRX	sole, rex	Glyptocephalus zachirus
SOLSD	sole, sand	Psettichthys melanostictus
SOLSL	sole, slender	Lyopsetta exilis
SOLST	turbot, spotted	Pleuronichthys ritteri
SOLYF	sole, yellowfin	Limanda aspera
SPBAR	surfperch, barred	Amphistichus argenteus
SPBLK	perch, black	Embiotoca jacksoni
SPCAL	surfperch, calico	Amphistichus koelzi
SPDPA	spadefish, Pacific	Chaetodipterus zonatus
SPDWF	perch, dwarf	Micrometrus minimus
SPFAM	surfperch family	Embiotocidae
SPKLP	perch, kelp	Brachyistius frenatus
SPPIL	perch, pile	Rhacochilus vacca
SPPNK	seaperch, pink	Zalembius rosaceus
SPRBW	seaperch, rainbow	Hypsurus caryi
SPREF	perch, reef	Micrometrus aurora
SPRTL	surfperch, redtail	Amphistichus rhodoterus
SPRUB	seaperch, rubberlip	Rhacochilus toxotes
SPSHN	seaperch, sharpnose	Phanerodon atripes
SPSHR	perch, shiner	Cymatogaster aggregata
SPSIL	surfperch, silver	Hyperprosopon ellipticum
SPSPF	surfperch, spotfin	Hyperprosopon anale
SPSTR	seaperch, striped	Embiotoca lateralis
SPWAL	surfperch, walleye	Hyperprosopon argenteum
SPWHT	seaperch, white	Phanerodon furcatus
SQTSE	squaretail, smalleye	Tetragonurus cuvieri
SQDJU	Humbolt/jumbo squid	Dosidicus gigas
SQDMK	market squid	Doryteuthis opalescens
SQTSE	squaretail, smalleye	Tetragonurus cuvieri
SQUID	squid	Cephalopoda
SRAGU	sierra, gulf	Scomberomorus concolor
SRAPA	sierra, Pacific	Scomberomorus sierra
SRDFS	swordfish	Xiphias gladius
SSTAR	sea star	Asterzoa
STBAS	bass, striped	Morone saxatilis
STGEN	sturgeon genus	Acipenser

Sorted by Species Code		
SP CODE	COMMON NAME	SCIENTIFIC NAME
STGRN	sturgeon, green	Acipenser medirostris
STMUL	mullet, striped	Mugil cephalus
STWHT	sturgeon, white	Acipenser transmontanus
SUNFM	mola family	Molidae
SUNOC	sunfish, ocean	Mola mola
SVRFM	silverside family	Atherinidae
TBESN	snout, tube	Aulorhynchus flavidus
TFPGE	tilefish, Pacific golden-	Caulolatilus affinis
	eyed	
THRBK	thornback	Platyrhinoidis triseriata
TNAAB	tuna, (albacore)	Thunnus alalunga
TNABE	tuna, bigeye	Thunnus obesus
TNABF	tuna, bluefin	Thunnus thynnus
TNASG	tunas (non-mackerel)	
TNASJ	tuna, skipjack	Euthynnus pelamis
TNASL	tuna, slender	Allothunnus fallai
TNAYF	tuna, yellowfin	Thunnus albacares
TNGCA	touguefish, California	Symphurus atricauda
TRTPA	tripletail, pacific	Lobotes pacificus
UNIFH	unidentified fish	
UNISF	unidentified (surface fish)	
URCHN	sea urchins	Diadematidae
WAHOO	wahoo	Acanthocybium solandri
WEKFS	weakfishes	Cynoscion
WOLFE	wolf-eel	Anarrhichthys ocellatus
WRABS	wrasse, blackspot	Decodon melasma
WRAFM	wrasse family	Labridae
WRARB	wrasse, rainbow	Thalassoma luvasanum
WRARK	wrasse, rock	Halichoeres semicinctus
YELTL	yellowtail	Seriola lalandi
18	shark, frill	Chlamydoselachus
		arguineus
22	shark, whale	Rhincodon typus
23	shark, ragged tooth	Odontaspis ferox
26	shark, basking	Cetorhinus maximus
29	shark, bigeye thresher	Alopias superciliosus
33	shark, longnose cat	Apristurus kampae
35	shark, filetail cat	Parmatyrus xaniurus
39	shark, Pacific sharpnose	Rhizoprionodon longurio
44	shark genus, gray	Carcharhinus
50	shark family,	Sphyrnidae
	hammerhead	
52	shark,smooth	Sphyrna zygaena
	hammerhead	
56	shark, prickly	Echinorhinus cookei

Sorted by Species Code		
SP CODE	COMMON NAME	SCIENTIFIC NAME
68	skate, sandpaper	Bathyraja interrupta
69	skate, black	Bathyraja trachura
70	skate, Alaska	Bathyraja parmifera
72	skate, flathead	Bathyraja rosispinis
74	skate, roughtail	Raja trachura
82	manta family	Mobulidae
84	mobula, spinetail	Mobula japanica
85	mobula, smoothtail	Mobula thurstoni
90	machete	Elops affinis
94	conger, Catalina	Gnathophis catalinensis
96	eel, Pacific worm	Myrophis vafer
97	eel, Pacific snake	Ophichthus triserialis
99	eel family, snipe	Nemichthyidae
100	eel, slender snake	Nemichthys scolopaceus
106	herring, middling thread	Opisthonema medirastre
107	herring, flatiron	Harengula thrissina
112	anchovy, slough	Anchoa delicatissima
113	anchoveta	Cetengraulis mysticetus
129	smelt, delta	Hypomesus transpacificus
131	smelt, rainbow	Osmerus mordax
139	spookfish family	Opisthoproctidae
140	barreleye	Macropinna microstoma
142	dragonfish, longfin	Tactostoma macropus
143	viperfish, Pacific	Chauliodus macouni
146	lancetfish family	Alepisauridae
148	daggertooth family	Anotopteridae
149	pearleye family	Scopelarchidae
150	pearleye, northern	Benthalbella dentata
151	lanternfish family	Myctophidae
152	lampfish, dogtooth	Ceratoscopelus townsendi
153	headlightfish, California	Diaphus theta
154	lampfish, pinpoint	Lampanyctus regalis
155	lampfish, patchwork	Notoscopelus resplendens
156	lampfish, northern	Stenobrachius leucopsarus
157	lanternfish, blue	Tarletonbeania crenularis
158	lampfish, diogenes	Diogenys lanternatus
159	flashlightfish	Protomyctophum crockeri
160	lampfish, Mexican	Triphoturus mexicanus
163	chihuil	Bagre panamensis
167	clingfish family	Gobiesocidae
169	clingfish, lined	Gobiesox eugrammus
170	clingfish, bearded	Gobiesox papillifer
171	clingfish, California	Gobiesox rhessondon
172	clingfish, kelp	Rimicola muscarum
173	clingfish, slender	Rimicola eigenmanni

Sorted by Species Code		
SP CODE	COMMON NAME	SCIENTIFIC NAME
174	frogfish, roughjaw	Antennarius avalonis
175	batfish, spotted	Zalieutes elater
176	seadevil, triplewart	Cryptopsaras couesi
183	brotula, red	Brosmophycis marginata
184	eel, spotted cusk	Chilara taylori
185	eel, basketweave cusk	Otophidium scrippsae
187	eelpout, bigfin	Lycodes cortezianus
188	eelpout, Alaska	Bothrocara pusillum
189	eelpout, pallid	Lycodapus mandibularis
190	eelpout, shortfin	Lycodes brevipes
191	eelpout, black	Lycodes diapterus
192	eelpout, wattled	Lycodes palearis
193	eelpout, Canadian	Lycodes polaris
194	eelpout, polar	Lycodes turneri
195	shulupaoluk	Lycodes jugoricus
196	eelpout, pale	Lycodes pallidus
197	eelpout, blackbelly	Lycodopsis pacifica
198	eelpout, bearded	Lyconema barbatum
201	halfbeak, longfin	Hemiramphus saltator
202	halfbeak	Hyporhamphus unifasciatus
203	halfbeak	Hyporhamphus rosae
204	halfbeak, ribbon	Euleptorhamphus viridis
205	flyingfish, sharpchin	Fodiator acutus
206	flyingfish, blackwing	Hirundichthys rondeleti
214	dory, mirror	Zenopsis nebulosa
216	crestfish	Lophotus lacepedei
217	ribbonfish family	Trachipteridae
219	ribbonfish, tapertail	Trachipterus fukuzaki
220	ribbonfish, scalloped	Zu cristatus
221	oarfish	Regalecus glesne
224	stickleback, ninespine	Pungitius pungitius
226	snipefish, slender	Macrorhamphosus gracilis
227	pipefish family	Sygnathidae
229	pipefish, barred	Syngnathus auliscus
230	pipefish, kelp	Syngnathus californiensis
231	seahorse, Pacific	Hippocampus ingens
293	rockfish, dwarf red	Sebastes rufinanus
298	searobin family	Triglidae
300	searobin, splitnose	Bellator xenisma
311	mackerel, Atka	Pleurogrammus
		monopterygius
314	skilfish	Erilepis zonifer
316	sculpin, twohorn	Icelus bicornis
317	sculpin, spatulate	Icelus spatula
320	hamecon	Artediellus scaber

Sorted by Species Code		
SP CODE	COMMON NAME	SCIENTIFIC NAME
323	sculpin, smoothhead	Artedius lateralis
324	sculpin, puget sound	Ruscarius meanyi
326	sculpin, corralline	Artedius corallinus
327	sculpin, roughcheek	Ruscarius creaseri
329	sculpin, crested	Blepsias bilobus
330	sculpin, silver spotted	Blepsias cirrhosus
332	sculpin, calico	Clinocottus embryum
333	sculpin, mosshead	Clinocottus glopiceps
338	sculpin, spinyhead	Dasycottus setiger
343	sculpin, armorhead	Gymnocanthus galeatus
347	lord, yellow Irish	Hemilepidotus jordani
349	sculpin, bigmouth	Hemitripterus bolini
354	sculpin, frogmouth	Icelinus oculatus
355	sculpin, pit head	Icelinus cavifrons
356	sculpin, fringed	Icelinus fimbriatus
357	sculpin, yellowchin	Icelinus quadriseriatus
360	sculpin, belligerent	Megalocottus platycephalus
361	sculpin, brightbelly	Microcottus sellaris
362	sculpin, plain	Myoxocephalus jaok
363	sculpin, warthead	Myoxocephalus niger
365	sculpin, fourhorn	Myoxocephalus
		quadricornis
366	sculpin, Arctic	Myoxocephalus scorpioides
367	sculpin, shorthorn	Myoxocephalus scorpius
369	sculpin, eyeshode	Nautichthys pribilovius
371	sculpin, saddleback	Oligocottus rimensis
372	sculpin, fluffy	Oligocottus snyderi
373	sculpin, thornback	Paricelinus hopliticus
374	sculpin, spineless	Phallocottus obtusus
375	sculpin, slim	Radulinus asprellus
376	sculpin, darter	Radulinus boleoides
377	sculpin, smoothgum	Radulinus vinculus
380	sculpin, kelp	Sigmistes caulias
381	sculpin, smithi	Sigmistes smithi
382	sculpin, monacled	Synchirus gilli
384 385	sculpin, roughspine	Triglops macellus
	sculpin, ribbed	Triglops pingeli
386 387	sculpin, spectacled sculpin, roughback	Triglops scepticus Chitonotus pugettensis
388	sculpin, roughback sculpin, spinynose	Asemichthys taylori
		Jordani zonope
389 390	sculpin, longfin sculpin, lavender	Leiocottus hirundo
391	sculpin, lavender sculpin, butterfly	Hemilepidotus papilio
392	sculpin, butterny sculpin, snubnose	Orthoropias triacis
393	sculpin, shubhose sculpin, tadpole	
<u> </u>	scuipin, tadpole	Psychrolutes paradoxus

Sorted by Species Code		
SP CODE	COMMON NAME	SCIENTIFIC NAME
394	sculpin, blob	Phychrolutes phrictus
395	sculpin, soft	Psychrolutes sigalutes
396	poacher family	Agonidae
397	poacher,northern	Agonopsis vulsa
	spearnose	
398	poacher,southern	Agonopsis sterletus
	spearnose	
399	alligatorfish, smooth	Anoplagonus inermis
400	alligatorfish, Aleutian	Aspidophoroides bartoni
401	alligatorfish, Arctic	Aspidophoroides olriki
402	starsnout, gray	Bathyagonus alascanus
403	starsnout, spinycheck	Bathyagonus infraspinatus
404	poacher, bigeye	Bathyagonus pentacanthus
405	poacher, blackfin	Bathyagonus nigripinnis
407	poacher, fourhorn	Hypsagonus quadricornis
408	poacher, Bering	Occella dodecaedron
409	poacher, warty	Occella verrucosa
410	poacher, pygmy	Odontopyxis trispinosa
411	poacher, tubenose	Pallasina barbata
412	poacher, blacktip	Xeneretmus latifrons
413	poacher, bluespotted	Xeneretmus triacanthus
414	poacher, pricklebreast	Stellerina xyosterna
415	snailfish family	Cyclopteridae
416	lumpsucker, smooth	Aptocyclus ventricosus
417	snailfish, blacktail	Careproctus melanurus
418	snailfish, blotched	Crystallichthys cyclopilus
419	lumpsucker, leatherfin	Eumicrotremus derjugini
420	lumpsucker, Pacific spiny	Eumicrotremus orbis
421	snailfish, spotted	Liparis callyodon
422	snailfish, ribbon	Liparis cyclopus
423	snailfish, polkadot	Liparis cyclostigma
424	snailfish, marbled	Liparis dennyi
425	snailfish, tidepool	Liparis florae
426	snailfish, slipskin	Liparis fucensis
427	seasnail, gelatinous	Liparis fabricii
428	snailfish, spiny	Liparis mucosus
429	snailfish, showy	Liparis pulchellus
430	snailfish, ringtail	Liparis rutteri
431	snailfish, tadpole	Nectoliparis pelagicus
432	snailfish, prickly	Paraliparis deani
433	snailfish, Bering	Liparis beringianus
434	snailfish, lobefin	Liparis greeni
442	bass, splittail	Hemanthias perunanus
448	seabass, pygmy	Serraniculus pumilio
451	bigeye family	Priacanthidae
	2.30,01011111	acananaac

Sorted by Species Code		
SP CODE	COMMON NAME	SCIENTIFIC NAME
453	cardinalfish, Guadalupe	Apogon guadalupensis
458	sucker, marlin	Remora osteochir
459	remora	Remora remora
460	remora, spearfish	Remora brachyptera
463	jack, green	Caranx caballus
464	bumper, Pacific	Chloroscombrus orqueta
465	leatherjacket	Oligoplites saurus
466	amberjack, Pacific	Seriola colburni
468	pompano, paloma	Trachinotus paitensis
469	pompano, gafftopsail	Trachinotus rhodopus
470	moonfish, Pacific	Selene peruviana
473	roosterfish	Nematistius pectoralis
474	dolphin family	Coryphaenidae
478	mojarra, spotfin	Eucinostomus argenteus
479	mojarra, Pacific flagfin	Eucinostomus gracilis
483	porgy, Pacific	Calamus brachysomus
495	goatfish, Mexican	Mulloidichthys dentatus
502	butterflyfish, threeband	Chaetodon humeralis
503	butterflyfish, scythe	Chaetodon falcifer
504	armorhead, pelagic	Pentaceros richardsoni
528	pomfret, Pacific	Brama japonica
529	pomfret, bigtooth	Brama orcini
530	pomfret, rough	Teractes asper
531	fanfish, Pacific	Pteraclis aesticola
532	pomfret, sickle	Taractichthys steindachneri
535	threadfin family	Polynemidae
536	bobo, blue	Polydactylus approximans
537	bobo, yellow	Polydactylus opercularis
543	sandfish, sailfin	Arctoscopus japonicus
548	searcher	Bathymaster signatus
550	stargazer, smooth	Kathetostoma averruncus
554	blenny, mussel	Hypsoblennius jenkinsi
560	kelpfish, scarlet	Gibbonsia erythra
562	kelpfish, island	Alloclinus holderi
563	pikeblenny, orangethroat	Chaenopsis alepidota
564	blenny, deepwater	Crypotrema corallinum
566	fringehead, yellowfin	Neoclinus stephensae
569	quillfish	Ptilichthys goodei
571	prickleback, pighead	Acantholumpenus mackayi
572	prickleback, lesser	Alectridium aurantiacum
573	prickleback, Y	Allolumpenus hypochrcmus
574	cockscomb, slender	Anoplarchus insignis
575	cockscomb, high	Anoplarchus purpurescens
576	warbonnet, matcheek	Chirolophis tarsodes
577	warbonnet, mosshead	Chirolophis nugator

Sorted by Species Code		
SP CODE	COMMON NAME	SCIENTIFIC NAME
578	warbonnet, decorated	Chirolophis decoratus
579	prickleback, nutcracker	Bryozoichthys lysimus
580	prickleback, trident	Gymnoclinus cristulatus
581	prickleback, longsnout	Lumpenella longirostris
582	eelblenny, slender	Lumpenus fabricii
584	shanny, daubed	Lumpenus maculatus
585	eelblenny, stout	Lumpenus medius
586	prickleback, ribbon	Phytichthys chirus
587	prickleback, bluebarred	Plectobranchus evides
588	prickleback, whitebarred	Poroclinus rothrocki
589	shanny, Arctic	Stichaeus punctatus
592	wrymouth, giant	Cryptacanthodes giganteus
593	wrymouth, dwarf	Cryptacanthodes aleutensis
594	snakeblenny, fourline	Eumesogrammus praecisus
595	cockscomb, stone	Alectrias alectrolophus
599	gunnel, longfin	Pholis clemensi
600	gunnel, stippled	Rhodymenichthys
		dolichogaster
601	gunnel, Bering	Pholis gilli
604	gunnel, red	Pholis schultzi
605	gunnel, rockweed	Apodichthys fucorum
606	gunnel, kelp	Ulvicola santaerosea
607	graveldiver	Scytalina cerdale
608	prowfish	Zaprora silenus
616	goby, cheekspot	llypnus gilberti
617	goby, halfblind	Lethops connetens
618	goby, zebra	Lythrypnus zebra
619	goby, shadow	Quietula ycauda
620	goby, trident	Tridentiger trigonocephalus
621	goby, blind	Typhlogobius californiensis
622	goby, tidewater	Eucyclogobius newberryi
623	sleeper, Pacific fat	Dormitator latifrons
625	mackerel family, snake	Trichiuridae
626	mackerel, snake	Gempylus serpens
627	escolar	Lepidocybium
		flavobrunneum
628	oilfish	Ruvettus pretiosus
630	scabbardfish, Pacific	Lepidopus fitchi
654	spearfish, shortbill	Tetrapturus angustirostris
656	cigarfish, longfin	Cubiceps paradoxus
680	dab, longhead	Pleuronectes proboscideus
699	puffer, oceanic	Lagocephalus lagocephalus
700	puffer, bullseye	Sphoeroides annulatus
701	burrfish, Pacific	Chilomycterus affinis
702	porcupinefish	Diodon hystrix

Sorted by Species Code		
SP CODE	COMMON NAME	SCIENTIFIC NAME
705	mola, slender	Ranzanic laevis
706	gerenadier, Pacific	Coryphaenoides acrolepis
707	rockfish, harlequin	Sebastes variegatus
708	rockfish, semaphore	Sebastes melanosema
709	flatnose, Pacific	Antimora microlepis
712	bass, hookthroat	Hemanthias signifer
715	gerenadier family	Macrouridae
716	sole, hybrids	Isopsetta
718	slickhead, California	Alepocephalus tenebrosus
719	gerenadier, giant	Albatrossia pectoralis

ABALO abalone Haliotis 400 alligatorfish, Aleutian Aspidophoroides bartoni 401 alligatorfish, Arctic Aspidophoroides otriki 399 alligatorfish, smooth Anoplagonus inermis 466 amberjack, Pacific Seriola colburni 113 anchoveta Cetengraulis mysticetus ANCFM anchovy family Engraulidae ANCGN anchovy genus Anchoa spp. ANCDB anchovy, deepbody Anchoa compressa ANCNO anchovy, northern Engraulis mordax 112 anchovy, slough Anchoa delicatissima ARGNT argentine, Pacific Argentina sialis 504 armorhead, pelagic Pentaceros richardsoni BARPA barracuda, Pacific Sphyraena argentea 140 barreleye Macropinna microstoma SBFAM bass family, sea Serranidae 712 bass, hookthroat Hemanthias signifer SBKLP bass, kelp Paralabrax clathratus 442 bass, splittail Hemanthias perunanus STBAS bass, striped Morone saxatilis SBTHF bass, threadfin Pronotogrammus multifasciatus 175 batfish, spotted Zalieutes elater 451 bigeye family Priacanthidae MARFM billifish family Istiophoridae BIVAL bivalves Bivalvia BLKSM blenny, deepwater Crypotrema corallinum FLNFM blenny family, rockpool Hypsoblennius gentilis KLPRB blenny, neef Paraclinus integripinnis BLNRP blenny, reef Paraclinus integripinnis BLNRP blenny, rockpool Hypsoblennius giberti 536 bobo, blue Polydactylus approximans 537 bobo, yellow Polydactylus approximans BONEF bonefish BONSP boxfish, spiny Ostracion diaphanum BULBR bullhead, brown Ictalurus nebulosus	Sorted by C	Sorted by Common Name		
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701 burrfish, Pacific Chilomycterus affinis				

Sorted by Common Name		
SP CODE	COMMON NAME	SCIENTIFIC NAME
BUTFM	butterfish family	Stromateidae
BFFFM	butterflyfish family	Chaetodontidae
503	butterflyfish, scythe	Chaetodon falcifer
502	butterflyfish,	Chaetodon humeralis
	threeband	
RYFLY	butterflyray, California	Gymnura marmorata
SCCAB	cabezon	Scorpaenichthys marmoratus
GRPSC	cabrilla, spotted	Epinephelus analogus
SMCAP	capelin	Mallotus villosus
453	cardinalfish,	Apogon guadalupensis
	Guadalupe	
CARPC	carp, common	Cyprinus carpio
CTFPE	catalufa, popeye	Pristigenys serrula
CATCN	catfish, channel	Ictalurus punctatus
163	chihuil	Bagre panamensis
SCBFM	chub family, sea	Kyphosidae
SCBBS	chub, bluestripped	Sectator ocyurus
656	cigarfish, longfin	Cubiceps paradoxus
CLAMS	clam,unspecified	Bivalvia
CLMBK	clam, basket cockle	Clinocardium nuttallii
CLMGD	clam, geoduck	Panopea generosa
CLMGP	clam, gaper	Tresus nuttallii
CLMLN	clam, common littleneck	Protothaca staminea
CLMNR	clam, northern razor	Siliqua patula
CLMPO	clam, pismo	Tivela stultorum
CLMWA	clam, common washington	Saxidomus nuttalli
167	clingfish family	Gobiesocidae
170	clingfish, bearded	Gobiesox papillifer
171	clingfish, California	Gobiesox rhessondon
172	clingfish, kelp	Rimicola muscarum
169	clingfish, lined	Gobiesox eugrammus
CLNGN	clingfish, nothern	Gobiesox maeandricus
173	clingfish, slender	Rimicola eigenmanni
KLPFM	clinid family	Clinidae
575	cockscomb, high	Anoplarchus purpurescens
574	cockscomb, slender	Anoplarchus insignis
595	cockscomb, stone	Alectrias alectrolophus
CODFM	cod family	Gadidae
CODPA	cod, Pacific	Gadus macrocephalus
CBFLS	combfish, longspine	Zaniolepis latipinnis
CBFSS	combfish, shortspine	Zaniolepis frenata
94	conger, Catalina	Gnathophis catalinensis
CRBCA	corbina, California	Menticirrhus undulatus

Sorted by Common Name		
SP CODE	COMMON NAME	SCIENTIFIC NAME
COROM	corvina, orangemouth	Cynoscion xanthulus
CORSF	corvina, shortfin	Cynoscion parvipinnis
CRBGN	crab genus, cancer	Cancer
CRABS	crab tribe, true	Brachyuratribe
CRBBR	crab, brown rock	Cancer antennarius
CRBDG	crab, Dungeness	Metacarcinus magister
CRBGR	crab, graceful rock	Cancer gracilis
CRBPR	crab, pelagic red	Pleuroncodes, palnipes
CRBRR	crab, red rock	Cancer productus
CRBSH	crab,sheep	Loxorhynchus grandis
CRBYR	crab, yellow rock	Cancer anthonyi
216	crestfish	Lophotus lacepedei
CRKBK	croaker, black	Cheilotrema saturnum
CRKSF	croaker, spotfin	Roncador stearnsi
CROWT	croaker, white	Genyonemus lineatus
CRKYF	croaker, yellowfin	Umbrina roncador
CRUST	crustaceans	Crustacea
CUTLP	cutlassfish, Pacific	Trichiurus nitens
680	dab, longhead	Pleuronectes proboscideus
148	daggertooth family	Anotopteridae
DAMFM	damselfish family	Pomacentridae
474	dolphin family	Coryphaenidae
POMDO	dolphin, pompano	Coryphaena equisetis
DRADO	dolphinfish	Coryphaena hippurus
214	dory, mirror	Zenopsis nebulosa
DRGFM	dragonfish family	Stomiidae
142	dragonfish, longfin	Tactostoma macropus
DRMFM	drum family	Sciaenidae
CSKFM	eel family, cusk	Ophidiidae
SELFM	eel family, snake	Ophichthidae
99	eel family, snipe	Nemichthyidae
EELOR	eel order	Anguilliformes
185	eel, basketweave cusk	Otophidium scrippsae
97	eel, Pacific snake	Ophichthus triserialis
96	eel, Pacific worm	Myrophis vafer
100	eel, slender snake	Nemichthys scolopaceus
184	eel, spotted cusk	Chilara taylori
SELYL	eel, yellow snake	Ophichthus zophochir
582	eelblenny, slender	Lumpenus fabricii
585	eelblenny, stout	Lumpenus medius
ELPFM	eelpout family	Zoarcidae
188	eelpout, Alaska	Bothrocara pusillum
198	eelpout, bearded	Lyconema barbatum
187	eelpout, bigfin	Lycodes cortezianus
191	eelpout, black	Lycodes diapterus

Sorted by Common Name		
SP CODE	COMMON NAME	SCIENTIFIC NAME
197	eelpout, blackbelly	Lycodopsis pacifica
193	eelpout, Canadian	Lycodes polaris
196	eelpout, pale	Lycodes pallidus
189	eelpout, pallid	Lycodapus mandibularis
194	eelpout, polar	Lycodes turneri
190	eelpout, shortfin	Lycodes brevipes
192	eelpout, wattled	Lycodes palearis
627	escolar	Lepidocybium flavobrunneum
SMEUL	eulachon	Thaleichthys pacificus
531	fanfish, Pacific	Pteraclis aesticola
159	flashlightfish	Protomyctophum crockeri
FLTOR	flatfish order	Pleuronectiformes
709	flatnose, Pacific	Antimora microlepis
FLLFN	flounder family, lefteye	Bothidae
FLRFM	flounder family, righteye	Pleuronectidae
SOLAF	flounder, Arctic	Pleuronectes glacialis
FLRAR	flounder, arrowtooth	Atheresthes stomias
SOLBF	flounder, Bering	Hippoglossoides robustus
FLRKM	flounder, Kamchatka	Atheresthes evermanni
FLRST	flounder, starry	Platichthys stellatus
FLYFM	flyingfish family	Exocoetidae
206	flyingfish, blackwing	Hirundichthys rondeleti
FLYCA	flyingfish, California	Cypselurus californicus
205	flyingfish, sharpchin	Fodiator acutus
KLPOF	fringehead, onespot	Neoclinus urinotatus
KLPSF	fringehead, sarcastic	Neoclinus blanchardi
566	fringehead, yellowfin	Neoclinus stephensae
174	frogfish, roughjaw	Antennarius avalonis
GARIB	garibaldi	Hypsypops rubicundus
715	gerenadier family	Macrouridae
719	gerenadier, giant	Albatrossia pectoralis
706	gerenadier, Pacific	Coryphaenoides acrolepis
495	goatfish, Mexican	Mulloidichthys dentatus
GOBFM	goby family	Gobiidae
GOBAR	goby, arrow	Clevelandia ios
BOGBY	goby, bay	Lepidogobius lepidus
GOBBE	goby, blackeye	Coryphopterus nicholsi
621	goby, blind	Typhlogobius californiensis
616	goby, cheekspot	llypnus gilberti
617	goby, halfblind	Lethops connetens
619	goby, shadow	Quietula ycauda
622	goby, tidewater	Eucyclogobius newberryi
620	goby, trident	Tridentiger trigonocephalus
BOGYL	goby, yellowfin	Acanthogobius flavimanus

Sorted by Common Name		
SP CODE	COMMON NAME	SCIENTIFIC NAME
618	goby, zebra	Lythrypnus zebra
607	graveldiver	Scytalina cerdale
GRNFM	greenlingfamily	Hexagrammidae
GRNGN	greenlinggenus	Hexagrammos
GRNKP	greenling, kelp	Hexagrammos decagrammus
GRNMA	greenling, masked	Hexagrammos octogrammus
GRNPT	greenling, painted	Oxylebius pictus
GRNRK	greenling, rock	Hexagrammos lagocephalus
GRNWT	greenling,	Hexagrammos stelleri
	whitespotted	
GRPGN	grouper,genus	Epinephelus
	(epinephelus)	
GRPBT	grouper, broomtail	Mycteroperca xenarcha
GRPGF	grouper, gulf	Mycteroperca jordani
GRPSS	grouper, star-studded	Hyporthodus niphobles
GRUCA	grunion, California	Leuresthes tenuis
GNTFM	gruntfamily	Haemulidae
GUIFM	guitarfish family	Rhinobatidae
GUIBD	guitarfish, banded	Zapteryx exasperata
GUISN	guitarfish, shovelnose	Rhinobatos productus
GRPGC	gulfconey	Hyporthodus acanthistius
GUNFM	gunnel family	Pholidae
601	gunnel, Bering	Pholis gilli
GUNCR	gunnel, crescent	Pholis laeta
606	gunnel, kelp	Ulvicola santaerosea
599	gunnel, longfin	Pholis clemensi
GUNPP	gunnel, penpoint	Apodichthys flavidus
604	gunnel, red	Pholis schultzi
605	gunnel, rockweed	Apodichthys fucorum
GUNSB	gunnel, saddleback	Pholis ornata
600	gunnel, stippled	Rhodymenichthys dolichogaster
HAGFM	hagfish order	Myxinidae
HAGBK	hagfish, black	Eptatretus deani
HAGPA	hagfish, Pacific	Eptatretus stouti
PHAKE	hake, Pacific	Merluccius productus
202	halfbeak	Hyporhamphus unifasciatus
203	halfbeak	Hyporhamphus rosae
201	halfbeak, longfin	Hemiramphus saltator
204	halfbeak, ribbon	Euleptorhamphus viridis
HALFM	halfmoon	Medialuna californiensis
HALCA	halibut, California	Paralichthys californicus
HALGL	halibut, Greenland	Reinhardtius hippoglossoides
HALPA	halibut, Pacific	Hippoglossus stenolepis
320	hamecon	Artediellus scaber

Sorted by Common Name		
SP CODE	COMMON NAME	SCIENTIFIC NAME
153	headlightfish,	Diaphus theta
	California	
HERFM	herring family	Clupeidae
107	herring, flatiron	Harengula thrissina
106	herring, middling	Opisthonema medirastre
	thread	
HERPA	herring, Pacific	Clupea pallasi
HERRD	herring, round	Etrumeus teres
JACFM	jack family	Carangidae
463	jack, green	Caranx caballus
KAWAK	kawakawa	Euthynnus affinis
KLPCR	kelpfish, crevice	Gibbonsia montereyensis
KLPGT	kelpfish, giant	Heterostichus rostratus
562	kelpfish, island	Alloclinus holderi
560	kelpfish, scarlet	Gibbonsia erythra
KLPSP	kelpfish, spotted	Gibbonsia elegans
KLPST	kelpfish, striped	Gibbonsia metzi
KLFCA	killifish, California	Fundalus parvipinnis
KOSAL	king-of-the-salmon	Trachipterus altivelis
158	lampfish, diogenes	Diogenys lanternatus
152	lampfish, dogtooth	Ceratoscopelus townsendi
160	lampfish, Mexican	Triphoturus mexicanus
156	lampfish, northern	Stenobrachius leucopsarus
155	lampfish, patchwork	Notoscopelus resplendens
154	lampfish, pinpoint	Lampanyctus regalis
LMPFM	lamprey family	Petromyzontidae
LMPAR	lamprey, Arctic	Lampetra japonica
LMPPA	lamprey, Pacific	Entosphenus tridentatus
SOLPA	lance, Pacific sand	Ammodytes hexapterus
146	lancetfish family	Alepisauridae
LANLN	lancetfish, longnose	Alepisaurus ferox
151	lanternfish family	Myctophidae
157	lanternfish, blue	Tarletonbeania crenularis
465	leatherjacket	Oligoplites saurus
LNGCD	lingcod	Ophiodon elongatus
LZDFM	lizardfish family	Synodontidae
LZDCA	lizardfish, California	Synodus lunioceps
LOBSP	lobster, spiny	Panulirus interruptus
SCILG	lord genus, Irish	Hemilepidotus
SCBIL	lord, brown Irish	Hemilepidotus spinosus
SCRIL	lord, red Irish	Hemilepidotus hemilepidotus
347	lord, yellow Irish	Hemilepidotus jordani
LUVAR	louvar	Luvarus imperialis
419	lumpsucker, leatherfin	Eumicrotremus derjugini

Sorted by Common Name		
SP CODE	COMMON NAME	SCIENTIFIC NAME
420	lumpsucker,Pacific spiny	Eumicrotremus orbis
416	lumpsucker, smooth	Aptocyclus ventricosus
90	machete	Elops affinis
MACFM	mackerel family	Scombridae
625	mackerel family, snake	Trichiuridae
311	mackerel, Atka	Pleurogrammus monopterygius
MACBL	mackerel, bullet	Auxis rochei
MACPA	mackerel, chub (Pacific)	Scomber japonicus
MACFR	mackerel, frigate	Auxis thazard
JACMK	mackerel, jack	Trachurus symmetricus
626	mackerel, snake	Gempylus serpens
MANTA	manta	Manta birostris
82	manta family	Mobulidae
MARBK	marlin, black	Makaira indica
MARBL	marlin, blue	Makaira nigricans
MARST	marlin, striped	Tetrapturus audax
MIDGN	midshipman genus	Porichthys
MIDPF	midshipman, plainfin	Porichthys notatus
MIDSP	midshipman, specklefin	Porichthys myriaster
85	mobula, smoothtail	Mobula thurstoni
84	mobula, spinetail	Mobula japanica
MOJFM	mojarra family	Gerreidae
479	mojarra, Pacific flagfin	Eucinostomus gracilis
478	mojarra, spotfin	Eucinostomus argenteus
SUNFM	mola family	Molidae
705	mola, slender	Ranzanic laevis
MOLLU	mollusks	Mollusca
470	moonfish, Pacific	Selene peruviana
MORAY	moray, California	Gymnothorax mordax
LJMUD	mudsucker, longjaw	Gillichthus mirabilis
STMUL	mullet, striped	Mugil cephalus
NEDCA	needlefish, California	Strongylura exilis
221	oarfish	Regalecus glesne
OCTOP	octopods	Octopoda
628	oilfish	Ruvettus pretiosus
OPAHS	opah	Lampris guttatus
OPALE RAJOR	opaleye	Girella nigricans
	order, skate and ray	Rajiformes
149 150	pearleye family	Scopelarchidae Benthalbella dentata
PERFM	pearleye, northern perch family	Percidae
FERFIN	регоптанину	reiciuae

Sorted by Common Name		
SP CODE	COMMON NAME	SCIENTIFIC NAME
SPBLK	perch, black	Embiotoca jacksoni
SPDWF	perch, dwarf	Micrometrus minimus
SPKLP	perch, kelp	Brachyistius frenatus
RFPOP	perch, Pacific ocean	Sebastes alutus
SPPIL	perch, pile	Rhacochilus vacca
SPREF	perch, reef	Micrometrus aurora
SPSHR	perch, shiner	Cymatogaster aggregata
PERZB	perch, zebra	Hermosilla azurea
563	pikeblenny,	Chaenopsis alepidota
	orangethroat	
PILTF	pilotfish	Naucrates ductor
227	pipefish family	Sygnathidae
229	pipefish, barred	Syngnathus auliscus
PIPEB	pipefish, bay	Syngnathus leptorhynchus
230	pipefish, kelp	Syngnathus californiensis
SOLPL	plaice, Alaska	Pleuronectes quadrituberculatus
396	poacher family	Agonidae
408	poacher, Bering	Occella dodecaedron
404	poacher, bigeye	Bathyagonus pentacanthus
405	poacher, blackfin	Bathyagonus nigripinnis
412	poacher, blacktip	Xeneretmus latifrons
413	poacher, bluespotted	Xeneretmus triacanthus
407	poacher, fourhorn	Hypsagonus quadricornis
397	poacher, northern	Agonopsis vulsa
	spearnose	
414	poacher, pricklebreast	Stellerina xyosterna
410	poacher, pygmy	Odontopyxis trispinosa
398	poacher,southern	Agonopsis sterletus
	spearnose	
411	poacher, tubenose	Pallasina barbata
409	poacher, warty	Occella verrucosa
POLWE	pollock, walleye	Theragra chalcogramma
POMFM	pomfret family	Bramidae
529	pomfret, bigtooth	Brama orcini
528	pomfret, Pacific	Brama japonica
530	pomfret, rough	Teractes asper
532	pomfret, sickle	Taractichthys steindachneri
469	pompano, gafftopsail	Trachinotus rhodopus
POMPA	pompano,Pacific	Peprilus simillimus
100	(butterfish)	T 1: 1 2 2 1
468	pompano, paloma	Trachinotus paitensis
702	porcupinefish	Diodon hystrix
483	porgy, Pacific	Calamus brachysomus
PRKFM	prickleback family	Stichaeidae
PRKBK	prickleback, black	Xiphister atropurpureus

Sorted by C	ommon Name	
SP CODE	COMMON NAME	SCIENTIFIC NAME
587	prickleback,	Plectobranchus evides
	bluebarred	
572	prickleback, lesser	Alectridium aurantiacum
581	prickleback, longsnout	Lumpenella longirostris
PRKMK	prickleback,	Cebidichthys violaceus
	monkeyface	
579	prickleback,	Bryozoichthys lysimus
	nutcracker	
571	prickleback, pighead	Acantholumpenus mackayi
586	prickleback, ribbon	Phytichthys chirus
PRKRK	prickleback, rock	Xiphister mucosus
PRKSN	prickleback, snake	Lumpenus sagitta
580	prickleback, trident	Gymnoclinus cristulatus
588	prickleback,	Poroclinus rothrocki
	whitebarred	
573	prickleback, Y	Allolumpenus hypochrcmus
608	prowfish	Zaprora silenus
PUFFM	puffer family	Tetraodontidae
700	puffer, bullseye	Sphoeroides annulatus
699	puffer, oceanic	Lagocephalus lagocephalus
QUEEN	queenfish	Seriphus politus
569	quillfish	Ptilichthys goodei
RAGFS	ragfish	lcosteus aenagmaticus
RATFS	ratfish, spotted	Hydrolagus colliei
RYBAT	ray, bat	Myliobatis californica
ERYPA	ray, Pacific electric	Torpedo californica
459	remora	Remora remora
REMFM	remora family	Echeneidae
460	remora, spearfish	Remora brachyptera
217	ribbonfish family	Trachipteridae
220	ribbonfish, scalloped	Zu cristatus
219	ribbonfish, tapertail	Trachipterus fukuzaki
RFGEN	rockfish genus	Sebastes
RFBOC	rockfish, (bocaccio)	Sebastes paucispinis
RFPEP	rockfish, (chilipepper)	Sebastes goodei
RFCOW	rockfish, (cowcod)	Sebastes levis
RFTRE	rockfish, (treefish)	Sebastes serriceps
RFAUR	rockfish, aurora	Sebastes aurora
RFBNK	rockfish, bank	Sebastes rufus
RFBLK	rockfish, black	Sebastes melanops
RFBAY	rockfish,black and	Sebastes chrysomelas
	yellow	
RFBKG	rockfish, blackgill	Sebastes melanostomus
RFBLU	rockfish, blue	Sebastes mystinus
RFBSP	rockfish,	Sebastes gilli
	bronzespotted	
		17-27

Sorted by Common Name		
SP CODE	COMMON NAME	SCIENTIFIC NAME
RFBRN	rockfish, brown	Sebastes auriculatus
RFCLO	rockfish, calico	Sebastes dalli
RFCAN	rockfish, canary	Sebastes pinniger
RFCMA	rockfish, chameleon	Sebastes phillipsi
RFCHN	rockfish, China	Sebastes nebulosus
RFCOP	rockfish, copper	Sebastes caurinus
RFDBL	rockfish, darkblotched	Sebastes crameri
RFDUS	rockfish, dusky	Sebastes ciliatus
293	rockfish, dwarf red	Sebastes rufinanus
RFFLG	rockfish, flag	Sebastes rubrivinctus
RFFRK	rockfish, freckled	Sebastes lentiginosus
RFGOP	rockfish, gopher	Sebastes carnatus
RFGRS	rockfish, grass	Sebastes rastrelliger
RFGBL	rockfish,	Sebastes rosenblatti
	greenblotched	
RFGRN	rockfish, greenspotted	Sebastes chlorostictus
RFGST	rockfish, greenstriped	Sebastes elongatus
RFHBD	rockfish, halfbanded	Sebastes semicinctus
707	rockfish, harlequin	Sebastes variegatus
RFHNC	rockfish, honeycomb	Sebastes umbrosus
RFKLP	rockfish, kelp	Sebastes atrovirens
RFMEX	rockfish, Mexican	Sebastes macdonaldi
RFOLV	rockfish, olive	Sebastes serranoides
RFPNK	rockfish, pink	Sebastes eos
RFPRS	rockfish, pinkrose	Sebastes simulator
RFPSD	rockfish, Puget Sound	Sebastes emphaeus
RFPYG	rockfish, pygmy	Sebastes wilsoni
RFQIL	rockfish, quillback	Sebastes maliger
RFRBD	rockfish, redbanded	Sebastes babcocki
RFRST	rockfish, redstripe	Sebastes proriger
RFRTN	rockfish, rosethorn	Sebastes helvomaculatus
RFROS	rockfish, rosy	Sebastes rosaceus
RFRGH	rockfish, rougheye	Sebastes aleutianus
708	rockfish, semaphore	Sebastes melanosema
RFSCN	rockfish, sharpchin	Sebastes zacentrus
RFSHB	rockfish, shortbelly	Sebastes jordani
RFSRK	rockfish, shortraker	Sebastes borealis
RFSLG	rockfish, silvergray	Sebastes brevispinis
RFSPK	rockfish, speckled	Sebastes ovalis
RFSNS	rockfish, splitnose	Sebastes diploproa
RFSQS	rockfish,	Sebastes hopkinsi
DECTA	squarespotted	Cabaataa aanatallatus
RFSTA	rockfish, starry	Sebastes constellatus
RFSTR	rockfish, stripetail	Sebastes saxicola
RFSDS	rockfish, swordspine	Sebastes ensifer

Sorted by Common Name		
SP CODE	COMMON NAME	SCIENTIFIC NAME
RFTIG	rockfish, tiger	Sebastes nigrocinctus
RFVER	rockfish, vermilion	Sebastes miniatus
RFWTB	rockfish, whitebelly	Sebastes vexillaris
RFWID	rockfish, widow	Sebastes entomelas
RFYEY	rockfish, yelloweye	Sebastes ruberrimus
RFYMN	rockfish, yellowmouth	Sebastes reedi
RFYTL	rockfish, yellowtail	Sebastes flavidus
ROCKH	rockhead	Bothragonus swani
RNQFM	ronquil family	Bathymasteridae
RNQBB	ronquil, bluebanded	Rathbunella hypoplecta
RNQNO	ronquil, northern	Ronqilus jordani
473	roosterfish	Nematistius pectoralis
SABLE	sablefish	Anoplopoma fimbria
SABFM	sablefish family	Anoplopomatidae
SAILF	sailfish	Istiophorus platypterus
SALEM	salema	Xenistius californiensis
SALFM	salmon family	Salmonidae
SALGN	salmon genus	Oncorhynchus spp.
SALAT	salmon, Atlantic	Salmo salar
SALCK	salmon, chinook	Oncorhynchus tshawytscha
SALCM	salmon, chum	Oncorhynchus keta
SALCO	salmon, coho	Oncorhynchus kisutch
SALPK	salmon, pink	Oncorhynchus gorbuscha
SALSE	salmon, sockeye	Oncorhynchus nerka
SBGEN	sandbass genus	Paralabrax
SBBAR	sandbass, barred	Paralabrax nebulifer
SBSPT	sandbass, spotted	Paralabrax maculatofascia
DABGN	sanddab genus	Citharichthys
DABLF	sanddab, longfin	Citharlchthys xanthostigma
DABPA	sanddab, Pacific	Citharichthys sordidus
DABSP	sanddab, speckled	Citharichthys stigmaeus
SNDFM	sandfish family	Trichodontidae
SNDPA	sandfish, Pacific	Trichodon trichodon
543	sandfish, sailfin	Arctoscopus japonicus
SARPA	sardine, Pacific	Sardinops sagax
SARGO	sargo	Anisotremus davidsoni
SAUPA	saury, Pacific	Cololabis saira
630	scabbardfish, Pacific	Lepidopus fitchi
SCBRZ	scabbardfish,	Assurger anzac
	razorback	
MSCAD	scad, Mexican	Decapterus scombrinus
SCPRO	scallop, giant rock	Crassadoma gigantea
SCPUS	scallop, unspecified	Pectinidae
SCRFM	scorpionfish family	Scorpaenidae

Sorted by Common Name		
SP CODE	COMMON NAME	SCIENTIFIC NAME
SCRCA	scorpionfish,	Scorpaena guttata
	California	
SCRRB	scorpionfish, rainbow	Scorpaenodes xyris
SCFAM	sculpin family	Cottidae
SCANT	sculpin, antlered	Enophrys diceraus
366	sculpin, Arctic	Myoxocephalus scorpioides
SCASH	sculpin, Arctic staghorn	Gymnocanthus tricuspis
343	sculpin, armorhead	Gymnocanthus galeatus
SCBLD	sculpin, bald	Clinocottus recalvus
360	sculpin, belligerent	Megalocottus platycephalus
349	sculpin, bigmouth	Hemitripterus bolini
SCBKF	sculpin, blackfin	Malacocottus kincaidi
394	sculpin, blob	Phychrolutes phrictus
SCBNH	sculpin, bonehead	Artedius notospilotus
361	sculpin, brightbelly	Microcottus sellaris
SCBUF	sculpin, buffalo	Enophrys bison
SCBUL	sculpin, bull	Enophrys taurina
391	sculpin, butterfly	Hemilepidotus papilio
332	sculpin, calico	Clinocottus embryum
SCCRG	sculpin, coastrange	Cottus aleuticus
326	sculpin, corralline	Artedius corallinus
329	sculpin, crested	Blepsias bilobus
376	sculpin, darter	Radulinus boleoides
SCDSK	sculpin, dusky	Icelinus burchani
369	sculpin, eyeshode	Nautichthys pribilovius
372	sculpin, fluffy	Oligocottus snyderi
365	sculpin, fourhorn	Myoxocephalus quadricornis
356	sculpin, fringed	Icelinus fimbriatus
354	sculpin, frogmouth	Icelinus oculatus
SCGRT	sculpin, great	Myoxocephalus
		polyacanthocephalus
SCGRU	sculpin, grunt	Rhamphocottus richardsoni
380	sculpin, kelp	Sigmistes caulias
390	sculpin, lavender	Leiocottus hirundo
SCLST	sculpin, leister	Enophrys lucasi
389	sculpin, longfin	Jordani zonope
382	sculpin, monacled	Synchirus gilli
333	sculpin, mosshead	Clinocottus glopiceps
SCNTH	sculpin, northern	Icelinus borealis
SCPSH	sculpin,Pacific	Leptocottus armatus
	staghorn	
SCPAD	sculpin, padded	Artedius fenestralis
355	sculpin, pit head	Icelinus cavifrons
362	sculpin, plain	Myoxocephalus jaok

Sorted by Common Name		
SP CODE	COMMON NAME	SCIENTIFIC NAME
SCPRK	sculpin, prickly	Cottus asper
324	sculpin, puget sound	Ruscarius meanyi
385	sculpin, ribbed	Triglops pingeli
SCRSL	sculpin, rosylip	Ascelichthys rhodorus
387	sculpin, roughback	Chitonotus pugettensis
327	sculpin, roughcheek	Ruscarius creaseri
384	sculpin, roughspine	Triglops macellus
371	sculpin, saddleback	Oligocottus rimensis
SCSFN	sculpin, sailfin	Nautichthys oculofasciatus
SCSCL	sculpin, scaled	Archaulus biseriatus
SCSLH	sculpin, scalyhead	Artedius harringtoni
SCSCT	sculpin, scissortail	Triglops forficata
SCSHN	sculpin, sharpnose	Clinocottus acuticeps
367	sculpin, shorthorn	Myoxocephalus scorpius
330	sculpin, silver spotted	Blepsias cirrhosus
375	sculpin, slim	Radulinus asprellus
381	sculpin, smithi	Sigmistes smithi
377	sculpin, smoothgum	Radulinus vinculus
323	sculpin, smoothhead	Artedius lateralis
392	sculpin, snubnose	Orthoropias triacis
395	sculpin, soft	Psychrolutes sigalutes
317	sculpin, spatulate	Icelus spatula
386	sculpin, spectacled	Triglops scepticus
374	sculpin, spineless	Phallocottus obtusus
338	sculpin, spinyhead	Dasycottus setiger
388	sculpin, spinynose	Asemichthys taylori
SCSPT	sculpin, spotfin	Icelinus tenuis
393	sculpin, tadpole	Psychrolutes paradoxus
373	sculpin, thornback	Paricelinus hopliticus
SCTRF	sculpin, threadfin	Icelinus filamentosus
SCTDP	sculpin, tidepool	Oligocottus maculosus
316	sculpin, twohorn	Icelus bicornis
363	sculpin, warthead	Myoxocephalus niger
SCWOL	sculpin, wolly	Clinocottus analis
357	sculpin, yellowchin	Icelinus quadriseriatus
CUCUM	sea cucumbers	Holothuroidea
URCHN	sea urchins	Diadematidae
GNTSB	seabass, giant	Stereolepis gigas
448	seabass, pygmy	Serraniculus pumilio
SBWHT	seabass, white	Atractoscion nobilis
176	seadevil, triplewart	Cryptopsaras couesi
231	seahorse, Pacific	Hippocampus ingens
SPPNK	seaperch, pink	Zalembius rosaceus
SPRBW	seaperch, rainbow	Hypsurus caryi
SPRUB	seaperch, rubberlip	Rhacochilus toxotes

Sorted by Common Name		
SP CODE	COMMON NAME	SCIENTIFIC NAME
SPSHN	seaperch, sharpnose	Phanerodon atripes
SPSTR	seaperch, striped	Embiotoca lateralis
SPWHT	seaperch, white	Phanerodon furcatus
548	searcher	Bathymaster signatus
298	searobin family	Triglidae
SERLT	searobin, limptail	Prionotus stephanophrys
300	searobin, splitnose	Bellator xenisma
GAPOD	sea slug, sea snail	Gastropoda
SSTAR	sea star	sea star
427	seasnail, gelatinous	Liparis fabricii
SENOR	senorita	Oxyjulis californica
SHADA	shad, American	Alosa sapidissima
589	shanny, Arctic	Stichaeus punctatus
584	shanny, daubed	Lumpenus maculatus
CTSFM	shark family, cat	Scyliorhinidae
CSHFM	shark family, cow	Hexanchidae
SHDFM	shark family, dogfish	Squalidae
FRSFM	shark family, frill	Chlamydoselachidae
50	Shark family,	Sphyrnidae
	hammerhead	
SHMFM	shark family, mackerel	Lamnidae
SHRFM	shark family, requiem	Carcharhinidae
44	shark genus, gray	Carcharhinus
26	shark, basking	Cetorhinus maximus
29	shark, bigeye thresher	Alopias superciliosus
SHBLU	shark, blue	Prionace glauca
SHBNH	shark, bonnethead	Sphyrna tiburo
SHBCS	shark, brown cat	Apristurus brunneus
SHBUL	shark, bull	Carcharhinus leucas
SHDKY	shark, dusky	Carcharhinus obscurus
35	shark, filetail cat	Parmatyrus xaniurus
18	shark, frill	Chlamydoselachus arguineus
SHHRN	shark, horn	Heterodontus francisci
SHLEP	shark, leopard	Triakis semifasciata
33	shark, longnose cat	Apristurus kampae
SHNTH	shark, narrowtooth	Carcharhinus brachyurus
SHANG	shark, Pacific angel	Squatina californica
39	shark,Pacific	Rhizoprionodon longurio
OLIOL D	sharpnose	0
SHSLP	shark, Pacific sleeper	Somniosus pacificus
56	shark, prickly	Echinorhinus cookei
23	shark, ragged tooth	Odontaspis ferox
SHSAL	shark, salmon	Lamna ditropis
SHSEV	shark, seven gill	Notorynchus maculatus
SHSMK	shark, shortfin mako	Isurus oxyrinchus

Sorted by Common Name		
SP CODE	COMMON NAME	SCIENTIFIC NAME
SHSIX	shark, six gill	Hexanchus griseus
52	shark,smooth	Sphyrna zygaena
	hammerhead	
SHFIN	shark, soupfin	Galeorhinus zyopterus
SHSDG	shark, spiny dogfish	Squalus acanthias
SHSWL	shark, swell	Cephaloscyllium ventriosum
SHTHR	shark, thresher	Alopias vulpinus
SHTIG	shark, tiger	Galeocerdo cuvieri
22	shark, whale	Rhincodon typus
SHWHT	shark, white	Carcharodon carcharias
SHEEP	sheephead, California	Semicossyphus pulcher
195	shulupaoluk	Lycodes jugoricus
SHRMP	shrimp	Caridea
SRAGU	sierra, gulf	Scomberomorus concolor
SRAPA	sierra, Pacific	Scomberomorus sierra
SVRFM	silverside family	Atherinidae
SKFAM	skate family	Rajidae
70	skate, Alaska	Bathyraja parmifera
SKALT	skate, Aleutian	Bathyraja aleutica
SKBIG	skate, big	Raja binoculata
69	skate, black	Bathyraja trachura
SKTCA	skate, California	Raja inornata
72	skate, flathead	Bathyraja rosispinis
SKLGN	skate, longnose	Raja rhina
74	skate, roughtail	Raja trachura
68	skate, sandpaper	Bathyraja interrupta
SKSTY	skate, starry	Raja stellulata
314	skilfish	Erilepis zonifer
SKBGN	skipback genus	Euthynnus
BLKSJ	skipjack, black	Euthynnus lineatus
623	sleeper, Pacific fat	Dormitator latifrons
718	slickhead, California	Alepocephalus tenebrosus
SMFAM	smelt family	Osmeridae
DSSFM	smelt family, deepsea	Bathylagidae
SMJAK	smelt, (jacksmelt)	Atherinopsis californiensis
SMTOP	smelt, (topsmelt)	Atherinops affinis
129	smelt, delta	Hypomesus transpacificus
SMLGF	smelt, longfin	Spirinchus thlaeichthys
SMNGT	smelt, night	Spirinchus starksi
131	smelt, rainbow	Osmerus mordax
SMSUR	smelt, surf	Hypomesus pretiosus
SMWTB	smelt, whitebait	Allosmerus elongatus
SHSGN	smoothhound genus	Mustelus
SHBSM	smoothhound, brown	Mustelus henlei
SHGSM	smoothhound, gray	Mustelus californicus

Sorted by Common Name		
SP CODE	COMMON NAME	SCIENTIFIC NAME
SHSSM	smoothhound,	Mustelus lunulatus
	sicklefin	
CASTG	smoothtongue,	Leuroglossus stilbius
	California	
415	snailfish family	Cyclopteridae
433	snailfish, Bering	Liparis beringianus
417	snailfish, blacktail	Careproctus melanurus
418	snailfish, blotched	Crystallichthys cyclopilus
434	snailfish, lobefin	Liparis greeni
424	snailfish, marbled	Liparis dennyi
423	snailfish, polkadot	Liparis cyclostigma
432	snailfish, prickly	Paraliparis deani
422	snailfish, ribbon	Liparis cyclopus
430	snailfish, ringtail	Liparis rutteri
429	snailfish, showy	Liparis pulchellus
426	snailfish, slipskin	Liparis fucensis
428	snailfish, spiny	Liparis mucosus
421	snailfish, spotted	Liparis callyodon
431	snailfish, tadpole	Nectoliparis pelagicus
425	snailfish, tidepool	Liparis florae
594	snakeblenny, fourline	Eumesogrammus praecisus
226	snipefish, slender	Macrorhamphosus gracilis
TBESN	snout, tube	Aulorhynchus flavidus
SOLBG	sole, bigmouth	Hippoglossina stomata
SOLBT	sole, butter	Isopsetta isolepis
SOLCO	sole, C-O	Pleuronichthys coenosus
SOLCF	sole, curlfin	Pleuronichthys decurrens
SOLDS	sole, deepsea	Embassichthys bathybius
SOLDV	sole, Dover	Microstomus pacificus
SOLEG	sole, English	Parophrys vetulus
SOLFT	sole, fantail	Xystreurys liolepis
SOLFH	sole, flathead	Hippoglossoides elassodon
716	sole, hybrids	Isopsetta
SOLPT	sole, petrale	Eopsetta jordani
SOLRX	sole, rex	Glyptocephalus zachirus
SOLRK	sole, rock	Lepidopsetta bilineatus
SOLSD	sole, sand	Psettichthys melanostictus
SOLSL	sole, slender	Lyopsetta exilis
SOLYF	sole, yellowfin	Limanda aspera
SPDPA	spadefish, Pacific	Chaetodipterus zonatus
654	spearfish, shortbill	Tetrapturus angustirostris
139	spookfish family	Opisthoproctidae
SQTSE	squaretail, smalleye	Tetragonurus cuvieri
SQUID	squid	Cephalopoda
SQDJU	squid, jumbo	Dosidicus gigas

Sorted by C	Sorted by Common Name		
SP CODE	COMMON NAME	SCIENTIFIC NAME	
SQDMK	squid, market	Doryteuthis opalescens	
550	stargazer, smooth	Kathetostoma averruncus	
402	starsnout, gray	Bathyagonus alascanus	
403	starsnout, spinycheck	Bathyagonus infraspinatus	
SKBFM	stickleback family	Gasterosteidae	
224	stickleback, ninespine	Pungitius pungitius	
SKBTS	stickleback,	Gasterosteus aculeatus	
	threespine		
SGFAM	stingray family	Dasyatidae	
SGGEN	stingray genus	Dasyatis spp.	
SGDIA	stingray, diamond	Dasyatis dipterura	
SGPEL	stingray, pelagic	Dasyatis violacea	
SGRND	stingray, round	Urolophus halleri	
STGEN	sturgeon genus	Acipenser	
STGRN	sturgeon, green	Acipenser medirostris	
STWHT	sturgeon, white	Acipenser transmontanus	
458	sucker, marlin	Remora osteochir	
SNFFM	sunfish family	Centrarchidae	
SUNOC	sunfish, ocean	Mola mola	
SPFAM	surfperch family	Embiotocidae	
SPBAR	surfperch, barred	Amphistichus argenteus	
SPCAL	surfperch, calico	Amphistichus koelzi	
SPRTL	surfperch, redtail	Amphistichus rhodoterus	
SPSIL	surfperch, silver	Hyperprosopon ellipticum	
SPSPF	surfperch, spotfin	Hyperprosopon anale	
SPWAL	surfperch, walleye	Hyperprosopon argenteum	
SRDFS	swordfish	Xiphias gladius	
THRBK	thornback	Platyrhinoidis triseriata	
RFLST	thornyhead, longspine	Sebastolobus altivelis	
RFSST	thornyhead,	Sebastolobus alascanus	
	shortspine		
535	threadfin family	Polynemidae	
TFPGE	tilefish, Pacific golden- eyed	Caulolatilus affinis	
CODTC	tomcod, Pacific	Microgadus proximus	
TNGCA	tonguefish, California	Symphurus atricauda	
FTRIG	triggerfish, finescale	Balistes polylepis	
TRTPA	tripletail, pacific	Lobotes pacificus	
SALAC	trout, Arctic char	Salvelinus alpinus	
SALCT	trout, cutthroat	Oncorhynchus clarki	
SALRB	trout, rainbow	Oncorhynchus mykiss	
SALTR	trouts, sea run		
TNAAB	tuna, (albacore)	Thunnus alalunga	
TNABE	tuna, bigeye	Thunnus obesus	
TNABF	tuna, bluefin	Thunnus thynnus	

Sorted by Common Name		
SP CODE	COMMON NAME	SCIENTIFIC NAME
TNASJ	tuna, skipjack	Katsuwonus pelamis
TNASL	tuna, slender	Allothunnus fallai
TNAYF	tuna, yellowfin	Thunnus albacares
TNASG	tunas (non-mackerel)	
SOLDT	turbot, diamond	Pleuronicthys guttulatus
SOLHT	turbot, hornyhead	Pleuronichthys verticalis
SOLST	turbot, spotted	Pleuronichthys ritteri
SHINS	Unidentified inshore	
	sharks	
SHOFF	Unidentified offshore	
	sharks	
UNISF	Unidentified	
UNIFH	unidentified fish	
SALDV	Varden, Dolly	Salvelinus malma
143	viperfish, Pacific	Chauliodus macouni
WAHOO	wahoo	Acanthocybium solandri
578	warbonnet, decorated	Chirolophis decoratus
576	warbonnet, matcheek	Chirolophis tarsodes
577	warbonnet, mosshead	Chirolophis nugator
WEKFS	weakfishes	Cynoscion
REMWS	whalesucker	Remora australis
OCWHT	whitefish, ocean	Caulolatilus princeps
WOLFE	wolf-eel	Anarrhichthys ocellatus
WRABS	wrasse, blackspot	
WRAFM	wrasse family	Labridae
WRARB	wrasse, rainbow	Thalassoma luvasanum
WRARK	wrasse, rock	Halichoeres semicinctus
593	wrymouth, dwarf	Cryptacanthodes aleutensis
592	wrymouth, giant	Cryptacanthodes giganteus
YELTL	yellowtail	Seriola lalandi

Sorted by AFS Common Name		
SP CODE	AFS COMMON NAME	SCIENTIFIC NAME
ABALO	abalone genus	Haliotis
188	Alaska eelpout	Bothrocara pusillum
SOLPL	Alaska plaice	Pleuronectes
		quadrituberculatus
70	Alaska skate	Bathyraja parmifera
TNAAB	albacore	Thunnus alalunga
400	Aleutian alligatorfish	Aspidophoroides bartoni
SKALT	Aleutian skate	Bathyraja aleutica
SHADA	American shad	Alosa sapidissima
113	anchoveta	Cetengraulis mysticetus
ANCFM	anchovy family	Engraulidae
ANCGN	anchovy genus	Anchoa spp.
SCANT	antlered sculpin	Enophrys diceraus
401	Arctic alligatorfish	Aspidophoroides olriki
SALAC	Arctic char	Salvelinus alpinus
SOLAF	Arctic flounder	Pleuronectes glacialis
LMPAR	Arctic lamprey	Lampetra japonica
366	Arctic sculpin	Myoxocephalus scorpioides
589	Arctic shanny	Stichaeus punctatus
SCASH	Arctic staghorn sculpin	Gymnocanthus tricuspis
343	armorhead sculpin	Gymnocanthus galeatus
GOBAR	arrow goby	Clevelandia ios
FLRAR	arrowtooth flounder	Atheresthes stomias
311	Atka mackerel	Pleurogrammus monopterygius
SALAT	Atlantic salmon	Salmo salar
RFAUR	aurora rockfish	Sebastes aurora
SCBLD	bald sculpin	Clinocottus recalvus
GUIBD	banded guitarfish	Zapteryx exasperata
RFBNK	bank rockfish	Sebastes rufus
229	barred pipefish	Syngnathus auliscus
SBBAR	barred sandbass	Paralabrax nebulifer
SPBAR	barred surfperch	Amphistichus argenteus
140	barreleye	Macropinna microstoma
185	basketweave cusk eel	Otophidium scrippsae
26	basking shark	Cetorhinus maximus
CLMBK	basket cockle	Clinocardium nuttallii
RYBAT	bat ray	Myliobatis californica
BLNBY	bay blenny	Hypsoblennius gentilis
BOGBY	bay goby	Lepidogobius lepidus
PIPEB	bay pipefish	Syngnathus leptorhynchus
170	bearded clingfish	Gobiesox papillifer
198	bearded eelpout	Lyconema barbatum
360	belligerentsculpin	Megalocottus platycephalus
SOLBF	Bering flounder	Hippoglossoides robustus

Sorted by AFS Common Name		
SP CODE	AFS COMMON NAME	SCIENTIFIC NAME
601	Bering gunnel	Pholis gilli
408	Bering poacher	Occella dodecaedron
433	Bering snailfish	Liparis beringianus
SKBIG	big skate	Raja binoculata
451	bigeye family	Priacanthidae
404	bigeye poacher	Bathyagonus pentacanthus
29	bigeye thresher shark	Alopias superciliosus
TNABE	bigeye tuna	Thunnus obesus
187	bigfin eelpout	Lycodes cortezianus
349	bigmouth sculpin	Hemitripterus bolini
SOLBG	bigmouth sole	Hippoglossina stomata
529	bigtooth pomfret	Brama orcini
MARFM	billfish family	Istiophoridae
BIVAL	bivalve class	Bivalvia
RFBAY	black and yellow rockfish	Sebastes chrysomelas
CRKBK	black croaker	Cheilotrema saturnum
191	black eelpout	Lycodes diapterus
HAGBK	black hagfish	Eptatretus deani
MARBK	black marlin	Makaira indica
SPBLK	black perch	Embiotoca jacksoni
PRKBK	black prickleback	Xiphister atropurpureus
RFBLK	black rockfish	Sebastes melanops
69	black skate	Bathyraja trachura
BLKSJ	black skipjack	Euthynnus lineatus
197	blackbelly eelpout	Lycodopsis pacifica
GOBBE	blackeye goby	Coryphopterus nicholsi
405	blackfin poacher	Bathyagonus nigripinnis
SCBKF	blackfin sculpin	Malacocottus kincaidi
RFBKG	blackgill rockfish	Sebastes melanostomus
BLKSM	blacksmith	Chromis punctipinnis
WRABS	blackspot wrasse	Decodon melasma
417	blacktail snailfish	Careproctus melanurus
412	blacktip poacher	Xeneretmus latifrons
206	blackwing flyingfish	Hirundichthys rondeleti
621	blind goby	Typhlogobius californiensis
394	blobsculpin	Phychrolutes phrictus
418	blotched snailfish	Crystallichthys cyclopilus
536	blue bobo	Polydactylus approximans
157	blue lanternfish	Tarletonbeania crenularis
MARBL	blue marlin	Makaira nigricans
RFBLU	blue rockfish	Sebastes mystinus
SHBLU	blue shark	Prionace glauca
RNQBB	bluebanded ronquil	Rathbunella hypoplecta
587	bluebarred prickleback	Plectobranchus evides
TNABF	bluefin tuna	Thunnus thynnus

Sorted by AFS Common Name		
SP CODE	AFS COMMON NAME	SCIENTIFIC NAME
413	bluespotted poacher	Xeneretmus triacanthus
SCBBS	bluestripped chub	Sectator ocyurus
SHSIX	bluntnose sixgill shark	Hexanchus griseus
RFBOC	bocaccio	Sebastes paucispinis
BONEF	bonefish	Albula vulpes
SCBNH	bonehead sculpin	Artedius notospilotus
SHBNH	bonnethead shark	Sphyrna tiburo
BOTOM	bottomfish (groundfish)	
361	brightbelly sculpin	Microcottus sellaris
RFBSP	bronzespotted rockfish	Sebastes gilli
GRPBT	broomtail grouper	Mycteroperca xenarcha
BULBR	brown bullhead	Ictalurus nebulosus
SHBCS	brown cat shark	Apristurus brunneus
SCBIL	brown Irish lord	Hemilepidotus spinosus
CRBBR	brown rock crab	Cancer antennarius
RFBRN	brown rockfish	Sebastes auriculatus
SHBSM	brown smoothhound	Mustelus henlei
SCBUF	buffalo sculpin	Enophrys bison
SCBUL	bull sculpin	Enophrys taurina
SHBUL	bull shark	Carcharhinus leucas
MACBL	bullet mackerel	Auxis rochei
700	bullseye puffer	Sphoeroides annulatus
SOLBT	butter sole	Isopsetta isolepis
BUTFM	butterfish family	Stromateidae
391	butterfly sculpin	Hemilepidotus papilio
BFFFM	butterflyfish family	Chaetodontidae
SCCAB	cabezon	Scorpaenichthys marmoratus
RFCLO	calico rockfish	Sebastes dalli
332	calico sculpin	Clinocottus embryum
SPCAL	calico surfperch	Amphistichus koelzi
RYFLY	California butterflyray	Gymnura marmorata
171	California clingfish	Gobiesox rhessondon
CRBCA	California corbina	Menticirrhus undulatus
FLYCA	California flyingfish	Cypselurus californicus
GRUCA	California grunion	Leuresthes tenuis
HALCA	California halibut	Paralichthys californicus
153	California headlightfish	Diaphus theta
KLFCA	California killifish	Fundalus parvipinnis
LZDCA	California lizardfish	Synodus lunioceps
MORAY	California moray	Gymnothorax mordax
NEDCA	California needlefish	Strongylura exilis
SCRCA	California scorpionfish	Scorpaena guttata
SHEEP	California sheephead	Semicossyphus pulcher
SKTCA	California skate	Raja inornata
718	California slickhead	Alepocephalus tenebrosus

Sorted by AFS Common Name		
SP CODE	AFS COMMON NAME	SCIENTIFIC NAME
CASTG	Californiasmoothtongue	Leuroglossus stilbius
TNGCA	California touguefish	Symphurus atricauda
193	Canadian eelpout	Lycodes polaris
RFCAN	canary rockfish	Sebastes pinniger
CRBGN	cancer genus	Cancer
SMCAP	capelin	Mallotus villosus
CTSFM	cat shark family	Scyliorhinidae
94	Catalina conger	Gnathophis catalinensis
620	chameleon goby	Tridentiger trigonocephalus
RFCMA	chameleon rockfish	Sebastes phillipsi
CATCN	channel catfish	Ictalurus punctatus
616	cheekspotgoby	llypnus gilberti
163	chihuil	Bagre panamensis
RFPEP	chilipepper	Sebastes goodei
RFCHN	Chinarockfish	Sebastes nebulosus
SALCK	chinook salmon	Oncorhynchus tshawytscha
MACPA	chub (Pacific) mackerel	Scomber japonicus
SALCM	chum salmon	Oncorhynchus keta
CLAMS	clams	Bivalvia
167	clingfish family	Gobiesocidae
KLPFM	clinid family	Clinidae
SOLCO	C-O sole	Pleuronichthys coenosus
SCCRG	coastrangesculpin	Cottus aleuticus
CODFM	cod family	Gadidae
SALCO	coho salmon	Oncorhynchus kisutch
FLNFM	combtooth blenny family	Blenniidae
CARPC	common carp	Cyprinus carpio
RFCOP	copper rockfish	Sebastes caurinus
326	corralline sculpin	Artedius corallinus
CSHFM	cow shark family	Hexanchidae
RFCOW	cowcod	Sebastes levis
GUNCR	crescentgunnel	Pholis laeta
329	crested sculpin	Blepsias bilobus
216	crestfish	Lophotus lacepedei
KLPCR	crevice kelpfish	Gibbonsia montereyensis
CRUST	crustacean subphylum	Crustacea
SOLCF	curlfin sole	Pleuronichthys decurrens
CSKFM	cusk eel family	Ophidiidae
SALCT	cutthroat trout	Oncorhynchus clarki
148	daggertooth family	Anotopteridae
DAMFM	damselfish family	Pomacentridae
RFDBL	darkblotched rockfish	Sebastes crameri
376	darter sculpin	Radulinus boleoides
584	daubed shanny	Lumpenus maculatus
578	decorated warbonnet	Chirolophis decoratus

Sorted by AFS Common Name		
SP CODE	AFS COMMON NAME	SCIENTIFIC NAME
ANCDB	deepbody anchovy	Anchoa compressa
DSSFM	deepsea smelt family	Bathylagidae
SOLDS	deepseasole	Embassichthys bathybius
564	deepwater blenny	Crypotrema corallinum
129	delta smelt	Hypomesus transpacificus
SGDIA	diamond stingray	Dasyatis dipterura
SOLDT	diamond turbot	Pleuronicthys guttulatus
158	diogenes lampfish	Diogenys lanternatus
SHDFM	dogfish shark family	Squalidae
152	dogtooth lampfish	Ceratoscopelus townsendi
SALDV	Dolly Varden	Salvelinus malma
474	dolphin family	Coryphaenidae
DRADO	dolphinfish	Coryphaena hippurus
SOLDV	Doversole	Microstomus pacificus
DRGFM	dragonfish family	Stomiidae
DRMFM	drum family	Sciaenidae
CRBDG	Dungeness crab	Metacarcinus magister
RFDUS	dusky rockfish	Sebastes ciliatus
SCDSK	dusky sculpin	Icelinus burchani
SHDKY	dusky shark	Carcharhinus obscurus
SPDWF	dwarf perch	Micrometrus minimus
293	dwarf red rockfish	Sebastes rufinanus
593	dwarf wrymouth	Cryptacanthodes aleutensis
EELOR	eel order	Anguilliformes
ELPFM	eelpout family	Zoarcidae
SOLEG	English sole	Parophrys vetulus
627	escolar	Lepidocybium flavobrunneum
SMEUL	eulachon	Thaleichthys pacificus
369	eyeshode sculpin	Nautichthys pribilovius
SOLFT	fantail sole	Xystreurys liolepis
35	filetail cat shark	Parmatyrus xaniurus
FTRIG	finescale triggerfish	Balistes polylepis
RFFLG	flag rockfish	Sebastes rubrivinctus
159	flashlightfish	Protomyctophum crockeri
FLTOR	flatfish order	Pleuronectiformes
72	flathead skate	Bathyraja rosispinis
SOLFH	flathead sole	Hippoglossoides elassodon
107	flatiron herring	Harengula thrissina
372	fluffy sculpin	Oligocottus snyderi
FLYFM	flyingfish family	Exocoetidae
407	fourhorn poacher	Hypsagonus quadricornis
365	fourhorn sculpin	Myoxocephalus quadricornis
594	fourline snakeblenny	Eumesogrammus praecisus
RFFRK	freckled rockfish	Sebastes lentiginosus
MACFR	frigate mackerel	Auxis thazard

Sorted by AFS Common Name		
SP CODE	AFS COMMON NAME	SCIENTIFIC NAME
18	frill shark	Chlamydoselachus arguineus
FRSFM	frill shark family	Chlamydoselachidae
356	fringed sculpin	Icelinus fimbriatus
354	frogmouth sculpin	Icelinus oculatus
469	gafftopsail pompano	Trachinotus rhodopus
CLMGP	gaper clam	Tresus nuttallii
GARIB	garibaldi	Hypsypops rubicundus
427	gelatinous seasnail	Liparis fabricii
CLMGD	geoduck, clam	Panopea generosa
719	giantgrenadier	Albatrossia pectoralis
KLPGT	giantkelpfish	Heterostichus rostratus
SCPRO	giant rock scallop	Crassadoma gigantea
GNTSB	giantseabass	Stereolepis gigas
592	giant wrymouth	Cryptacanthodes giganteus
GOBFM	goby family	Gobiidae
RFGOP	gopher rockfish	Sebastes carnatus
CRBGR	graceful rock crab	Cancer gracilis
RFGRS	grass rockfish	Sebastes rastrelliger
607	graveldiver	Scytalina cerdale
44	gray shark genus	Carcharhinus
SHGSM	gray smooth hound	Mustelus californicus
402	gray starsnout	Bathyagonus alascanus
SCGRT	great sculpin	Myoxocephalus
		polyacanthocep
463	green jack	Caranx caballus
STGRN	green sturgeon	Acipenser medirostris
RFGBL	greenblotchedrockfish	Sebastes rosenblatti
HALGL	Greenland halibut	Reinhardtius hippoglossoides
GRNFM	greenling family	Hexagrammidae
GRNGN	greenlinggenus	Hexagrammos
RFGRN	greenspotted rockfish	Sebastes chlorostictus
RFGST	greenstriped rockfish	Sebastes elongatus
715	grenadier family	Macrouridae
GRPGN	grouper,genus	Epinephelus
	(epinephelus)	
GNTFM	gruntfamily	Haemulidae
SCGRU	gruntsculpin	Rhamphocottus richardsoni
453	Guadalupe cardinalfish	Apogon guadalupensis
GUIFM	guitarfish family	Rhinobatidae
GRPGC	gulfconey	Hyporthodus acanthistius
GRPGF	gulfgrouper	Mycteroperca jordani
SRAGU	gulfsierra	Scomberomorus concolor
GUNFM	gunnel family	Pholidae
HAGFM	hagfish order	Myxinidae
RFHBD	halfbanded rockfish	Sebastes semicinctus

Sorted by AFS Common Name		
SP CODE	AFS COMMON NAME	SCIENTIFIC NAME
202	halfbeak	Hyporhamphus unifasciatus
203	halfbeak	Hyporhamphus rosae
617	halfblind goby	Lethops connetens
HALFM	halfmoon	Medialuna californiensis
320	hamecon	Artediellus scaber
50	hammerhead shark	Sphyrnidae
	family	
707	harlequin rockfish	Sebastes variegatus
HERFM	herring family	Clupeidae
575	high cockscomb	Anoplarchus purpurescens
RFHNC	honeycomb rockfish	Sebastes umbrosus
712	hookthroat bass	Hemanthias signifer
SHHRN	horn shark	Heterodontus francisci
SOLHT	hornyhead turbot	Pleuronichthys verticalis
716	hybrid soles	Isopsetta
SCILG	Irish lord genus	Hemilepidotus
562	island kelpfish	Alloclinus holderi
JACFM	jack family	Carangidae
JACMK	jack mackerel	Trachurus symmetricus
SMJAK	jacksmelt	Atherinopsis californiensis
SQDJU	jumbo squid	
FLRKM	Kamchatka flounder	Atheresthes evermanni
KAWAK	kawakawa	Euthynnus affinis
SBKLP	kelp bass	Paralabrax clathratus
172	kelp clingfish	Rimicola muscarum
GRNKP	kelp greenling	Hexagrammos decagrammus
606	kelp gunnel	Ulvicola santaerosea
SPKLP	kelp perch	Brachyistius frenatus
230	kelp pipefish	Syngnathus californiensis
RFKLP	kelp rockfish	Sebastes atrovirens
380	kelp sculpin	Sigmistes caulias
KOSAL	king-of-the-salmon	Trachipterus altivelis
LMPFM	lamprey family	Petromyzontidae
146	lancetfish family	Alepisauridae
151	lanternfish family	Myctophidae
390	lavender sculpin	Leiocottus hirundo
419	leatherfin lumpsucker	Eumicrotremus derjugini
465	leatherjacket	Oligoplites saurus
FLLFN	lefteye flounder family	Bothidae
SCLST	leister sculpin	Enophrys lucasi
SHLEP	leopard shark	Triakis semifasciata
572	lesser prickleback	Alectridium aurantiacum
169	lined clingfish	Gobiesox eugrammus
LNGCD	lingcod	Ophiodon elongatus
CLMLN	littleneck clam	Protothaca staminea

Sorted by AFS Common Name		
SP CODE		SCIENTIFIC NAME
LZDFM	lizardfish family	Synodontidae
434	lobefin snailfish	Liparis greeni
656	longfin cigarfish	Cubiceps paradoxus
142	longfin dragonfish	Tactostoma macropus
599	longfin gunnel	Pholis clemensi
201	longfin halfbeak	Hemiramphus saltator
DABLF	longfin sanddab	Citharlchthys xanthostigma
389	longfin sculpin	Jordani zonope
SMLGF	longfin smelt	Spirinchus thlaeichthys
680	longhead dab	Pleuronectes proboscideus
LJMUD	longjaw mudsucker	Gillichthus mirabilis
33	longnose cat shark	Apristurus kampae
LANLN	longnose lancetfish	Alepisaurus ferox
SKLGN	longnose skate	Raja rhina
581	Iongsnoutprickleback	Lumpenella longirostris
CBFLS	longspine combfish	Zaniolepis latipinnis
RFLST	longspine thornyhead	Sebastolobus altivelis
LUVAR	louvar	Luvarus imperialis
SERLT	lumptail searobin	Prionotus stephanophrys
90	machete	Elops affinis
MACFM	mackerel family	Scombridae
SHMFM	mackerel shark family	Lamnidae
MANTA	manta	Manta birostris
82	manta family	Mobulidae
424	marbled snailfish	Liparis dennyi
458	marlin sucker	Remora osteochir
SQDMK	market squid	Doryteuthis opalescens
GRNMA	masked greenling	Hexagrammos octogrammus
576	matcheek warbonnet	Chirolophis tarsodes
495	Mexican goatfish	Mulloidichthys dentatus
160	Mexican lampfish	Triphoturus mexicanus
RFMEX	Mexican rockfish	Sebastes macdonaldi
MSCAD	Mexican scad	Decapterus scombrinus
106	middling thread herring	Opisthonema medirastre
MIDGN	midshipman genus	Porichthys
214	mirror dory	Zenopsis nebulosa
MOJFM	mojarra family	Gerreidae
SUNFM	mola family	Molidae
MOLLU	mollusk phylum	Mollusca
382	monacled sculpin	Synchirus gilli
PRKMK	monkeyface prickleback	Cebidichthys violaceus
333	mosshead sculpin	Clinocottus glopiceps
577	mosshead warbonnet	Chirolophis nugator
554	mussel blenny	Hypsoblennius jenkinsi
SHNTH	narrowtooth shark	Carcharhinus brachyurus

Sorted by AFS Common Name		
SP CODE	AFS COMMON NAME	SCIENTIFIC NAME
SMNGT	nightsmelt	Spirinchus starksi
224	ninespine stickleback	Pungitius pungitius
ANCNO	northern anchovy	Engraulis mordax
156	northern lampfish	Stenobrachius leucopsarus
150	northern pearleye	Benthalbella dentata
RNQNO	northern ronquil	Rongilus jordani
SCNTH	northern sculpin	Icelinus borealis
397	northern spearnose	Agonopsis vulsa
	poacher	
CLNGN	nothern clingfish	Gobiesox maeandricus
579	nutcracker prickleback	Bryozoichthys lysimus
221	oarfish	Regalecus glesne
SUNOC	ocean sunfish	Mola mola
OCWHT	ocean whitefish	Caulolatilus princeps
699	oceanic puffer	Lagocephalus lagocephalus
OCTOP	octopus order	Octopoda
628	oilfish	Ruvettus pretiosus
RFOLV	olive rockfish	Sebastes serranoides
KLPOF	onespot fringehead	Neoclinus urinotatus
OPAHS	opah	Lampris guttatus
OPALE	opaleye	Girella nigricans
COROM	orangemouth corvina	Cynoscion xanthulus
563	orangethroat pikeblenny	Chaenopsis alepidota
466	Pacific amberjack	Seriola colburni
SHANG	Pacific angel shark	Squatina californica
ARGNT	Pacific argentine	Argentina sialis
BARPA	Pacific barracuda	Sphyraena argentea
BONPA	Pacific bonito	Sarda chiliensis
464	Pacific bumper	Chloroscombrus orqueta
701	Pacific burrfish	Chilomycterus affinis
CODPA	Pacific cod	Gadus macrocephalus
CUTLP	Pacific cutlassfish	Trichiurus nitens
ERYPA	Pacific electric ray	Torpedo californica
531	Pacific fanfish	Pteraclis aesticola
623	Pacific fat sleeper	Dormitator latifrons
479	Pacific flagfin mojarra	Eucinostomus gracilis
709	Pacific flatnose	Antimora microlepis
706	Pacific grenadier	Coryphaenoides acrolepis
TFPGE	Pacific golden-eyed	Caulolatilus affinis
1140004	tilefish	
HAGPA	Pacific hagfish	Eptatretus stouti
PHAKE	Pacific hake	Merluccius productus
HALPA	Pacific halibut	Hippoglossus stenolepis
HERPA	Pacific herring	Clupea pallasi
LMPPA	Pacific lamprey	Entosphenus tridentatus

Sorted by AFS Common Name		
SP CODE	AFS COMMON NAME	SCIENTIFIC NAME
470	Pacific moonfish	Selene peruviana
RFPOP	Pacific ocean perch	Sebastes alutus
528	Pacific pomfret	Brama japonica
POMPA	Pacific pompano	Peprilus simillimus
	(butterfish)	
483	Pacific porgy	Calamus brachysomus
SOLPA	Pacific sand lance	Ammodytes hexapterus
DABPA	Pacific sanddab	Citharichthys sordidus
SNDPA	Pacific sandfish	Trichodon trichodon
SARPA	Pacific sardine	Sardinops sagax
SAUPA	Pacific saury	Cololabis saira
630	Pacific scabbardfish	Lepidopus fitchi
231	Pacific seahorse	Hippocampus ingens
39	Pacific sharpnose shark	Rhizoprionodon longurio
SRAPA	Pacific sierra	Scomberomorus sierra
SHSLP	Pacific sleeper shark	Somniosus pacificus
97	Pacific snake eel	Ophichthus triserialis
SPDPA	Pacific spadefish	Chaetodipterus zonatus
420	Pacific spiny lumpsucker	Eumicrotremus orbis
SCPSH	Pacific staghorn sculpin	Leptocottus armatus
CODTC	Pacific tomcod	Microgadus proximus
TRTPA	Pacific tripletail	Lobotes pacificus
143	Pacific viperfish	Chauliodus macouni
96	Pacific worm eel	Myrophis vafer
SCPAD	padded sculpin	Artedius fenestralis
GRNPT	painted greenling	Oxylebius pictus
315	painted greenling	Oxylebius pictus
196	pale eelpout	Lycodes pallidus
189	pallid eelpout	Lycodapus mandibularis
468	palomapompano	Trachinotus paitensis
155	patchwork lampfish	Notoscopelus resplendens
149	pearleye family	Scopelarchidae
504	pelagic armorhead	Pentaceros richardsoni
CRBPR	pelagic red crab	Pleuroncodes palnipes
SGPEL	pelagic stingray	Dasyatis violacea
GUNPP	penpoint gunnel	Apodichthys flavidus
PERFM	perch family	Percidae
SOLPT	petrale sole	Eopsetta jordani
571	pighead prickleback	Acantholumpenus mackayi
SPPIL	pile perch	Rhacochilus vacca
PILTF	pilotfish	Naucrates ductor
RFPNK	pink rockfish	Sebastes eos
SALPK	pink salmon	Oncorhynchus gorbuscha
SPPNK	pink seaperch	Zalembius rosaceus
RFPRS	pinkrose rockfish	Sebastes simulator

Sorted by AFS Common Name		
SP CODE	AFS COMMON NAME	SCIENTIFIC NAME
154	pinpoint lampfish	Lampanyctus regalis
227	pipefish family	Sygnathidae
CLMPO	pismo clam	Tivela stultorum
355	pit head sculpin	Icelinus cavifrons
362	plain sculpin	Myoxocephalus jaok
MIDPF	plainfin midshipman	Porichthys notatus
396	poacher family	Agonidae
194	polar eelpout	Lycodes turneri
423	polkadotsnailfish	Liparis cyclostigma
POMFM	pomfret family	Bramidae
POMDO	pompano dolphin	Coryphaena equisetis
CTFPE	popeye catalufa	Pristigenys serrula
702	porcupinefish	Diodon hystrix
PRKFM	prickleback family	Stichaeidae
414	pricklebreastpoacher	Stellerina xyosterna
SCPRK	prickly sculpin	Cottus asper
56	prickly shark	Echinorhinus cookei
432	prickly snailfish	Paraliparis deani
608	prowfish	Zaprora silenus
PUFFM	puffer family	Tetraodontidae
RFPSD	Puget Sound rockfish	Sebastes emphaeus
324	Puget Sound sculpin	Ruscarius meanyi
410	pygmy poacher	Odontopyxis trispinosa
RFPYG	pygmy rockfish	Sebastes wilsoni
448	pygmy seabass	Serraniculus pumilio
QUEEN	queenfish	Seriphus politus
RFQIL	quillback rockfish	Sebastes maliger
569	quillfish	Ptilichthys goodei
RAGFS	ragfish	Icosteus aenagmaticus
23	ragged tooth shark	Odontaspis ferox
SCRRB	rainbowscorpionfish	Scorpaenodes xyris
SPRBW	rainbow seaperch	Hypsurus caryi
131	rainbow smelt	Osmerus mordax
SALRB	rainbow trout	Oncorhynchus mykiss
WRARB	rainbow wrasse	Thalassoma luvasanum
SCBRZ	razorback scabbardfish	Assurger anzac
CLMNR	razor clam	Siliqua patula
183	red brotula	Brosmophycis marginata
604	red gunnel	Pholis schultzi
SCRIL	red Irish lord	Hemilepidotus hemilepidotus
CRBRR	red rock crab	Cancer productus
RFRBD	redbanded rockfish	Sebastes babcocki
RFRST	redstripe rockfish	Sebastes proriger
SPRTL	redtail surfperch	Amphistichus rhodoterus
KLPRB	reef blenny	Paraclinus integripinnis

Sorted by AFS Common Name		
SP CODE	AFS COMMON NAME	SCIENTIFIC NAME
SPREF	reef perch	Micrometrus aurora
459	remora	Remora remora
REMFM	remora family	Echeneidae
SHRFM	requiem shark family	Carcharhinidae
SOLRX	rex sole	Glyptocephalus zachirus
385	ribbed sculpin	Triglops pingeli
204	ribbon halfbeak	Euleptorhamphus viridis
586	ribbon prickleback	Phytichthys chirus
422	ribbon snailfish	Liparis cyclopus
217	ribbonfish family	Trachipteridae
FLRFM	righteye flounder family	Pleuronectidae
430	ringtail snailfish	Liparis rutteri
GRNRK	rock greenling	Hexagrammos lagocephalus
PRKRK	rock prickleback	Xiphister mucosus
SOLRK	rock sole	Lepidopsetta bilineatus
WRARK	rock wrasse	Halichoeres semicinctus
RFGEN	rockfish genus	Sebastes
ROCKH	rockhead	Bothragonus swani
BLNRP	rockpool blenny	Hypsoblennius gilberti
605	rockweed gunnel	Apodichthys fucorum
RNQFM	ronquil family	Bathymasteridae
473	roosterfish	Nematistius pectoralis
RFRTN	rosethorn rockfish	Sebastes helvomaculatus
RFROS	rosy rockfish	Sebastes rosaceus
SCRSL	rosylip sculpin	Ascelichthys rhodorus
530	rough pomfret	Teractes asper
387	roughback sculpin	Chitonotus pugettensis
327	roughcheek sculpin	Ruscarius creaseri
RFRGH	rougheye rockfish	Sebastes aleutianus
174	roughjaw frogfish	Antennarius avalonis
384	roughspinesculpin	Triglops macellus
74	roughtail skate	Raja trachura
HERRD	round herring	Etrumeus teres
SGRND	round stingray	Urolophus halleri
SPRUB	rubberlip seaperch	Rhacochilus toxotes
SABLE	sablefish	Anoplopoma fimbria
SABFM	sablefish family	Anoplopomatidae
GUNSB	saddleback gunnel	Pholis ornata
371	saddleback sculpin	Oligocottus rimensis
543	sailfin sandfish	Arctoscopus japonicus
SCSFN	sailfin sculpin	Nautichthys oculofasciatus
SAILF	sailfish	Istiophorus platypterus
SALEM	salema	Xenistius californiensis
SALFM	salmon family	Salmonidae
SALGN	salmon genus	Oncorhynchus spp.

Sorted by AFS Common Name		
SP CODE	AFS COMMON NAME	SCIENTIFIC NAME
SHSAL	salmon shark	Lamna ditropis
SOLSD	sand sole	Psettichthys melanostictus
SBGEN	sandbass genus	Paralabrax
DABGN	sanddab genus	Citharichthys
SNDFM	sandfish family	Trichodontidae
68	sandpaper skate	Bathyraja interrupta
KLPSF	sarcastic fringehead	Neoclinus blanchardi
SARGO	sargo	Anisotremus davidsoni
SCSCL	scaled sculpin	Archaulus biseriatus
220	scalloped ribbonfish	Zu cristatus
SCPUS	scallops	Pectinidae
SCSLH	scalyhead sculpin	Artedius harringtoni
560	scarlet kelpfish	Gibbonsia erythra
SCSCT	scissortail sculpin	Triglops forficata
SCRFM	scorpionfish family	Scorpaenidae
SCFAM	sculpin family	Cottidae
503	scythe butterflyfish	Chaetodon falcifer
SBFAM	sea bass family	Serranidae
SCBFM	sea chub family	Kyphosidae
CUCUM	sea cucumber class	Holothuroidea
SALTR	sea run trouts	
SSTAR	sea star	Asterzoa
GAPOD	sea slug, sea snail	Gastropoda
URCHN	sea urchin family	Diadematidae
548	searcher	Bathymaster signatus
298	searobin family	Triglidae
708	semaphore rockfish	Sebastes melanosema
SENOR	senorita	Oxyjulis californica
SHSEV	seven gill shark	Notorynchus maculatus
619	shadow goby	Quietula ycauda
205	sharpchin flyingfish	Fodiator acutus
RFSCN	sharpchin rockfish	Sebastes zacentrus
SCSHN	sharpnose sculpin	Clinocottus acuticeps
SPSHN	sharpnose seaperch	Phanerodon atripes
CRBSH	sheep crab	Loxorhynchus grandis
SPSHR	shiner perch	Cymatogaster aggregata
RFSHB	shortbelly rockfish	Sebastes jordani
654	shortbill spearfish	Tetrapturus angustirostris
CORSF	shortfin corvina	Cynoscion parvipinnis
190	shortfin eelpout	Lycodes brevipes
SHSMK	shortfin mako shark	Isurus oxyrinchus
367	shorthorn sculpin	Myoxocephalus scorpius
RFSRK	shortraker rockfish	Sebastes borealis
CBFSS	shortspine combfish	Zaniolepis frenata
RFSST	shortspine thomyhead	Sebastolobus alascanus

Sorted by AFS Common Name		
SP CODE		SCIENTIFIC NAME
GUISN	shovelnose guitarfish	Rhinobatos productus
429	showy snailfish	Liparis pulchellus
SHRMP	shrimp	Caridea
195	shulupaoluk	Lycodes jugoricus
532	sickle pomfret	Taractichthys steindachneri
SHSSM	sicklefin smoothhound	Mustelus lunulatus
330	silver spotted sculpin	Blepsias cirrhosus
SPSIL	silver surfperch	Hyperprosopon ellipticum
RFSLG	silvergray rockfish	Sebastes brevispinis
SVRFM	silverside family	Atherinidae
RAJOR	skate and ray order	Rajiformes
SKFAM	skate family	Rajidae
314	skilfish	Erilepis zonifer
SKBGN	skipback genus	Euthynnus
TNASJ	skipjack tuna	Euthynnus pelamis
173	slender clingfish	Rimicola eigenmanni
574	slender cockscomb	Anoplarchus insignis
582	slender eelblenny	Lumpenus fabricii
705	slender mola	Ranzanic laevis
CRBGR	slender rockcrab	Cancer gracillis
100	slender snake eel	Nemichthys scolopaceus
226	slendersnipefish	Macrorhamphosus gracilis
SOLSL	slendersole	Lyopsetta exilis
TNASL	slendertuna	Allothunnus fallai
375	slimsculpin	Radulinus asprellus
426	slipskin snailfish	Liparis fucensis
112	slough anchovy	Anchoa delicatissima
SQTSE	smalleye squaretail	Tetragonurus cuvieri
SMFAM	smelt family	Osmeridae
381	smithi sculpin	Sigmistes smithi
399	smooth alligatorfish	Anoplagonus inermis
52	smooth hammerhead	Sphyrna zygaena
	shark	
416	smooth lumpsucker	Aptocyclus ventricosus
550	smooth stargazer	Kathetostoma averruncus
377	smoothgumsculpin	Radulinus vinculus
323	smoothhead sculpin	Artedius lateralis
SHSGN	smoothhound genus	Mustelus
85	smoothtail mobula	Mobula thurstoni
415	snailfish family	Cyclopteridae
SELFM	snake eel family	Ophichthidae
626	snake mackerel	Gempylus serpens
625	snake mackerel family	Trichiuridae
PRKSN	snake prickleback	Lumpenus sagitta
99	snipe eel family	Nemichthyidae

Sorted by AFS Common Name SP CODE AFS COMMON NAME 392 snubnose sculpin Orthoropias triacis SALSE sockeye salmon Oncorhynchus nerka 395 soft sculpin Psychrolutes sigalutes SHFIN soupfin shark Galeorhinus zyopterus 398 Southern Agonopsis sterletus 398 Southern Agonopsis sterletus 398 Southern Agonopsis sterletus 398 Southern Agonopsis sterletus 317 spatulate sculpin Icelus spatula 460 spearfish remora Remora brachyptera RFSPK speckled rockfish Sebastes ovalis DABSP speckled sanddab Citharichthys stigmaeus MIDSP specklefin midshipman Porichthys myriaster 386 spectacled sculpin Triglops scepticus 374 spineless sculpin Phallocottus obtusus 84 spinetail mobula Mobula japanica BOXSP spiny boxfish Ostracion diaphanum SHSDG spiny dogfish shark Squalus acanthias LOBSP spiny lobster Panulirus interruptus 428 spiny snailfish Liparis mucosus 403 spinycheck starsnout Bathyagonus infraspinatus 338 spinynose sculpin Dasycottus setiger 388 spinynose sculpin Asemichthys taylori RFSNS splitnose rockfish Sebastes diploproa 300 splitnose searobin Bellator xenisma 442 splittail bass Hemanthias perunanus 139 spookfish family Opisthoproctidae CRKSF spotfin croaker Roncador stearnsi 478 spotfin sculpin Icelinus tenuis
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SCSPT spotfin sculpin Icelinus tenuis
SPSPF spotfin surfperch Hyperprosopon anale
175 spotted batfish Zalieutes elater
GRPSC spotted cabrilla Epinephelus analogus
184 spotted cusk eel <i>Chilara taylori</i>
KLPSP spotted kelpfish Gibbonsia elegans
RATFS spotted ratfish Hydrolagus colliei
SBSPT spotted sandbass Paralabrax maculatofascia
421 spotted snailfish Liparis callyodon
SOLST spotted turbot Pleuronichthys ritteri
RFSQS squarespot rockfish Sebastes hopkinsi
SQUID squid class Cephalopoda
FLRST starry flounder Platichthys stellatus
RFSTA starry rockfish Sebastes constellatus
SKSTY starry skate Raja stellulata
GRPSS star-studded grouper Hyporthodus niphobles
SKBFM stickleback family Gasterosteidae

Sorted by AFS Common Name		
SP CODE	AFS COMMON NAME	SCIENTIFIC NAME
SGFAM	stingray family	Dasyatidae
SGGEN	stingray genus	Dasyatis spp.
600	stippled gunnel	Rhodymenichthys
		dolichogaster
595	stone cockscomb	Alectrias alectrolophus
585	stout eelblenny	Lumpenus medius
STBAS	striped bass	Morone saxatilis
KLPST	striped kelpfish	Gibbonsia metzi
MARST	striped marlin	Tetrapturus audax
STMUL	striped mullet	Mugil cephalus
SPSTR	striped seaperch	Embiotoca lateralis
RFSTR	stripetail rockfish	Sebastes saxicola
STGEN	sturgeon genus	Acipenser
SNFFM	sunfish family	Centrarchidae
SMSUR	surf smelt	Hypomesus pretiosus
SPFAM	surfperch family	Embiotocidae
SHSWL	swell shark	Cephaloscyllium ventriosum
SRDFS	swordfish	Xiphias gladius
RFSDS	swordspine rockfish	Sebastes ensifer
393	tadpolesculpin	Psychrolutes paradoxus
431	tadpolesnailfish	Nectoliparis pelagicus
219	tapertail ribbonfish	Trachipterus fukuzaki
THRBK	thornback	Platyrhinoidis triseriata
373	thornback sculpin	Paricelinus hopliticus
SBTHF	threadfin bass	Pronotogrammus multifasciatus
535	threadfin family	Polynemidae
SCTRF	threadfin sculpin	Icelinus filamentosus
502	threeband butterflyfish	Chaetodon humeralis
SKBTS	threespine stickleback	Gasterosteus aculeatus
SHTHR	threshershark	Alopias vulpinus
SCTDP	tidepool sculpin	Oligocottus maculosus
425	tidepool snailfish	Liparis florae
622	tidewater goby	Eucyclogobius newberryi
RFTIG	tigerrockfish	Sebastes nigrocinctus
SHTIG	tiger shark	Galeocerdo cuvieri
SMTOP	topsmelt	Atherinops affinis
RFTRE	treefish	Sebastes serriceps
580	trident prickleback	Gymnoclinus cristulatus
176	triplewart seadevil	Cryptopsaras couesi
CRABS	true crabs	Brachyuratribe
TBESN	tube snout	Aulorhynchus flavidus
411	tubenose poacher	Pallasina barbata
TNASG	tunas (non-mackerel)	
316	twohorn sculpin	Icelus bicornis

SHINS unidentified inshore sharks SHOFF Unidentified offshore sharks UNISF unidentified (surface fish) UNISF unidentified (surface fish) UNIFH unidentified fish RFVER vermilion rockfish Sebastes miniatus WAHOO wahoo Acanthocybium solandri POLWE walleye pollock Theragra chalcogramma SPWAL walleye surfperch Hyperprosopon argenteum 363 warthead sculpin Myoxocephalus niger 409 warty poacher Occella verrucosa CLIMWA washington, clam Saxidomus nuttalli 192 wattled eelpout Lycodes palearis WEKFS weakfishes Cynoscion 22 whale shark Rhincodon typus REMWS whalesucker Remora australis CROWT white croaker Genyonemus lineatus SBWHT white seabass Atractoscion nobilis SPWHT white shark Carcharodon carcharias STWHT white sturgeon Acipenser transmontanus SMWTB whitebalt smelt Allosmerus elongatus 588 whitebarred prickleback Poroclinus rothrocki RFWTB whitebelly rockfish Sebastes vexillaris GRNWT whitespotted greenling Hexagrammos stelleri RFWID widow rockfish Sebastes vexillaris GRNWT whitespotted greenling Hexagrammos stelleri RFWID widow rockfish Sebastes vexillaris SWOLFE wolf-eel Anarrhichthys ocellatus SCWOL wolly sculpin Clinocottus analis WRAFM wrasse family Labridae SELYL yellow bobo Polydactylus opercularis SFYEY yellow rock crab Cancer anthonyi SELYL yellowfin sculpin Icelinus quadriseriatus RFYEY yellowfin roaker Umbrina roncador SEMST Sebastes ruberrimus CRKYF yellowfin roaker Umbrina roncador SEMST Sebastes ruberrimus CRKYF yellowfin roaker Umbrina roncador SEMST Sebastes ruberrimus CRKYF yellowfin roaker Umbrina roncador SEMST Sebastes rediiny Hexagramous SOLYF yellowfin sole Limanda aspera Thannus albacares RFYMN yellowmouth rockfish Sebastes redii YELTL yellowtail Seriola lalandi	Sorted by A	AFS Common Name	
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WEKFS weakfishes Cynoscion 22 whale shark Rhincodon typus REMWS whalesucker Remora australis CROWT white croaker Genyonemus lineatus SBWHT white seabass Atractoscion nobilis SPWHT white seaperch Phanerodon furcatus SHWHT white shark Carcharodon carcharias STWHT white sturgeon Acipenser transmontanus SMWTB whitebait smelt Allosmerus elongatus 588 whitebarred prickleback Poroclinus rothrocki RFWTB whitespotted greenling Hexagrammos stelleri RFWID widowrockfish Sebastes vexillaris GRNWT whitespotted greenling Hexagrammos stelleri RFWID widowrockfish Sebastes entomelas WOLFE wolf-eel Anarrhichthys ocellatus SCWOL wolly sculpin Clinocottus analis WRAFM wrasse family Labridae 537 yellow bobo Polydactylus opercularis 347 yellow lrish lord Hemilepidotus jordani CRBYR yellow rock crab Cancer anthonyi SELYL yellow snake eel Ophichthus zophochir 357 yellowchin sculpin Icelinus quadriseriatus RFYEY yellowfin croaker Umbrina roncador 566 yellowfin fringehead Neoclinus stephensae BOGYL yellowfin sole Limanda aspera TNAYF yellowfin tuna Thunnus albacares RFYMN yellowtail Seriola lalandi	CLMWA	washington, clam	Saxidomus nuttalli
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REMWS whalesucker Remora australis CROWT white croaker Genyonemus lineatus SBWHT white seabass Atractoscion nobilis SPWHT white seaperch Phanerodon furcatus SHWHT white shark Carcharodon carcharias STWHT white sturgeon Acipenser transmontanus SMWTB whitebait smelt Allosmerus elongatus 588 whitebarred prickleback Poroclinus rothrocki RFWTB whitebelly rockfish Sebastes vexillaris GRNWT whitespotted greenling Hexagrammos stelleri RFWID widow rockfish Sebastes entomelas WOLFE wolf-eel Anarrhichthys ocellatus SCWOL wolly sculpin Clinocottus analis WRAFM wrasse family Labridae 573 Y prickleback Allolumpenus hypochrcmus 537 yellow bobo Polydactylus opercularis 347 yellow Irish Iord Hemilepidotus jordani CRBYR yellow rock crab Cancer anthonyi SELYL yellow snake eel Ophichthus zophochir 357 yellowchin sculpin Icelinus quadriseriatus RFYEY yelloweye rockfish Sebastes ruberrimus CRKYF yellowfin fringehead Neoclinus stephensae BOGYL yellowfin sole Limanda aspera TNAYF yellowmouth rockfish Sebastes reedi YELTL yellowtail Seriola lalandi	WEKFS	weakfishes	Cynoscion
CROWT white croaker Genyonemus lineatus SBWHT white seabass Atractoscion nobilis SPWHT white seaperch Phanerodon furcatus SHWHT white shark Carcharodon carcharias STWHT white sturgeon Acipenser transmontanus SMWTB whitebait smelt Allosmerus elongatus 588 whitebarred prickleback Poroclinus rothrocki RFWTB whitebelly rockfish Sebastes vexillaris GRNWT whitespotted greenling Hexagrammos stelleri RFWID widow rockfish Sebastes entomelas WOLFE wolf-eel Anarrhichthys ocellatus SCWOL wolly sculpin Clinocottus analis WRAFM wrasse family Labridae 573 Y prickleback Allolumpenus hypochrcmus 537 yellow bobo Polydactylus opercularis 347 yellow lrish lord Hemilepidotus jordani CRBYR yellow rock crab Cancer anthonyi SELYL yellowsnake eel Ophichthus zophochir 357 yellowchin sculpin Icelinus quadriseriatus RFYEY yellowfin croaker Umbrina roncador 566 yellowfin fringehead Neoclinus stephensae BOGYL yellowfin sole Limanda aspera TNAYF yellowmouth rockfish Sebastes reedi YELTL yellowtail Seriola lalandi		whale shark	Rhincodon typus
SBWHT white seabass Atractoscion nobilis SPWHT white seaperch Phanerodon furcatus SHWHT white shark Carcharodon carcharias STWHT white sturgeon Acipenser transmontanus SMWTB whitebait smelt Allosmerus elongatus 588 whitebarred prickleback Poroclinus rothrocki RFWTB whitebelly rockfish Sebastes vexillaris GRNWT whitespotted greenling Hexagrammos stelleri RFWID widow rockfish Sebastes entomelas WOLFE wolf-eel Anarrhichthys ocellatus SCWOL wolly sculpin Clinocottus analis WRAFM wrasse family Labridae 573 Y prickleback Allolumpenus hypochrcmus 537 yellow bobo Polydactylus opercularis 347 yellow lrish lord Hemilepidotus jordani CRBYR yellow rock crab Cancer anthonyi SELYL yellowsnake eel Ophichthus zophochir 357 yellowchin sculpin Icelinus quadriseriatus RFYEY yellowfin croaker Umbrina roncador 566 yellowfin fringehead Neoclinus stephensae BOGYL yellowfin sole Limanda aspera TNAYF yellowfin tuna Thunnus albacares RFYMN yellowmouth rockfish Sebastes reedi YELTL yellowtail Seriola lalandi	REMWS	whalesucker	Remora australis
SPWHT white seaperch Phanerodon furcatus SHWHT white shark Carcharodon carcharias STWHT white sturgeon Acipenser transmontanus SMWTB whitebait smelt Allosmerus elongatus 588 whitebarred prickleback Poroclinus rothrocki RFWTB whitebelly rockfish Sebastes vexillaris GRNWT whitespotted greenling Hexagrammos stelleri RFWID widow rockfish Sebastes entomelas WOLFE wolf-eel Anarrhichthys ocellatus SCWOL wolly sculpin Clinocottus analis WRAFM wrasse family Labridae 573 Y prickleback Allolumpenus hypochrcmus 537 yellow bobo Polydactylus opercularis 347 yellow Irish Iord Hemilepidotus jordani CRBYR yellow rock crab Cancer anthonyi SELYL yellow snake eel Ophichthus zophochir 357 yellowchin sculpin Icelinus quadriseriatus RFYEY yellowfin croaker Umbrina roncador 566 yellowfin fringehead Neoclinus stephensae BOGYL yellowfin goby Acanthogobius flavimanus SOLYF yellowfin tuna Thunnus albacares RFYMN yellowmouth rockfish Sebastes reedi YELTL yellowtail Seriola lalandi	CROWT	white croaker	Genyonemus lineatus
SHWHT white shark Carcharodon carcharias STWHT white sturgeon Acipenser transmontanus SMWTB whitebait smelt Allosmerus elongatus 588 whitebarred prickleback Poroclinus rothrocki RFWTB whitebelly rockfish Sebastes vexillaris GRNWT whitespotted greenling Hexagrammos stelleri RFWID widow rockfish Sebastes entomelas WOLFE wolf-eel Anarrhichthys ocellatus SCWOL wolly sculpin Clinocottus analis WRAFM wrasse family Labridae 573 Y prickleback Allolumpenus hypochrcmus 537 yellow bobo Polydactylus opercularis 347 yellow Irish Iord Hemilepidotus jordani CRBYR yellow rock crab Cancer anthonyi SELYL yellow snake eel Ophichthus zophochir 357 yellowchin sculpin Icelinus quadriseriatus RFYEY yellowfin croaker Umbrina roncador 566 yellowfin fringehead Neoclinus stephensae BOGYL yellowfin sole Limanda aspera TNAYF yellowmouth rockfish Sebastes reedi YELTL yellowtail Seriola lalandi	SBWHT	white seabass	Atractoscion nobilis
STWHT white sturgeon Acipenser transmontanus SMWTB whitebait smelt Allosmerus elongatus 588 whitebarred prickleback Poroclinus rothrocki RFWTB whitebelly rockfish Sebastes vexillaris GRNWT whitespotted greenling Hexagrammos stelleri RFWID widow rockfish Sebastes entomelas WOLFE wolf-eel Anarrhichthys ocellatus SCWOL wolly sculpin Clinocottus analis WRAFM wrasse family Labridae 573 Y prickleback Allolumpenus hypochrcmus 537 yellow bobo Polydactylus opercularis 347 yellow Irish lord Hemilepidotus jordani CRBYR yellow rock crab Cancer anthonyi SELYL yellow snake eel Ophichthus zophochir 357 yellowchin sculpin Icelinus quadriseriatus RFYEY yelloweye rockfish Sebastes ruberrimus CRKYF yellowfin croaker Umbrina roncador 566 yellowfin fringehead Neoclinus stephensae BOGYL yellowfin sole Limanda aspera TNAYF yellowfin tuna Thunnus albacares RFYMN yellowmouth rockfish Sebastes reedi YELTL yellowtail Seriola lalandi	SPWHT	white seaperch	Phanerodon furcatus
SMWTB whitebait smelt Allosmerus elongatus 588 whitebarred prickleback Poroclinus rothrocki RFWTB whitebelly rockfish Sebastes vexillaris GRNWT whitespotted greenling Hexagrammos stelleri RFWID widow rockfish Sebastes entomelas WOLFE wolf-eel Anarrhichthys ocellatus SCWOL wolly sculpin Clinocottus analis WRAFM wrasse family Labridae 573 Y prickleback Allolumpenus hypochrcmus 537 yellow bobo Polydactylus opercularis 347 yellow Irish lord Hemilepidotus jordani CRBYR yellow rock crab Cancer anthonyi SELYL yellow snake eel Ophichthus zophochir 357 yellowchin sculpin Icelinus quadriseriatus RFYEY yelloweje rockfish Sebastes ruberrimus CRKYF yellowfin goby Acanthogobius flavimanus SOLYF yellowfin tuna Thunnus albacares RFYMN yellowmouth rockfish Sebastes reedi YELTL yellowtail Seriola lalandi	SHWHT	white shark	Carcharodon carcharias
588whitebarred pricklebackPoroclinus rothrockiRFWTBwhitebelly rockfishSebastes vexillarisGRNWTwhitespotted greenlingHexagrammos stelleriRFWIDwidow rockfishSebastes entomelasWOLFEwolf-eelAnarrhichthys ocellatusSCWOLwolly sculpinClinocottus analisWRAFMwrasse familyLabridae573Y pricklebackAllolumpenus hypochrcmus537yellow boboPolydactylus opercularis347yellow lrish lordHemilepidotus jordaniCRBYRyellow rock crabCancer anthonyiSELYLyellow snake eelOphichthus zophochir357yellowchin sculpinIcelinus quadriseriatusRFYEYyelloweye rockfishSebastes ruberrimusCRKYFyellowfin croakerUmbrina roncador566yellowfin fringeheadNeoclinus stephensaeBOGYLyellowfin gobyAcanthogobius flavimanusSOLYFyellowfin tunaThunnus albacaresRFYMNyellowmouth rockfishSebastes reediYELTLyellowtailSeriola lalandi	STWHT	white sturgeon	Acipenser transmontanus
RFWTB whitebelly rockfish Sebastes vexillaris GRNWT whitespotted greenling Hexagrammos stelleri RFWID widow rockfish Sebastes entomelas WOLFE wolf-eel Anarrhichthys ocellatus SCWOL wolly sculpin Clinocottus analis WRAFM wrasse family Labridae 573 Y prickleback Allolumpenus hypochrcmus 537 yellow bobo Polydactylus opercularis 347 yellow lrish lord Hemilepidotus jordani CRBYR yellow rock crab Cancer anthonyi SELYL yellow snake eel Ophichthus zophochir 357 yellowchin sculpin Icelinus quadriseriatus RFYEY yelloweye rockfish Sebastes ruberrimus CRKYF yellowfin croaker Umbrina roncador 566 yellowfin fringehead Neoclinus stephensae BOGYL yellowfin goby Acanthogobius flavimanus SOLYF yellowfin tuna Thunnus albacares RFYMN yellowmouth rockfish Sebastes reedi YELTL yellowtail Seriola lalandi	SMWTB	whitebait smelt	Allosmerus elongatus
GRNWT whitespotted greenling Hexagrammos stelleri RFWID widow rockfish Sebastes entomelas WOLFE wolf-eel Anarrhichthys ocellatus SCWOL wolly sculpin Clinocottus analis WRAFM wrasse family Labridae 573 Y prickleback Allolumpenus hypochrcmus 537 yellow bobo Polydactylus opercularis 347 yellow Irish lord Hemilepidotus jordani CRBYR yellow rock crab Cancer anthonyi SELYL yellow snake eel Ophichthus zophochir 357 yellowchin sculpin Icelinus quadriseriatus RFYEY yelloweye rockfish Sebastes ruberrimus CRKYF yellowfin croaker Umbrina roncador 566 yellowfin fringehead Neoclinus stephensae BOGYL yellowfin goby Acanthogobius flavimanus SOLYF yellowfin tuna Thunnus albacares RFYMN yellowmouth rockfish Sebastes reedi YELTL yellowtail Seriola lalandi	588	whitebarred prickleback	Poroclinus rothrocki
RFWID widowrockfish Sebastes entomelas WOLFE wolf-eel Anarrhichthys ocellatus SCWOL wolly sculpin Clinocottus analis WRAFM wrasse family Labridae 573 Y prickleback Allolumpenus hypochrcmus 537 yellow bobo Polydactylus opercularis 347 yellow Irish lord Hemilepidotus jordani CRBYR yellow rock crab Cancer anthonyi SELYL yellow snake eel Ophichthus zophochir 357 yellowchin sculpin Icelinus quadriseriatus RFYEY yelloweye rockfish Sebastes ruberrimus CRKYF yellowfin croaker Umbrina roncador 566 yellowfin fringehead Neoclinus stephensae BOGYL yellowfin goby Acanthogobius flavimanus SOLYF yellowfin tuna Thunnus albacares RFYMN yellowmouth rockfish Sebastes reedi YELTL yellowtail Seriola lalandi		whitebelly rockfish	Sebastes vexillaris
WOLFE wolf-eel Anarrhichthys ocellatus SCWOL wolly sculpin Clinocottus analis WRAFM wrasse family Labridae 573 Y prickleback Allolumpenus hypochrcmus 537 yellow bobo Polydactylus opercularis 347 yellow Irish lord Hemilepidotus jordani CRBYR yellow rock crab Cancer anthonyi SELYL yellow snake eel Ophichthus zophochir 357 yellowchin sculpin Icelinus quadriseriatus RFYEY yelloweye rockfish Sebastes ruberrimus CRKYF yellowfin croaker Umbrina roncador 566 yellowfin fringehead Neoclinus stephensae BOGYL yellowfin goby Acanthogobius flavimanus SOLYF yellowfin tuna Thunnus albacares RFYMN yellowmouth rockfish Sebastes reedi YELTL yellowtail Seriola lalandi	GRNWT	whitespotted greenling	Hexagrammos stelleri
SCWOL wolly sculpin Clinocottus analis WRAFM wrasse family Labridae 573 Y prickleback Allolumpenus hypochrcmus 537 yellow bobo Polydactylus opercularis 347 yellow Irish Iord Hemilepidotus jordani CRBYR yellow rock crab Cancer anthonyi SELYL yellow snake eel Ophichthus zophochir 357 yellowchin sculpin Icelinus quadriseriatus RFYEY yelloweye rockfish Sebastes ruberrimus CRKYF yellowfin croaker Umbrina roncador 566 yellowfin fringehead Neoclinus stephensae BOGYL yellowfin goby Acanthogobius flavimanus SOLYF yellowfin tuna Thunnus albacares RFYMN yellowmouth rockfish Sebastes reedi YELTL yellowtail Seriola lalandi	RFWID	widow rockfish	Sebastes entomelas
WRAFM wrasse family Labridae 573 Y prickleback Allolumpenus hypochrcmus 537 yellow bobo Polydactylus opercularis 347 yellow Irish lord Hemilepidotus jordani CRBYR yellow rock crab Cancer anthonyi SELYL yellow snake eel Ophichthus zophochir 357 yellowchin sculpin Icelinus quadriseriatus RFYEY yelloweye rockfish Sebastes ruberrimus CRKYF yellowfin croaker Umbrina roncador 566 yellowfin fringehead Neoclinus stephensae BOGYL yellowfin goby Acanthogobius flavimanus SOLYF yellowfin tuna Thunnus albacares RFYMN yellowmouth rockfish Sebastes reedi YELTL yellowtail Seriola lalandi	WOLFE	wolf-eel	Anarrhichthys ocellatus
573 Y prickleback Allolumpenus hypochrcmus 537 yellow bobo Polydactylus opercularis 347 yellow Irish Iord Hemilepidotus jordani CRBYR yellow rock crab Cancer anthonyi SELYL yellow snake eel Ophichthus zophochir 357 yellowchin sculpin Icelinus quadriseriatus RFYEY yelloweye rockfish Sebastes ruberrimus CRKYF yellowfin croaker Umbrina roncador 566 yellowfin fringehead Neoclinus stephensae BOGYL yellowfin goby Acanthogobius flavimanus SOLYF yellowfin tuna Thunnus albacares RFYMN yellowmouth rockfish Sebastes reedi YELTL yellowtail Seriola lalandi	SCWOL	wolly sculpin	Clinocottus analis
537 yellow bobo Polydactylus opercularis 347 yellow Irish Iord Hemilepidotus jordani CRBYR yellow rock crab Cancer anthonyi SELYL yellow snake eel Ophichthus zophochir 357 yellowchin sculpin Icelinus quadriseriatus RFYEY yelloweye rockfish Sebastes ruberrimus CRKYF yellowfin croaker Umbrina roncador 566 yellowfin fringehead Neoclinus stephensae BOGYL yellowfin goby Acanthogobius flavimanus SOLYF yellowfin sole Limanda aspera TNAYF yellowfin tuna Thunnus albacares RFYMN yellowmouth rockfish Sebastes reedi YELTL yellowtail Seriola lalandi	WRAFM	wrasse family	
537 yellow bobo Polydactylus opercularis 347 yellow Irish Iord Hemilepidotus jordani CRBYR yellow rock crab Cancer anthonyi SELYL yellow snake eel Ophichthus zophochir 357 yellowchin sculpin Icelinus quadriseriatus RFYEY yelloweye rockfish Sebastes ruberrimus CRKYF yellowfin croaker Umbrina roncador 566 yellowfin fringehead Neoclinus stephensae BOGYL yellowfin goby Acanthogobius flavimanus SOLYF yellowfin sole Limanda aspera TNAYF yellowfin tuna Thunnus albacares RFYMN yellowmouth rockfish Sebastes reedi YELTL yellowtail Seriola lalandi	573	Y prickleback	Allolumpenus hypochrcmus
347 yellow Irish Iord Hemilepidotus jordani CRBYR yellow rock crab Cancer anthonyi SELYL yellow snake eel Ophichthus zophochir 357 yellowchin sculpin Icelinus quadriseriatus RFYEY yelloweye rockfish Sebastes ruberrimus CRKYF yellowfin croaker Umbrina roncador 566 yellowfin fringehead Neoclinus stephensae BOGYL yellowfin goby Acanthogobius flavimanus SOLYF yellowfin sole Limanda aspera TNAYF yellowfin tuna Thunnus albacares RFYMN yellowmouth rockfish Sebastes reedi YELTL yellowtail Seriola lalandi	537	yellow bobo	
SELYL yellow snake eel Ophichthus zophochir 357 yellowchin sculpin Icelinus quadriseriatus RFYEY yelloweye rockfish Sebastes ruberrimus CRKYF yellowfin croaker Umbrina roncador 566 yellowfin fringehead Neoclinus stephensae BOGYL yellowfin goby Acanthogobius flavimanus SOLYF yellowfin sole Limanda aspera TNAYF yellowfin tuna Thunnus albacares RFYMN yellowmouth rockfish Sebastes reedi YELTL yellowtail Seriola Ialandi	347	yellow Irish lord	Hemilepidotus jordani
SELYL yellow snake eel Ophichthus zophochir 357 yellowchin sculpin Icelinus quadriseriatus RFYEY yelloweye rockfish Sebastes ruberrimus CRKYF yellowfin croaker Umbrina roncador 566 yellowfin fringehead Neoclinus stephensae BOGYL yellowfin goby Acanthogobius flavimanus SOLYF yellowfin sole Limanda aspera TNAYF yellowfin tuna Thunnus albacares RFYMN yellowmouth rockfish Sebastes reedi YELTL yellowtail Seriola Ialandi	CRBYR	yellow rock crab	Cancer anthonyi
RFYEY yelloweye rockfish Sebastes ruberrimus CRKYF yellowfin croaker Umbrina roncador 566 yellowfin fringehead Neoclinus stephensae BOGYL yellowfin goby Acanthogobius flavimanus SOLYF yellowfin sole Limanda aspera TNAYF yellowfin tuna Thunnus albacares RFYMN yellowmouth rockfish Sebastes reedi YELTL yellowtail Seriola lalandi	SELYL		Ophichthus zophochir
CRKYF yellowfin croaker Umbrina roncador 566 yellowfin fringehead Neoclinus stephensae BOGYL yellowfin goby Acanthogobius flavimanus SOLYF yellowfin sole Limanda aspera TNAYF yellowfin tuna Thunnus albacares RFYMN yellowmouth rockfish Sebastes reedi YELTL yellowtail Seriola lalandi	357	yellowchin sculpin	Icelinus quadriseriatus
CRKYF yellowfin croaker Umbrina roncador 566 yellowfin fringehead Neoclinus stephensae BOGYL yellowfin goby Acanthogobius flavimanus SOLYF yellowfin sole Limanda aspera TNAYF yellowfin tuna Thunnus albacares RFYMN yellowmouth rockfish Sebastes reedi YELTL yellowtail Seriola lalandi	RFYEY		
BOGYL yellowfin goby Acanthogobius flavimanus SOLYF yellowfin sole Limanda aspera TNAYF yellowfin tuna Thunnus albacares RFYMN yellowmouth rockfish Sebastes reedi YELTL yellowtail Seriola lalandi		yellowfin croaker	Umbrina roncador
BOGYL yellowfin goby Acanthogobius flavimanus SOLYF yellowfin sole Limanda aspera TNAYF yellowfin tuna Thunnus albacares RFYMN yellowmouth rockfish Sebastes reedi YELTL yellowtail Seriola lalandi		yellowfin fringehead	Neoclinus stephensae
TNAYF yellowfin tuna Thunnus albacares RFYMN yellowmouth rockfish Sebastes reedi YELTL yellowtail Seriola lalandi	BOGYL		
TNAYF yellowfin tuna Thunnus albacares RFYMN yellowmouth rockfish Sebastes reedi YELTL yellowtail Seriola lalandi	SOLYF	yellowfin sole	Limanda aspera
YELTL yellowtail Seriola lalandi	TNAYF	yellowfin tuna	
YELTL yellowtail Seriola lalandi	RFYMN	yellowmouth rockfish	Sebastes reedi
	YELTL	yellowtail	Seriola lalandi
RFYTL yellowtail rockfish Sebastes flavidus	RFYTL	yellowtail rockfish	Sebastes flavidus

Sorted by AFS Common Name		AFS Common Name	
	SP CODE	AFS COMMON NAME	SCIENTIFIC NAME
	618	zebra goby	Lythrypnus zebra
	PERZB	zebra perch	Hermosilla azurea

OTHER CODES

PR Non-Fishing Codes

Target Activity

NFCOM NF commercial fishing (does not include CPFVs)

NFPC6 NF Commercial Passenger Fishing Vessels (includes open

party, charter and "6 pack" vessels)

NFOTH NF other (all other boating activity)

California Island Codes / Saltwater Cutoffs

Island	Code
Coronado	1
San Clemente	2
Catalina	3
Santa Barbara	4
San Nicolas	5
Anacapa	6
Santa Cruz	7
Santa Rosa	8
San Miguel	9
Farallon	F

CRFS Priority Species

Highest Pr	Highest Priority:							
Ad-clipped salmon (both Chinook and Coho), length only					thresher shark			
Higher Pri	ority: Species of Concer	n (in no par	ticular orde	r)				
yelloweye, cowcod, bronzespotted and canary rockfishes			Pacific halibut		bluefin tuna			
High Prior	ity: Species with Harves	t Limits (in	no particula	r order)				
cabezon	California sheephead	greenlings		black, black-and-yellow, blue, bocaccio, brown, copper, calid				
lingcod	California scorpionfish	(Hexagram	nmos spp.)	China, gopher, grass, kelp, olive, quillback, treefish, widow, and yellowtail rockfishes				

PR1 Port Codes

PKIPU	rt Codes						
District	Cnty	Site	Mode	Port	Site Name	MjPort	SubPort
6	15	301	PR1	CRD	Crescent City Inner Boat Basin Docks Crescent City Harbor Launch	CR	CR
6	15	400	PR1	CRL	Ramp	CR	CR
6	23	102	PR1	TRH	Trinidad Hoist	EU	EU
6	23	103	PR1	FLD	Fields Landing Launch Ramp Eureka Marina	EU	EU
6	23	120	PR1	EUR	Launch Ramp	EU	EU
6	23	307	PR1	TRD	Trinidad Docks (water taxi)	EU	EU
5	23	106	PR1	SHC	Shelter Cove Launch Noyo River	FB	SH
5	45	100	PR1	FTB	Launch Ramp	FB	FB
4	1	100	PR1	BER	Berkeley Marina Launch Ramp Sausalito Clipper	SF	SF
4	41	100	PR1	SAU	Launch Ramp Princeton-Pillar	SF	SF
4	81	100	PR1	PRI	Point Launch Ramp Bodega Westside	SF	SF
4	97	100	PR1	BOD	Launch Ramp	SF	SF
3	87	101	PR1	SCR	Santa Cruz Marina Launch Ramp Coast Guard Jetty	МО	SM
3	53	102	PR1	MOC	Launch Ramp	МО	SM
3	53	104	PR1	MOS	Moss Landing Launch Ramp	МО	SM
3	53	107	PR1	мон	Monterey Marina Launch Ramp Morro Bay Launch	МО	SM
3	79	100	PR1	MOR	Ramp	МО	MA
3	79	101	PR1	AVI	Avila Boat Sling	MO	MA
2	83	400	PR1	SBA	Santa Barbara Launch Ramp Ventura Launch	SB	SB
2	111	103	PR1	VEN	Ramp Channel Islands	SB	VN
2	111	104	PR1	OXN	Launch Ramp	SB	VN
1	37	10	PR1	MDR	Marina Del Rey Launch Ramp Dave's Launch	LA	LA
1	37	105	PR1	DLR	Ramp	LA	LA
1	37	110	PR1	CLR	Cabrillo Launch Ramp South Shores	LA	LA
1	37	201	PR1	SSL	Launch Ramp	LA	LA

District	Cnty	Site	Mode	Port	Site Name	MjPort	SubPort
1	59	101	PR1	WAR	Dana Point Launch Ramp Sunset Aquatic	LA	OR
1	59	104	PR1	SUN	Launch Ramp	LA	OR
1	59	106	PR1	NEW	Newport Dunes Launch Ramp	LA	OR
1	73	104	PR1	SHL	Shelter Island Launch Ramp Glorietta Launch	LA	SD
1	73	112	PR1	GLO	Ramp Oceanside	LA	SD
1	73	113	PR1	OCN	Launch Ramp Dana Basin	LA	SD
1	73	204	PR1	DBN	Launch Ramp South Shores	LA	SD
1	73	205	PR1	SSH	Launch Ramp	LA	SD

PC Port Codes

District	Cnty	Site	Mode	Port	Site Name	MjPort	SubPort
6	15	301	PC	CRC	Inner Boat Basin	CR	CR
6	23	121	PC	EUR	Woodley Isl Marina	EU	EU
6	23	307	PC	TRD	Trinidad Pier	EU	EU
5	23	106	PC	SHC	Shelter Cove	FB	SH
5	45	400	PC	FTB	North Noyo Harbor	FB	FB
4	1	400	PC	BER	Berkeley PC	SF	SF
4	1	401	PC	EME	Emeryville PC	SF	SF
4	13	400	PC	СКТ	Crockett PC	SF	SF
4	13	403	PC	SPB	San Pablo PC	SF	SF
4	13	405	PC	RCH	Richmond PC	SF	SF
4	41	400	PC	SAU	Sausalito PC	SF	SF
4	41	402	PC	LMD	Loch Lomond PC SF Fisherman's	SF	SF
4	75	400	PC	SNF	Wharf PC	SF	SF
4	81	400	PC	PRI	Princeton-Pillar Point PC	SF	SF
4	97	400	PC	BOD	Porto Bodega PC	SF	SF
3	53	104	PC	MOS	Moss Landing PC	MO	SM
3	87	101	PC	SCR	Santa Cruz PC	МО	SM

District	Cnty	Site	Mode	Port	Site Name	MjPort	SubPort
3	53	402	PC	мон	Randy's Sportfishing	МО	SM
3	53	403	PC	МОН	Chris' Sportfishing	МО	SM
3	79	100	PC	MOR	Morro Bay PC	MO	MA
3	79	101	PC	AVI	Patriot's Landing	МО	MA
2	83	400	PC	SBA	Sea Landing	SB	SB
2	111	43	PC	CIS	Channel Island/Ciscos Hook's	SB	VN
2	111	45	PC	CAP	Sportfishing Ventura	SB	VN
2	111	103	PC	VEN	Sportfishing	SB	VN
1	37	10	PC	MDR	Marina Del Rey Sportfishing	LA	LA
1	37	13	PC	LBS	Long Beach Sportfishing 22nd Street	LA	LA
1	37	14	PC	TWE	Sportfishing LA Harbor	LA	LA
1	37	15	PC	LAH	Sportfishing Long Beach	LA	LA
1	37	17	PC	LBM	Marina Sportfishing	LA	LA
1	37	202	PC	PPT	Pier Point Landing Redondo Beach	LA	LA
1	37	303	PC	RED	Sportfishing Malibu	LA	LA
1	37	401	PC	MAL	Sportfishing Rocky Point	LA	LA
1	37	405	PC	ROC	Charters	LA	LA
1	59	101	PC	WAR	Dana Wharf Sportfishing Newport	LA	OR
1	59	106	PC	NEW	Sportfishing Davey's Locker	LA	OR
1	59	111	PC	LOC	Sportfishing	LA	OR
1	73	18	PC	SEA	Seaforth Sportfishing	LA	SD
1	73	19	PC	HMS	H&M Sportfishing	LA	SD
1	73	20	PC	LOM	Sportfishing Fisherman's	LA	SD
1	73	21	PC	FIS	Landing	LA	SD
1	73	113	PC	OCN	Helgren's Sportfishing Dana Landing	LA	SD
1	73	119	PC	DAN	Charters	LA	SD

California Saltwater Angling Records as of January 1, 2017 (excludes diving records)

diving records) Species	Cmarin			
Common Name	Species Scientific Name	Weight	County	Date
Barracuda, California	Sphyraena argentea	15 lb 15 oz	San Diego	August 24, 1957
Bass, Barred Sand	Paralabrax nebulifer	13 lb 3 oz	Orange	August 29, 1988
Bass, Giant Sea*	Stereolepis gigas	563 lb 8 oz	Ventura	August 20, 1968
Bass, Kelp	Paralabrax clathratus	14 lb 7 oz	Los Angeles	July 30, 1958
Bass, Spotted Sand	Paralabrax maculatofasciatus	6 lb 12 oz	Orange	October 1, 1994
Bonito, Pacific	Sarda chiliensis	21 lb 5 oz	San Diego	October 19, 2003
Cabezon	Scorpaenichthys marmoratus	23 lb 4 oz	Los Angeles	April 20, 1958
Corbina, California	Menticirrhus undulatus	7 lb 1 oz	Orange	May 30, 2005
Croaker, Spotfin	Roncador stearnsii	14 lb 0 oz	Los Angeles	September 24, 1951
Croaker, Yellowfin	Umbrina roncador	3 lb 14 oz	Los Angeles	October 8, 2000
Dolphinfish	Coryphaena hippurus	66 lb 0 oz	Orange	September 9, 1990
Flounder, Starry	Platichthys stellatus	11 lb 4 oz	San Luis Obispo	August 29, 1993
Greenling, Kelp	Hexagrammos decagrammus	3 lb 4 oz	Monterey	September 17, 2014
Halibut, California	Paralichthys californicus	67 lb 4 oz	Santa Barbara	July 1, 2011
Jacksmelt	Atherinopsis californiensis	1 lb 8 oz	Ventura	June 12, 1998
Lingcod	Ophiodon elongatus	56 lb 0 oz	Del Norte	July 12, 1992
Mackerel, Jack	Trachurus symmetricus	5 lb 8 oz	Orange	September 1, 1988
Mackerel, Pacific (Chub)	Scomber japonicus	2 lb 8 oz	Los Angeles	November 5, 1995

Species Common Name	Species Scientific Name	Weight	County	Date
Mackerel, Pacific (Chub)	Scomber japonicus	2 lb 8 oz	San Diego	November 11, 2005
Marlin, Blue	Makaira nigricans	692 lb 0 oz	Orange	August 18, 1931
Marlin, Striped	Tetrapturus audax	339 lb 0 oz	Los Angeles	July 4, 1985
Opah	Lampris guttatus	163 lb 0 oz	San Luis Obispo	October 8, 1998
Opaleye	Girella nigricans	6 lb 4 oz		May 13, 1956
Perch, Black	Embiotoca jacksoni	2 lb 9 oz	Monterey	February 20, 2011
Perch, Calico	Amphistichus koelzi	1 lb 8 oz	Santa Cruz	February 23, 2013
Perch, Pile	Rhacochilus vacca	2 lb 4 oz	Monterey	July 31, 2013
Prickleback, Monkeyface	Cebidichthys violaceus	6 lb 1 oz	San Mateo	February 7, 2005
Ray, Bat	Myliobatis californica	181 lb 0 oz	Orange	July 24, 1978
Rockfish, Black	Sebastes melanops	9 lb 2 oz	San Francisco	September 3, 1988
Rockfish, Blue	Sebastes mystinus	3 lb 14 oz	San Luis Obispo	October 14, 1993
(Rockfish), Bocaccio	Sebastes paucispinis	17 lb 8 oz	Del Norte	October 25, 1987
Rockfish, Bronzespotted*	Sebastes gilli	14 lb 8 oz	Los Angeles	February 22, 1997
Rockfish, Brown	Sebastes auriculatus	6 lb 15 oz	San Mateo	September 29, 2008
Rockfish, Canary*	Sebastes pinniger	6 lb 15 oz	Mendocino	September 30, 2001
Rockfish, China	Sebastes nebulosus	3 lb 4 oz	Sonoma	July 24, 1998
Rockfish, Copper	Sebastes caurinus	8 lb 5 oz	Monterey	August 18, 1985
Rockfish, Cowcod*	Sebastes levis	21 lb 14 oz	Ventura	August 10, 1998
Rockfish, Grass	Sebastes rastrelliger	6 lb 7 oz	San Mateo	June 30, 2012

Species Common Name	Species Scientific Name	Weight	County	Date
Rockfish, Greenspotted	Sebastes chlorostictus	2 lb 5 oz	San Luis Obispo	June 24, 2005
Rockfish, Olive	Sebastes serranoides	5 lb 14 oz	Santa Barbara	November 21,1991
Rockfish, Treefish	Sebastes serriceps	4 lb 3 oz	Los Angeles	August 9, 2003
Rockfish, Vermilion	Sebastes miniatus	14 lb 9 oz	San Luis Obispo	July 31, 1996
Rockfish, Yelloweye*	Sebastes ruberrimus	18 lb 3 oz	San Luis Obispo	April 15, 1994
Rockfish, Yellowtail	Sebastes flavidus	5 lb 8 oz	Monterey	August 4, 1991
Salmon, Chinook (King)	Oncorhynchus tshawytscha	65 lb 4 oz	Del Norte	August 21, 2002
Sargo	Anisotremus davidsonii	3 lb 3 oz	Los Angeles	December 28, 2010
Scorpionfish, California	Scorpaena guttata	3 lb 0 oz	San Diego	December 26, 1997
Seabass, White	Atractoscion nobilis	79 lb 0 oz	Santa Cruz	October 14, 2011
Seaperch, Rubberlip	Rhacochilus toxotes	5 lb 0 oz	Monterey	June 18, 2009
Seaperch, Striped	Embiotoca lateralis	2 lb 6 oz	Monterey	January 20, 2011
Shark, Blue	Prionace glauca	258 lb 8 oz	Santa Barbara	August 29, 2008
Shark, Leopard	Triakis semifasciata	47 lb 1 oz	Los Angeles	July 18, 2007
Shark, Sevengill	Notorynchus cepedianus	276 lb 0 oz	Humboldt	October 17, 1996
Shark, Shortfin Mako	Isurus oxyrinchus	1,098 lb 12 oz	Ventura	July 24, 2010
Shark, Thresher	Alopias vulpinus	575 lb 0 oz	San Diego	May 26, 2007
Sheephead, California	Semicossyphus pulcher	30 lb 8 oz	Orange	August 29, 2009
Sole, Fantail	Xystreurys liolepis	8 lb 8 oz	Los Angeles	June 6, 2001

Species Common Name	Species Scientific Name	Weight	County	Date
Squid, Humboldt	Dosidicus gigas	OPEN – Minimum Size Requirement: 40 pounds		
Surfperch, Barred	Amphistichus argenteus	4 lb 2 oz	San Luis Obispo	November 8, 1995
Surfperch, Barred	Amphistichus argenteus	4 lb 2 oz	Ventura	March 30, 1996
Surfperch, Calico	Amphistichus koelzi	1 lb 14 oz	San Mateo	April, 9, 2016
Surfperch, Rainbow	Hypsurus caryi	OPEN – Minimum Size Requirement: 1 pound		
Surfperch, Redtail	Amphistichus rhodoterus	3 lb 7 oz	Del Norte	April 23, 2012
Surfperch, Walleye	Hyperprosopon argenteum	OPEN – Minimum Size Requirement: 1 pound		
Swordfish	Xiphias gladius	452 lb 8 oz	Los Angeles	September 30, 2003
Tuna, Albacore	Thunnus alalunga	90 lb 0 oz	Santa Cruz	October 21, 1997
Tuna, Bigeye	Thunnus obesus	240 lb 0 oz	San Diego	August 1, 1987
Tuna, Bluefin	Thunnus orientalis	245 lb 0 oz	San Diego	August 14, 2016
Tuna, Skipjack	Katsuwonus pelamis	26 lb 0 oz	San Diego	August 28, 1970
Tuna, Yellowfin	Thunnus albacares	239 lb 0 oz	Los Angeles	November 24, 1984
Whitefish, Ocean	Caulolatilus princeps	13 lb 12 oz	San Diego	April 23, 1988
Whitefish, Ocean	Caulolatilus princeps	13 lb 12 oz	Ventura	July 3, 2010
Yellowtail	Seriola lalandi	63 lb 1 oz	Santa Barbara	June 18, 2000

* State law presently prohibits the take of giant (black) sea bass, cowcod, and yelloweye rockfish off California, Section 28.10 and 28.55 (b), Title 14, California Administrative code.

California Saltwater Diving Records as of January 1, 2017

Species Common Name	Species Scientific Name	Weight/ Length	County	Date
Abalone, Flat*	Haliotis walallensis	6 7/8 in	Sonoma	June 20, 1994
Abalone, Green*	Haliotis fulgens	8 1/4 in	Los Angeles	August 5, 1986
Abalone, Red**	Haliotis rufescens	12 1/3 in	Humboldt	September 5, 1993
Barracuda, California	Sphyraena argentea	14 lb 7 oz	Los Angeles	August 1, 1957
Bass, Barred Sand	Paralabrax nebulifer	12 lb 12 oz	Los Angeles	August 9, 2004
Bass, Giant Sea*	Stereolepis gigas	545 lb 0 oz	Santa Barbara	September 1, 1968
Bass, Kelp	Paralabrax clathratus	12 lb 6 oz	Orange	October 30, 2012
Bonito, Pacific	Sarda chiliensis	10 lb 4 oz	Los Angeles	July 1, 1967
Cabezon	Scorpaenichthys marmoratus	18 lb 6 oz	Sonoma	May 1, 1984
Clam, Pismo	Tivela stultorum	6 3/4 in (tie)	San Diego	June 13, 2010
Ciam, i ismo	riveia staitoram	0 3/4 III (tie)	Ventura	December 5, 2014
Corbina, California	Menticirrhus undulatus	5 lb 15 oz	Orange	July 1, 1982
Croaker, Spotfin	Roncador stearnsii	8 lb 12 oz	Orange	January 1, 1968
Dolphinfish	Coryphaena hippurus	24 lb 8 oz	Orange	July 26, 2009
Flounder, Starry	Platichthys stellatus	OPEN – Minimum Size Requirement: 8 pounds		
Halibut, California	Paralichthys californicus	72 lb 8 oz	Santa Barbara	August 1, 1982

Species Common Name	Species Scientific Name	Weight/ Length	County	Date
Lingcod	Ophiodon elongatus	37 lb 0 oz	Mendocino	August 2, 2012
Lobster, Spiny	Panulirus interruptus	16 lb 1 oz	Los Angeles	Febuary 1968
Opaleye	Girella nigricans	13 lb 7 oz	Orange	October 18, 1964
Perch, Pile	Rhacochilus vacca	2 lb 8 oz	Monterey	May 7, 2011
Prickleback, Monkeyface	Cebidichthys violaceus	7 lb 5 oz	Monterey	September 7, 2013
Rockfish, Black	Sebastes melanops	8 lb 3 oz	Sonoma	November 19, 2008
Rockfish, Blue	Sebastes mystinus	3 lb 6 oz	Humboldt	August 18, 2010
Rockfish, Copper	Sebastes caurinus	9 lb 5 oz	Mendocino	October 14, 1972
Rockfish, Grass	Sebastes rastrelliger	6 lb 3 oz	Marin	August 1, 2014
Rockfish, Olive	Sebastes serranoides	6 lb 1 oz	Monterey	January 2, 2012
Rockfish, Vermilion	Sebastes miniatus	10 lb 6 oz	Mendocino	August 1, 1983
Scallop, Rock	Crassedoma giganteum	11 1/8 in	Los Angeles	June 1, 1972
Seabass, White	Atractoscion nobilis	93 lb 4 oz	Los Angeles	September 17, 2007
Seaperch, Rubberlip	Rhacochilus toxotes	4 lb 10 oz	Monterey	January 2, 2012
Shark, Blue	Prionace glauca	231 lb 0 oz	Santa Barbara	August 1, 1974
Shark, Shortfin Mako	Isurus oxyrinchus	426 lb 0 oz	San Diego	August 28, 1999
Sheephead, California	Semicossyphus pulcher	40 lb 7 oz	Santa Barbara	August 9, 1992
Tuna, Albacore	Thunnus alalunga	30 lb 11 oz	Monterey	October 14, 1998
Tuna, Bluefin	Thunnus thynnus	269 lb 11 oz	San Diego	June 22, 2016

Species Common Name	Species Scientific Name	Weight/ Length	County	Date
Tuna, Yellowfin	Thunnus albacares	66 lb 7 oz		August 27, 2016
Yellowtail	Seriola lalandi	65 lb 0 oz	San Diego	October 11, 1988

^{*} State law presently prohibits the take of Giant (black) Sea Bass, Cowcod and Yelloweye Rockfish, statewide. Abalone may not be taken south of the mouth of San Francisco Bay.

^{**} Restrictions apply to the take of red abalone (Section 28.10, 28.55 [b], 29.05, 29.15 of Title 14, California Administrative Code), which may only be taken north of the mouth of San Francisco Bay.

Code Foreign Country
FAF Afghanistan
FAL Albania

FDZ Algeria

FAS American Samoa

FAD Andorra FAO Angola FAI Anguilla FAQ Antarctica

FAG Antigua and Barbuda

FAR Argentina FAM Armenia FAW Aruba

FAC Ascension Island

FAU Australia FAT Austria FAZ Azerbaijan **FBS** Bahamas FBH Bahrain FBD Bangladesh FBB Barbados **FBY** Belarus FBF Belgium FB7 Belize FBJ. Renin FBM Bermuda **FBT** Bhutan FBO Bolivia

FBA Bosnia and Herzegovina

FBW Botswana FBV Bouvet Island

FBR Brazil

FIO British Indian Ocean Territory

FBN Brunei Darussalam

FBG Bulgaria FBF Burkina Faso FBI Burundi FKH Cambodia FCM Cameroon FCA Canada FCV Cap Verde FKY Cayman Islands

FCF Central African Republic

FTD Chad FCL Chile FCN China

FCX Christmas Island

FCC Cocos (Keeling) Islands

Code Foreign Country

FCO Colombia
FKM Comoros
FCK Cook Islands
FCR Costa Rica
FCI Cote d'Ivoire
FHR Croatia/Hrvatska

FCU Cuba FCY Cyprus

FCZ Czech Republic

FCD Democratic Republic of the Congo

FDK Denmark FDJ Djibouti FDM Dominica

FDO Dominican Republic

FTP East Timor
FEC Ecuador
FEG Egypt
FSV El Salvador
FGQ Equatorial Guinea

FER Eritrea FEE Estonia FET Ethiopia

FFK Falkland Islands (Malvina)

FFO Faroe Islands

FFM Federal State of Micronesia

FFJ Fiji FFI Finland

FMK Former Yugoslav Republic Macedonia

FFR France

FGF French Guiana FPF French Polynesia

FTF French Southern Territories

FGA Gabon FGM Gambia FGE Georgia FDE Germany FGH Ghana FGI Gibraltar FGR Greece FGL Greenland FGD Grenada FGP Guadeloupe FGU Guam

FGU Guam
FGT Guatemala
FGG Guernsey
FGN Guinea

FGW Guinea-Bissau

FGY Guyana

Code Foreign Country

FHT Haiti

FHM Heard and McDonald Islands FVA Holy See (City Vatican State)

FHN Honduras
FHK Hong Kong
FHU Hungary
FIS Iceland
FIN India
FID Indonesia

FIR Iran (Islamic Republic of)

FIQ Iraq FIF Ireland FIM Isle of Man FII Israel FIT Italy FJM Jamaica **FJP** Japan FJE Jersey FJO Jordan FKZ Kazakhstan FKF Kenva FKI Kiribati **FKP** Korea, North FKR Korea. South

FKW Kuwait FKG Kyrgyzstan

FLA Lao People's Democratic Republic

FLV Latvia FLB Lebanon FLS Lesotho FLR Liberia

FLY Libyan Arab Jamahiriya

FLI Liechtenstein
FLT Lithuania
FLU Luxembourg
FMO Macau
FMG Madagascar
FMW Malawi

FMW Malawi FMY Malaysia FMV Maldives FML Mali FMT Malta

FMH Marshall Islands
FMQ Martinique
FMR Mauritania
FMU Mauritius

FYT Mayotte FMX Mexico

Code **Foreign Country**

FMC Monaco FMN Mongolia **FMS** Montserrat FMA Morocco Mozambique FM7 **FMM** Myanmar FNA Namibia **FNR** Nauru FNP Nepal

Netherlands FNL

FAN Netherlands Antilles **FNC** New Caledonia FN7 New Zealand FNI Nicaragua FNF Niger **FNG** Nigeria

Niue FNF Norfolk Island

FMP Northern Mariana Islands

Norway FNO FOM Oman FPK Pakistan **FPW** Palau

FNU

FPS Palestinian Territories

FPA Panama

FPG Papua New Guinea

Paraguay FPY **FPE** Peru FPH **Philippines** FPN Pitcairn Island

FPI Poland FPT Portugal FPR Puerto Rico

FQA Qatar

FCG Republic of Congo Republic of Moldova FMD FRE Reunion Island FRO Romania

FRU Russian Federation

FRW Rwanda

FKN Saint Kitts and Nevis

FLC Saint Lucia

FVC Saint Vincent and the Grenadines

San Marino FSM

FST Sao Tome and Principe

FSA Saudi Arabia FSN Senegal **FSC** Seychelles

Code Foreign Country
FSL Sierra Leone
FSG Singapore
FSK Slovak Republic

FSI Slovenia

FSB Solomon Islands

FSO Somalia FZA South Africa

FGS South Georgia and the South Sandwich Islands

FES Spain FLK Sri Lanka FSH St. Helena

FPM St. Pierre and Miquelon

FSD Sudan FSR Suriname

FSJ Svalbard and Jan Mayen Islands

FSZ Swaziland FSE Sweden FCH Switzerland

FSY Syrian Arab Republic

FTW Taiwan
FTJ Tajikistan
FTZ Tanzania
FTH Thailand
FTG Togo
FTK Tokelau
FTO Tonga

FTT Trinidad and Tobago

FTN Tunisia FTR Turkey

FTM Turkmenistan

FTC Turks and Caicos Islands
FTV Tuvalu

FUG Uganda FUA Ukraine

FAE United Arab Emirates
FUK United Kingdom
FUS United States
FUY Uruguay

FUM US Minor Outlying Islands

FUZ Uzbekistan FVU Vanuatu FVE Venezuela FVN Vietnam

FVG Virgin Islands (British)
FVI Virgin Islands (USA)
FWF Wallis and Futuna Islands

FEH Western Sahara FWS Western Samoa

Alpha Foreign Country Codes Code Foreign Country

FYE Yemen
FYU Yugoslavia
FZM Zambia
FZW Zimbabwe

Angler Slang Names

Common Name	Scientific Name	Slang Name(s)
Albacore	Thunnus alalunga	German, abrego, albie, longfin, pigfish, football, longfin tuna
Bank Rockfish	Sebastes rufus	Florida red, bank perch, red widow, Louisiana ridge runner
Barred Sand Bass	Paralabrax nebulifer	California rock bass, California sandbass, ground bass, grumpy, rock bass, sandy, sand perch, turd roller
Barred Surfperch	Amphistichus argenteus	barred perch, sand perch
Bigeye Tuna	Thunnus obesus	albacore, bigeye, gorilla
Black and Yellow Rockfish	Sebastes chrysomelas	cefalutano, China cod, gopher, gopher cod, zurndicky
Black Croaker	Cheilotrema saturnum	Chinese croaker, China croaker, black bass, black perch
Black Rockfish	Sebastes melanops	black bass, black snapper, black rock cod, black sea bass
Black Surfperch	Embiotoca jacksoni	bay perch, black perch, black seaperch, buttermouth perch, porgy, pogie, buttermouth
Blackgill Rockfish	Sebastes melanostomus	blackmouth rockfish, deepsea rockfish
Blacksmith	Chromis punctipinnis	black garibaldi, black perch, kelp perch, blacksmith perch, rockperch
Blue Rockfish	Sebastes mystinus	blue bass, blue snapper, blue rock cod, blue fish, priestfish
Blue Shark	Prionace glauca	blue dog, blue pointer, blue whaler, great blue shark
Bluefin Tuna	Thunnus orientalis	football, great albacore, oriental tuna, shortfin tuna

Common Name	Scientific Name	Slang Name(s)
Bocaccio	Sebastes paucispinis	andygumps, wormbag, sewer trout, salmon grouper, grouper, jack grouper, red snapper, Pacific red snapper, rock perch, sewer salmon, longjaw, salmon rockfish
Bronzespotted Rockfish	Sebastes gilli	Arkansas red, warthog
Brown Irish Lord	Hemilepidotus spinosus	bullhead
Brown Rockfish	Sebastes auriculatus	bolina, brown bass, brown bomber, chocolate bass
Brown Smoothhound	Mustelus henlei	sand shark, smooth hound shark
Cabezon	Scorpaenichthys marmoratus	biggyhead, bull cod, cab, cabby, cab driver, giant sculpin, scaleless sculpin, sculpin, scorpion
Calico Surfperch	Amphistichus koelzi	California porgie, porgie, humpback perch
California Batray	Myliobatis californica	bat sting ray, batfish, eagle ray, mud marlin, sting ray
California Corbina	Menticirrhus undulatus	California king croaker, whiting, sucker, surf fish
California Halibut	Paralichthys californicus	California flounder, Monterey halibut, alabato, barn door, bastard halibut, door mat, flatty, hali, fly swatter, Southern halibut
California Lizardfish	Synodus lucioceps	barracuda, candlefish, gar, lizardfish, snakefish
California Scorpionfish	Scorpaena guttata	rattlesnake, sculpin, scorpion
California Sheephead	Semicossyphus pulcher	California redfish, billy goat, goat, humpy, redfish, sheepie, sheephead, sheepshead
Canary Rockfish	Sebastes pinniger	canary, fantail, orange rockfish, red rockfish, red snapper, swallowtail
Chameleon Rockfish	Sebastes phillipsi	orange rockfish
Chilipepper	Sebastes goodei	chili, johnnies, johnny cod, red snapper
China Rockfish	Sebastes nebulosus	black and yellow rockcod, cefalutano, China cod, Chinafish, Chinese rockfish, garrupa, yellowspotted rockfish, yellowstripe rockfish

Common Name	Scientific Name	Slang Name(s)
Chinook Salmon	Oncorhynchus tshawytscha	Columbia river salmon, Sacramento river salmon, black jaw, black mouth, king salmon, spring salmon, tshawytscha, tyee
Coho Salmon	Oncorhynchus kisutch	Pacific salmon, silver salmon, white salmon, blueback, blueback salmon, kisutch
Common Thresher Shark	Alopias vulpinus	blue thresher, fox shark, longtail shark, sea fox, thresher, whiptail shark, swingtail shark, thintail shark
Copper Rockfish	Sebastes caurinus	bariaga branca, chucklehead, neverdies, fighting bob, garrupa, whitebelly, white gopher
Cowcod	Sebastes levis	calf, cow, cow rockfish, cowfish, red snapper, rooster, roosterfish
Darkblotched Rockfish	Sebastes crameri	blackblotched rockfish, blotchie, blackmouth rockfish
Dolphinfish	Coryphaena hippurus	common dolphinfish, dolphin, dorado, mahi mahi
Drum Family	Sciaenidae	croakers, drums, roncadores, ronkies
Eulachon	Thaleichthys pacificus	candlefish, hooligan
Flag Rockfish	Sebastes rubrivinctus	barber pole, convictfish, hollywood, spanish flag, tiger
Garibaldi	Hypsypops rubicundus	goldfish
Giant Kelpfish	Heterostichus rostratus	butterfish, eel, iodine fish, kelpfish, kelp blenny
Giant Sea Bass	Stereolepis gigas	California black sea bass, black seabass, jewfish, freight train
Gopher Rockfish	Sebastes carnatus	butter bass, butterball, gopher, gopher cod, rock bass, garrupa
Grass Rockfish	Sebastes rastrelliger	garrupa, grass bass, grassie, pepper bass, rock bass, green bomber, kelp bass
Gray Smoothhound	Mustelus californicus	sand shark, smoothhound shark, dogfish
Green Sturgeon	Acipenser medirostris	golden sturgeon
Greenblotched Rockfish	Sebastes rosenblatti	bosco, chucklehead, santa maria, starry eyes, warthog, bolina

Common Name	Scientific Name	Slang Name(s)
Greenspotted Rockfish	Sebastes chlorostictus	bosco, chucklehead, santa maria, starry eyes, warthog, bolina
Greenstriped Rockfish	Sebastes elongatus	belinda bass, chilipepper, cucumber, striped rockfish, watermelon, reina
Halfbanded Rockfish	Sebastes semicinctus	inspector
Halfmoon	Medialuna californiensis	California halfmoon, blue perch, Catalina blue perch, blue bass
Honeycomb Rockfish	Sebastes umbrosus	dusky rockfish, speckled rockfish
Horn Shark	Heterodontus francisci	California horn shark, Port Jackson shark, bullhead shark, horned shark
Jack Mackerel	Trachurus symmetricus	Spaniard, Spanish mackerel
Kelp Bass	Paralabrax clathratus	bull bass, cabrilla, calico, calico bass, checkerboard, kelp salmon, rock bass
Kelp Greenling	Hexagrammos decagrammus	seatrout, rock trout, rockfish, green ling, kelp trout, speckled sea trout, spotted rock trout
Kelp Rockfish	Sebastes atrovirens	dumb bass, sugar bass, garrupa, oogly-googly, grass bass, green rockfish, kelp rock cod
Leopard Shark	Triakis semifasciata	cat shark, tiger shark
Lingcod	Ophiodon elongatus	blue cod, ling, linger, lingasaurus, cultus cod, greenlinger, slinky linky, gator, dragon fish
Longspine Thornyhead	Sebastolobus altivelis	anglefin rockfish, bonehead, channel rockfish, hardhead, idiot, idiot fish, spinycheeked rockfish, thomhead
Mexican Rockfish	Sebastes macdonaldi	Arkansas black, Arkansas red, coral cod, coral red, salmon grouper
Monkeyface Prickleback	Cebidichthys violaceus	California monkeyface eel, monkeyface eel, giant monkeyface eel, monkeyface blenny
Moray Eel	Gymnothorax mordax	conger eel, moray
Night Smelt	Spirinchus starksi	nightfish, sand smelt, whitebait
Northern Anchovy	Engraulis mordax	California anchoveta, California anchovy, Pacific anchovy, pinhead, chovy

Common Name	Scientific Name	Slang Name(s)
Ocean Sunfish	Mola Mola	mola mola, sunfish, mola
Ocean Whitefish	Caulolatilus princeps	ocean tilefish, poor man's yellowtail, bottom dorado, whitefish
Olive Rockfish	Sebastes serranoides	bass rockfish, jonathan, johnny bass, kelp bass, sugar bass, yellowtail rockfish
Opah	Lampris regius	Jerusalem haddock, moonfish
Opaleye	Girella nigricans	California opaleye, Catalina perch, blue bass, blue-eye, blue-eyed perch, green perch, jack benny, opaleye perch
Pacific Angel Shark	Squantina californica	California angel shark, monkfish, northern angel shark, squat, squato
Pacific Barracuda	Sphyraena argenta	alligator gar, barracuda, barry, cuda, log, pencil, scooter, skinny, snake, stovepipe, Pacific barracuda
Pacific Bonito	Sarda chilensis	California bonito, bone, bonehead, boney, boner, little tuna, micronito, striped tuna, bonita
Pacific Butterfish	Peprilus simillimus	butterfish, pompano
Pacific Hake	Merluccius productus	California hake, Pacific whiting, haddock, oatmeal fish, whiting, silver hake
Pacific Halibut	Hippoglossus stenolepis	barn door, chicken, flatty
Pacific Herring	Clupea pallasi	Easter herring, California herring, sardine
Pacific Mackerel	Scomber japonicus	American mackerel, mac, greenback mackerel, striped mackerel, tiny tuna, chub mackerel
Pacific Ocean Perch	Sebastes alutus	longjaw rockfish, redfish, rosefish, pop
Pacific Sanddab	Citharichthys sordidus	Catalina sanddab sand dab, soft flounder, sole
Pacific Sardine	Sardinops sagax	fire crackers, dines, pilchards
Pacific Staghorn Sculpin	Leptocottus armatus	smooth cabezon, smooth sclulpin, bullhead
Petrale Sole	Eopsetta jordani	California sole, Jordan's flounder, brill, cape sole

Common Name	Scientific Name	Slang Name(s)
Pile Surfperch	Rhacochilus vacca	dusky perch, forktail perch, porgy, splittail perch, silver perch, white perch
Pink Rockfish	Sebastes eos	bosco, chucklehead, santa maria, starry eyes
Plainfin Midshipman	Porichthys notatus	bullhead, grunter, midshipman, toadfish, singing fish
Pygmy Rockfish	Sebastes wilsoni	Wilson's rockfish, dwarf rockfish, slender rockfish
Queenfish	Seriphus politus	brown bait, herring, herring croaker, kingfish, sea trout, shiner
Quillback Rockfish	Sebastes maliger	frecklebelly, gopher, orangespotted rockfish, speckled rockfish, yellowbacked rockfish
Rainbow Surfperch	Hypsurus caryi	blue perch, rainbow perch, striped seaperch, striped surf fish
Rainbow Trout	Salmo gairdnerii	steelhead trout
Red Irish Lord	Hemilepidotus hemilepidotus	bullhead, red sculpin, spotted Irish lord
Redbanded Rockfish	Sebastes babcocki	bandit, barber pole, convict, flag, hollywood, red bandit
Redtail Surfperch	Amphistichus rhodoterus	Oregon porgie, porgy, redtail seaperch, rosy surf fish
Rock Greenling	Hexagrammos lagocephalus	Pacific red rock trout, fringed greenling, sea trout, kelp cod, kelp trout, red sea trout, rock trout, spotted rock trout
Rock Sole	Lepidopsetta bilineata	broadfin sole, double-lined flounder, rock flounder, roughback sole, gravel sole, two-lined flounder
Rock Wrasse	Halichoeres semicinctus	California wrasse, iodine fish, parrot fish, wrasse
Rockfish Genus	Sebastes spp.	crotch cricket (small), snapper, rock cod
Rosethorn Rockfish	Sebastes helvomaculatus	deepwater scacciatale, orange-red rockfish, rosy
Rosy Rockfish	Sebastes rosaceus	avocado rockfish, corsair, rinky dink, rosy, strawberry
Rougheye Rockfish	Sebastes aleutianus	blackthroat rockfish, buoy keg

Common Name	Scientific Name	Slang Name(s)
Rubberlip Seaperch	Rhacochilus toxotes	alfione, bigmouth surf fish, buttermouth, liverlip, porgy
Sablefish	Anoplopoma fimbria	black candlefish, blackcod, coal cod, coal fish, deep sea trout, sable
Salema	Xenistius californiensis	California salema, striped bass, bigeye bass
Sand Sole	Psettichthys melanostictus	flounder, fringe sole, sand flounder, spotted flounder
Sargo	Anisotremus davidsoni	California sargo, grunt, China croaker, black croaker, perch
Señorita	Oxyjulis californica	butterfish, iodine fish, kelp wrasse, kelpfish
Sevengill Shark	Notorynchus cepedians	bluntnose sevengill shark, broadnose sevengill shark, broadsnouted shark, cow shark, spotted cow shark
Sharpchin Rockfish	Sebastes zacentrus	bigeye rockfish
Shiner Surfperch	Cymatogaster aggregata	bay perch, perch, shiner, seven eleven perch, shiner perch, shiner seaperch
Shortbelly Rockfish	Sebastes jordani	slender rockfish
Shortfin mako Shark	Isurus oxyrinchus	pacific mako, porbeagle, salmon shark, mackerel shark
Shortraker Rockfish	Sebastes borealis	blackthroated rockfish, buoy keg, red snapper
Shortspine Thornyhead	Sebastolobus alascanus	bonehead, channel rockfish, hardhead, idiot, idiotfish, thomhead, scorpion, spinycheeked rockfish
Shovelnose Guitarfish	Rhinobatos productus	guitarfish, sand shark, shovelnose shark
Silver Surfperch	Hyperprosopon ellipticum	porgy, shiner, silver perch
Silvergray Rockfish	Sebastes brevispinis	greenie, longjaw, rock grouper, rock salmon
Sixgill Shark	Hexanchus griseus	bluntnose sixgill shark, bulldog, bull shark, cow shark, grey shark, sixgill cow shark, mud shark
Skipjack	Katsuwonus pelamis	arctic bonito, lesser tuna, skippy, striped tuna
Soupfin Shark	Galeorhinus zyopterus	oil shark, tope, tope shark, soupfin

Common Name	Scientific Name	Slang Name(s)
Speckled Rockfish	Sebastes ovalis	belinda cod, bank perch, cinnamon, widow
Speckled Sandddab	Citharichthys stigmaeus	Catalina sanddab, sanddab, soft flounder
Spiny Dogfish	Squalus acanthias	California dogfish, dog shark, horned shark, piked dogfish, sand shark, spikey jack, pinback, pinback shark
Splitnose Rockfish	Sebastes diploproa	banjo, channel cod, red rock cod, redfish, splitlips
Spotfin Croaker	Roncador stearnsi	black croaker, roncador, spotfin drum, golden croaker
Spotted Sand Bass	Paralabrax maculatofasciatus	bay bass, spotted bay bass, spotted bass, spotty, cabrilla
Squarespot Rockfish	Sebastes hopkinsi	belinda bass, smallmouth rockfish, spotted rockfish
Starry Flounder	Platichithys stellatus	California flounder, diamond flounder, emery flounder, grindstone, roughjacket, sand paper flounder, swamp flounder
Starry Rockfish	Sebastes constellatus	spotted corsair, spotted rockfish, starry eyes, santa maria, whitespotted rockfish, red snapper
Striped Bass	Morone saxatilis	greenhead, rockfish, rock bass, striper
Striped Marlin	Tetrapturus audax	marlin, Pacific marlin, spearfish, spikefish, striper
Striped Seaperch	Embiotoca lateralis	blue perch, rainbow perch, pogy, rainbow seaperch
Stripetail Rockfish	Sebastes saxicola	big-eye rockfish, popeye rockfish,
Surfsmelt	Hypomesus pretiosus	Pacific surf smelt, surf fish, day smelt, day fish, silver smelt
Swell Shark	Cephaloscyllium ventriosum	California swell shark, balloon shark, puffer shark
Swordfish	Xipius gladius	billfish, broadbill, broadbill swordfish
Swordspine Rockfish	Sebastes ensifer	flyfish, hanky panky
Thornback	Platyrhinoidis triseriata	California thornback, banjo shark, round skate
Tiger Rockfish	Sebastes nigrocinctus	barred rockfish, blackbanded, candystripe

Common Name	Scientific Name	Slang Name(s)
Tomcod	Microgadus proximus	California tomcod, piciata
Topsmelt	Atherinops affinis	San Francisco topsmelt, bay smelt, capron, panzarotti, rainbow smelt
Treefish	Sebastes serriceps	barber pole, convict bass, garrupa, lipstick bass, lipstick fish
Vermilion Rockfish	Sebastes miniatus	borracho, red rock cod, red snapper, Pacific red snapper, red
Walleye Surfperch	Hyperprosopon argenteum	China pompano, silver perch, walleye seaperch, walleye surf fish
White Croaker	Genyonemus lineatus	California silver bass, Pasadena trout, herring, kingfish, tomcod, sewer trout, tommy croaker, brown bait,
White Seabass	Atractoscion nobilis	bull tomcod, king croaker, sea trout, weakfish
White Shark	Carcharodon carcharias	blue pointer, great white shark, white pointer, tax man, man in the gray suit
White Sturgeon	Acipenser transmontanus	Columbia sturgeon, Oregon sturgeon, Pacific sturgeon, Sacramento sturgeon
White Surfperch	Phanerodon furcatus	Pacific white perch, forktail perch, splittail perch, white perch
Widow Rockfish	Sebastes entomelas	beccafico, belind bass, brown bomber, brownie, cinnamon, red snapper, widowfish
WolfEel	Anarrhichthys ocellatus	moray eel
Yelloweye Rockfish	Sebastes ruberrimus	cowfish, goldeneye, rasphead, red snapper, red rock cod, turkey red
Yellowfin Croaker	Umbrina roncador	Catalina croaker, golden croaker, yellowfinned roncador, yellowtailed croaker
Yellowfin Tuna	Thunnus albacares	Pacific yellowfin, allison tuna, autumn albacore
Yellowtail	Seriola lalandi	California yellowtail, ahi, amberjack, kingfish, forktail, mossback, yellowtail tuna, yellowjack
Yellowtail Rockfish	Sebastes flavidus	cherne, green snapper, johnny bass, jonathan, red snapper
Zebra Perch	Hermosilla azurea	convict fish, perch

GLOSSARY

TERM	DEFINITION
Ad Clip	A salmonid with its adipose fin missing, for
Ad Clip	salmont his signifies the fish has a coded-wire
	tag (CWT) inserted in its head.
Adipose fin	A fleshy, dorsal fin without rays, located toward
(Ad-fin)	the caudal fin. Found most notably in Salmonids.
AFS	American Fisheries Society – the oldest and
	largest professional society for fisheries
	scientists in the world.
ALDS	The Automated License Data System (ALDS) is
	CDFW's enterprise customer licensing system.
	All sport-fishing, hunting and special permits
	sales are housed in this database. It is a multi-
	user server where enforcement, fish and wildlife
	managers and point of sales can be
	automatically linked to the most up to date
	license sales data.
Anaphylactic shock	Hyper-immune response to foreign proteins or
(anaphylaxis)	drugs producing excessive histamine resulting in
`	swelling, dilated blood vessels and lowered
	blood pressure. If the condition is left untreated
	anaphylactic shock could occur. Typical
	symptoms include hives, swelling and redness of
	the skin, swelling of the eyelids, tongue and
	throat. In very few occurrences anaphylaxis has
	been documented from proteins introduced by
	being pierced by the spines on some fish.
Anchor	A type of fishing code used on the onboard
Alichoi	location form (PCO sampling). Anchor occurs
	when the boat deploys its anchor so that its
	position relative to the bottom is constant,
	without having to use the boat's engines to hold
	position.
Angler license	Telephone survey based on angler contact
directory telephone	information (ALDS) collected during license
survey (ALDTS)	purchase. Designed to identify effort data
Survey (ALDIS)	, ,
	needed to estimate total number of marine
	recreational fishing trips taken by license
Angles	holders.
Angler	A person fishing or who has caught fish, includes
	persons releasing their catch.
Angler eligibility	Determination of whether a person is eligible (as
	an angler) to be interviewed by the Sampler.
Angler survey	A survey conducted by intercepting anglers upon
	completion of fishing to obtain catch and fishing
	effort information (see creel survey).
Arrival Time	When the Sampler arrives on site (a specific time

TERM	DEFINITION
	coded to the nearest minute).
ASF	Assignment Summary Form, the cover page
	used to track sampled assignments.
Assignment	An appointment scheduled to sample a specific
	site or group of sites and issued to a Sampler(s)
	to collect data.
Assignment ID	The specific six digit code used to identify all
	sample assignments issued in a given month.
Avidity	How often an angler fishes in a 12 month period,
	in CA ocean waters, not including today.
Bank	The slope of elevated land adjoining the ocean
	or bay. Can be rock or an overhanging cliff, and
	may be reinforced by materials placed there by
	humans.
Beach	An expanse of pebble, sand, or rock along a
	shore of an ocean that is affected by tidal action.
Beach and Bank (BB)	A cluster assignment survey conducted on
	beaches and bank sites primarily for catch data.
Bias	In statistics, a biased sample is a sample that
	contains members of a population that are not
	equally likely to be chosen as other members of
	the population.
Bio data	Survey data such as lengths, weights, tag wand
	scan results, sex, and headtags.
Boat mode	A mode of fishing from a boat (skiff, vessel,
	kayak, etc.) Includes PR and PC modes.
Bow	The exterior of the forward end of a vessel.
California Code of	The set of administrative rules issued by an
Regulations (CCR)	agency such as Title 14 issued by the
	Resources Agency for the management of fish
OF Newshar	and wildlife resources in the state.
CF Number	The CF number is a vessel registration number
	issued by the Department of Motor Vehicles. A
	CF number is required for every sail-powered
	vessel over eight feet in length and every motor-
	driven vessel (regardless of length) that is not
	documented by the U.S. Coast Guard which is
California Fish and	used or on the waters of this state.
Game Code	The set of laws (statutes) enacted by the California State Legislature and signed by the
Gaine Code	Governor that governs the management of fish
	and wildlife resources in the state.
California Fish and	A separate entity from CDFW. Body composed
Game Commission	of five Commissioners appointed by the
(FGC)	Governor. Responsible for setting seasons, bag
(FGC)	limits and other regulations for game animals,
	sport fishing and some commercial fishing.

TERM	DEFINITION
CDFW permit #	CDFW's identification number for CPFVs. This
	number is usually found on the CPFV's wheel
	house in prominent lettering. Also on the District
	CPFV list provided by your Lead.
California	An integrated state and federally funded finfish
Recreational	sampling program for California marine
Fisheries Survey	recreational fisheries. Conducted since January
(CRFS)	2004. Replaced the MRFSS.
Calipers	A calibrated instrument used for measuring
	distance or thickness, usually with a sliding
	adjustable piece. CRFS Samplers can use
	calipers to measure certain species of sport-
Catal	caught invertebrates. Fish that are caught. Includes kept and released
Catch	fish.
Catch estimate (see	An expanded number based on a statistical
total catch estimate)	sample with inference to the population.
Catch per unit of	The quantity of fish caught per unit of fishing
effort (CPUE)	effort, such as number of fish per angler day or
	pounds of released catch per boat hour.
Caudal fin	The unpaired fin at the posterior end of the fish
	body which may be forked.
California Department	A state natural resource agency department that
of Fish and Wildlife	in part is responsible for marine resource
(CDFW)	management. Formerly known as the California Department of Fish and Game (CDFG) – name
	changed as of January 2013.
Census	A complete count of all members of a population.
Charter boat	A CPFV reserved for a specific group; usually
Charter boat	means the boat is closed to anyone not in the
	group.
Cluster	A grouping of sample sites specific to a single
	mode that are scheduled to be sampled together
	as one sample unit, usually for geographic and
	economic efficiency.
Coded wire tag (CWT)	Small pieces of stainless steel wire that are
	injected into the snouts of juvenile salmon. Each
	tag is etched with a code that relates to certain
	life history information about each release group.
Commercial fishing	Fishing in which the fish harvested, either whole
	or in part, are intended to enter commerce
	through sale, barter, or trade.
Commercial	Commercially registered vessels which
Passenger Fishing	participate in recreational passenger trips.
Vessel (CPFV)	1
County code	A specific code assigned to each California
	county. For sample sites it is numeric.

TERM	DEFINITION
Courtesy headtag	A head tag that is prepared and attached to a
(see headtag also)	salmon head which was voluntarily brought to
	the Sampler by an angler outside of a CRFS
	sampling assignment.
CPFV	Commercial passenger fishing vessel (party or
	charter boat).
Complete interview	An interview that has all the necessary
	information to be used in the CRFS estimates.
	Also, an interview obtained from a shore-based
	angler that has completed his/her fishing trip.
Creel survey	A survey conducted by intercepting anglers upon
	completion of fishing to obtain catch and fishing
	effort information. The term creel refers to an
	interwoven basket that anglers use to retain fish
	in (see angler survey).
CRFS	California Recreational Fisheries Survey
CWT	See coded wire tag
Deadhead (see also	Non-paying angler on a party/charter vessel.
pinhead)	
Derby	A fishing tournament that is non-specific for a
	date and/or location; fishing derbies are usually
	conducted over a long period of time, sometimes
	for the entire fishing season. Derby participants
	are eligible for CRFS interviews. This differs from
	a jackpot or tournament.
Descending device	A device used to return rockfish suffering from
(DD)	barotrauma to depth. Includes inverted crate,
	inverted hook, and commercially available
	devices. Does NOT include venting or "fizzing"
Dan antuna tima	the fish. When the Sampler physically leaves the site (a
Departure time	
Depth	specific time coded to the nearest minute). For boat modes, this is the average bottom
Deptii	depth in feet where the majority of catch was
	taken or where the majority of effort occurred if
	no catch.
Discard	Fish not retained by angler and returned to the
Diodaid	ocean. Fish may be classified as released alive
	or dead. For PCO sampling, the location of catch
	and lengths are obtained if possible.
Disposition (Assn)	On the ASF: Assignment disposition is either
	complete (1), reassigned (2), or canceled (6).
Disposition (Site)	On the ASF: Site disposition is either complete
	(1), roving (7), pressure check (0), low effort (4),
	or other (5).
District	The six geographical areas the CRFS divides
	California into for survey estimation purposes.

TERM	DEFINITION
	District boundaries tend to follow certain county
	lines.
Dock	A floating platform with land access used
	primarily for boat moorage, loading, or fishing.
Dockside sampling	Sampling of PCs at their berth or slip when they
	return from their fishing trip.
Drift	A type of fishing code used on the onboard
	location form (PCO sampling). A drift occurs
	when the CPFV shuts down engines or takes
	them out of gear, so that the boat drifts with the
	prevailing currents or winds.
Effort	A unit of measure of fishing activity. This could
	be angler(s) hours or trip.
Eligible Angler	A recreational angler who meets CRFS interview
	criteria: must be an angler who leaves the boat
	with fish or intended to leave the boat with fish
	(PR and PC mode), or be at least 50% done with
	their fishing trip (MM mode) or has fished for at
	least 30 minutes (BB mode). Anglers who do not
	meet these criteria are considered ineligible and
	should not be interviewed.
Essential fish habitat	Those waters and substrate necessary for fish
(EFH)	spawning, breeding, feeding or growth to
	maturity.
Estimate	An expanded number based on a statistical
Fatimated discond	sample with inference to the population.
Estimated discard	An estimate of the proportion of discarded fish
mortality	that do not survive release.
Examined catch	Catch that the CRFS Sampler was able to see,
Fathom	count and identify. Also called observed catch. A unit of measurement used chiefly in measuring
Fathoni	marine depth. A fathom equals six feet.
Field check	Also called a Quality Control (QC) check, when a
Field Check	Lead or Fish and Wildlife Technician visits an
	assignment to evaluate Samplers, provide
	feedback, or train.
Finfish	Pertains to marine fish with fins for the purposes
	of CRFS. Does not include invertebrates
	(crustaceans and mollusks which are designated
	"shellfish").
Fish and Game Code	Legal form of California Law pertaining to fish
	and wildlife.
Fish and Game	A separate entity from the Department of Fish
Commission	and Wildlife that has been involved in the
	management and use of California's fish and
	wildlife resources since 1870. It is composed of
	up to five members, appointed by the Governor

TERM	DEFINITION
	and confirmed by the Senate. The Legislature
	delegated a variety of powers to the
	Commission, some general in nature and some
	very specific. A major responsibility is the
	formulation of general policies for the conduct of
	the Department of Fish and Wildlife and the
	interpretation of laws into regulations.
Fishery management	A fisheries management body established by the
council	Magnuson Stevens Act to manage fishery
	resources in designated regions of the United
	States. Membership varies in size depending on
	the number of states involved. There are eight
	regional Councils, including the Pacific Fishery
	Management Council (PFMC).
Fishing AREA	The water area or island where the anglers
	fished.
Fishing boat	A boat, either privately owned or rented, upon
	which fishing effort (for finfish OR invertebrates)
	occurred. Boats that targeted invertebrates only
	are considered fishing boats. See non-fishing
	boat.
Fishing mode	The method of access to the fisheries. The major
	modes are man-made structures (MM), beach
	and bank fishing (BB), party and charter boat
	fishing (PC), and private and rental boat fishing
	(PR).
Fishing pressure	Number of anglers or boats at a fishing site; a
Fighting to the	gauge of effort.
Fishing type	The type of fishing performed by CPFVs: Drift,
	Station keeping, Anchored or Troll.
Flag	A letter qualifier recorded after the sample
	number on the PR form, used to denote kayaks,
	personal watercraft, sailboats, etc. See also sample flag.
Flat	A gear type used to take invertebrates primarily
FIAL	(although some species of finfish can legally be
	taken with it), usually lobster. Refers to a hoop
	net with collapsible sides that lies flat on the
	bottom when deployed; when retrieved, the
	sides of the hoop net are raised which makes
	escape difficult during retrieval.
Fork length	A measurement used frequently for fish length
I OIK IEIIGUI	when the tail has a fork shape. Projected straight
	distance between the tip of the fish's closed
	mouth and the medial caudal fin ray.
Free diving	A gear type used to take invertebrates. Coded
Free aiving	when the diver is not using SCUBA, and is
	when the diver is not using SCODA, and is

TERM	DEFINITION
	breath-hold diving to take invertebrates. For
	spearfishermen using breath-hold techniques to
	take finfish use the gear code Spear.
Gear	The fishing equipment used to target fish, such
Geographic	as hook-and-line, pots, spear, snare, hand, etc. A method of collecting and presenting graphic
information system	data that allows for replication. This is used by
(GIS)	CRFS to reference effort and/or catch to a
(0.0)	specific location.
GPS Format	In reference to onboard sampling locations, the
	type of GPS format used to report latitude and
	longitude. Can be degrees, minutes and
	seconds of latitude and longitude OR degrees
	and decimal minutes of latitude and longitude.
Groundfish	A group of over 90 generally benthic species
	managed through the policies of the Pacific
	Fishery Management Council's Groundfish
	Fishery Management Plan and under the Magnuson Stevens Fishery Conservation and
	Management Act and other Federal laws.
	Groundfish include all species of rockfish,
	several species of flatfish, some species of
	sharks and skates, and several species of
	roundfish like lingcod, greenlings, sablefish and
	Pacific cod. See Section 1.91 of the Ocean Sport
	Fishing Regulations booklet for a complete
	listing of all species included in the Groundfish
	FMP.
Headtag	A uniquely numbered tag affixed to the lower jaw
	of an adipose fin clipped salmon that tracks the
	salmon head collected on the docks through
	coded wire tag processing in the lab and entry into project databases.
Headtag report	Data sheet that records the date, port and
l leading report	sample mode for all headtags used in ocean
	salmon sampling.
Hook and Line	A gear type used to take finfish. A hook or
	multiple hooks tied to a line that is attached to a
	reel mounted on a fishing rod, or the line can be
	tied directly to the rod itself.
Incomplete	A Shore Forminterview where the angler's
interview/sample	fishing trip is partially complete. For BB, anglers
	need to have completed at least 30 minutes of
	fishing. For MM, anglers need to have completed
	at least 50% of their fishing trip and incomplete
	interviews MUST be obtained after the stop count for that site. See also partial interview and
	countror that site. See also partial interview and complete interview.
	Complete interview.

TERM	DEFINITION
Ineligible angler	An angler who does not meet the criteria as an
]	eligible angler for an interview. See eligible
	angler.
Initial refusal	An angler that refuses the CRFS interview from
1	the beginning.
Inland marine waters	A body of saltwater enclosed by land or barriers
	with a mouth that allows access to the ocean:
	e.g. San Francisco Bay, Morro Bay, Monterey
	Harbor, etc.
In-season	Regulatory changes that affect an ongoing
management	fishery during its open season.
Intercept	To approach/encounter an angler or a boat in
	the field to interview for the survey.
Inventory tag	A tag that is affixed externally to an inventory
	bag containing one or more tagged and
	individually bagged salmon heads. The inventory
	tag documents the Sampler's name, the date the
	inventory bag was taken to the drop off location,
	and the head tag series present in the bag.
Invertebrate trips	Trips that target invertebrates. CRFS interviews
	anglers/boats targeting crab, squid, lobster and
In also a f	abalone.
Jackpot	A fishing competition aboard a CPFV, usually
	largest fish wins the pot. Jackpot participants are
	eligible for CRFS interviews. See also derby and
Jetty	tournament. A narrow man-made structure that projects into
Jelly	the water from land to reduce wave action in a
	waterway or harbor.
Key refusal	An angler who refuses the CRFS interview by
i Noy iciusal	not answering a key question.
Key questions	Key questions must be answered for the data to
1.05 quodilono	be used in the statistical programs to compute
	estimates.
KOD	Kind of day – weekend day or weekday day.
	Some holidays are considered weekend days.
Landing	Within a port there are one or more specific sites
	where anglers can fish. Landings tend to refer to
	where CPFVs and commercial boats dock.
Language barrier	Occurs when the Sampler and angler cannot
	communicate due to the lack of a common
	language. As a result, the interview is terminated
	and a "B" is recorded in the sample # field.
Latitude	An angular distance north or south of the
	equator. These measurements are parallel to the
	equator.
Launch ramp	A sloping roadway into a body of water that

TERM	DEFINITION
	allows vehicles towing boats on trailers to back
	the trailer down into the water until the boat can
	float off the trailer.
Logbook	A log of each fishing trip is required by the
	CDFW to be completed and returned for each
	commercial passenger fishing vessel or
	commercial fishing trip. The log captures
	location, catch and effort information.
Longitude	An angular distance east or west of the Prime
_	Meridian (in England). These measurements are
	perpendicular to the equator from pole to pole.
Magnuson Stevens	The MSFCMA, sometimes known as the
Fishery Conservation	"Magnuson Stevens Act," established the 200
and Management Act	mile fishery conservation zone, the regional
	fishery management council system, and other
	provisions of U.S. marine fishery law.
Marine Mammal	Federal law prohibiting the harvest or
Protection Act	harassment of marine mammals, although
(MMPA)	permits for incidental take of marine mammals
	while commercial fishing may be issued subject
	to regulation. (See "incidental take" for a
	definition of "take").
Marine Recreational	A national survey developed in 1979 by the
Fisheries Statistics	National Oceanic and Atmospheric
Survey (MRFSS)	Administration and conducted by National
	Marine Fisheries Service to estimate the impact
	of recreational fishing on marine resources.
	Conducted in California through 2003.
Man-Made (MM)	A shore fishing mode. A structure built by
	humans that anglers can potentially fish from:
	jetty, pier, dock, wharf.
Missed boat	A boat, either in the PR1 or PR2 survey, fishing
	or not, that was observed at the site but not
	sampled. Can be on-site or off-site.
Marine Protected Area	Named, discrete geographic marine or estuarine
(MPA)	areas set aside primarily to protect or conserve
	marine life and habitat.
Mode (see Fishing	Type of access to water for angling.
mode)	
Mooch	A gear type used to take salmon. Fishing with
	bait while the vessel is stationary.
Mooring buoy	An anchor station for boats to be stored in the
	harbor. A type of private access boat.
National Marine	A division of the U.S. Department of Commerce,
Fisheries Service	National Ocean and Atmospheric Administration
(NMFS) a.k.a. NOAA	(NOAA). NMFS is responsible for conservation
Fisheries	and management of offshore fisheries (and

TERM	DEFINITION
	inland salmon). The NMFS Regional Director is a
	voting member of the Council. Recently renamed
	to NOAA Fisheries.
NMFS Economic	In some years, NMFS requests that CRFS
Survey	interviews include additional questions (e.g.
	name, telephone, mail, home address) directed
	at shore-mode anglers and sometimes PC
h	anglers.
National Oceanic &	The parent agency of the National Marine
Atmospheric Administration	Fisheries Service (NOAA Fisheries).
,	
(NOAA) Non-fishing (NF) boat	There are three types of NF boats: NFCOM
Non-rishing (NF) boat	(commercial finfish or invertebrate fishing),
	NFPC6 (Commercial Passenger Fishing
	Vessels, a.k.a. party/charter boats, including 6-
	pack boats) and NFOTH (all other non-fishing
	boats, including sailing, whale watching, burials
	at sea, cruises, enforcement, research, etc.).
	See fishing boat.
Non-Recovered	A coded-wire tagged salmon head which cannot
Species (NRS)	be recovered for some reason.
Ocean Salmon Project	The Department of Fish and Wildlife's program
(OSP)	to determine recreational and commercial catch,
	effort, and hatchery contributions to California's
	ocean salmon fisheries.
Onboard sampling	Sampling PC boats by riding the boat throughout
(PCO)	the whole fishing trip.
Open bay	A wide bend or curve in a shoreline where a
	wide unenclosed portion of the ocean is formed.
	Also known as a bight. California examples:
	Santa Monica Bay, Monterey Bay, etc. Not a true
Opportunistic	bay. Interviews for party/charter trips completed
interviews	outside of a scheduled PC assignment. Can be
interviews	salmon or non-salmon trips.
Optimum yield (OY)	The amount of fish that will provide the greatest
	overall benefit to the Nation, particularly with
	respect to food production and recreational
	opportunities, and taking into account the
	protection of marine ecosystems.
Otolith	A bone in the inner ear of vertebrates.
	Movement of otoliths, caused by a change in
	position of the animal, stimulates sensory hair
	cells which convey the information to the brain.
	In some species of fish, can be used to
	determine the age of the fish.

Overfished Any stock or stock complex whose size is sufficiently small that a change in manageme	
I was a the second and the selection of	
practices is required to achieve an appropriate	
level and rate of rebuilding. Under current fish	
management practices, a stock is considered	d to
be overfished if the current population is	
estimated to be at or below 20% of the pre-	
fished population.	
Pacific States Marine The PSMFC is a non-regulatory agency that	
Fisheries serves Alaska, California, Idaho, Oregon and	
Commission (PSMFC) Washington. The PSMFC provides information	
the form of data services for various fisheries	i
Party boat A CPFV boat on which fishing space and	
privilege are provided for a fee per angler.	.1
Usually refers to open party, or non-chartered	a
trips.	
PC Party and charter boats (see CPFV)	
PC Effort check (PEC) A sample of CPFV activity based on checking	g
sites for docked status and type of activity if n	ot
docked.	
PC Onboard forms Includes the Onboard Angler Form, Onboard	
Location Form, and Onboard Catch and Disc	ard
from.	0
Pacific Fishery A fisheries management body established by	tne
Management Council Magnuson Stevens Act to manage fishery	J
(PFMC) resources in designated regions of the United States. Membership varies in size depending	
the number of states involved. There are eigh	
regional Councils, including the Pacific Councils	
Partial A Shore Forminterview where the angler's	JII.
interview/sample interview/sample fishing trip is partially complete. For BB, angle	ore
need to have completed at least 30 minutes of	
fishing and incomplete interviews can be	′1
obtained at any time. For MM, anglers need t	0
have completed at least 50% of their fishing t	
and incomplete trip interviews MUST be	۳.۳
obtained after the stop count for that site. See)
also incomplete interview and complete	
interview.	
Pier A man-made structure made with pilings	
projecting from the bottom out of the water an	d
covered with a platform on top so that waves	
may pass under the platform.	
Pinhead (see Non-paying angler on a party/charter vessel.	
deadhead also)	
Pinnipeds Seals or sea lions.	
Port A specific area where people access the fish	ery;

TERM	DEFINITION
	usually landings grouped to gether
	geographically. Ports are given 3-letter codes.
	Ports are made up of one or more landings.
Port	Facing the bow of a vessel, the left side of the vessel.
Pot	A gear type used to take invertebrates, primarily
	crab. An enclosed, trap-like device with ports
	constructed to allow entry to access bait and
	then prohibit legal sized animals from escaping.
B. (15 (1	Also called trap.
Private and Rental	Private and rental boat mode of fishing. A type of
boats (PR)	Boat mode.
PR Form	This form is used when sampling PR1 and PR2
DD4 Duimour muistata	assignments. Primary private and rental boat survey that
PR1 – Primary private boat survey	samples sites where 90% of the catch of
boat survey	important species is landed.
PR2 Secondary	Secondary private and rental boat survey that
private boat survey	samples sites where 10% of the catch of
private boat survey	important species is landed.
Pressure check (see	Site visit for the purpose of estimating angler
site check)	effort (numbers of anglers and/or boats).
Private access fishery	The private or rental boats that access the water
	from marinas, moorings and slips (private areas
	not accessed by CRFS).
Private boat	A boat belonging to an individual not for rent or
	with paying passengers.
PWC	Personal water craft (e.g. jet ski).
Ramp (launch ramp)	Roadway leading down into the water for the
	purpose of launching a boat from a trailer.
Random	With no pattern. Occurring sporadically or
	intermittently in an unpredictable way.
Random sampling	A method of selecting a sample from a
	population in such a way that every possible
	sample that could be selected has an equal
B. FIN	probability of being selected.
RecFIN	Recreational Fishery Information Network. A
	database managed by the Pacific States Marine Fisheries Commission that provides recreational
	fishery information for Washington, Oregon, and
	California.
Recreational fishing	In California, fishing for recreation, sport, or
	personal, non-commercial use. Conducted under
	the authority of a California sport fishing license.
	Sport-caught fish may not be bought, sold,
	bartered or traded. See also sport fishing
Refugia	An area in the water where living things or their

TERM	DEFINITION
	habitat are managed to minimize anthropogenic
	impacts. May be a place where fishing is not
	allowed so that fish can reproduce, grow and
	migrate.
Region	An area of interest. In CRFS, California is split
	into two subregions; North and South. The split
	occurs at San Luis Obispo/Santa Barbara county
	line. This is based on historical fishery related
	differences.
Refusal	A denial on the part of the angler to be
	interviewed by the Sampler or to refuse a key
	item during the interview.
Rental boat	A boat that is rented but without crew or a guide.
Rigid	A gear type used to take invertebrates primarily
	(although some species of finfish can legally be
	taken with it), usually lobster. A hoop net with
	sides that are fixed in place with rigid supports,
	making the net stand erect when deployed on the bottom. The diameter of the opening at the
	top is less than the diameter of the bottom,
	making the angle at the base convex and
	thereby making escape from the net very
	difficult.
Roving	In reference to cluster sampling, when the
Koving	Sampler travels among multiple sites within an
	assignment looking for recreational anglers to
	interview.
Salmon Refusal (RS)	A flag on the PR form (RS) indicating that the
	anglers on the boat refused to participate in the
	CRFS survey, but all data elements needed for
	salmon management were collected.
Sample boat	A boat intercepted in the PR survey for which a
	sequential number is given and specific data
	collected.
Sample Flags	On the PR form, a code that provides additional
	information about a specific boat that was
	sampled. Flags include kayaks (K), sailboats (S),
	personal watercraft (P), boats participating in a
	tournament (T), and refused boats (R). See also
	flags.
Sampler Location	In reference to Onboard CPFV sampling, it is the
	location on the boat where the Sampler
	observed anglers during stops (e.g. bow, stern,
	side).
SCUBA	Acronym for self-contained underwater breathing
	apparatus. Also a gear type used to take
	invertebrates, coded when SCUBA gear is used

TERM	DEFINITION
	by the diver to take invertebrates. For
	spearfishermen using SCUBA gear to take finfish
	use the gear code Spear.
Seal take	Fish lost to seals/sea lions (pinnipeds).
Shellfish	Animals with shells such as clams, lobsters,
	squid and abalone (crustaceans and mollusks).
Shore Form	This form is used when sampling MM or BB
	cluster assignments.
Shore trip	A fishing trip conducted from the shore (BB and
	MM modes).
Site check	A visit to a fishing site to check for effort or CPFV
	boat status.
Site code	The numeric code used to distinguish specific
	fishing areas within a CRFS District.
Site disposition	The code on the ASF which indicates the status
	of the site visit and the reason for leaving the
	site.
Site effort check	A count of the number of finfish anglers or boat
(SEC)	trailers at all sites adjacent to a CRFS
	assignment. Effort if recorded on the ASF in the
	Pressure Check Count column. Not all
	assignments will have a SEC.
Site name	The name of a CRFS sampling site.
Site register/list	A complete list of sites with names, codes and
	descriptions for a given District.
Six pack	An informal term applied to a commercial
	passenger fishing vessel which has a license to
	take not more than six paying passengers at a
	time. Term also used to describe CPFVs that
	carry six or fewer anglers.
Sling	A sling or hoist that is used to lower and lift boats
	from the water.
Snare	A gear type used to take invertebrates, usually
	crab. A small cage-like structure that contains
	bait to attract crab, with up to six monofilament
	loops on the outside of the structure. Usually
	deployed with a rod and reel, similar to hook and
	line fishing for finfish, when retrieved, the loops
	constrict, trapping the legs of any crab that are
	attempting to reach the bait cage.
State site code	A location on the water that has been issued a
	code to match a name so that map coordinates
	are automatically found in the database.
Spear	A gear type used to take finfish; either an arrow-
	like projectile fired from a gun-like launcher,
	powered by one or more elastic bands, or a two
	or three pronged fork launched by a single

TERM	DEFINITION
	elastic band (Hawaiian sling).
Species code	A specific five letter code used to record fish
	taxon on the survey forms.
Sport fishing	In California, fishing for recreation, sport, or
	personal, non-commercial use. Conducted under
	the authority of a California sport fishing license.
	Sport-caught fish may not be bought, sold,
Ot and a small	bartered or traded. See also recreational fishing.
Starboard	Facing the bow of a vessel, the right side of the vessel.
Start time	A time after the arrival time onsite when the
	Sampler actually begins sampling (a specific
01.71	time coded to the nearest minute).
Stat	A type of fishing code used on the onboard location form (PCO sampling), short for station
	keeping. Stat occurs when the CPFV uses its
	engines to hold the boat in a stationary position
	relative to the ocean bottom.
Stern	The rear or aft part of a vessel, opposite the
	bow.
Stop time	A time when the Sampler actually stops
·	sampling, but before they depart (a specific time
	coded to the nearest minute).
Systematic	A regular predictable pattern. Used in statistical
	sampling to promote sampling simplicity and to
	even out the sample.
Systematic sampling	Any sample drawn from a list using a random
	start and a fixed sampling interval (e.g. every Nth boat). An efficient and functional substitute for
	random sampling.
Target (fishing)	Fishing for the primary purpose of catching a
raiget (iisiiiig)	particular species or species group (the target
	species).
Target (mode)	The specific fishing mode(s) that the Sampler
• '	should be monitoring at a given site. Listed on
	the Site list.
Total catch estimate	An expanded number based on a statistical
	sample with inference to the population for all
T::1 44	modes combined.
Title 14	Regulations adopted by the Fish and Game
	Commission, through their regulatory powers
	function, are printed in the California Code of Regulations (a.k.a. CCR), Title 14, Natural
	Resources. There are 28 separate California
	Code of Regulations "Titles" containing
	regulations proposed by over 200 state
	agencies. Title 14 is the section of the California

TERM	DEFINITION
	Code of Regulations concerning natural
	resources. Regulations are printed in the
	California Code of Regulations after they are
	adopted by the rulemaking agency, approved by
	Office of Administrative Law and filed with the
	Secretary of State.
Tournament	A fishing contest for which participants register
	and compete. For the purposes of CRFS, a
	tournament is site and date specific. Generally
	speaking, CRFS does not sample tournaments.
	Speak with your Lead when you encounter a
	tournament. See also derby and jackpot.
Trailer counts	Usually done at arrival and departure from boat-
	mode sites as a way to gauge effort.
Troll	A gear type used to take finfish, primarily
	salmon, although trolling is frequently used to
	fish over large distances such as when fishing
	for pelagic species like tunas. A baited hook or
	lure is pulled behind a vessel under power. Also
	a fishing code used on the onboard location form
	(PCO sampling) coded when the boat is trolling.
Unavailable catch	Catch that is not available for the Sampler to
	observe. Includes fish used as bait, given away,
	thrown back alive or dead, filleted, or
	immediately consumed. This type of catch is
	angler-reported – kept/unobserved, released
	alive and released dead.
Unbiased	Free of non-random effects that tend to move an
	estimate higher or lower in prediction of the true
	population.
Validate	Independent verification, generally by field
	sampling, of information received through the
	submission of fishing activity logs, especially
	CPFV logs.
Wand	A device which can detect the presence of a
	metallic object, such as an internal tag, when
	passed over the surface of the fish. Used for
	such species as White Seabass.
WD	Weekday
WE	Weekend and some holidays
Weekly Report	Weekly Reports are Excel files sent to your Lead
	every Monday morning by 8 AM that show all the
	assignments worked the previous week.
Wet gear hours	The time spent fishing with line in the water
	(wet).
Wharf	A fixed platform that originates on land and
	projects into a harbor, ocean, etc., so that

TERM	DEFINITION
	vessels may be moored alongside. See Pier.
Wildlife Officer	(Formerly titled Warden) An officer that
	represents the enforcement branch of CDFW.

Appendix A.

CRFS Sampler Guidelines

Samplers, upon hire, are required to read and sign the Sampler Guidelines document which becomes part of their personnel file. The purpose of this paperwork is to document in writing the importance of understanding CRFS Sampler job duties and expectations. The Sampler Guidelines are a critical part of the Sampler hiring and training process; it sets clear boundaries and outlines the major roles of the Sampler. Supervisors and Leads hold their Samplers accountable for these guidelines. The Guidelines are listed below, followed by a detailed description of each of the items.

The purpose of this guidance is to provide common understanding, clarity and fairness regarding work performance expectations and the workplace environment. The intent is to provide guidelines that help create a positive and productive work environment. Other guidance is available from the Department of Fish and Wildlife's Operations Manual, Bargaining Unit contracts, and the Marine Region's intranet site. Note: These guidelines supersede the Marine Region Standard Operating Procedures and Staff Expectations document. For conflicting procedures, always refer to this document.

Safety

- Your safety is your number one priority
- Notify your Lead if you have ANY health issues that may affect your safety or performance in the field
- You are to call your Lead AS SOON AS POSSIBLE when you are injured on the job
- Report criminal activity (non-fishery violations) to local law enforcement –
 if applicable, call 911 and then call your Lead when it is safe to do so
- If you feel threatened, leave the site and then call your Lead when it is safe to do so
- Lock your car, keep valuables with you
- Carry a cell phone or know where pay phones are located
- Know which fish are potentially dangerous and how to handle them; ask your Lead for gloves if you want them
- Do not sample after sunset unless you are sampling a twilight or overnight
 PC trip
- Wear sunscreen and hat for sun protection
- Do not wear dangling jewelry
- · Long hair must be contained and out of the sampler's face
- Be aware of swell conditions while at sea to avoid falls and injury
- Be careful climbing up or down into and out of boats

Data Quality

- **NEVER** falsify data; times on site are considered data
- If you make a mistake, notify your Lead do not try to cover it up
- You are responsible for the quality of your data
- When your Lead notifies you of an error you are making, take the time to make sure you understand the nature of the error and take steps to stop making the same error
- Refer to your manual with questions on protocol. Ask your Lead if you are
 unable to find your answer there come into the office, call or send an
 email. Ask the Fish and Wildlife Technician (FWT) if your Lead is not
 available.
- Take detailed notes regarding field conditions
- Take detailed notes on situations that you are unclear on how to handle when you are unable to contact your Lead or the FWT for guidance
- Make continuous improvements to your interviewing skills, fish identification skills, data collection techniques and understanding of the sampling protocol

Data Deadlines

- Weekly reports are due to your Lead and OSP by 8 AM every Monday whether you worked the previous week or not
- Head tag reports, courtesy tag reports, CRFS-OSP PC Dockside forms and PC Effort Check forms are due to OSP by 8 AM Monday (either faxed, photographed or scanned and emailed) whether you worked the previous week or not
- The data week is Monday through Sunday. All original data sheets are due
 to your Lead by close of business on the following Wednesday. If mailing
 data, the envelope shall be postmarked on Monday, unless Monday is a
 holiday.
- Quizzes are mandatory and are due on the date and time specified by your Lead

Time Sheets and Travel Claims

- The work week is Sunday through Saturday. You are not to exceed 40 hours in a week without prior approval from your Supervisor.
- You are to call your Lead AS SOON AS POSSIBLE if you believe that you
 will not be able to complete all assignments in a week without exceeding
 40 hours for the week
- Time sheets are due on the date and time specified by your Lead each month. A delay on your end may result in a late paycheck.
- Be accurate in your time reporting claim only time that you work
- For time sheet purposes, round daily hours worked to the nearest quarter hour
- Travel expense claims are due on the date specified by your Lead each month. You are required to submit your claims monthly.

- You may claim the cost of dinner IF you traveled more than 50 miles in one direction AND your day ended after 1900 hrs
- You may claim the cost of breakfast **IF** you traveled more than 50 miles in one direction **AND** your day began before 0600 hrs
- Do not purchase anything for the job with the expectation of reimbursement (other than meals, parking, and fuel) without prior approval from your Supervisor. Include the amount of the purchase when making a request.
- · Retain receipts and be accurate in your expense reporting

Assignment Management

- You are to call or text your Lead BEFORE THE ASSIGNMENT START TIME when you are unable to complete an assignment
 - If using sick leave, notify your Supervisor by email, text or phone on the same day
- Assignments are required to be worked on the date they are assigned unless other arrangements have been made with your Lead
- Do not cancel an assignment without prior approval from your Lead
- Do not reassign an assignment without prior approval from your Lead
- Do not give an assignment to or take an assignment from another sampler without prior approval from your Lead
- Leaving an assignment early for reasons such as traffic, social obligations, a second job, school, etc. are not acceptable
- Do not perform work for any other DFW project without prior approval from your Supervisor

Appearance

- You are to wear the CRFS attire issued to you when in the field at all times
- Do not wear your CRFS attire when you are not working
- You are to wear closed-toe shoes in the field at all times
- You may wear shorts or jeans. Shorts shall not be "short shorts", no more than 3 inches above the knee; jeans shall not have holes or be "cut-offs". No sweatpants. No skinny pants or tight jeans.
- No non-DFW logos visible on any clothing except shoes
- Wear appropriate clothing in the office
- Make an effort to look presentable and official

Vehicles

- If a state vehicle is not available for your use, you are responsible for providing your own reliable transportation. Mileage is reimbursed at the current state rate.
- Report all automobile accidents that occur while working to your Lead AS SOON AS POSSIBLE. Complete the appropriate vehicle accident report form AS SOON AS POSSIBLE and follow all instructions on the form.

- A copy of the State Driver Accident Review STD274 and Vehicle Accident Report STD270 must be kept in the glove box of your personal vehicle when using your vehicle for state business.
- Do not use a cell phone without a hands-free device while driving on the job
- You are responsible for all traffic violations and citations while driving on the job
- Do not park on red curbs or in handicapped parking spots
- Wear your CRFS attire while driving or riding in a state vehicle
- Do not carry unauthorized people in a state vehicle
- Do not make changes to the vehicle assignment schedule without prior approval from your Lead
- Do not use your DFW shield parking placard except on official CRFS business
- Vehicle logs are to be completed at the time of vehicle use
- Do not take a state vehicle home without prior approval from your Lead
- Do not conduct personal business while driving a state vehicle
- Leave the state vehicle with a full tank of gas for the next user
- Do not purchase fuel for your personal vehicle using the Voyager fuel card
- Do not use the Voyager fuel card to purchase anything except authorized items
- Your Voyager PIN should only be used to fill state vehicles used for CRFS work
- Remove all trash and sampling gear from state vehicles after use
- Return all state vehicles and keys to the proper location after use. Make sure windows are rolled up and all doors locked.
- Notify your Lead of any observed deficiencies in state vehicles
- Do not attempt to drive to a site during extremely hazardous weather; notify your Lead
- Be a courteous, safe driver while driving on the job

Party/Charter (PC) Boat Sampling

- You are to notify your Lead AS SOON AS POSSIBLE of all PC refusals
- Do not perform the duties of a crew member while onboard a PC
- Do not fish while onboard a PC
- Do not sleep while onboard a PC, except when sampling an overnight trip during transit
- Do not discuss PC activities or your opinions of specific PC operations with the public
- Do not share the sampling schedule with any crew members

General Onsite Procedures

- Have your state identification card with you while working at all times
- Do not allow the public to believe you are a Wildlife Officer
- Report egregious or repeat fishing regulation violations to your Lead do not contact enforcement directly
- Refer reports of pollution or poaching to CalTIP have the person reporting it to you make the call
- Do not claim to represent the Department when not conducting CRFS
- Do not discuss your opinions of other Department employees with the public
- Do not have unauthorized people (family, friends, children) accompany you on assignment
- Do not trespass on private property
- Be respectful of other's property
- Introduce yourself don't expect anglers to know who you are
- Ask permission before boarding any boat
- Do not assist with the launching or recovery of boats at launch ramps
- Do not collect fish from anglers except yelloweye rockfish, salmon heads and white seabass heads
- Work the sites in an MM cluster in the assigned order
- Visit all sites in an MM or BB cluster before considering the assignment complete
- Be sure to have enough forms with you in the field to complete the day's assignment
- Be productive during slow times in the field review your manual, regulations booklet, fish identification materials, site descriptions, edit data, etc.
- Do not disclose information obtained in a CRFS interview with anyone outside of CRFS
- Do not speak to the media refer them to your Lead
- Do not discuss your personal opinions on natural resource management with the public while on the job
- Educate the angling public on fishing regulations, fish identification and the role of the Department and CRFS in resource management
- Do not guess at answers when asked questions that you don't know the answers to-refer them to your Lead
- Do not accept gifts of any kind fish, free fishing trips, etc.
- Do not smoke while conducting interviews, or where the public may observe you
- Do not use alcoholor cannabis while on the job, or be under the influence of alcoholor cannabis while on the job
- Do not use illegal drugs while on the job, or be under the influence of drugs while on the job
- Do not attempt to aid stranded or injured marine mammals or birds call your local wildlife care center

- Refrain from using vulgar words or negative body language
- Represent the Department in a professional, friendly, courteous manner
- Be aware of diversity

Communication

- Respond to all communications with your fellow samplers, Lead and Supervisor in a timely manner. At the minimum, read your email and all attachments and respond as needed before you begin each scheduled work day. Email messages from your Supervisor and Lead must be promptly opened and acknowledged. Return all phone calls and text messages within 24 hours
- When working with another sampler, coordinate arrivals and departures before the assignment
- Coordinate with your port leads regarding PC activities and salmon PC sampling rates
- You are to notify your Lead AS SOON AS POSSIBLE of changing site conditions (road construction/closures, launch ramp closures, safety issues, etc.)
- Be professional in your communications use correct grammar and punctuation
- Use your DFW email address for all work-related correspondence
- Set up an automated "out of the office" reply for your DFW email account for absences greater than one week

Sampling Gear

- You are to have all sampling gear that is issued to you and one set of CRFS attire with you and ready to be used at every sampling assignment and office assignment that you work
- You are to have your knife in sheath on your belt at every PR1 assignment during salmon season
- Do not lend your knife to anyone other than another sampler
- Have a time keeping device on your person in the field at all times. Verify its accuracy.
- You are responsible for maintaining your sampling gear in proper working order
 - o Clean your gear while sampling if possible
 - o Do not rinse scales in saltwater
 - o At the minimum, calibrate your scales monthly
 - Immediately notify your Lead of any lost/broken gear
 - You may be responsible for the cost to replace lost/broken gear
- You are to return your sampling gear to your Lead AS SOON AS POSSIBLE on request or after your employment ends
- Keep your eyes on your gear do not walk out of view of your gear

Arrival and Departure

- Arrival times at sites:
 - PR1: if you are the first sampler, plan to arrive before the first boat returns. The second sampler arrives at a predetermined time coordinated with the first sampler.
 - o MM and PR2: arrive at the time specified by your Lead
 - PC onboard: plan to arrive at least 30 minutes prior to boat departure, or 45 minutes if you need to talk with the charter master
 - o PC dockside: plan to arrive before the first boat returns
- Departure times at sites:
 - PR1: after the last boat returns, sunset, or relieved by another sampler. If no effort or the last boat returns early, stay on site for two hours or until expected peak fishing time has passed.
 - BB: stay on site for the minimum amount of time prescribed by your Lead; after that, stay until you believe that the goal of obtaining at least one interview per hour cannot be met
 - MM and PR2: stay on site for an 8 hour day (w/travel) or two hours of sampling time if there is no effort (no anglers at any MM site and no trailers at the PR2 site)
 - PC onboard: after the boat has returned to the dock and collection of required data
 - PC dockside: after the last boat has returned or sunset, or at a time prescribed by your Lead

Administration/Personnel

- Arrive at your scheduled work location at the time set by your Lead or Supervisor
 - Show up to/call in to meetings/conference calls on time and prepared to participate. Attendance is mandatory unless prior arrangements have been made with your Lead
 - Show up to office assignments on time
 - Follow all DFW office building security procedures
 - Do not lend your building key to anyone
 - o Do not give your building alarm code to anyone
 - You may be responsible for the cost of law enforcement response for false alarms
 - Relinquish your building key at the end of employment or on request
 - You may be responsible for the cost of re-keying the building if you lose your key
 - Do not admit non-DFW personnel into the building without approval
 - Relinquish your state identification card at the end of employment or on request
 - Notify your Lead if your contact information, address or emergency contact information changes

- You cannot work more than 1,500 hours in any calendar year or 189 days
 from your date of appointment. If you exceed either, you will be
 separated from state service for a minimum of three months after
 which you may be re-hired if a position is available.
- Weekend and holiday work is mandatory. Your Lead will consider CRFS scheduling needs prior to approving requests for time off.
- Schedule requests are to be submitted via email to your Lead by the 15th of the preceding month. Submit requests in a separate email with an appropriate subject.
- Hours are not guaranteed. Hours can be affected each pay period by assignment scheduling, fishing effort (weather), sampler availability, and sampler work performance.

Appendix B.

Marine Mammal Protection Act of 1972

The MMPA established a moratorium, with certain exceptions, on the "taking" of marine mammals in U.S. waters and by U.S. citizens on the high seas, and on the importing of marine mammals and marine mammal products into the United States. It also charged NOAA Fisheries with providing guidelines for deterring marine mammals.

The term "take" is statutorily defined to mean "to harass, hunt, capture, or kill, or attempt to harass, hunt, capture or kill any marine mammal." Under the 1994 amendments, the Congress statutorily defined and divided the term "harassment" to mean any act of pursuit, torment, or annoyance which -- 1. (Level A Harassment) has the potential to injure a marine mammal or marine mammal stock in the wild; or 2. (Level B Harassment) has the potential to disturb a marine mammal or marine mammal stock in the wild by causing disruption or behavioral patterns, including, but not limited to, migration, breathing, nursing, breeding, feeding, or sheltering.

Section 101(a)(4) of the MMPA authorizes the intentional interaction of private citizens with marine mammals. Recreational fishers may now deter marine mammals from damaging fishing gear or catch; property owners or their agents may now deter marine mammals from damaging their property; and the general public may now deter marine mammals from endangering personal safety, provided such deterrence does not cause a marine mammal's death or serious injury. The proposed guidelines and prohibited measures set forth activities that are not likely to cause a marine mammal death or serious injury and specifically prohibit activities determined, using the best scientific information available, to have a significant adverse effect on marine mammals. Actions by the public to deter non-ESA listed marine mammals consistent with such guidelines would not be a violation of the MMPA.

Approved Deterrence Measures

NOAA Fisheries Guidelines for Intentional Interaction (Marine Mammal Deterrence)

Samplers should be familiar with these guidelines in order to inform your Lead of any illegal or unusual actions taken by anglers or party/charter boat crews. The following is copied verbatim from the NOAA web site at:

http://www.westcoast.fisheries.noaa.gov/publications/protected_species/ marine_mammals/pinnipeds/sea_lion_removals/112515_potential_deterr ence_methods.pdf

The following list of "potential methods" and "deterrents to avoid" is not an exhaustive list of non-lethal methods or techniques. If you have questions about protecting your property and/or fishing gear and catch from nuisance Pacific harbor seals, California sea lions, and Eastern U.S. stock Steller sea lions please contact our marine mammal specialists: mammal specialists: (Seattle, WA) Brent Norberg, 206- 526-6550; Lynne Barre, 206-526-4745; (Long Beach, CA) Monica DeAngelis, 562-980-3232; Penny Ruvelas, 562-980-4197.

Note: Some of the methods listed (such as loud noise or pyrotechnics) may not be appropriate for use in some areas, or are subject to prohibition under federal, state or local ordinances. The presence of ESA listed species (marine mammals or fish) in some areas may advise against the use of certain methods. Please consult with appropriate authorities to determine if such prohibitions exist in your area, or if ESA-listed species may be encountered.

Potential methods for use by private property owners to deter Pacific harbor seals, California sea lions, and Eastern U.S. stock Steller sea lions from damaging property (developed waterfront, decks, docks, floats, piers, bait receivers, vessels at an chor, etc.):

Barriers & Exclusion Devices:

- fencing (e.g., plastic construction/snow fence, chain link)
- · closely spaced posts
- · bull rails
- electric livestock fencing
- netting
- · swim step protector

Visual Repellents:

- flags, pinwheels, or streamers
- · flashing lights or strobes
- balloons
- human attendants/monitors

Noise Makers:

- · horns, whistles, bells
- electronic acoustic devices (Acoustic Harassment Devices)
- clapping, banging on pots, pans, drums; empty aluminum cans on a string banging together
- music
- starter pistols
- pyrotechnics (e.g., bird screamers, bangers, firecrackers, propane canons)
- propane canons

Physical Contact:

- high or low pressure water hoses
- · sprinklers, sprayers
- crowder boards
- bull poles (blunttip), brooms
- cattle prod (these products produce only a mild electric shock designed for handling livestock and are in no way related to "stun guns" designed for self-defense)
- toy water guns (e.g., "Super Soaker©")
- non-toxic and water soluble paint ball or air soft guns
- slingshot
- chemical irritants (e.g., non-toxic pepper spray, mace) used for animal control (there are many municipal and state ordinances controlling the use and possession of these irritants)

Note: Guard dogs are not included on the list of suggested measures because of risks to both dogs and marine mammals, including the potential risk of disease transmission between them.

Potential methods for use by fishers to deter Pacific harbor seals, California sea lions, and Eastern U.S. stock Steller sea lions from damaging gear or catch (anglers must be actively fishing with gear deployed).

Visual Repellents/Noise Makers: • boat hazing, circling • pounding on hull • pyrotechnics (e.g., bird screamers, bangers, underwater firecrackers, cracker shells) • starter pistols • horns, bells, whistles

Physical Contact: • slingshots • non-toxic and water soluble paint ball guns • non-lethal ammunition (e.g., rubber bullets, sabot rounds, game stingers)

Methods to Avoid – The following methods and techniques have an increased likelihood of causing injury or death and should be avoided.

- No firearms with "live" (lethal) ammunition
- No devices with injurious projectiles (e.g., archery gear, crossbows, spear guns, bangsticks)
- No sharp/pointed objects (e.g., harpoons, spears, gaffs, nail studded bats/poles/clubs)
- No entangling devices (e.g., loose webbing, snares, concertina wire)
- No aggressive tactile methods (e.g., striking animals with bats, hammers etc., impact with vehicles or boats)
- No tainted baits or poisons

Act responsibly & use common sense

Regardless of method or intent, the property owner or fisher may be subject to prosecution should a marine mammal be seriously injured or killed as a result of deterrence efforts for the protection of property, gear or catch.

Remember personal safety

Attempts by property owners and/or fishers to deter nuisance animals from engaging in unwanted behaviors using non-lethal means is a personal choice and not without risk (to the person doing the deterring and anyone around them). Sea lions and seals are wild animals that may react unpredictably to non-lethal deterrence measures, resulting in personal injury or additional damage to property. Sea lions are large and powerful animals that can move as quickly as a person on land.

Be aware of people around you and be courteous

The safe use of some of the above-listed potential methods (e.g., cracker shells, non-lethal ammunition) requires considerable skill and experience. The use of some of these methods may precipitate undesirable social interactions. If you are in

possession of a firearm, law enforcement officers approaching your property or vessel will assume that your firearm is loaded with lethal ammunition.

Individuals attempting to deter nuisance sea lions and seals, using the above the listed potential methods are similar techniques, do so at their own risk.